# Molecular Modeling Based Delivery System Enhances Everolimus-Induced Apoptosis in Caco-2 Cells

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**1. Assessment of FGF7:β-CD:EV complex stability and EV release profile in Dulbecco's Modified Eagle Medium (DMEM)**



**Figure S1. Dialysis tubing procedure**

# **2. Cytotoxicity of FGF7:β-CD:EV complex on normal cells**



**Figure S2.** Cytotoxic effect of EV on FHs 74 Int cells as displayed by the RTCA DP instrument. Cells were seeded overnight to reach the log phase, then incubated with 3.26  $\mu$ M EV (A), 6.52  $\mu$ M EV (B), or 13.04  $\mu$ M EV (C). IC<sub>50</sub> value of EV on FHs 74 Int cells was found to be  $19.25 \pm 1.35$   $\mu$ M.



**Figure S3.** Cytotoxic effect of FGF7:EV on FHs 74 Int cells as displayed by the RTCA DP instrument. Cells were seeded overnight to reach the log phase, then incubated with FGF7:EV complex (contains 1.58 μM FGF7 and 3.26 μM EV) (A), FGF7:EV (contains 1.58 μM FGF7 and 6.52 μM EV) (B), or FGF7-EV (contains 1.58 μM FGF7 and 13.04 μM EV) (C). IC<sub>50</sub> value of EV on FHs 74 Int cells was found to be  $15.9 \pm 0.95$   $\mu$ M.



**Figure S4.** Cytotoxic effect of β-CD:EV on FHs 74 Int cells as displayed by the RTCA DP instrument. Cells were seeded overnight to reach the log phase, then incubated with β-CD:EV inclusion complex (contains 3.26 μM EV) (A), β-CD:EV inclusion complex (contains 6.52 μM EV) (B), or β-CD:EV inclusion complex (contains 13.04 μM EV) (C). IC<sub>50</sub> value of EV on FHs 74 Int cells was found to be  $16.55 \pm 1.05$   $\mu$ M.



**Figure S5.** Cytotoxic effect of FGF7:β-CD:EV complex on FHs 74 Int cells as displayed by the RTCA DP instrument. Cells were seeded overnight to reach the log phase, then incubated with FGF7:β-CD:EV complex (contains 1.58 μM FGF7, 55.07 μM β-CD and 3.26 μM EV) (A), FGF7:β-CD:EV complex (contains 1.58 μM FGF7, 110.13 μM β-CD and 6.52 μM EV) (B) or FGF7:β-CD:EV complex (contains 1.58 μM FGF7, 220.27 μM β-CD and 13.04 μM EV) (C). IC<sub>50</sub> value of EV on FHs 74 Int cells was found to be  $34.11 \pm 1.9 \mu M$ .

#### **3. Assessment of FGF7:β-CD:EV complex retention in Caco-2 cells**

#### *Chromatographic conditions*

The method is modified from Carpentier et al.<sup>1</sup> and Spandana et al. <sup>2</sup>. Briefly, treated cells were detached and centrifuged at 400g x 7 minutes, then 0.1 ml of tetraborate buffer (pH 9.8), 0.1 ml of EV solution (10 mg/L) as internal standard, and 1.8 ml of dichloromethane/methanol 4:l (v/v) were immediately added to the cell pellet. After a vigorous agitation, the organic phase was removed and evaporated to dryness. The dry residue was dissolved in the mobile phase: acetonitrile/double ultra-pure water 95:05 (v/v), and injected onto an Ascentis® C18 column (Supelco, Bellefonte PA, USA). The flow rate was 1 ml/min, the peaks were detected by PDA detection and wave length was set at 278 nm.

The results indicated that the penetration of EV from both samples was time-dependent, thus the accumulation was significantly increased with FGF7:β-CD:EV complex application, only 3% of free EV was detected. The retention of EV was enhanced with the complex application as well. After 24 hours of exposure, ~3.5% of free drug was detected from cells treated with complex sample, and ~7% from cells treated with EV only sample.



**Figure S6.** The influence of FGF7:β-CD complex on the time-dependent rate of Caco-2 cells uptake and retention of EV for an administered dose of 6.52 μM. The free EV concentration was measured by HPLC.



**Figure S7.** Standard chromatogram of EV (6.25  $\mu$ g/mL).

## **4. Target Identification of β-CD and EV**

Swiss target prediction software has been used for predicting molecular targets of β-CD and EV  $^3$ . SMILES of β-CD and EV were obtained from PubChem (Table S1). The consensus molecular targets predicted by Swiss for β-CD and EV are listed in Table S3 and S4.

### **Table S1. Canonical SMILES of β-CD and EV**



Swiss predicts the molecular targets of small molecules based on their 2D and 3D similarity by comparing the query molecule to a library of 280 thousand compounds. Table S2 and S3 listed the molecular targets of β-CD and EV respectively. With high probability Swiss predicted that β-CD could target FGF-7 and FGF-10 (also known as KGF-1 and KGF-2, respectively) based on its 3D similarity to CHEMBL198643 (Figure S6) with a similarity score of 0.758 out of 1 EV targets Serine/threonine-protein kinase mTOR.



**Figure S8.** 2D structure of A) β-CD, B) CHEMBL198643

It has been reported that CHEMBL198643 strongly binds with FGF-1 and  $2<sup>4</sup>$ . Based on the shape-similarity theory which states that molecules possessing similar 3D structure might exhibit analogous biological activity, we thought that β-CD would be able to binds to fibroblast growth factors. This is could be supported by the strong affinity of a similar structure (*β-CD* Tetradecasulfate) toward FGF<sup>5</sup>. Heparin is essential for FGF to generate a biological response through binding to FGF receptor. Heparin functions by binding to several FGF molecules forming FGF oligomerization. Then the FGF-heparin complex bind to couple of FGFRs, this leads to FGFR dimerization which

activates tyrosine kinase pathway. A study showed that synthetic heparin analog which can only bind to one FGF blocks the dimerization of FGFR, thus stopping its activation. We propose that β-CD antagonizes the action of heparin by binding to only one FGF, therefore it cannot induce FGF oligomerization thus preventing FGFR dimerization and activation  $6, 7$ .

# **5. Binding ability of β-CD and EV to FGF7**

Molecular docking was performed via the molecular operating environment (MOE.2014) software for β-CD and EV in the heparin binding site of basic fibroblast growth factor (1BFB.pdb)<sup>8</sup> with scoring affinity London dG and GBVI/WSA dG. The obtained docking affinity scores are shown in Figure S9.



**Figure S9.** Docking scores for binding ability of β-CD (PubChem CID: 444041) and EV (PubChem CID: 6442177) to FGF7, obtained by MOE.2014 software.

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