Supporting Information

Design, Synthesis and Biological Evaluations of Asymmetric Bow-Tie PAMAM Dendrimer-Based Conjugates for Tumor-Targeted Drug Delivery

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Figure S2. ¹³C NMR spectrum of 1



Figure S3. ¹H NMR spectrum of 2



Figure S4. ¹³C NMR spectrum of 2





Figure S6. ¹³C NMR spectrum of 3







Figure S8. ¹³C NMR spectrum of 5



Figure S10. ¹³C NMR spectrum of 6



Figure S12. ¹³C NMR spectrum of 7



Figure S14. ¹³C NMR spectrum of 8





Figure S16. ¹³C NMR spectrum of 9



Figure S18. ¹³C NMR spectrum of 12





Compound Name	TW-B05-P25	-	
Processed by	twang	2	
Solvent	CDC13		
Temperature	298.2		
Number of Scans	203		
Pulse Sequence	zgpg30		
Relaxation Delay	2.0000		
Spectral Width	29761.9		
Spectrometer Freque	ney 125.71		
H	H_{1}		
210 200 100 180			 · · · · · · ·

Figure S20. ¹³C NMR spectrum of 14





Figure S22. ¹³C NMR spectrum of 16







Figure S24. ¹³C NMR spectrum of 19







Figure S26. ¹³C NMR spectrum of 20







Figure S28. ¹³C NMR spectrum of 22



Figure S30. ¹³C NMR spectrum of 24







Figure S32. ¹³C NMR spectrum of 25







Figure S34. ¹³C NMR spectrum of 28



Figure S35. MALDI-TOF analysis of fully biotinylated G3 PAMAM dendrimer 7.



Figure S36. MALDI-TOF analysis of fully alkynylated G1-PAMAM dendrimer 8.



Figure S37. LC-UV-TOF analysis of fully alkynylated G1-PAMAM dendrimer 8.



Figure S38. LC-TOF deconvolution result for fully alkynylated G1-PAMAM dendrimer 8.



Figure S39. MALDI-TOF analysis of G1-PAMAM half-dendron-linker 12.



Figure S40. LC-UV-TOF analysis of G1-PAMAM half-dendron-linker 12.



Figure S41. LC-UV-TOF deconvolution result for G1-PAMAM half-dendron-linker 12.



Figure S42. MALDI-TOF analysis of (G1 half-dendron)-linker-(G3 half-dendron) 6.



Figure S43. LC-UV-TOF mass analysis of (G1 half-dendron)-linker-(G3 half-dendron) 6.



Figure S44. LC-UV-TOF deconvolution result of (G1 half-dendron)-linker-(G3 half-dendron) 6.



Figure S45. GPC analysis of (A) ABTD-TTC-1 (1) and (B) ABTD-TTC-1 (1) + click-ready ABTD 6. Conditions: UltrahydrogelTM 500 column (7.8 mm x 300 mm, 10 μ m); MeCN/H₂O (20/80); 0.5 mL/min; 55 μ M; UV detector at 215 nm.



Figure S46. GPC analysis of ABTD-TTC-2 (2). Conditions: UltrahydrogelTM 500 column (7.8 mm x 300 mm, 10 μ m); MeCN/H₂O (20/80); 0.5 mL/min; 50 μ M; UV detector at 254 nm. The anticipated t_R of **6** is also shown.



Figure S47. GPC analysis of (A) ABTD-TTC-3 (**3**) and (B) ABTD-TTC-2 (**2**) + click-ready ABTD **6**. Conditions: UltrahydrogelTM 500 column (7.8 mm x 300 mm); MeCN/H₂O (20/80); 0.5 mL/min; 64 μ M; UV detector at 215 nm.



Figure S48. Internalization of fluorescent probe ABTD-TTC-2 (**2**) (20 μ M) in ID-8 (ovarian cancer) cells by confocal fluorescence microscopy (CFM) and flow cytometry after 1 h of incubation at 37 °C (B) as compared to the control ID8 cells (A).



Figure S49. Assessment of the internalization of fluorescent probe ABTD-TTC-3 (3) (20 μ M) in ID-8 (ovarian cancer), MX-1 (breast cancer) and WI-38 (normal lung fibroblast) cells by confocal fluorescence microscopy (CFM) and flow cytometry at 0 h, 1 h, and 3 h periods, at 37 °C.



Figure S50. Assessment of the multi-binding effect by flow cytometry based on the internalization of fluorescent probe **28** (10 μ M, top) and ABTD-TTC-3 (**3**) (10 μ M, bottom) in ID-8 cells at 0, 1, 2, 4, 6 and 16 h periods, at 37 °C.



Figure S51. Assessment of the multi-binding effect by flow cytometry based on the internalization of fluorescent probe **28** (10 μ M, top) and ABTD-TTC-3 (**3**) (10 μ M, bottom) in MX-1 cells at 0, 1, 2, 4, 6 and 16 h periods, at 37 °C.