

Supplemental Information

Table S1: Diffraction data and crystallographic refinement statistical values

| Structure | CaMKII kinase domain /dEAG _{long} / Mg ²⁺ -ADP | CaMKII kinase domain /dEAG _{long} / Mg ²⁺ -AMPPN | CaMKII ^{D136N} kinase domain /dEAG _{long} ^P / Mg ²⁺ -ADP |
|---|--|--|--|
| PDB entry | 5FG8 | 5H9B | 5HU3 |
| Diffraction data | | | |
| Space group | P12 ₁ 1 | P12 ₁ 1 | P12 ₁ 1 |
| Unit cell parameters a, b, c (Å), α=γ, β | 36.50, 59.22, 70.76, 90.00, 97.23 | 35.82, 59.13, 70.34, 90.00, 99.02 | 36.91, 59.22, 70.30, 90.00, 96.68 |
| Wavelength (Å) | 0.98403 | 0.98406 | 0.97625 |
| Resolution range (Å) | 30.19-1.88 (1.95-1.88) | 45.03-2.25 (2.33-2.25) | 30.08-1.82 (1.88-1.82) |
| No. of unique reflections | 23,764 (2,099) | 13,806 (1,342) | 26,540 (2,412) |
| No. of measured reflections | 77,000 (5,111) | 47,277 (4,686) | 87,981 (5,854) |
| Multiplicity | 3.2 (2.4) | 3.4 (3.5) | 3.3 (2.4) |
| Completeness (%) | 97.2 (88.0) | 99.4 (99.5) | 97.9 (91.0) |
| I/σI | 12.4 (1.5) | 10.4 (1.7) | 19.3 (1.5) |
| R _{meas} (%) | 6.0 (86.2) | 8.0 (93.9) | 4.1 (73.6) |
| Refinement data | | | |
| Resolution range (Å) | 30.19-1.96 (2.04-1.96) | 45.03-2.25 (2.42-2.25) | 30.08-1.89 (1.96-1.89) |
| No. of reflections | 21,328 | 13,780 | 24,034 |
| R _{work} /R _{free} (%) | 19.1/22.1 (29.9/30.3) | 20.3/22.2 (32.2/37.8) | 19.2/22.8 (39.7/42.0) |
| No. of atoms in model: | | | |
| Protein | 2,304 | 2,288 | 2,271 |
| Solvent | 69 | 22 | 79 |
| Nucleotide | 27 | 27 | 27 |
| Mg ²⁺ | 1 | 1 | 1 |
| Average B value (Å ²): | | | |
| Protein | 58 | 76 | 49 |
| Solvent | 44 | 49 | 41 |
| Nucleotide | 43 | 69 | 39 |
| Mg ²⁺ | 46 | 72 | 39 |
| RMSD bond length (Å) | 0.010 | 0.004 | 0.017 |
| RMSD bond angle (°) | 1.171 | 0.758 | 1.347 |

RMSD: root-mean-square deviation. High-resolution bin statistics are shown in parentheses.

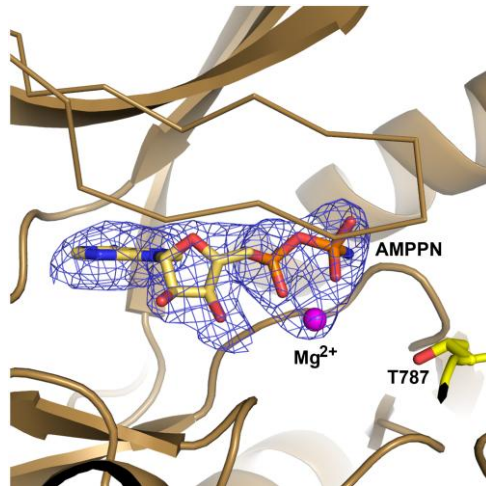


Figure S1: CaMKII kinase domain bound with AMPPN

View of active site of native CaMKII kinase domain (residues 1-283) crystallized in the presence of *d*EAG_{long} and AMPPNP. Mesh shows omit density map revealing the presence of AMPPN. Kinase shown as dark-wheat cartoon and T787 in channel fragment as yellow stick. AMPPN and Mg²⁺ are indicated.

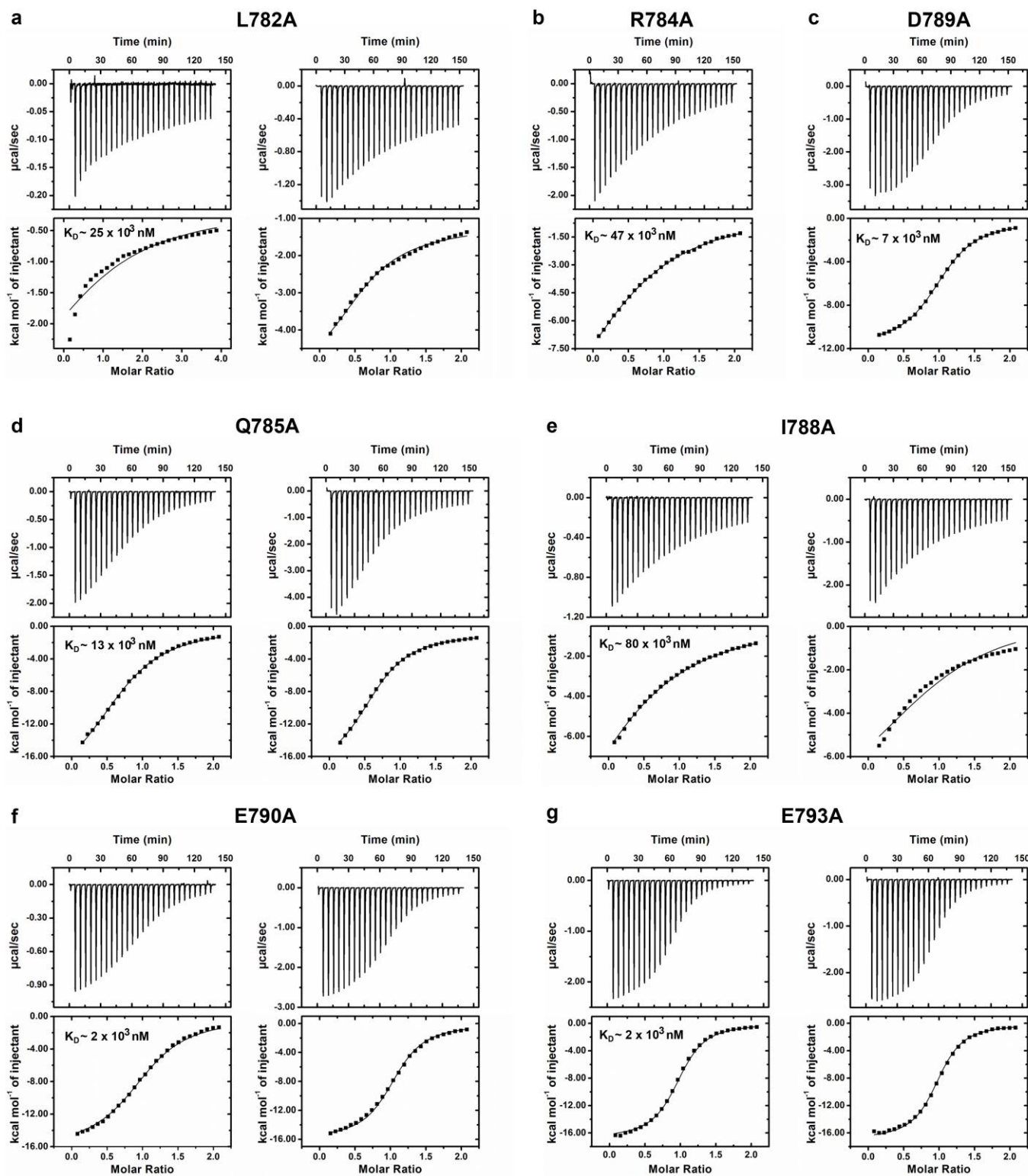


Figure S2: *dEAG* channel fragment alanine scanning experiments.

ITC experiments of wild type CaMKII kinase domain and mutants of *dEAG*_{long} channel fragment in the presence of Mg²⁺-AMPPNP. Mutants as indicated. Titration injection heats are shown in the upper panel and binding isotherm of integrated binding enthalpies in the lower panel. K_D values are shown for each mutant in lower panels. The large decrease in affinity and/or decrease in protein expression levels for some mutants led us to perform just 2 independent calorimetric assays for L782A, Q785A, I788A, E790A and E793A. In these cases we executed assays at two different

concentrations and performed a global fit of the data using the Affinimeter package. Data and fit for both these experiments is shown. For R784A and D789A one of the three replicas performed is shown.

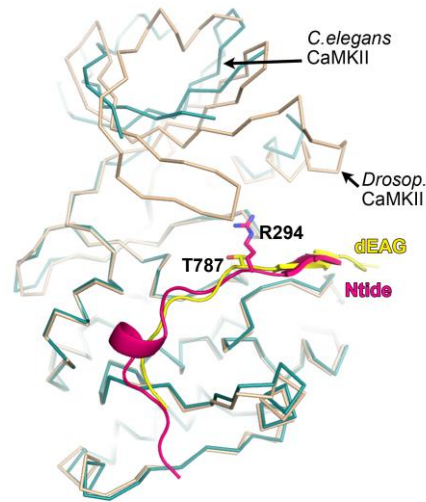


Figure S3: Structural similarity between kinase domain bound to *dEAG* and bound to the CaMKII inhibitor Ntide.

CaMKII kinase domain/*dEAG*_{long} complex (in wheat color) with bound Mg²⁺-ADP superposed on CaMKII kinase domain/Ntide complex (shown in cyan; PDB entry: 3KL8) from *C. elegans*. Channel fragment (*dEAG*) in yellow and Ntide (Ntide) in magenta. Residues T787 in *dEAG* and R294 in Ntide shown in stick. Superposition matched position of C α -atoms in the C-lobe of the two kinase domains (residues 97-275).

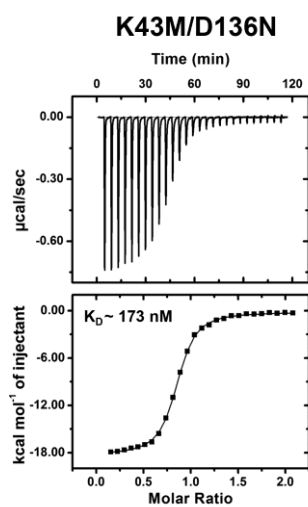


Figure S4: Determination of affinity of channel fragment for CaMKII kinase domain mutant.

ITC experiment of kinase domain K43M/D136N double mutant titrated with $dEAG_{\text{short}}$ channel fragment in the presence of Mg^{2+} -AMPPNP. Titration injection heats are shown in the upper panel and binding isotherm of integrated binding enthalpies in the lower panel. K_D value is shown in lower panel.

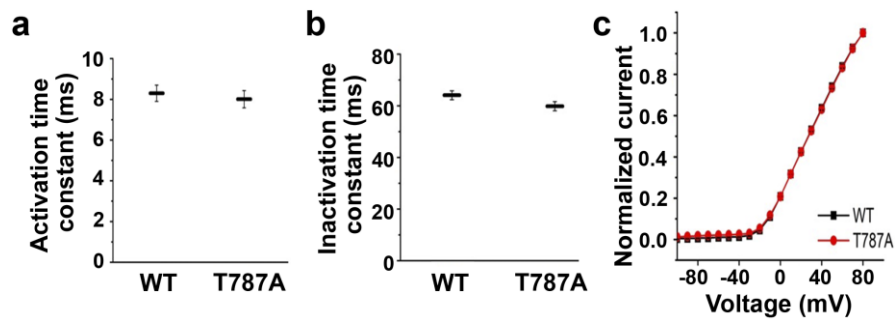


Figure S5: Current characteristics for WT and T787A *dEAG* channels

Comparison of **a)** activation time constant and **b)** inactivation time constant of WT and T787A *dEAG* channel current evoked at 60 mV. **c)** Comparison of the WT and T787A *dEAG* channel current-voltage relationship. Data are mean \pm SEM and $n=7-8$.

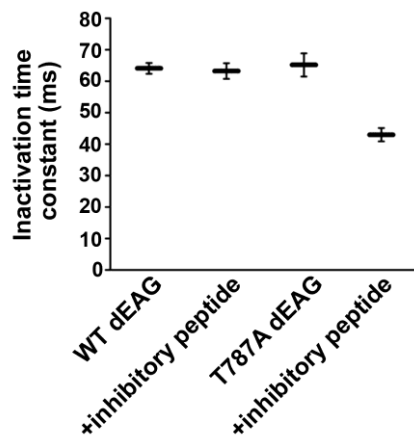


Figure S6: Inactivation time constant for *dEAG* channel

Inactivation time constant of current at 60 mV mediated by WT *dEAG* channel alone and exposed to the inhibitory peptide, T787A *dEAG* channel alone and exposed to the inhibitory peptide. Data are mean \pm SEM and $n=7-10$.

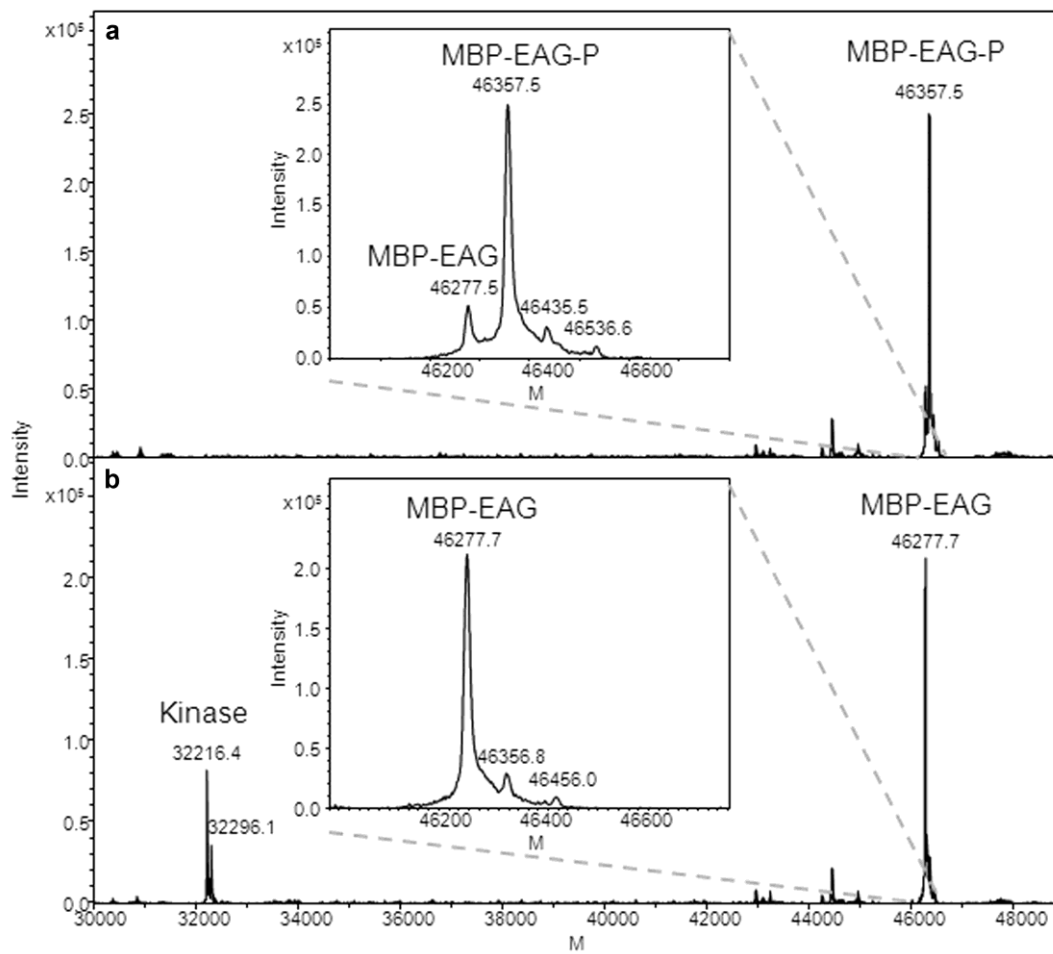


Figure S7: Mass spectrometry spectra of samples before and after ITC experiments showing dephosphorylation.

Deconvoluted UHR-QTOF mass spectra of phosphorylated MBP-*dEAG*_{short} before **a)** and after **b)** ITC experiment in the presence of wild type CaMKII kinase domain and Mg²⁺-AMPCP.

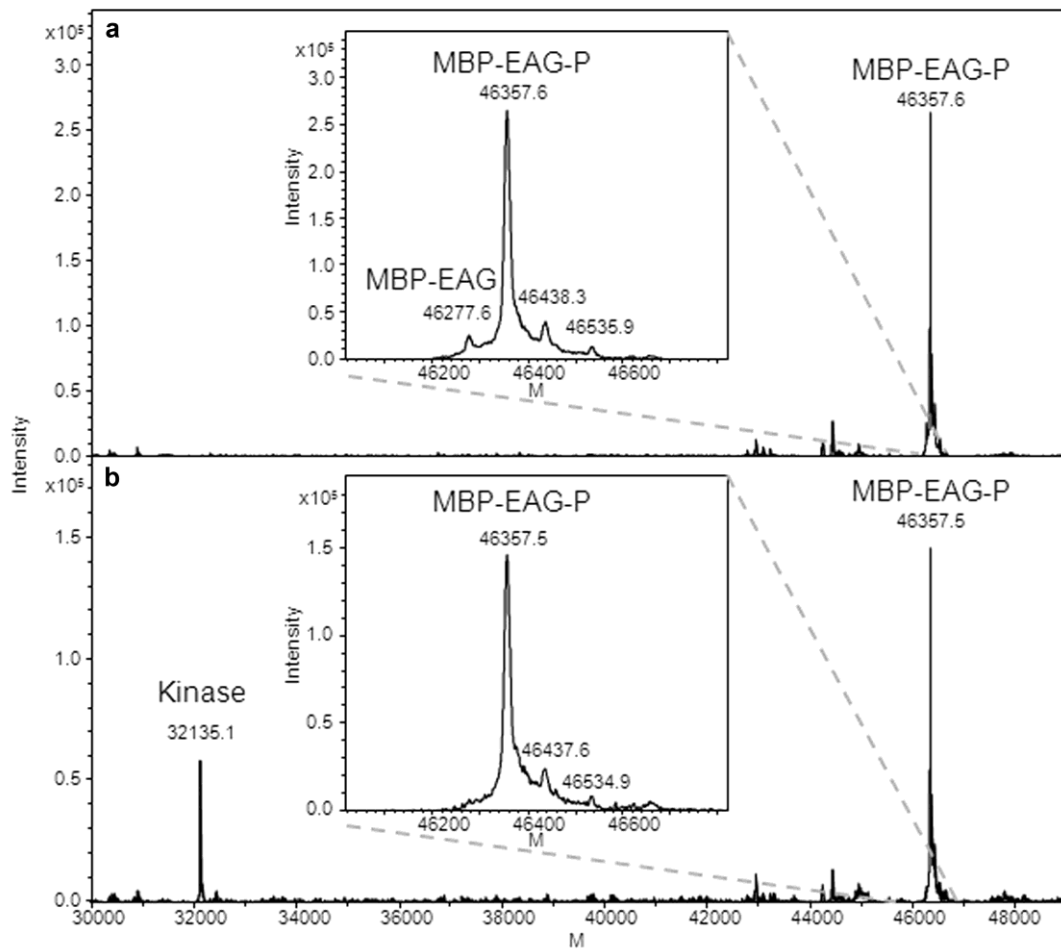


Figure S8: Mass spectrometry spectra of samples before and after ITC experiments showing no change in phosphorylation.

Deconvoluted UHR-QTOF mass spectra of phosphorylated MBP-*d*EAG before **a**) and after **b**) ITC experiment in the presence of D136N CaMKII kinase domain and Mg^{2+} -ADP.