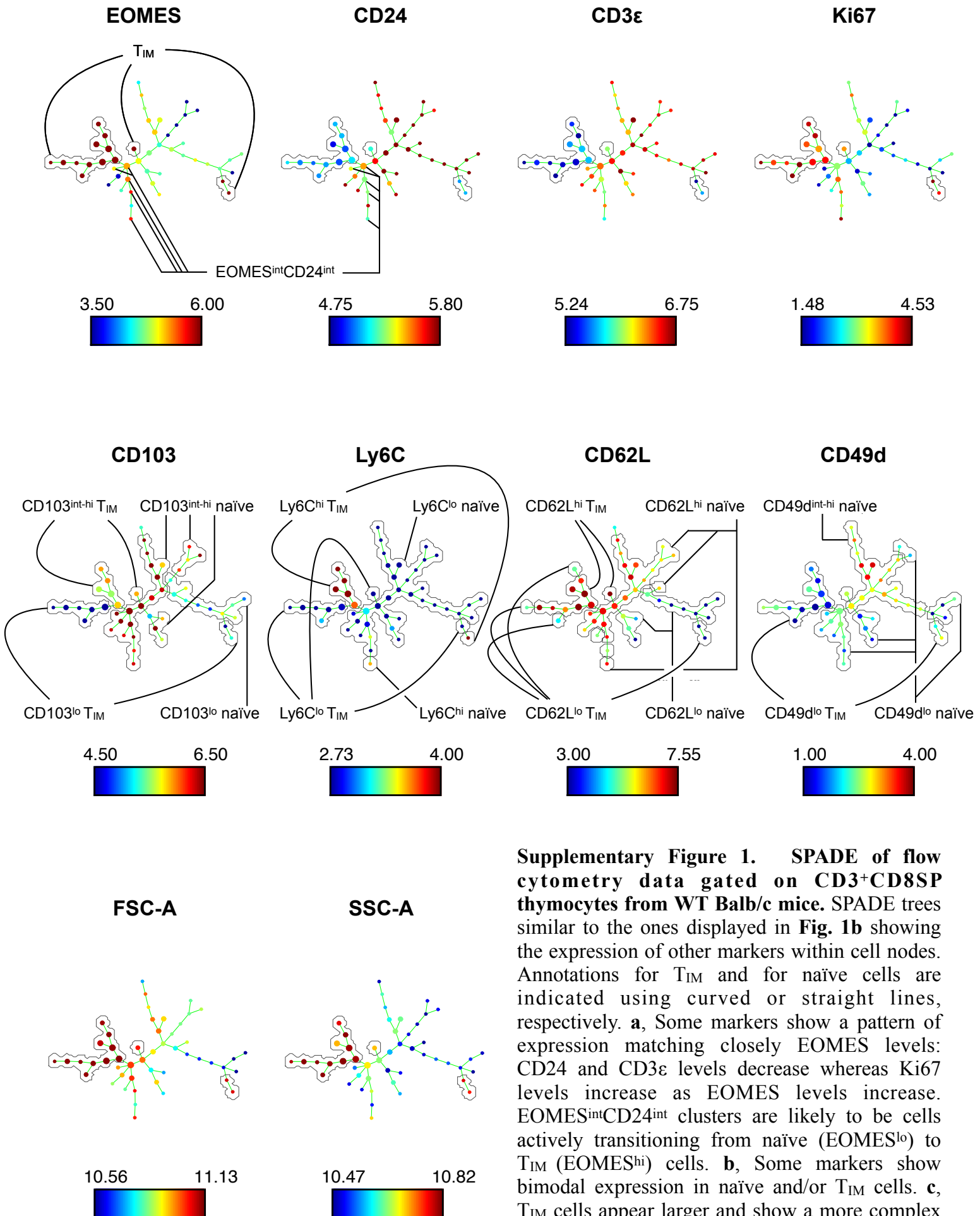


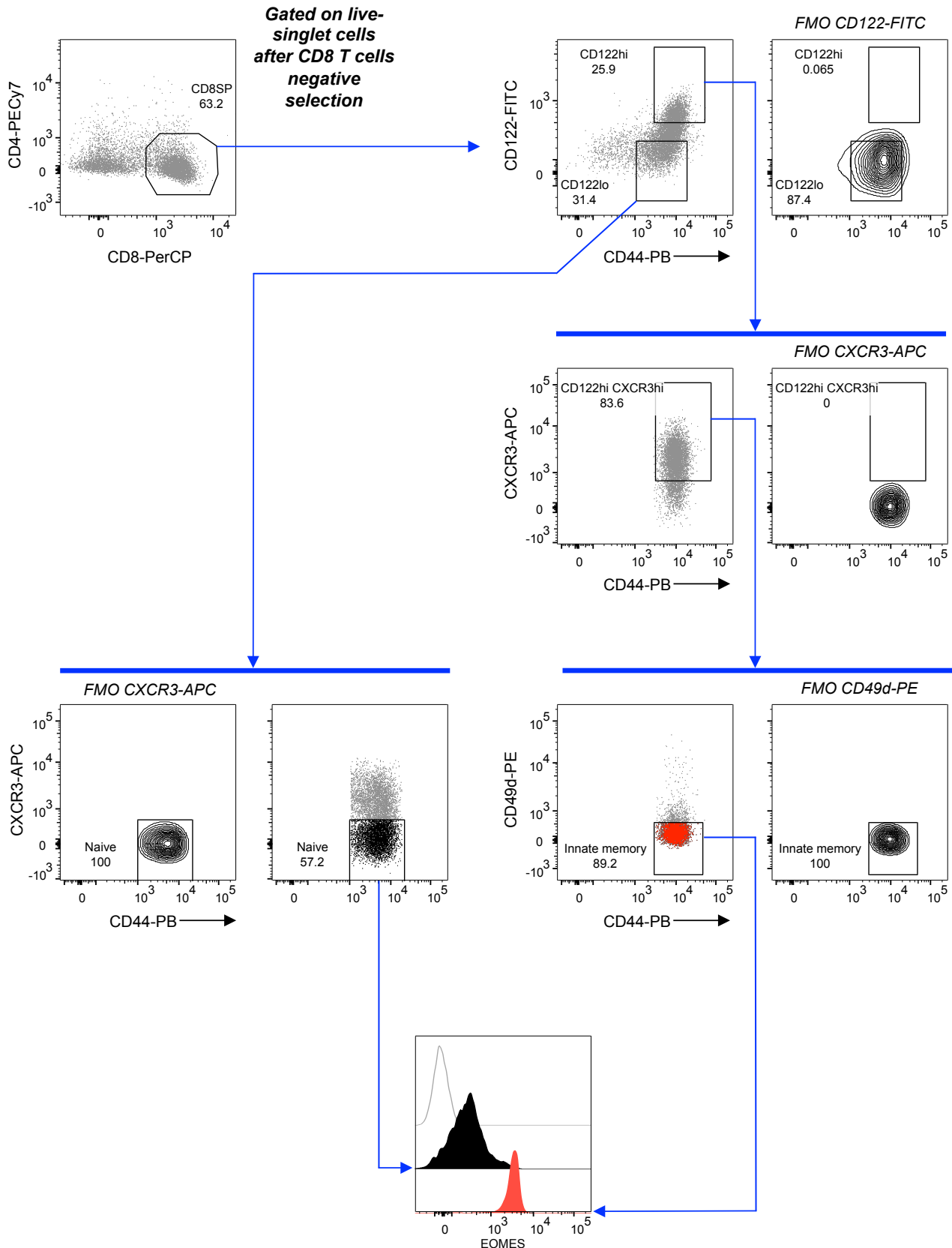
Supplementary information

EOMES interacts with RUNX3 and BRG1 to promote innate memory cell formation through epigenetic reprogramming

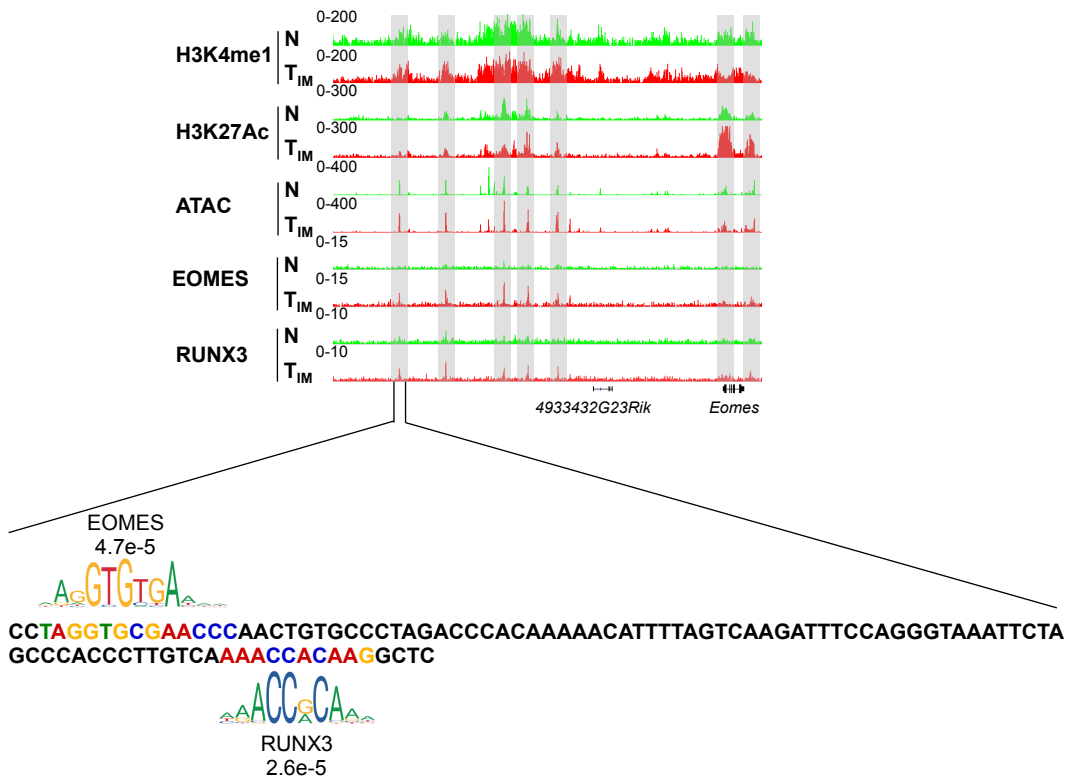
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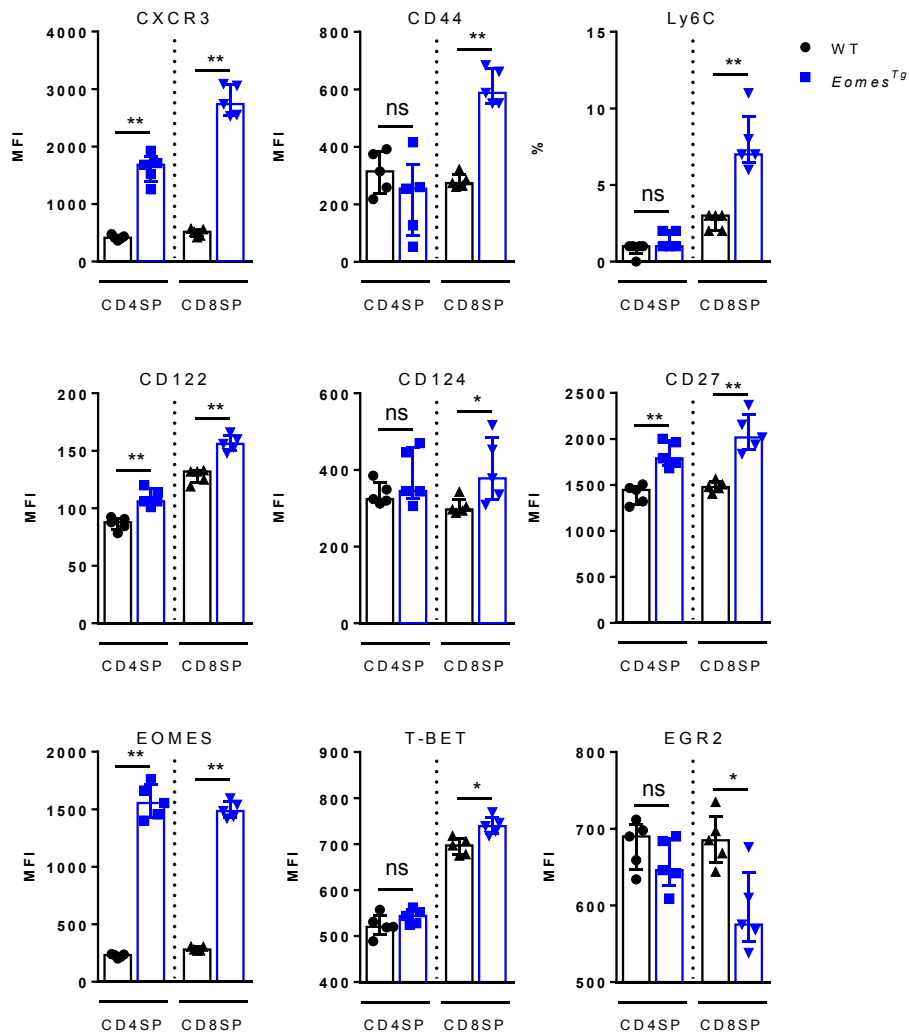
Supplementary Figure 1. SPADE of flow cytometry data gated on CD3⁺CD8^{SP} thymocytes from WT Balb/c mice. SPADE trees similar to the ones displayed in Fig. 1b showing the expression of other markers within cell nodes. Annotations for T_{IM} and for naïve cells are indicated using curved or straight lines, respectively. **a**, Some markers show a pattern of expression matching closely EOMES levels: CD24 and CD3 ϵ levels decrease whereas Ki67 levels increase as EOMES levels increase. EOMES^{int}CD24^{int} clusters are likely to be cells actively transitioning from naïve (EOMES^{lo}) to T_{IM} (EOMES^{hi}) cells. **b**, Some markers show bimodal expression in naïve and/or T_{IM} cells. **c**, T_{IM} cells appear larger and show a more complex granularity than naïve cells.



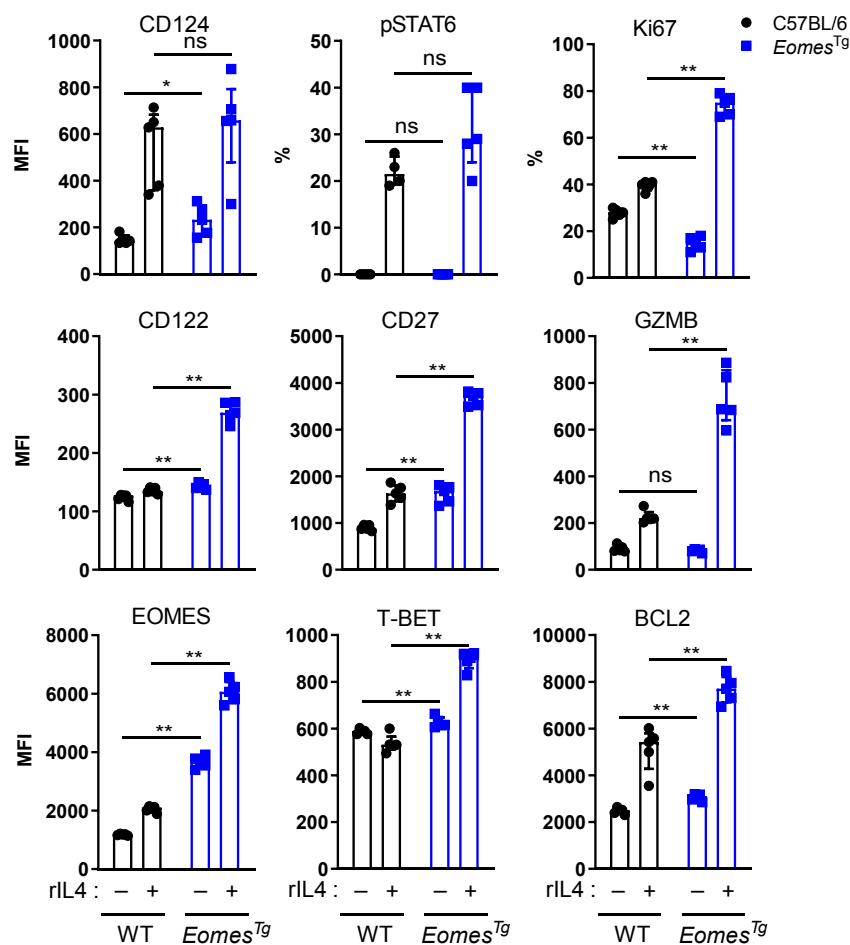
Supplementary Figure 2. Sorting strategy of naïve and T_{IM} CD8SP thymocytes from WT Balb/c mice. After negative selection of CD8⁺ cells, untouched cells were stained to exclude dead cells and incubated with Fc receptor-blocking antibodies and a surface staining antibody mix to be sorted as T_{IM} (CD8⁺CD44^{hi}CXCR3⁺CD122⁺CD49d^{lo}) or naïve (CD8⁺CD44^{lo}CXCR3⁻CD122⁻) cells. The obtained T_{IM} cells are EOMES^{hi}, whereas naïve cells are EOMES^{lo-int}.



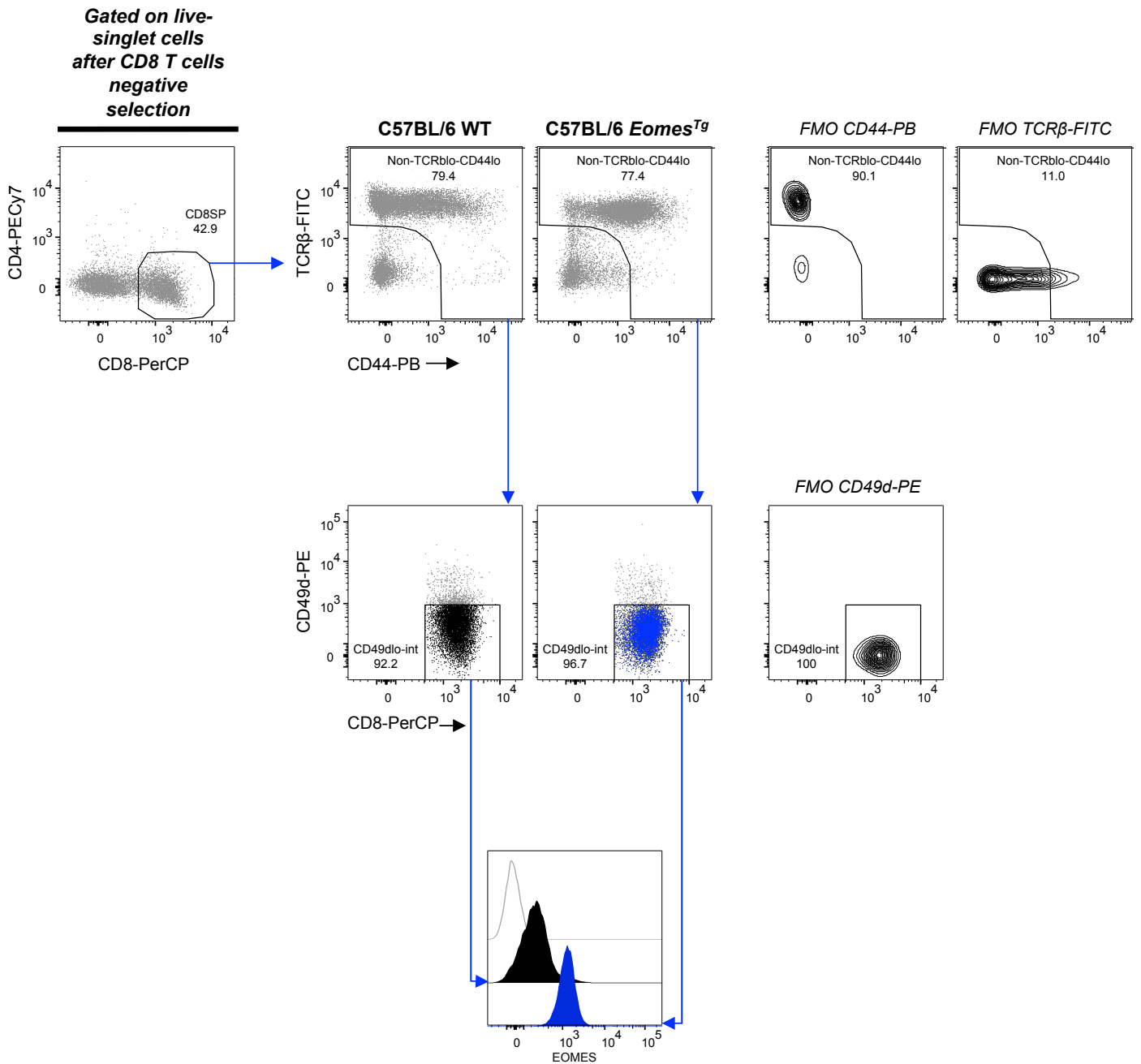
Supplementary Figure 3. Representative EOMES, RUNX3, H3K4me1, H3K27Ac ChIP-seq and ATAC-seq tracks at the *Eomes* locus in naïve and T_{IM} CD8SP thymocytes showing the co-localisation of EOMES and RUNX3 (highlighted in grey, top). Sequence of the indicated region and the location of EOMES and RUNX motifs with their position p-value are shown (down).



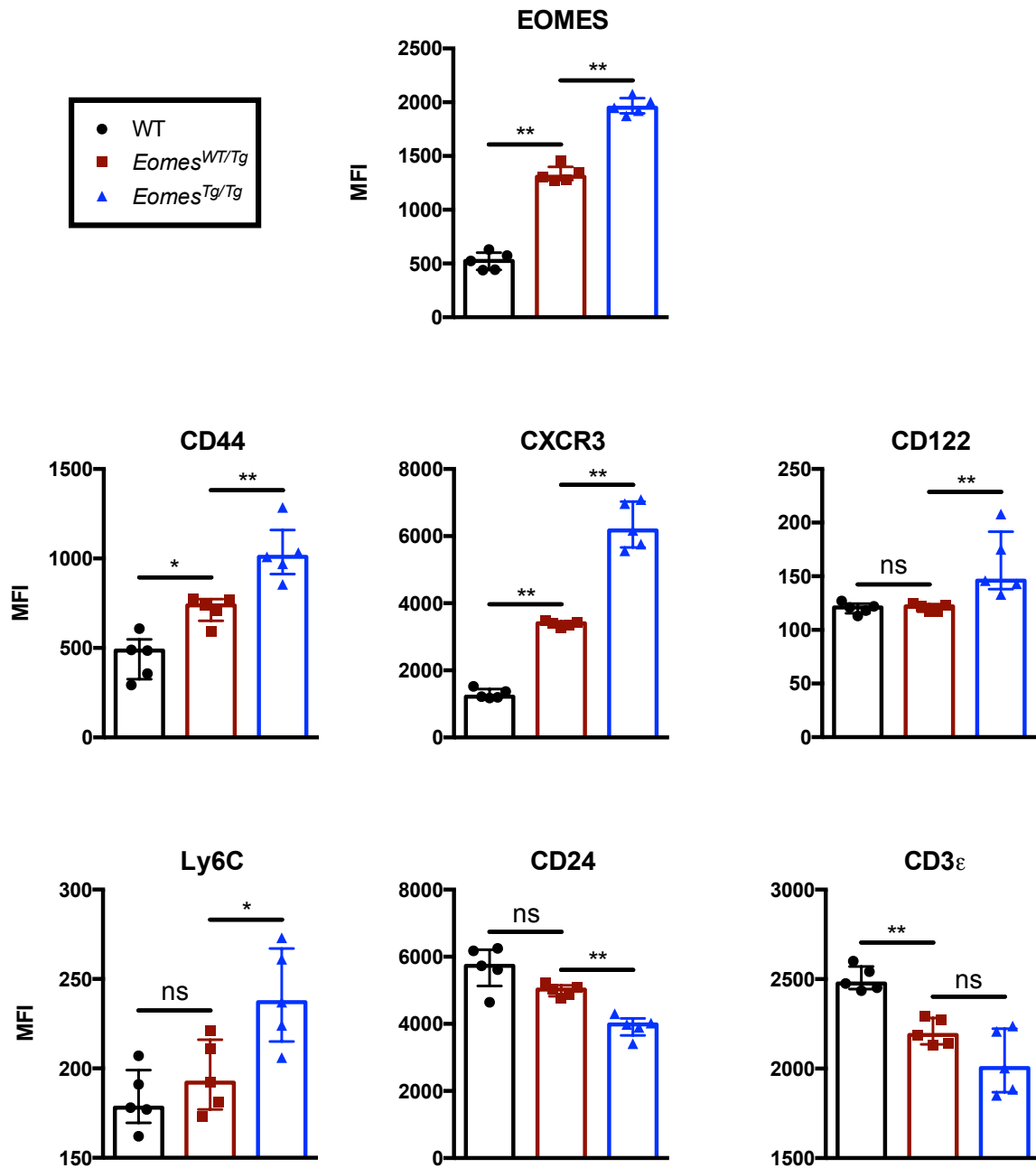
Supplementary Figure 4. Distinct effects of EOMES ectopic expression in CD4SP and CD8SP thymocytes. Thymocytes from WT or *Eomes*^{Tg} mice were analyzed by flow cytometry for the indicated protein. Although some markers follow similar patterns between CD3⁺CD4SP and CD3⁺CD8SP thymocytes upon EOMES overexpression (CXCR3, CD122, CD27), other markers are only impacted in CD3⁺CD8SP (CD44, Ly6C, CD124, T-BET, EGR2), despite similar EOMES levels. Horizontal bars indicate median ± interquartile range (MFI or frequency). Each point represents a single mouse, n=5 mice per group). Statistics were calculated using Mann-Whitney test. ns not significant, * P < 0.05, ** P < 0.01.



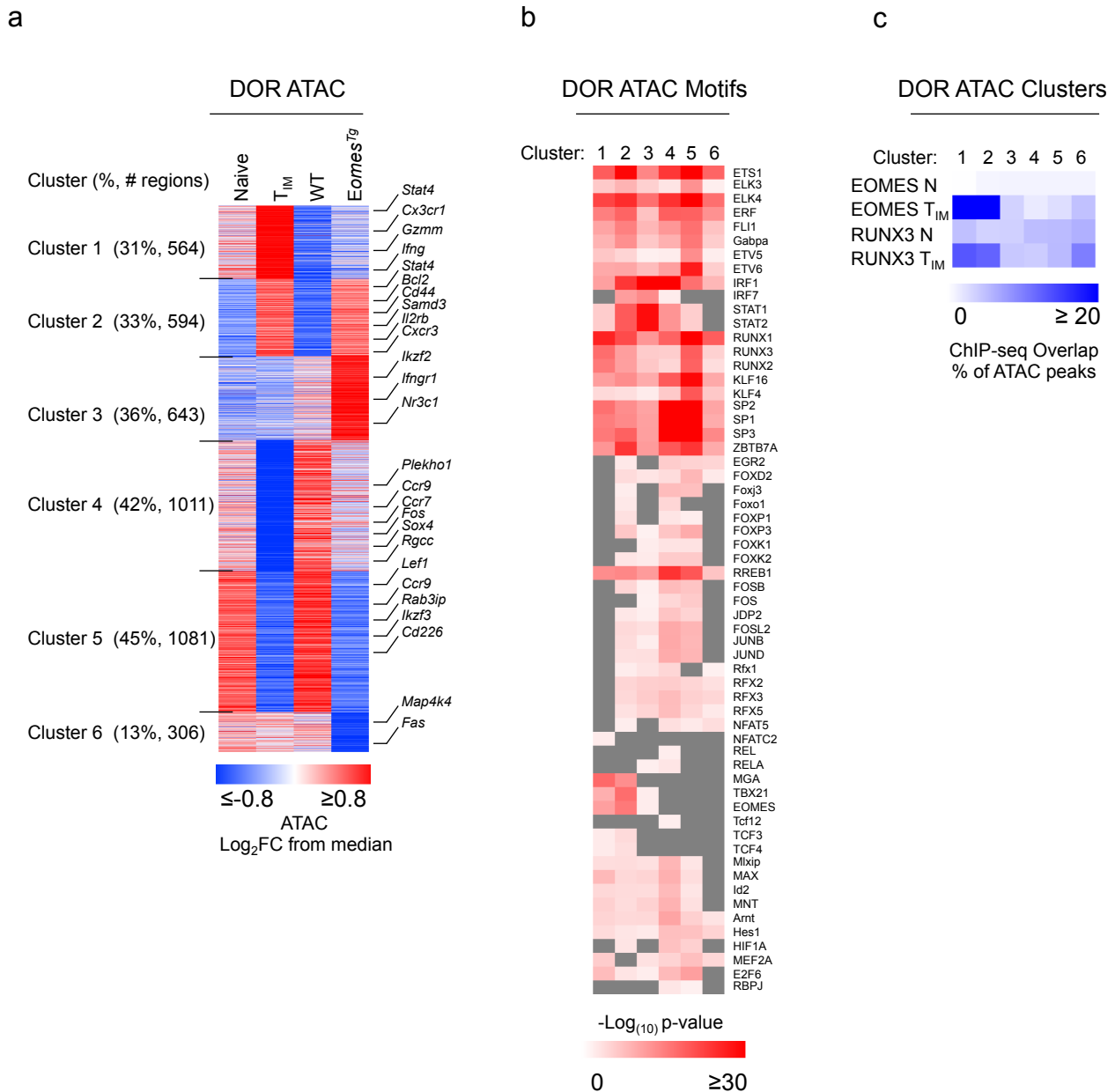
Supplementary Figure 5. CD8SP thymocytes from *Eomes^{Tg}* mice display heightened responsiveness upon *ex vivo* stimulation with rIL-4. Thymocytes from WT and *Eomes^{Tg}* mice were cultured in complete medium with or without rmIL-4 (20 ng ml⁻¹) for 72h, except for phospho-STAT6 staining (30 min culture with or without rmIL-4 at 30 ng ml⁻¹). Although CD124 basal levels are slightly higher in CD3⁺CD8SP thymocytes from *Eomes^{Tg}* mice, early signalling (STAT6 phosphorylation) and latter CD124 upregulation (72h *ex vivo* IL-4 stimulation) are not different as compared to WT cells. Nonetheless, CD3⁺CD8SP thymocytes from *Eomes^{Tg}* mice show stronger responses to IL-4 than WT mice, as they more readily proliferate (Ki67) and upregulate surface receptors (CD122, CD27) along with T-box transcription factors (EOMES, T-BET), effector (GZMB) and anti-apoptotic (BCL2) molecules. Horizontal bars indicate median ± interquartile range (MFI or frequency). Each point represents a single mouse, n=5 mice per group. Statistics were calculated using Mann-Whitney test. ns not significant, * P < 0.05, ** P < 0.01



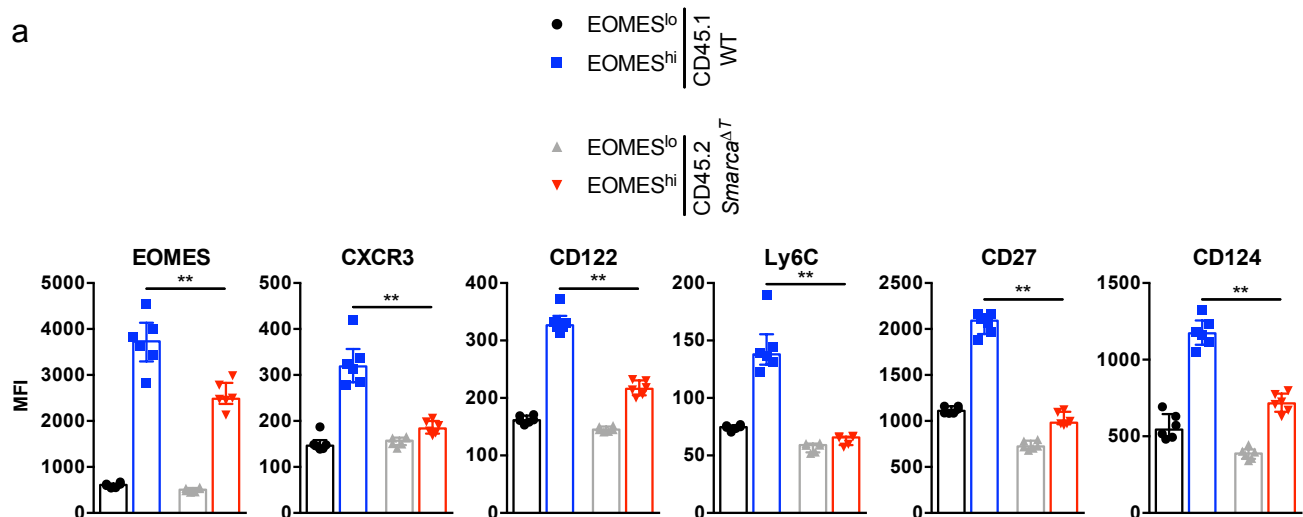
Supplementary Figure 6. Sorting strategy of CD8SP thymocytes from WT and *Eomes*^{Tg} C57BL/6 mice. After CD8⁺ negative selection, untouched cells were stained to exclude dead cells and incubated with Fc receptor-blocking antibodies and a surface staining antibody mix to be sorted as WT or *Eomes*^{Tg} CD8SP thymocytes. In both cases, a TCRβ^{lo}CD44^{lo} population was removed from CD8⁺ cells, as were CD49^{dhi} cells.



Supplementary Figure 7. EOMES ectopic expression in CD3⁺CD8SP thymocytes impacts the expression of several surface markers in a dose-dependent fashion. CD3⁺CD8SP thymocytes from WT, heterozygote (*Eomes^{WT/Tg}*) or homozygote transgenic (*Eomes^{Tg/Tg}*) mice expressing different levels of EOMES were analysed by flow cytometry for the indicated protein. Horizontal bars indicate median ± interquartile range (MFI). Each point represents a single mouse, n=5 mice per group. Statistics were calculated using Mann-Whitney test. ns not significant, * P < 0.05, ** P < 0.01



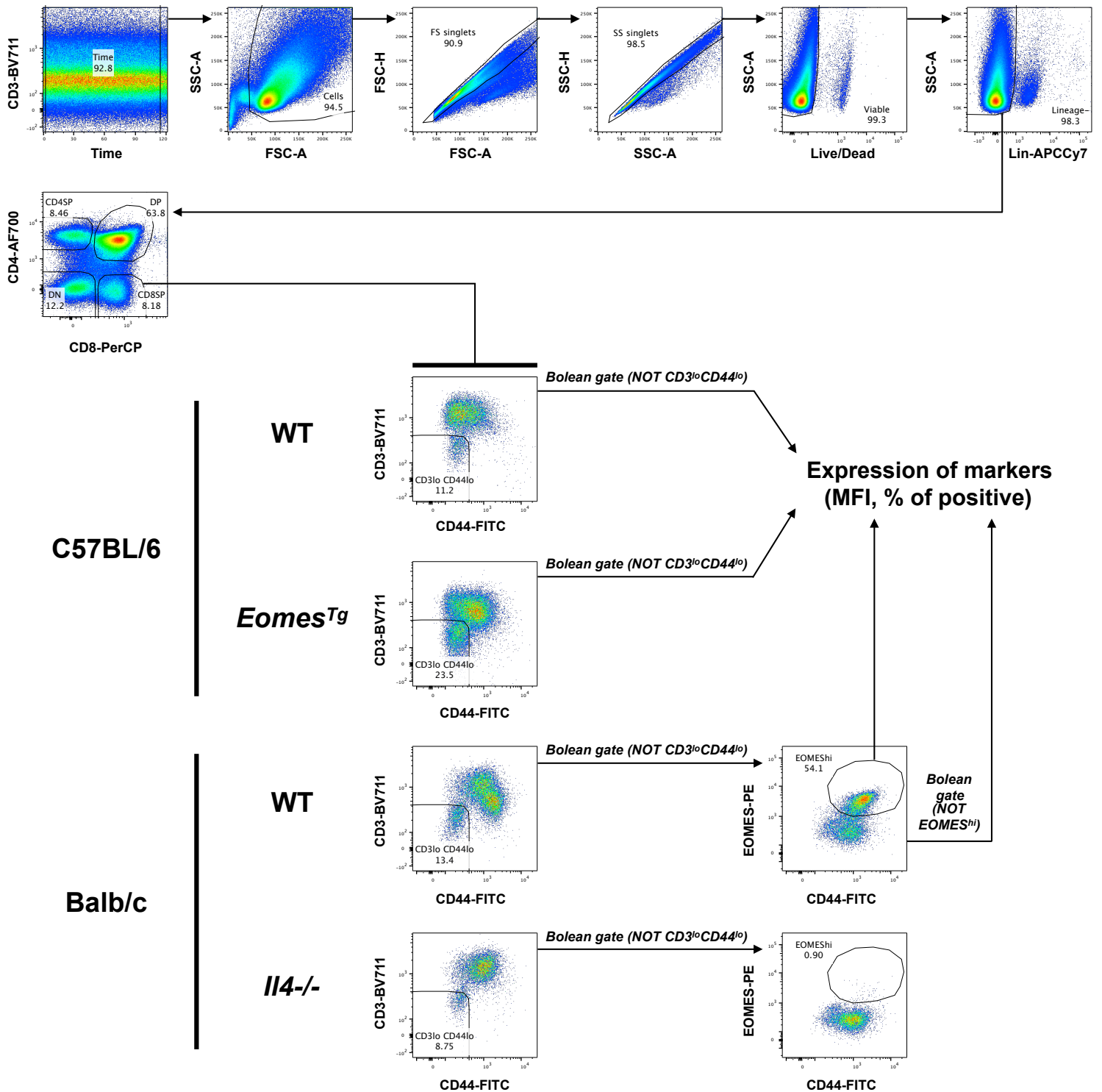
Supplementary Figure 8. T_{IM} and Eomes^{Tg} CD8SP thymocytes share differentially open regions in enhancers characterised by T-box motifs enrichment. **a**, Differentially open regions clustering within enhancers based on ATAC-seq signal in CD8SP thymocytes (Naïve, T_{IM}, WT and Eomes^{Tg}). Clusters in 1-3 and 4-6 are more or less open, respectively. Selected genes associated with each cluster are displayed in the right margin. **b**, Motif enrichment analysis of differentially active enhancers for clusters shown in **a** using AME and presented as -Log₁₀ of p values. **c**, Percentage of regions within each enhancers' cluster that overlaps with ChIP-seq peaks for EOMES and RUNX3. ChIP-seq was performed on 3 independent immunoprecipitations (n=5 mice per sample).



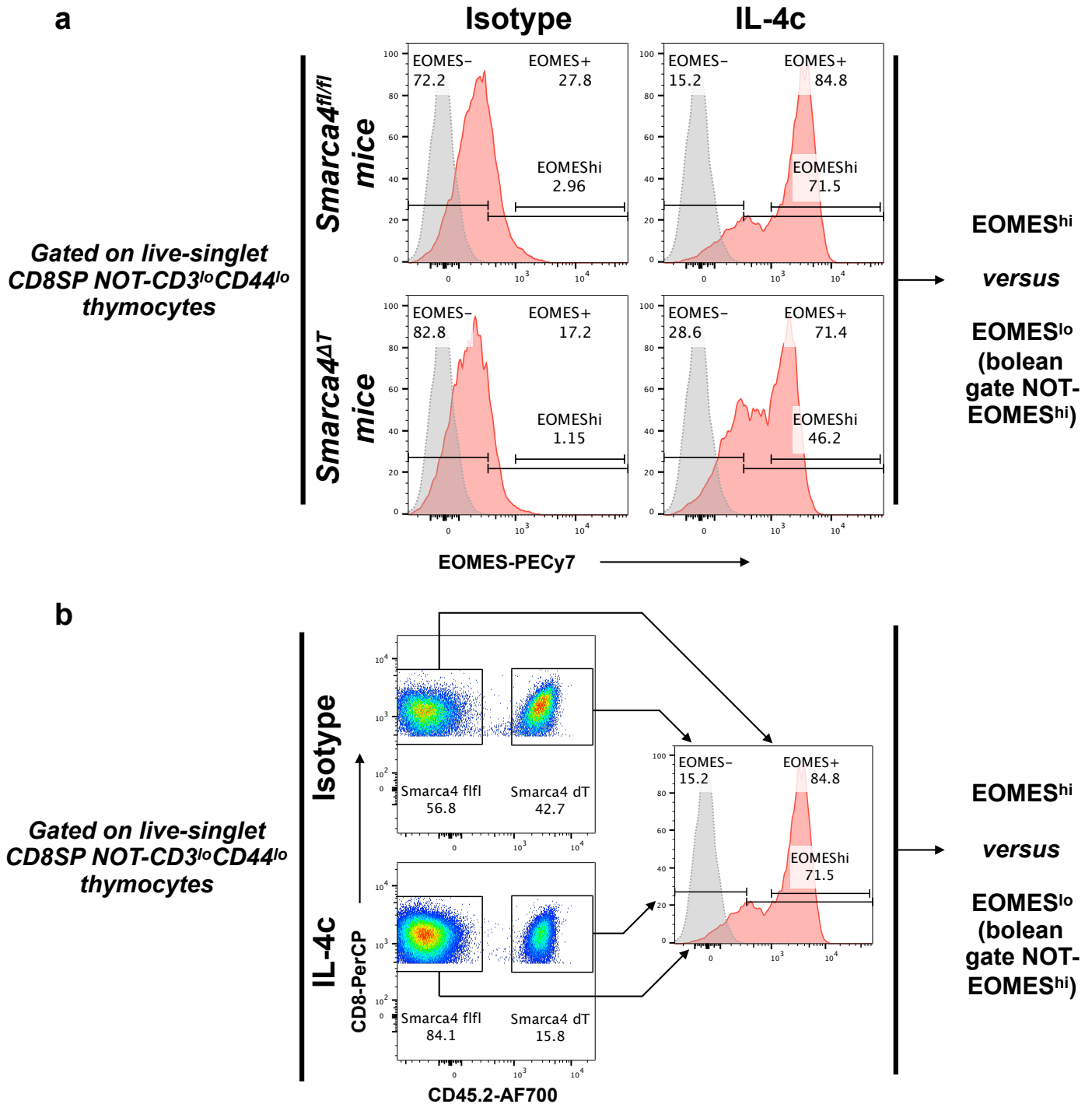
b

	scEOMES (x)	scCXCR3 (y)	scCD122 (y)	scLy6C (y)	scCD27 (y)	scCD124 (y)
Kendall's Tau	<i>Smarca4</i> ^{fl/fl}	0.33	0.43	0.26	0.32	0.30
	<i>Smarca4</i> ^{ΔT}	0.06	0.19	0.08	0.13	0.24
Fisher's Z transformation (p-value)	Are correlations different?	Yes Z=28.94 (p<0.001)	Yes Z=27.23 (p<0.001)	Yes Z=19.33 (p<0.001)	Yes Z=9.67 (p<0.001)	Yes Z=8.07 (p<0.001)

Supplementary Figure 9. a, Data from the mixed bone marrow chimera experiment showing the expression (MFI) of cell markers in WT (CD45.1) or *Smarca4*^{ΔT} (CD45.2) EOMES^{lo} and EOMES^{hi} CD3⁺CD8SP thymocytes after IL-4c injections. Horizontal bars indicate median ± interquartile range (MFI). Each point represents a single mouse, n=6 mixed bone marrow chimera (CD3ε^{-/-} mice previously irradiated and injected with a mix of 50% bone marrow from CD45.1 WT mice and 50% bone marrow from CD45.2 *Smarca4*^{ΔT} mice) that all received 4 daily intraperitoneal IL-4c injections (see **Fig. 7d** experimental scheme). Statistics were calculated using Mann-Whitney test. ** P < 0.01. **b**, Kendall's Tau from correlations between single-cell (sc) fluorescences for EOMES and scCXCR3, scCD122, scLy6C, scCD27, and scCD124 are shown for the two representative IL-4c-injected mice (*Smarca4*^{fl/fl} and *Smarca4*^{ΔT}) displayed in **Fig. 7c** and compared using Fisher's Z transformation.



Supplementary Figure 10. Sequential gating strategy used to select, within a stable acquisition time frame, live-singlet non-CD3^{lo}CD44^{lo} CD8SP thymocytes (more generally referred to throughout the manuscript as CD3⁺CD8SP thymocytes) and Balb/c EOMES^{hi} and EOMES^{lo} (Bolean gate NOT EOMES^{hi}) CD3⁺CD8SP thymocytes. This strategy was universally applied to select CD3⁺CD8SP thymocytes (Fig. 1a-c, Fig. 5a-c, Fig. 7c-f, Supplementary Figures 1, 4, 5, 7, 9 and 11), with the particularity that the selection of Lineage-negative cells (Lin: CD11b-CD11c-CD19-Gr1-NK1.1) was only applied in Fig. 5a to assess the frequencies of double negative-positive (DN-DP) and CD4-CD8 single positive (SP) thymocytes. CD8⁺ splenocytes (Fig. 7g,h) were selected as shown here for CD8SP thymocytes (without the Lineage gate) and naïve (N) CD8⁺ splenocytes were defined with a boolean gate NOT-TM AND NOT-VM (gating of VM and TM cells is shown in Fig. 7g).



Supplementary Figure 11. a, Gating strategy used for the comparison of EOMES^{lo} and EOMES^{hi} CD3⁺CD8SP thymocytes from *Smarca4^{fl/fl}* and *Smarca4^{AT}* mice after isotype control or IL-4 complex (IL-4c) injection (this gating strategy was used for **Fig. 7c,e,f** and **Supplementary Figure 9b**). **b**, Gating strategy used for the comparison of EOMES^{lo} and EOMES^{hi} cells among *Smarca4^{fl/fl}* (CD45.1⁺CD45.2⁻) or *Smarca4^{AT}* (CD45.1⁻CD45.2⁺) CD3⁺CD8SP thymocytes from mixed bone marrow chimeras (see experimental scheme, **Fig. 7d**) after isotype control or IL-4 complex (IL-4c) injection (this gating strategy was used for **Fig. 7d** and **Supplementary Figure 9a**). Gray histograms represent fluorescence minus one controls, red histograms represent EOMES expression on gated cells. Percentages of cell populations are shown.