# Molecular Modeling Based Delivery System Enhances Everolimus-Induced Apoptosis in Caco-2 Cells

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Figure S1. Dialysis tubing procedure

## 2. Cytotoxicity of FGF7:β-CD:EV complex on normal cells



**Figure S2.** Cytotoxic effect of EV on FHs 74 Int cells as displayed by the RTCA DP instrument. Cells were seeded overnight to reach the log phase, then incubated with 3.26  $\mu$ M EV (A), 6.52  $\mu$ M EV (B), or 13.04  $\mu$ M EV (C). IC<sub>50</sub> value of EV on FHs 74 Int cells was found to be 19.25  $\pm$  1.35  $\mu$ M.



**Figure S3.** Cytotoxic effect of FGF7:EV on FHs 74 Int cells as displayed by the RTCA DP instrument. Cells were seeded overnight to reach the log phase, then incubated with FGF7:EV complex (contains 1.58  $\mu$ M FGF7 and 3.26  $\mu$ M EV) (A), FGF7:EV (contains 1.58  $\mu$ M FGF7 and 6.52  $\mu$ M EV) (B), or FGF7-EV (contains 1.58  $\mu$ M FGF7 and 13.04  $\mu$ M EV) (C). IC<sub>50</sub> value of EV on FHs 74 Int cells was found to be 15.9 ± 0.95  $\mu$ M.



**Figure S4.** Cytotoxic effect of  $\beta$ -CD:EV on FHs 74 Int cells as displayed by the RTCA DP instrument. Cells were seeded overnight to reach the log phase, then incubated with  $\beta$ -CD:EV inclusion complex (contains 3.26  $\mu$ M EV) (A),  $\beta$ -CD:EV inclusion complex (contains 6.52  $\mu$ M EV) (B), or  $\beta$ -CD:EV inclusion complex (contains 13.04  $\mu$ M EV) (C). IC<sub>50</sub> value of EV on FHs 74 Int cells was found to be 16.55 ± 1.05  $\mu$ M.



**Figure S5.** Cytotoxic effect of FGF7: $\beta$ -CD:EV complex on FHs 74 Int cells as displayed by the RTCA DP instrument. Cells were seeded overnight to reach the log phase, then incubated with FGF7: $\beta$ -CD:EV complex (contains 1.58  $\mu$ M FGF7, 55.07  $\mu$ M  $\beta$ -CD and 3.26  $\mu$ M EV) (A), FGF7: $\beta$ -CD:EV complex (contains 1.58  $\mu$ M FGF7, 110.13  $\mu$ M  $\beta$ -CD and 6.52  $\mu$ M EV) (B) or FGF7: $\beta$ -CD:EV complex (contains 1.58  $\mu$ M FGF7, 220.27  $\mu$ M  $\beta$ -CD and 13.04  $\mu$ M EV) (C). IC<sub>50</sub> value of EV on FHs 74 Int cells was found to be 34.11  $\pm$  1.9  $\mu$ M.

#### **3.** Assessment of FGF7:β-CD:EV complex retention in Caco-2 cells

#### Chromatographic conditions

The method is modified from Carpentier et al.<sup>1</sup> and Spandana et al.<sup>2</sup>. Briefly, treated cells were detached and centrifuged at 400g x 7 minutes, then 0.1 ml of tetraborate buffer (pH 9.8), 0.1 ml of EV solution (10 mg/L) as internal standard, and 1.8 ml of dichloromethane/methanol 4:1 (v/v) were immediately added to the cell pellet. After a vigorous agitation, the organic phase was removed and evaporated to dryness. The dry residue was dissolved in the mobile phase: acetonitrile/double ultra-pure water 95:05 (v/v), and injected onto an Ascentis<sup>®</sup> C18 column (Supelco, Bellefonte PA, USA). The flow rate was 1 ml/min, the peaks were detected by PDA detection and wave length was set at 278 nm.

The results indicated that the penetration of EV from both samples was time-dependent, thus the accumulation was significantly increased with FGF7: $\beta$ -CD:EV complex application, only 3% of free EV was detected. The retention of EV was enhanced with the complex application as well. After 24 hours of exposure, ~3.5% of free drug was detected from cells treated with complex sample, and ~7% from cells treated with EV only sample.



**Figure S6.** The influence of FGF7: $\beta$ -CD complex on the time-dependent rate of Caco-2 cells uptake and retention of EV for an administered dose of 6.52  $\mu$ M. The free EV concentration was measured by HPLC.



**Figure S7.** Standard chromatogram of EV (6.25  $\mu$ g/mL).

## 4. Target Identification of β-CD and EV

Swiss target prediction software has been used for predicting molecular targets of  $\beta$ -CD and EV <sup>3</sup>. SMILES of  $\beta$ -CD and EV were obtained from PubChem (Table S1). The consensus molecular targets predicted by Swiss for  $\beta$ -CD and EV are listed in Table S3 and S4.

#### Table S1. Canonical SMILES of β-CD and EV

Name	SMILE
CD (PUBCHEM CID: 444041)	C(C1C2C(C(C(01)0C3C(OC(C(C30)0)0C4C(OC(C(C40)0)0C5C(OC(C(C50)0)0C6 C(OC(C(C60)0)0C7C(OC(C(C70)0)0C8C(OC(02)C(C80)0)C0)C0)C0)C0)C0)C0) 0)0)0
EV (PUBCHEM CID: 6442177)	CC1CCC2CC(C(=CC=CC=CC(CC(C(=O)C(C(C(=CC(C(=O)CC(OC(=O)C3CCCCN3C(= O)C(=O)C1(O2)O)C(C)CC4CCC(C(C4)OC)OCCO)C)C)O)OC)C)C)C)C)OC

Swiss predicts the molecular targets of small molecules based on their 2D and 3D similarity by comparing the query molecule to a library of 280 thousand compounds. Table S2 and S3 listed the molecular targets of  $\beta$ -CD and EV respectively. With high probability Swiss predicted that  $\beta$ -CD could target FGF-7 and FGF-10 (also known as KGF-1 and KGF-2, respectively) based on its 3D similarity to CHEMBL198643 (Figure S6) with a similarity score of 0.758 out of 1 EV targets Serine/threonine-protein kinase mTOR.



Figure S8. 2D structure of A) β-CD, B) CHEMBL198643

It has been reported that CHEMBL198643 strongly binds with FGF-1 and 2<sup>4</sup>. Based on the shape-similarity theory which states that molecules possessing similar 3D structure might exhibit analogous biological activity, we thought that  $\beta$ -CD would be able to binds to fibroblast growth factors. This is could be supported by the strong affinity of a similar structure ( $\beta$ -CD Tetradecasulfate) toward FGF <sup>5</sup>. Heparin is essential for FGF to generate a biological response through binding to FGF receptor. Heparin functions by binding to several FGF molecules forming FGF oligomerization. Then the FGF-heparin complex bind to couple of FGFRs, this leads to FGFR dimerization which

activates tyrosine kinase pathway. A study showed that synthetic heparin analog which can only bind to one FGF blocks the dimerization of FGFR, thus stopping its activation. We propose that  $\beta$ -CD antagonizes the action of heparin by binding to only one FGF, therefore it cannot induce FGF oligomerization thus preventing FGFR dimerization and activation <sup>6,7</sup>.

# 5. Binding ability of $\beta$ -CD and EV to FGF7

Molecular docking was performed via the molecular operating environment (MOE.2014) software for  $\beta$ -CD and EV in the heparin binding site of basic fibroblast growth factor (1BFB.pdb)<sup>8</sup> with scoring affinity London dG and GBVI/WSA dG. The obtained docking affinity scores are shown in Figure S9.

	mol	rseq	mseq	S	rmsd_refine	E_conf	E_place	E_score1	E_refine	E_score2
1	6442177	1	1	-5.0092	5.3896	249.3156	-27.0249	-8.0561	-17.0150	-5.0092
2	6442177	1	1	-4.8737	3.2159	238.9634	-41.8554	-8.3144	-14.0658	-4.8737
3	6442177	1	1	-4.8022	1.6303	256.8998	-21.7023	-8.5919	-15.1917	-4.8022
4	6442177	1	1	-4.7126	2.8163	245.0996	-23.7716	-8.0905	-13.0527	-4.7126
5	6442177	1	1	-4.7104	2.2270	238.7883	-38.0680	-9.0713	-12.1594	-4.7104
6	6442177	1	1	-4.6170	2.7003	254.6145	-3.1844	-7.9129	-17.1669	-4.6170
7	6442177	1	1	-4.4615	1.7729	238.0406	-30.6175	-9.5801	-14.7607	-4.4615
8	6442177	1	1	-4.2688	2.2017	237.3359	-54.2724	-8.1642	-11.7695	-4.2688
9	6442177	1	1	-4.0870	4.2871	247.2750	-40.7168	-8.3377	-8.9479	-4.0870
10	6442177	1	1	-4.0772	2.0780	256.3575	7.1687	-8.6806	-7.7938	-4.0772
11	444041	1	2	-5.2439	1.9786	624.7195	-64.3874	-9.1450	-18.8291	-5.2439
12	444041	1	2	-4.8375	2.8882	621.0886	-7.2920	-9.5388	-15.1178	-4.8375
13	444041	1	2	-4.5602	2.9452	615.8110	-78.4285	-9.1247	-11.2252	-4.5602
14	444041	1	2	-4.1812	2.3501	631.4261	-66.3765	-9.6591	-9.7925	-4.1812
15	444041	1	2	-4.0467	4.3664	631.7632	-51.2175	-9.3040	-10.8069	-4.0467
16	444041	1	2	-4.0261	3.1508	625.8408	-50.4002	-10.1711	-11.2005	-4.0261
17	444041	1	2	-3.6537	1.6517	623.4816	-80.9887	-9.6476	-9.3564	-3.6537
18	444041	1	2	-3.6457	2.4040	617.8505	-61.2162	-9.5400	-8.0208	-3.6457
19	444041	1	2	-3.6078	3.0788	624.0917	-39.1023	-9.8357	-9.6159	-3.6078
20	444041	1	2	-3.5234	2.3457	624.4732	-56.9427	-9.2841	-10.6297	-3.5234

**Figure S9.** Docking scores for binding ability of  $\beta$ -CD (PubChem CID: 444041) and EV (PubChem CID: 6442177) to FGF7, obtained by MOE.2014 software.

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