

Supporting Information

Two-step fractionation of a model technical lignin by combined organic solvent extraction and membrane ultrafiltration

Chiara Allegretti,^a Simon Fontanay,^b Klaus Rischka,^c Alberto Strini,^d Julien Troquet,^b
Stefano Turri,^a Gianmarco Griffini,^{a*} Paola D'Arrigo^{a,e*}

^a*Department of Chemistry, Materials and Chemical Engineering "Giulio Natta", Politecnico of Milano, p.zza L. da Vinci 32, Milano, 20133, Italy*

^b*Biobasic Environnement, Biopôle Clermont-Limagne, Saint-Beauzire, 63360, France*

^c*Fraunhofer Institute for Manufacturing Technology and Advanced Materials IFAM, Wiener Str. 12, 28359 Bremen, Germany*

^d*Construction Technologies Institute - National Research Council of Italy (ITC-CNR), via Lombardia 49, San Giuliano Mil., 20098, Italy*

^e*Istituto di Chimica del Riconoscimento Molecolare, CNR, via Mario Bianco 9, Milano, 20131, Italy.*

*Corresponding authors:

gianmarco.griffini@polimi.it; phone: +39 2 2399 3213; fax: +39 2 23993280

paola.darrigo@polimi.it; phone: +39 2 23993075; fax: +39 2 23993180

CONTENT

Figure S1: Apparatus used in the Fractionation process of a solution of Protobind 1000.	S3
Table S2: Operational parameters for the membrane assisted ultrafiltration process	S4
Figure S3: Comparison between the extraction yields (% w/w) of Protobind 1000 and Kraft Lignins in different solvents.	S5
Figure S4: Molecular weight distribution of all extracted lignin fractions as obtained from GPC analysis.	S6
Figure S5: Calibration Curve of Folin-Ciocalteu phenol titration with gallic acid.	S7
Figure S6: GC/MS Chromatogram of LF-S.	S8
Table S7: GC/MS Results of LF-S.	S9
Figure S8: GC/MS Chromatogram of LF-R5.	S10
Table S9: GC/MS Results of LF-R5.	S11
Figure S10: GC/MS Chromatogram of LF-P5.	S12
Table S11: GC/MS Results of LF-P5.	S13
Figure S12 GC/MS Chromatogram of LF-R2.	S14
Table S13: GC/MS Results of LF-R2.	S15
Figure S14: GC/MS Chromatogram of LF-P2.	S16
Table S15: GC/MS Results of LF-P2.	S17
Figure S16: TGA curves of the lignins LF-0 and LF-S	S18
Figure S17: Identification of the main compounds identified in Pyr-GC/MS analysis of LF-0 at 600°C.	S19
Table S18: Peak Identification of Protobind 1000 pyrolysed at 450 °C.	S20



Panel A



Panel B



Panel C

Figure S1: Apparatus used in the Fractionation process of a solution of Protobind 1000. (Panel A: Soxhlet extraction, Panel B: Details of solvent beakers, Panel C: Ultrafiltration apparatus; Photograph courtesy of Julien Troquet Copyright 2018).

	Ultrafiltration process data	Unit	Average values from 3 successive filtration trials	
			Step 1	Step 2
Membrane data	Membrane type		SARTORIUS STEDIM Hydrosart (305 144 29 01 E-SW)	SARTORIUS STEDIM Hydrosart (305 144 19 01 E-SW)
	Membrane material		Stabilized cellulose	Stabilized cellulose
	Membrane area	m ²	0.1	0.1
	Cut-off	kDa	5	2
	Clean water flow (CWF)	L min ⁻¹ m ⁻²	24	18
Process data*	Cross flow	L min ⁻¹	1	0.5
	Feed pressure	bar	2.5	2.5
	TMP	bar	1.92	1.74
	Permeate flow	mL min ⁻¹ L min ⁻¹ m ⁻²	0.2 0.002	2.2 0.022
	Time to process 500 mL	min	900	80

* Data obtained from 3 successive trials each realized with a starting volume of 1.5 L of the soluble extracted fraction (LF-S). The 3 trials have been runned with the same membranes which have been cleaned and tested (clean water flow measurement) between each trial.

Table S2: Operational parameters for the membrane assisted ultrafiltration process.

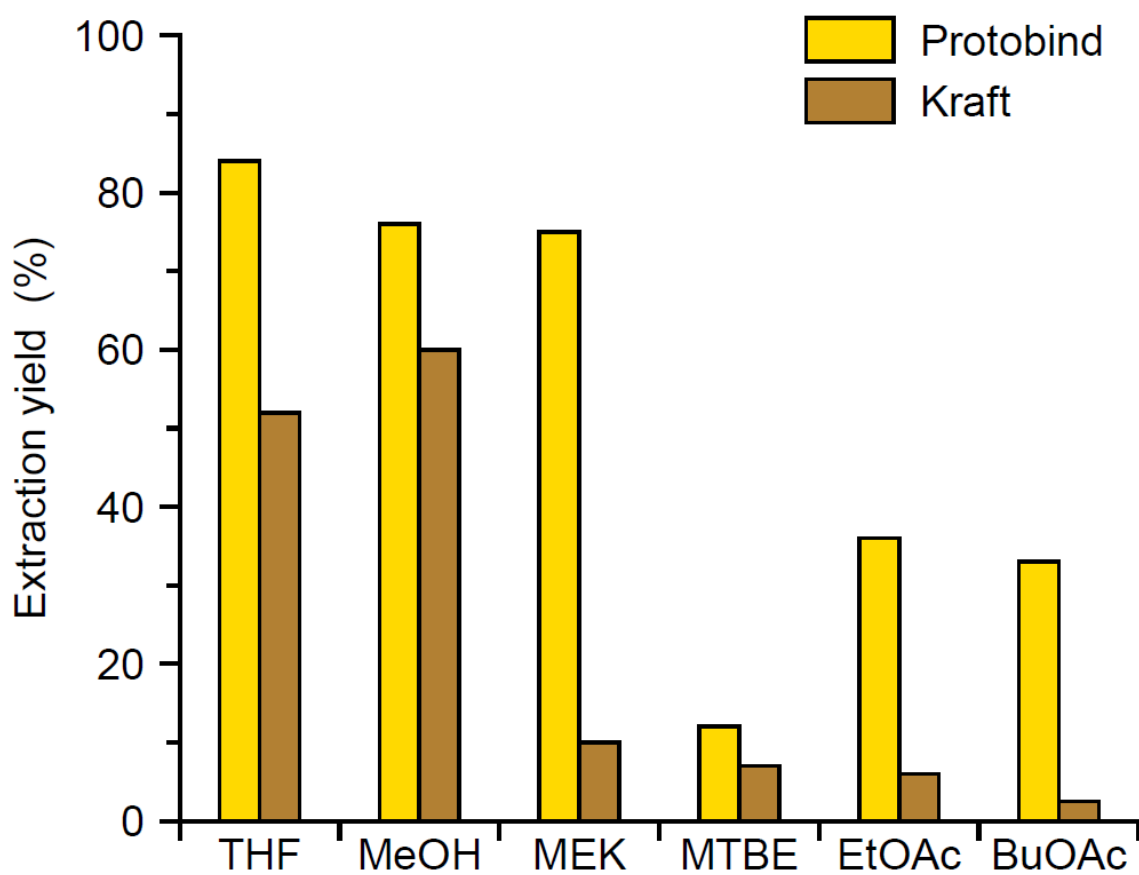


Figure S3: Comparison between the extraction yields (% w/w) of Protobind 1000 and Kraft Lignins in different solvents.

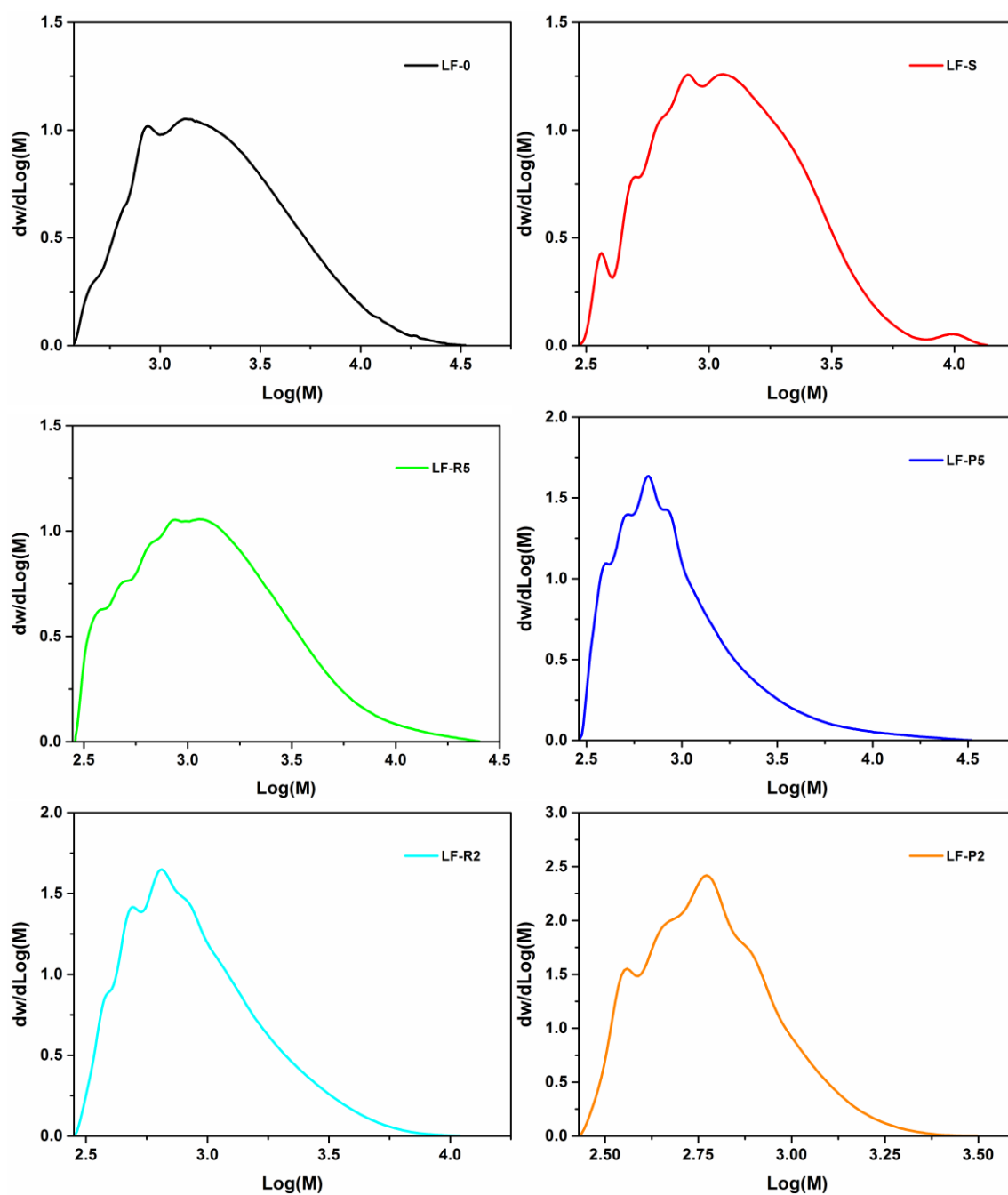


Figure S4: Molecular weight distributions of all extracted lignin fractions as obtained from GPC analysis (the parent material LF-0 is also reported as reference). Samples were acetylated prior to analysis.

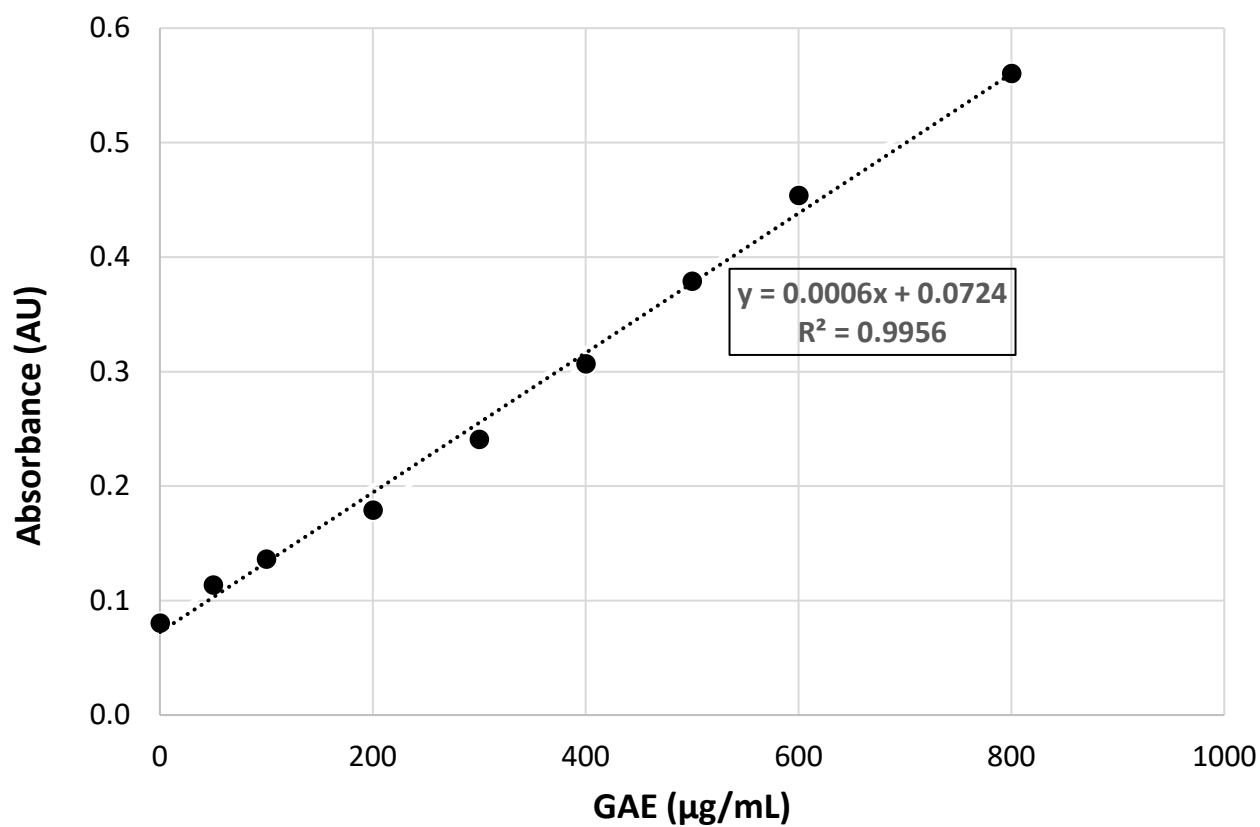


Figure S5: Calibration Curve of Folin-Ciocalteu phenol titration with gallic acid.

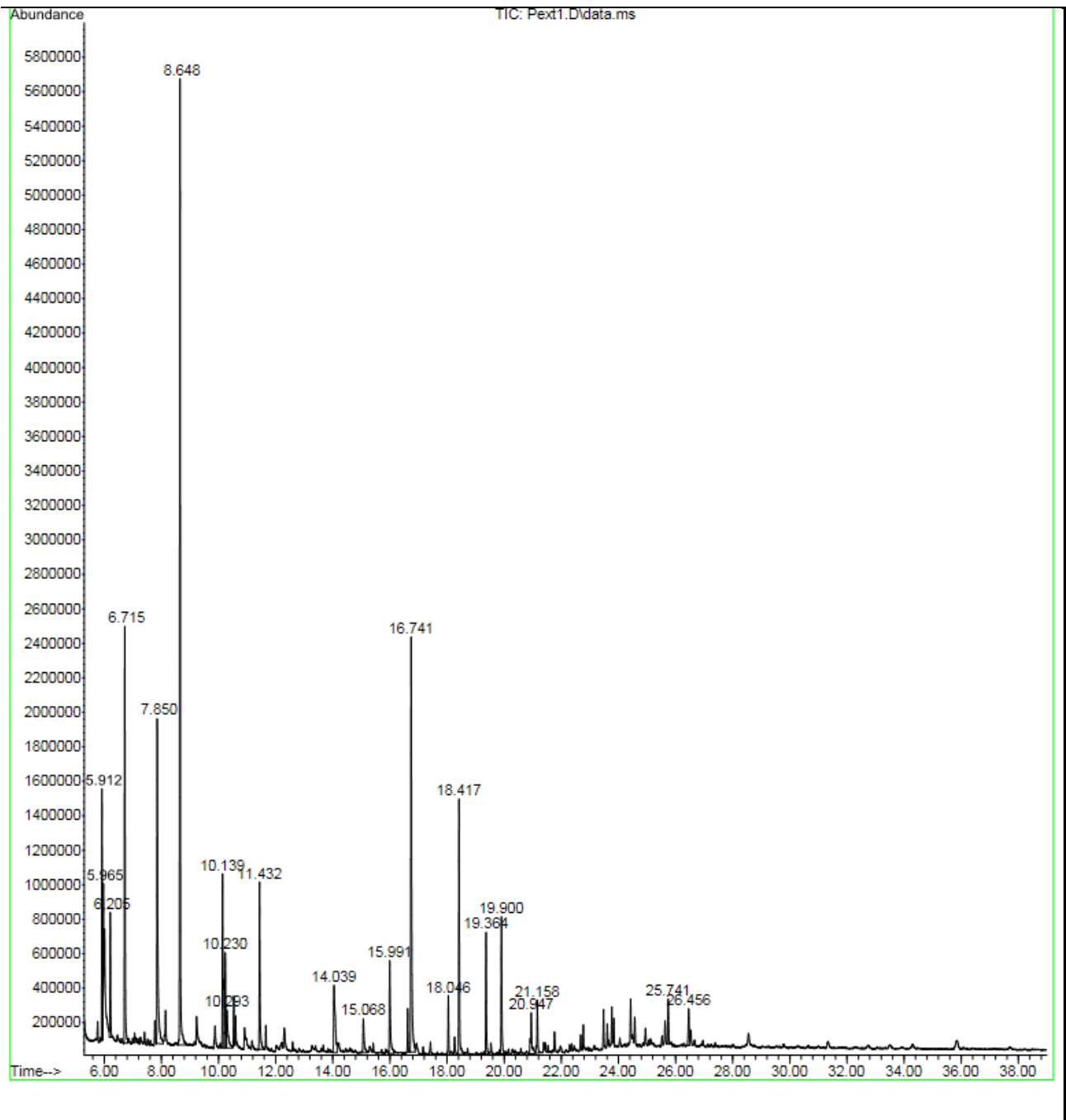


Figure S6: GC/MS chromatogram of LF-S.

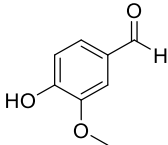
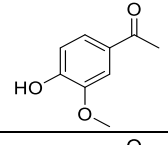
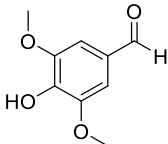
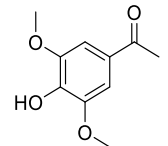
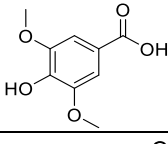
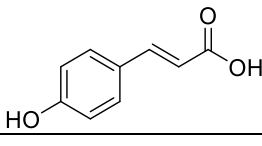
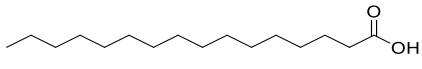
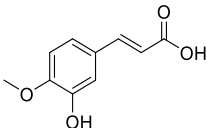
Retention time (min)	Molecule	Name	% in the analysis
14		4-hydroxy-3-methoxybenzaldehyde	3.4
15		1-(4-hydroxy-3-methoxyphenyl)ethan-1-one	1.3
15.9		4-hydroxy-3,5-dimethoxybenzaldehyde	3
16.74		1-(4-hydroxy-3,5-dimethoxyphenyl)ethan-1-one	11
18		4-hydroxy-3,5-dimethoxybenzoic acid	1.3
18.4		(E)-3-(4-hydroxyphenyl)acrylic acid	5.9
19.3		hexadecanoic acid	2.6
19.9		(E)-3-(3-hydroxy-4-methoxyphenyl)acrylic acid	3.38

Table S7: GC/MS Results of LF-S.

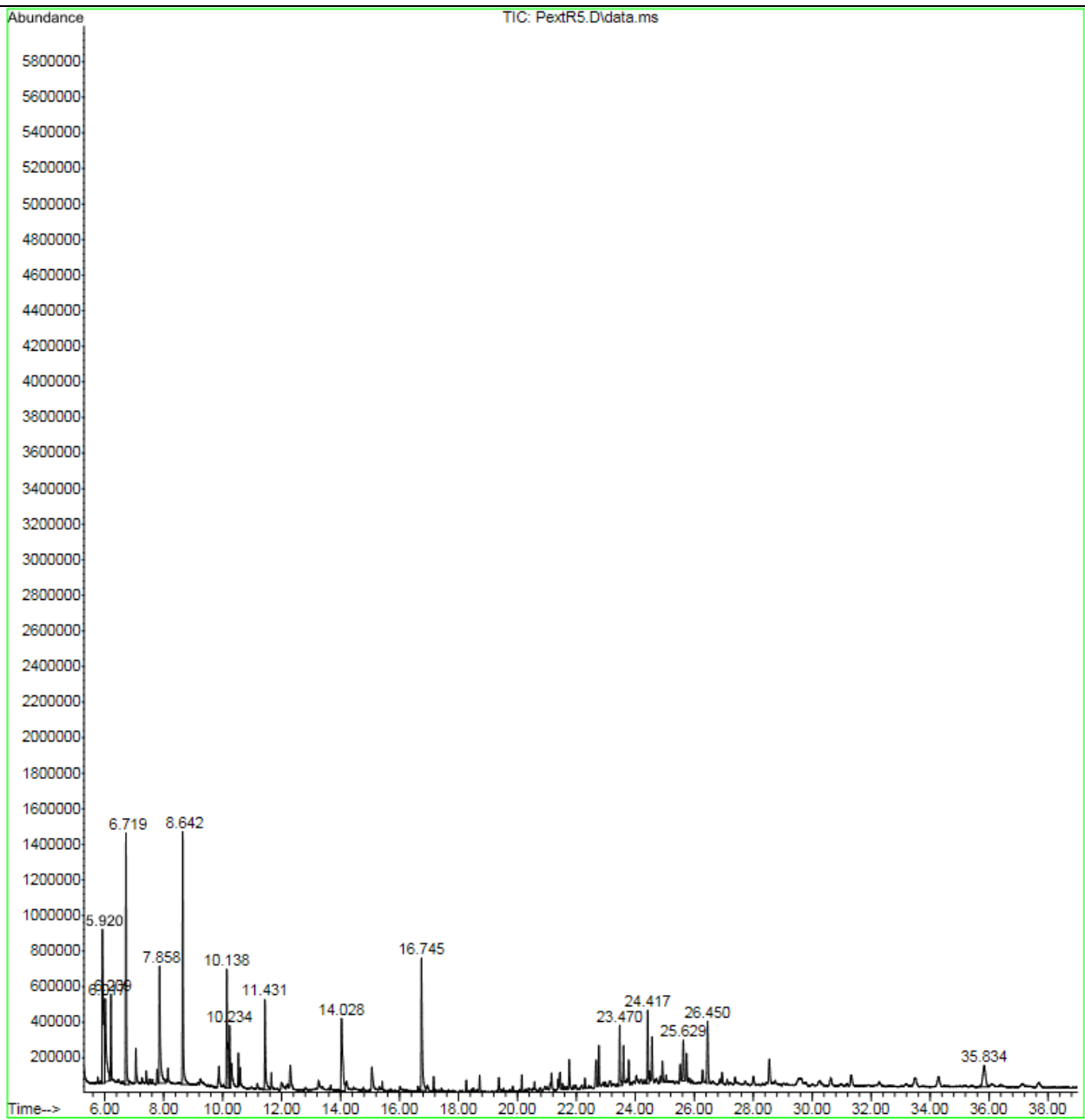


Figure S8: GC/MS chromatogram of LF-R5.

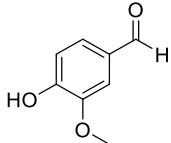
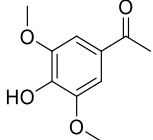
Retention time (min)	Molecule	Name	% in the analysis
14		4-hydroxy-3-methoxybenzaldehyde	14.9
16.7		1-(4-hydroxy-3,5-dimethoxyphenyl)ethan-1-one	18.1

Table S9: GC/MS Results of LF-R5.

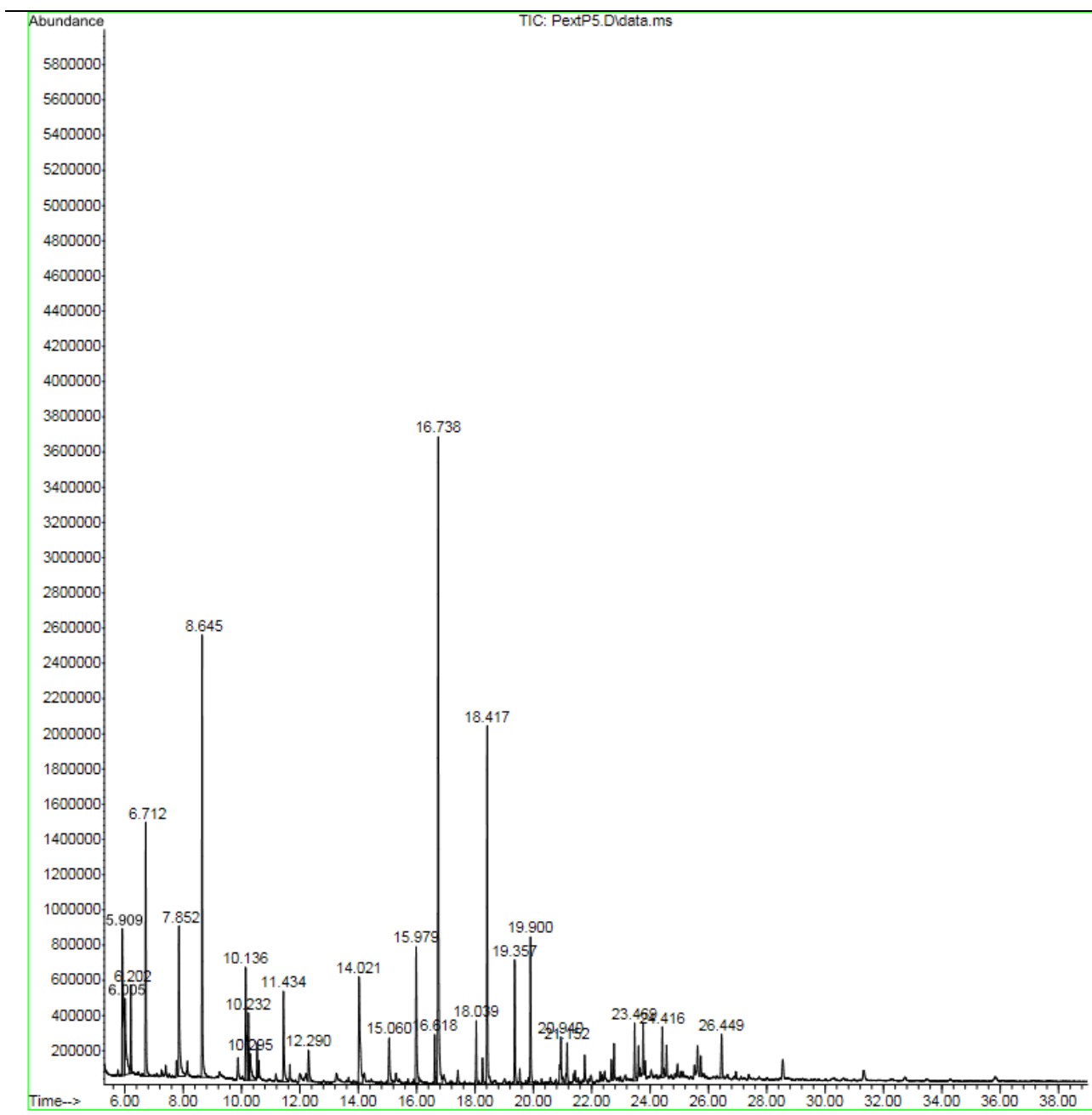


Figure S10: GC/MS chromatogram of LF-P5.

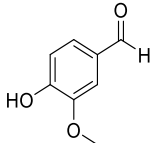
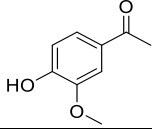
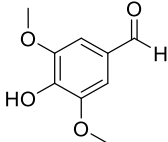
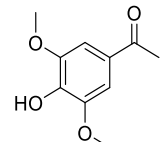
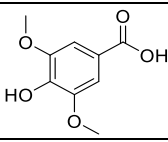
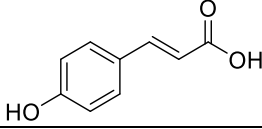
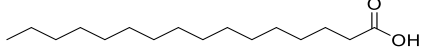
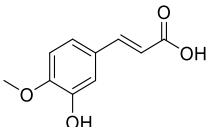
Retention time (min)	Molecule	Name	% in the analysis
14		4-hydroxy-3-methoxybenzaldehyde	4.9
15		1-(4-hydroxy-3-methoxyphenyl)ethan-1-one	1.9
15.9		4-hydroxy-3,5-dimethoxybenzaldehyde	4.3
16.74		1-(4-hydroxy-3,5-dimethoxyphenyl)ethan-1-one	17.5
18		4-hydroxy-3,5-dimethoxybenzoic acid	1.5
18.4		(E)-3-(4-hydroxyphenyl)acrylic acid	8.7
19.3		hexadecanoic acid	3.2
19.9		(E)-3-(3-hydroxy-4-methoxyphenyl)acrylic acid	3.6

Table S11: GC/MS Results of LF-P5.

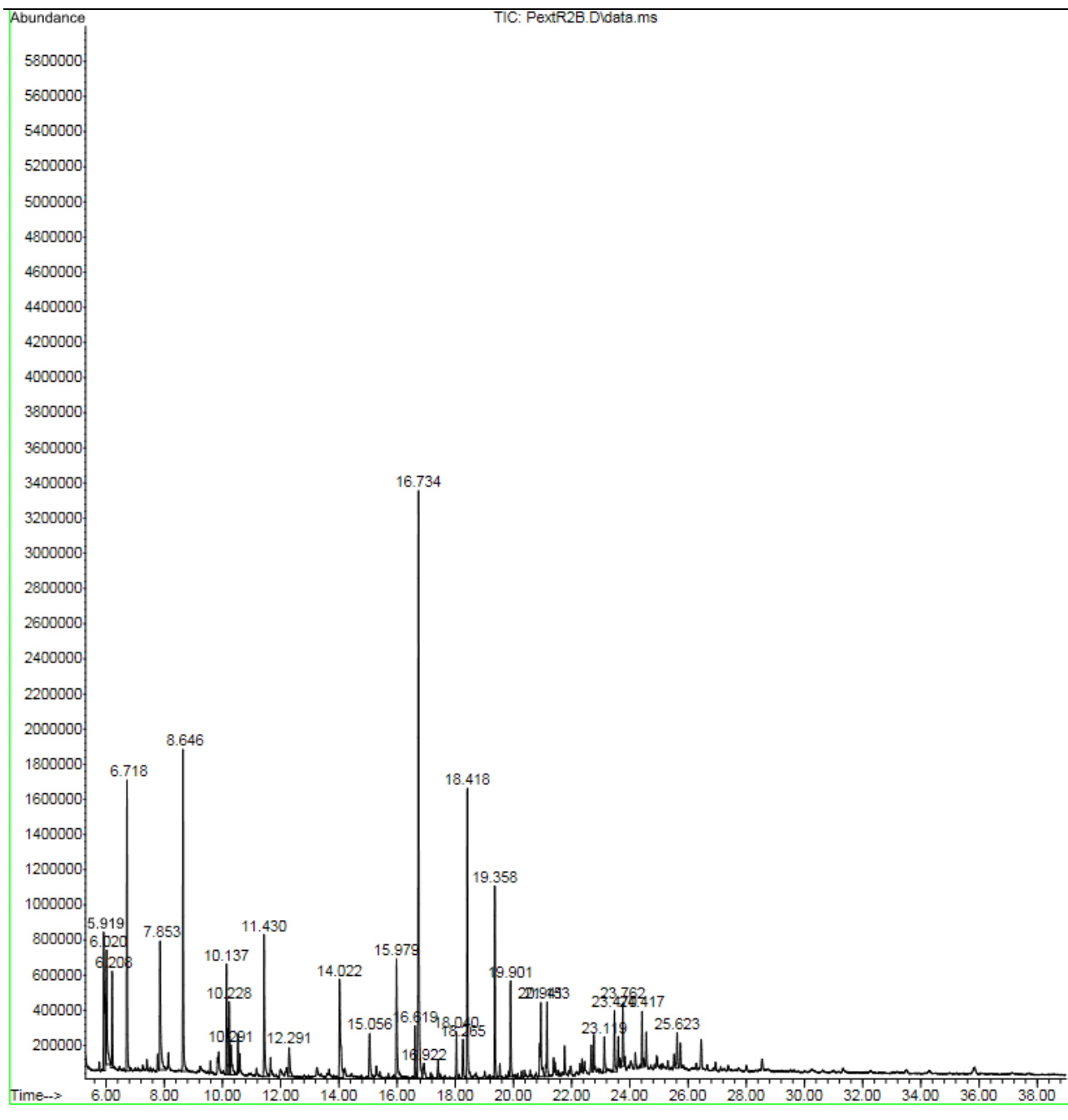


Figure S12: GC/MS chromatogram of LF-R2.

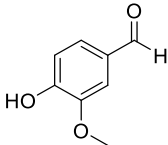
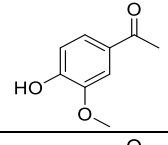
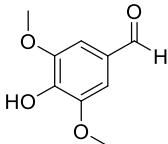
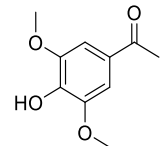
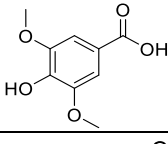
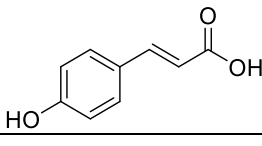
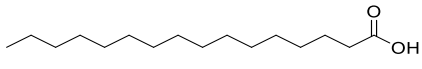
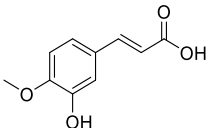
Retention time (min)	Molecule	Name	% in the analysis
14		4-hydroxy-3-methoxybenzaldehyde	6.3
15		1-(4-hydroxy-3-methoxyphenyl)ethan-1-one	2.7
15.9		4-hydroxy-3,5-dimethoxybenzaldehyde	5.7
16.74		1-(4-hydroxy-3,5-dimethoxyphenyl)ethan-1-one	22
18		4-hydroxy-3,5-dimethoxybenzoic acid	1.6
18.4		(E)-3-(4-hydroxyphenyl)acrylic acid	10.6
19.3		hexadecanoic acid	6.2
19.9		(E)-3-(3-hydroxy-4-methoxyphenyl)acrylic acid	3.4

Table S13: GC/MS Results of LF-R2.

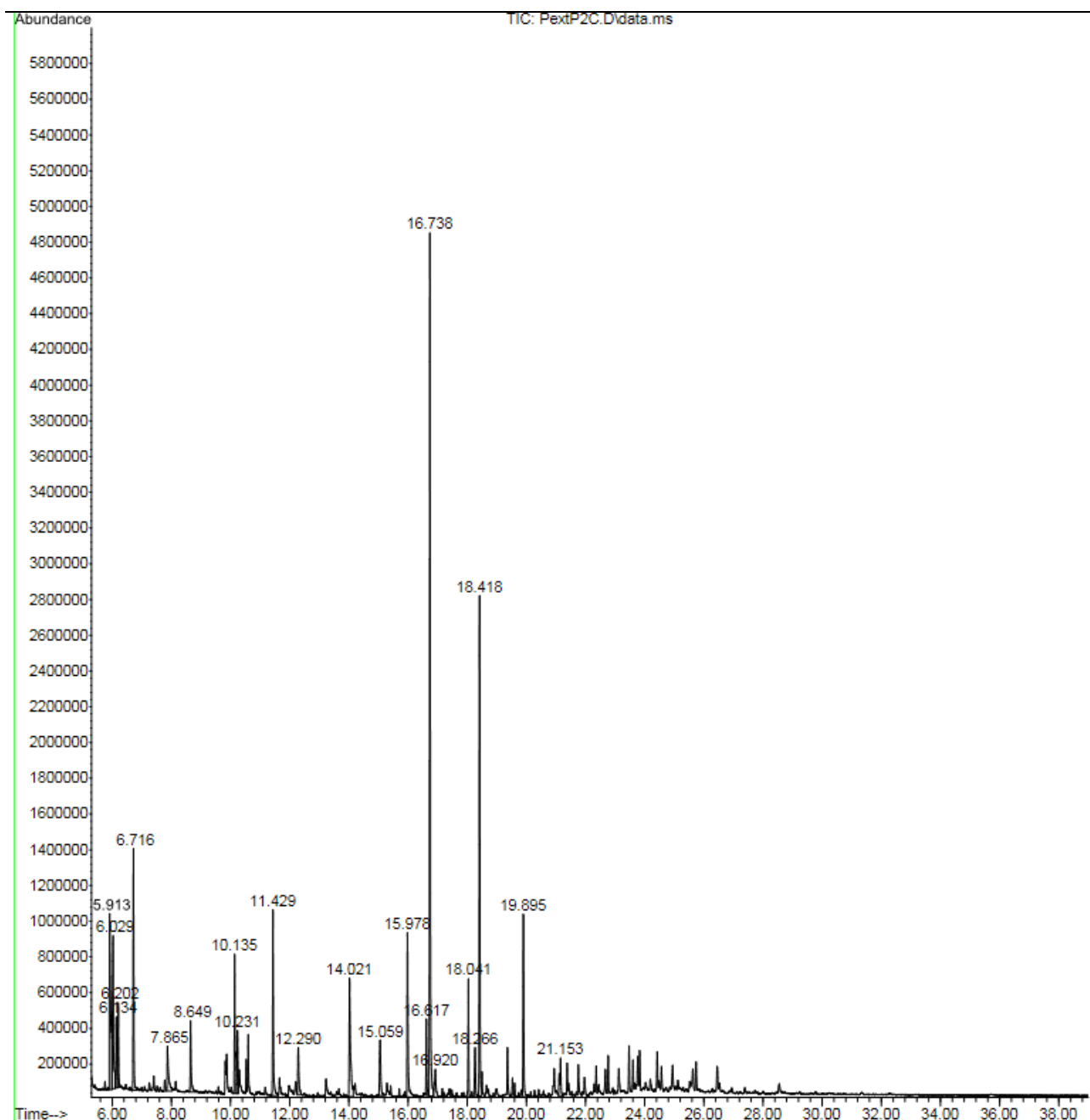


Figure S14: GC/MS chromatogram of LF-P2.

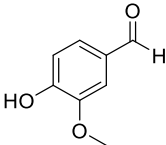
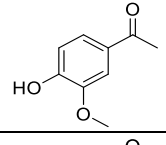
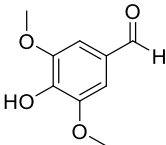
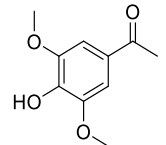
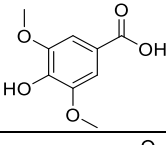
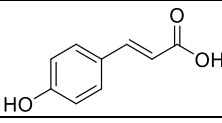
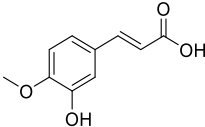
Retention time (min)	Molecule	Name	% in the analysis
14		4-hydroxy-3-methoxybenzaldehyde	6.9
15		1-(4-hydroxy-3-methoxyphenyl)ethan-1-one	3.2
15.9		4-hydroxy-3,5-dimethoxybenzaldehyde	7.3
16.74		1-(4-hydroxy-3,5-dimethoxyphenyl)ethan-1-one	30
18		4-hydroxy-3,5-dimethoxybenzoic acid	3.7
18.4		(E)-3-(4-hydroxyphenyl)acrylic acid	15.4
19.9		(E)-3-(3-hydroxy-4-methoxyphenyl)acrylic acid	6.4

Table S15: GC/MS Results of LF-P2.

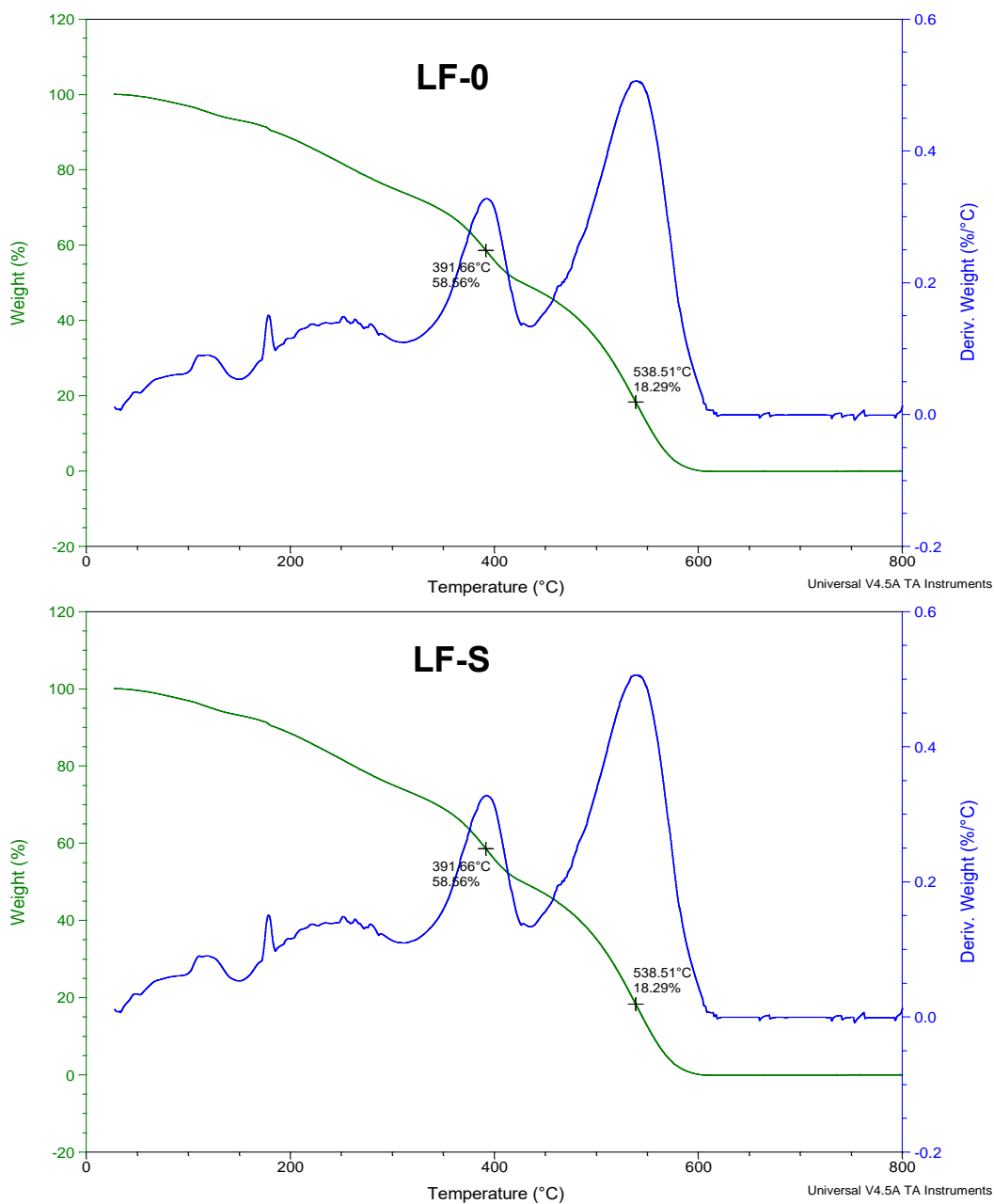


Figure S16: TGA curves of the lignin samples LF-0 (above) and LF-S (below) evaluated by Pyr-GC/MS. The weight loss is indicated by the green line (left green scale) The first derivation of the weight loss is indicated by the blue line (right blue scale).

RT: 0.00 - 78.85

NL:
1.10E9
TIC MS
Proto1000

Protobind 1000 @ 600°C

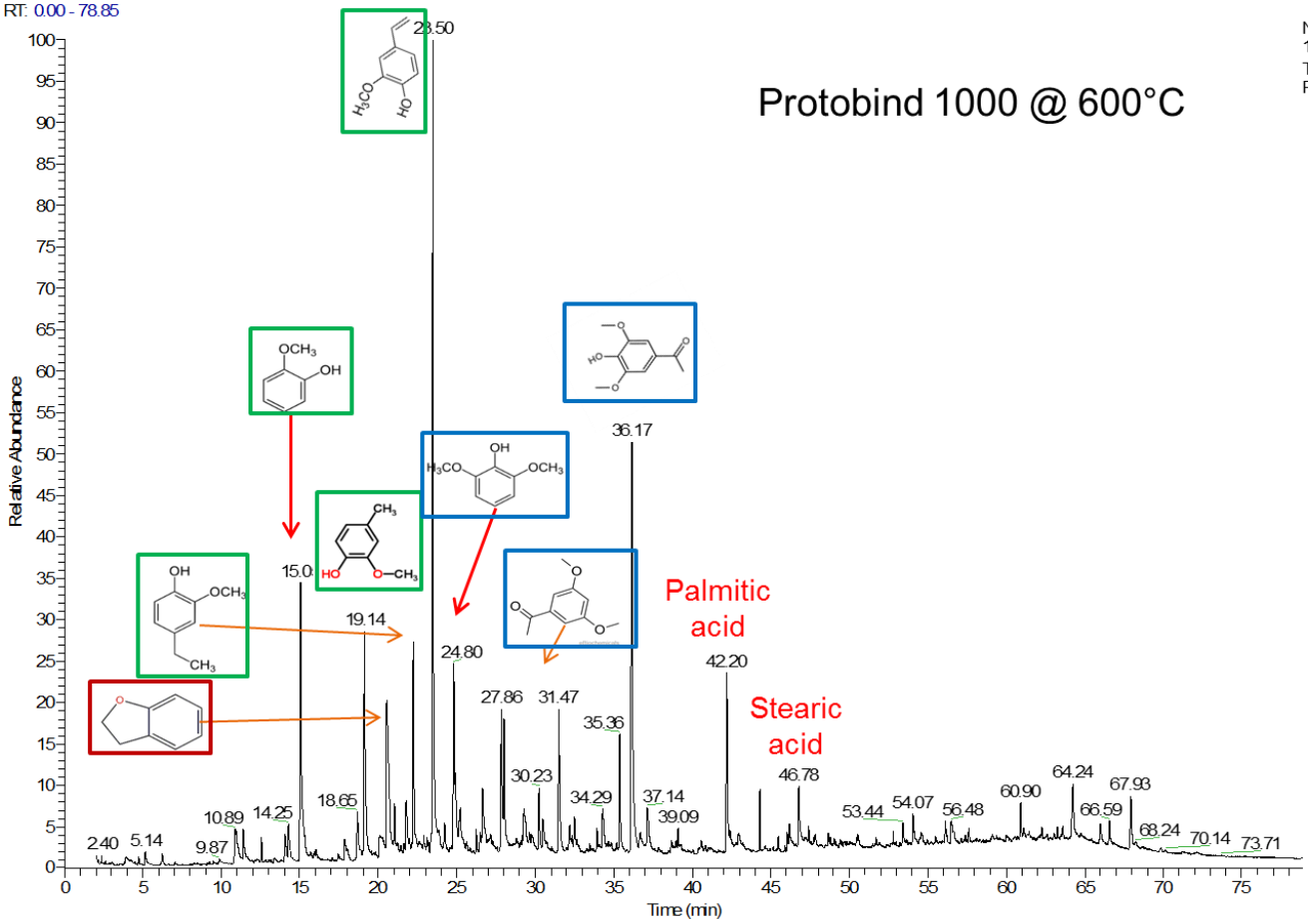


Figure S17: Identification of the main compounds identified in Pyr-GC/MS analysis of LF-0 at 600 °C.

Apex RT [min]	CAS-Nr.	Name of Compound	Unit
2.22	74-93-1	Methanthiol	
4.26	937-61-1	Benzylpropylether	C
5.95	98-01-1	Furfural	C
7.08	106-42-3	p-Xylene	C
10.63	20602-79-3	acryloylurea	C
11.54	108-95-2	Phenol	H
13.11	1461-27-4	m-Mentha-6,8-diene	
14.35	95-48-7	o-Cresol	H
15.2	106-44-5	p-Cresol	H
15.35	90-05-1	o-Methoxyphenol	G
18.57	123-07-9	p-Ethylphenol	H
18.72	18102-31-3	2-Methoxy-3-methylphenol	G
19.23	93-51-3	Creosol	G
20.04	120-80-9	Catechol	
20.51	496-16-2	Coumaran	
21.64	934-00-9	3-Methoxy-pyrocatechol	S
22.22	2785-89-9	p-Ethylguaiaicol	G
23.25	488-17-5	3-Methylpyrocatechol	
23.52	7786-61-0	2-Methoxy-4-vinylphenol	G
24.07	3209-13-0	3-Methoxy-5-Methylphenol	
24.75	91-10-1	2,6-Dimethoxyphenol (Syringol)	S
24.88	97-53-0	Eugenol	G
25.01	2033-89-9	3,4-Dimethoxyphenol	S
25.2	2785-87-7	2-Methoxy-4-propylphenol	G
26.15	6380-23-0	3,4-Dimethoxystyrene	G
26.35	121-33-5	Vanillin	G
26.56	5932-68-3	trans-Isoeugenol	G
27.79	135-77-3	1,2,4-Trimethoxybenzene	S
27.95	5932-68-3	trans-Isoeugenol	G
29.05	6100-74-9	1-(3-Hydroxy-4-methoxyphenyl)ethanone (apocynin)	G
30.14	6443-69-2	3,4,5-Trimethoxytoluene	S
30.29	2503-46-0	Guaiacylacetone	G
32.4	6627-88-9	4-allyl-2,6-dimethoxyphenol	S
34.1	134-96-3	3,5-Dimethoxygallaldehyde	S
35.27	6627-88-9	4-allyl-2,6-dimethoxyphenol	S
36.11	2478-38-8	4'-Hydroxy-3',5'-Dimethoxyacetophenone	S
36.35	458-35-5	4-[(1E)-3-Hydroxy-1-propenyl]-2-methoxyphenol	
36.97	437-72-9	1-(2,6-Dihydroxy-4-methoxyphenyl)-1-butanone	

Table S18: Peak Identification of Protobind 1000 pyrolysed at 450 °C.