# Increased lymphocyte activation and atherosclerosis in CD47-deficient mice

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### Supplemental Figure 1. Plaque and immune phenotype in Cd47<sup>-/-</sup> mice

Experimental outline (A). Aortic sinus macrophage area (B) and percent macrophage area (C) as determined by immunohistochemistry. TUNEL staining at either 10x or 40x magnification (D). Aortic CD4<sup>+</sup> (E) and CD8<sup>+</sup> (F) T cells determined by flow cytometry. Splenocyte count (G). Percentage of naïve T cells (H) and IFN- $\gamma$  production of CD4<sup>+</sup> and CD8<sup>+</sup> T cells (I). Levels of effector (J) and naïve (K) T cells in aorta-draining LN. Plasma cytokine and chemokine analysis (L; n=6-7 mice/group). Gating example for MHC-II<sup>+</sup>CD11c<sup>hi</sup> dendritic cells (M) and splenic DC quantification (N). \* p<0.05, \*\* p<0.01, \*\*\* p<0.001



# **Supplemental Figure 2**. Immune perturbations observed in Cd47<sup>-/-</sup> mice are not contingent on hypercholesterolemia

Splenovytes from chow-fed *Cd47<sup>-/-</sup>* and WT mice were analyzed by flow cytometry. Percentage (A) and numbers (B) of CD44<sup>hi</sup>CD62L<sup>low</sup> CD4<sup>+</sup> and CD8<sup>+</sup> T-effector cells. Percentage (C) and numbers (D) of IFN-γ-producing CD4<sup>+</sup> and CD8<sup>+</sup> T cells. Percentage of DCs (E; MHC-II<sup>+</sup>CD11c<sup>hi</sup>) and SIRPα<sup>+</sup>CD11b<sup>+</sup> DCs (F) of total DCs. DCs were analyzed for expression of SIRPα vs. CD4 (G) and SIRPα vs. CD8 (H). \* p<0.05, \*\* p<0.01, \*\*\* p<0.01



**Supplemental Figure 3.** Depletion of T-cells after anti-CD4/anti-CD8 treatment WT and *Cd47<sup>-/-</sup>* mice (n=7-9) injected with depleting anti-CD4/anti-CD8 mAb were analyzed for T-cells in blood (A-B) and digested aorta (C).



#### Supplemental Figure 4. CD47-deficiency affects NK phenotype

Cd47<sup>-/-</sup> and WT mice (n=7-8) were fed high-cholesterol diet for 8 weeks. Percentage of NK cells in blood and liver (A). Gating (B) and quantification of CD25<sup>+</sup> NK cells in spleen (C). Analysis of CD27, CD127, CD11b and KLRG1 expression on splenic NK cells (D). Numbers of splenic IFNy<sup>+</sup> NK cells (E) and levels of Granzyme B<sup>+</sup> NK cells (F) after stimulation with with Brefeldin A or PMA/ionomycin/Brefeldin A. *Cd47<sup>-/-</sup>* and WT mice (n=7-8) were fed chow diet and spleen and liver analyzed for NK cell subsets using flow cytometry (G-J). Violet labeled RMA and RMA-S cells (1:1 ratio) were co-injected i.p. into *Cd47<sup>-/-</sup>* and WT mice (n=3-5) and recovered 48h later by peritoneal lavage (K). Quantification of %RMA-S (L) #RMA-S (M) and %NK1.1 (N) in peritoneal lavage. \* p<0.05, \*\* p<0.01, \*\*\* p<0.01



#### Supplemental Figure 5. Flow cytometry analysis of murine aorta

Aortas from LDLr-/- mice were digested, treated with PMA/ionomycin/Brefeldin A for 4 hours and stained for expression of surface markers and intracellular IFN- $\gamma$ . Gating for IFN- $\gamma$  production in CD90<sup>+</sup> and CD90<sup>-</sup> NK cells (CD45<sup>+</sup>Zombie<sup>-</sup>NK1.1<sup>+</sup>CD3<sup>-</sup>) shown for .



## Supplemental Figure 6. Immune phenotype of WT mice treated with anti-CD47 (MIAP410)

Experimental design (A) of anti-CD47 treatment of WT mice given AAV-PCSK9<sup>DY</sup> and fed high-fat diet (n=10/group). Percentage of splenic DCs (B). Levels of leukocyte subsets in spleen (C) and blood (D). Numbers of NK cells in spleen (E). \* p<0.05, \*\* p<0.01, \*\*\* p<0.001