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Supplemental Information

**Systematic Discovery of RNA Binding
Proteins that Regulate MicroRNA Levels**

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Figure S1. Increasing the stringency of the fold enrichment and padj cutoffs allows filtering of most robust candidates and reveals cell-type specificity of miR-RBP interactions. Related to Figure 1A-E and Figure 3.

(A) eCLIP IP/SMInput \log_2 (fold enrichment) does not correlate with mature guide miRNA expression (RPM) for DROSHA or DGCR8.

(B) Heatmap of the number of RBPs with at least 1 (n=1) miRNA loci eCLIP clusters exceeding a fold enrichment (x-axis) and p-adjusted (y-axis) cutoff. Dark blue = 100% (126) RBPs passing (at least 1 pre-miR eCLIP cluster exceeding the fold enrichment and p-adjusted cutoffs), white = 0% passing.

(C) Heatmaps as in (B) of the number of RBPs with at least 1 (n=1), 5 (n=5) or 10 (n=10) miRNA loci eCLIP clusters passing the fold enrichment and p-adjusted cutoffs. Selected final cutoff of \log_2 (fold enrichment) = 2 and $-\log_{10}(\text{padj}) = 3$ denoted by yellow text. The percent of RBPs passing the cutoffs: n=1 92% pass, n=5 51% pass, and n=10 25% pass.

(D) Comparison of DGCR8 eCLIP cluster distribution across genic (excluding intergenic) and non-coding (excluding scRNA) regions with and without \log_2 (fold enrichment) and $-\log_{10}(\text{padj})$ cutoffs. Distributions are compared to previously published HITS-CLIP experiments (Macias *et al.*, 2012). Values are percentages, except where the true count is denoted for introns and miRNAs in parenthesis. Below the HITS-CLIP and more stringent eCLIP non-coding RNA distributions are the number of unique miRNA loci represented by the total clusters in parenthesis.

(E) Comparison of unique DGCR8 eCLIP and HITS-CLIP miR targets showing fewer targets identified by eCLIP but better overlap between cell types.

(F) Kim *et al.* further filtered their DROSHA fCLIP using a novel cut-site algorithm; we compare these more stringently filtered miRNA targets with our eCLIP clusters passing cutoffs.

(G) Some RBP-miRNA loci interactions are cell-type specific, even when target is expressed in both contexts. Table quantifies the intersection of miRNA loci with at least 1 eCLIP cluster passing the fold enrichment and p-adj cutoffs in all encode RBPs screened. Clusters in HepG2 and K562 are considered shared if they intersect the same miR locus even if the clusters don't intersect. RBPs not displayed either had no eCLIP clusters passing cutoff in one or both of the cell lines, or only one cell line was subjected to eCLIP. The last two columns show the number of loci that are expressed in the other cell line but weren't bound.

Figure S1

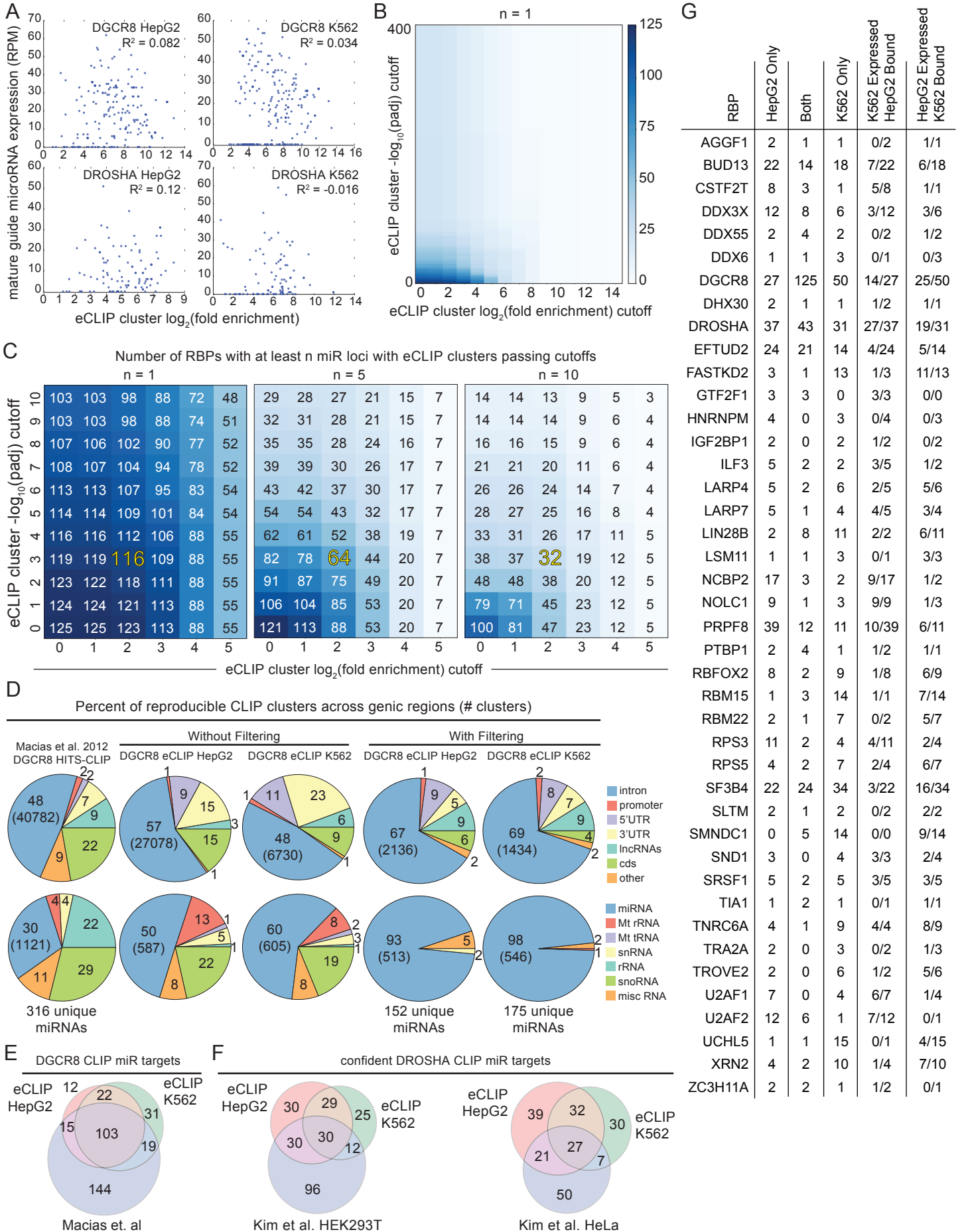


Figure S2. Volcano plots of pre-miR eCLIP clusters in candidate RBPs. Related to Figure 3B.

(A-B) Volcano plots of eCLIP clusters at miRNA loci in HepG2 and K562 cell lines for RBP candidates. Cutoffs of eCLIP IP/SMInput $\log_2(\text{fold enrichment}) = 2$ and $-\log_{10}(\text{padj}) = 3$ are denoted by a dashed red line. Clusters are colored as in Figure 1. RBP and cell line indicated above each panel.

Figure S2

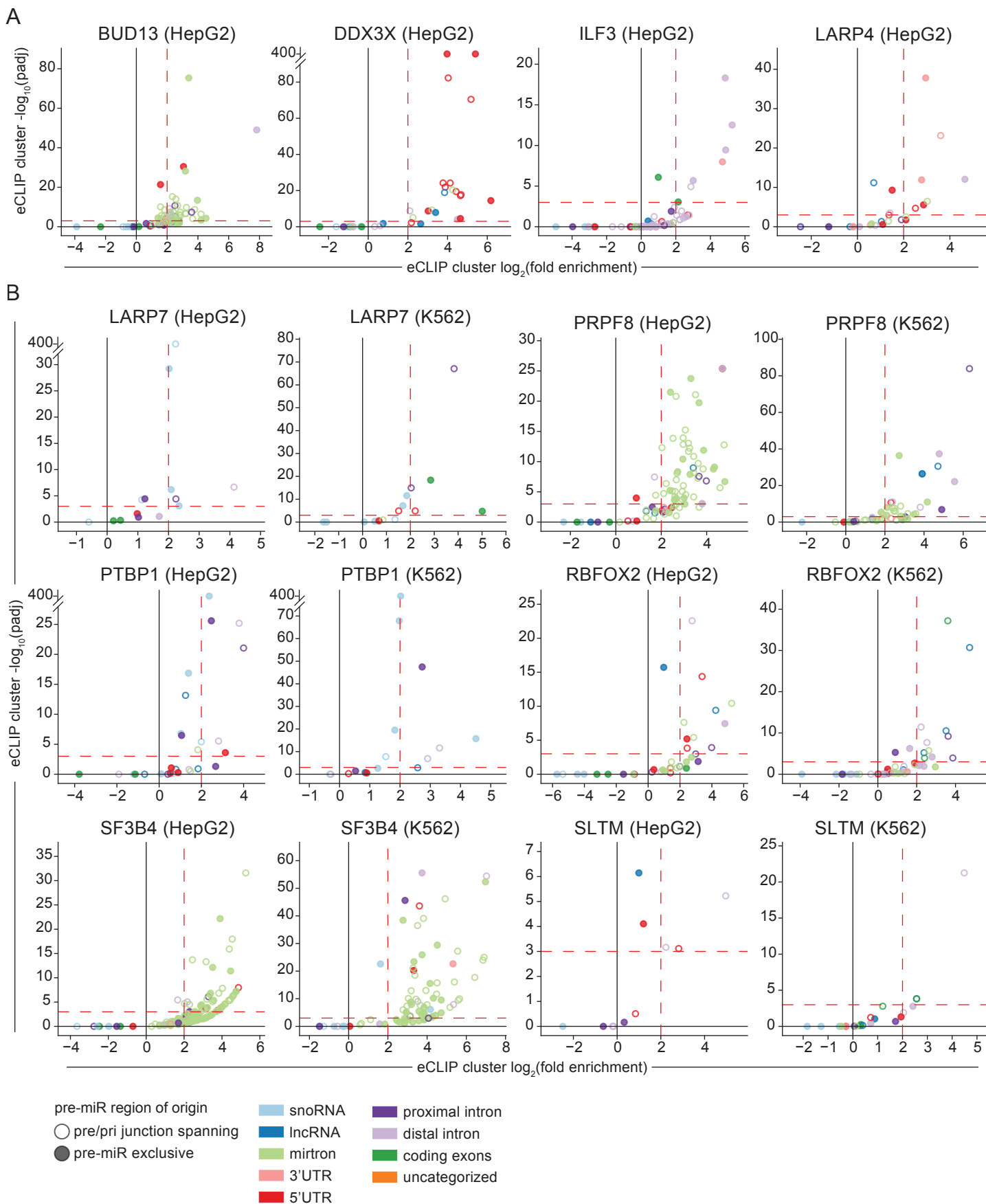


Figure S3. Western Blots of RBP knockdown and small RNA-Seq replicate correlation. Related to Figure 2 and 3.

(A-C) Western blot of RBP knockdowns in HepG2 and K562 cells by lentiviral transduction of shRNA. Controls were performed in quadruplicate and shRNA knockdowns in duplicate, controls are representative; C=control, sh=RBP knockdown. GAPDH and α -tubulin were used as loading controls where indicated.

(D) Distribution of the R^2 Pearson correlation values for shControl replicates of small RNA-Seq samples.

(E) Representative correlation plot for (D).

(F) Distribution of the R^2 Pearson correlation values for shRBP knockdown replicates of small RNA-Seq samples.

(G) Representative correlation plot for (F).

Figure S3

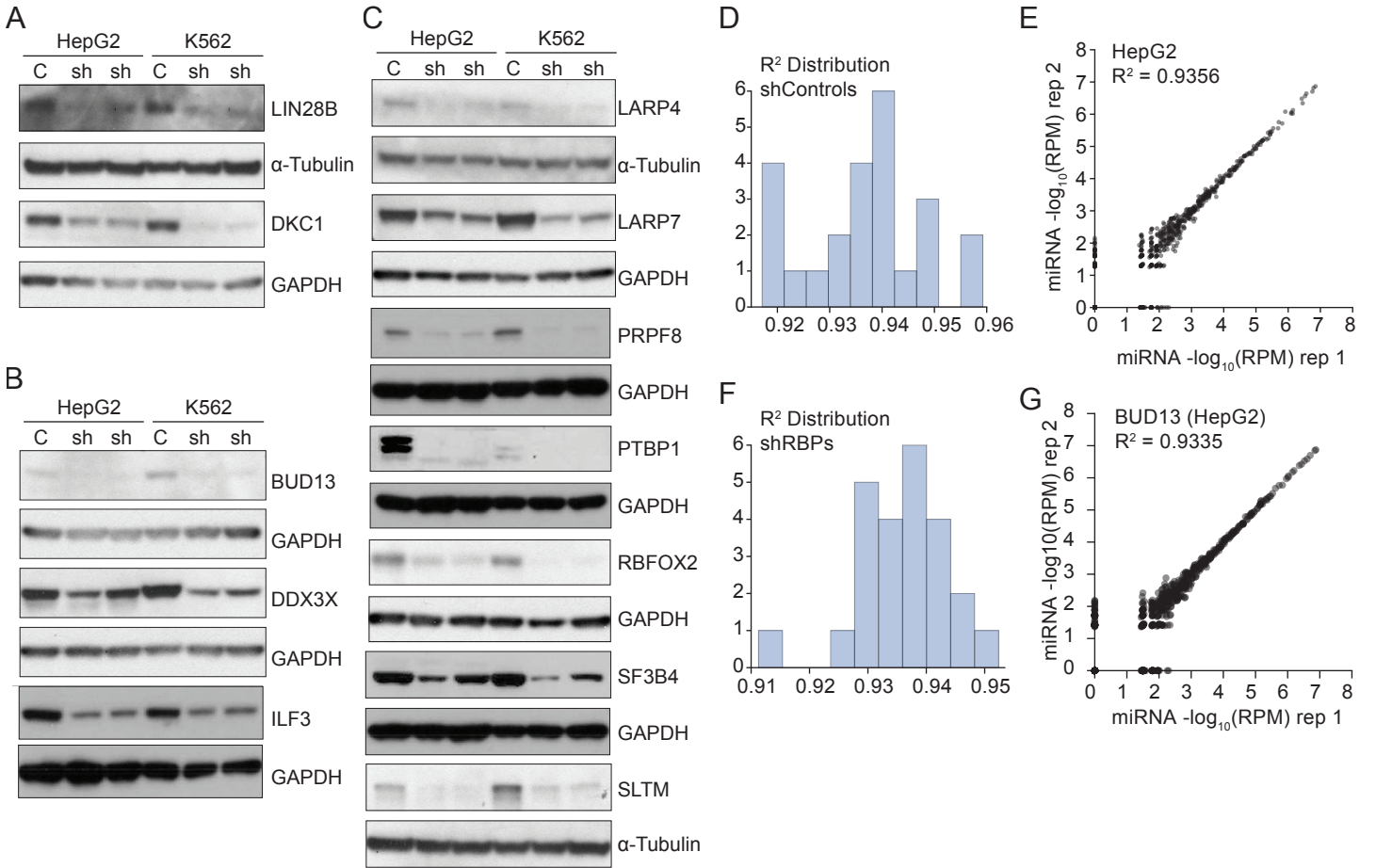


Figure S4. eCLIP and small RNA-Seq integration. Related to Figure 4.

(A-B) Integration of eCLIP and small RNA-Seq data. eCLIP clusters at miRNA loci were further filtered for those intersecting miRNAs detected by small RNA-Seq. For these clusters, eCLIP cluster IP/smInput \log_2 (fold enrichment) was plotted vs. small RNA-Seq miRNA \log_2 (fold change). miRNAs with significant eCLIP clusters (padj < 0.05) that also changed significantly upon RBP knockdown (padj < 0.05; Benjamini-Hochberg correction) are highlighted and colored as in Figure 1.

Figure S4

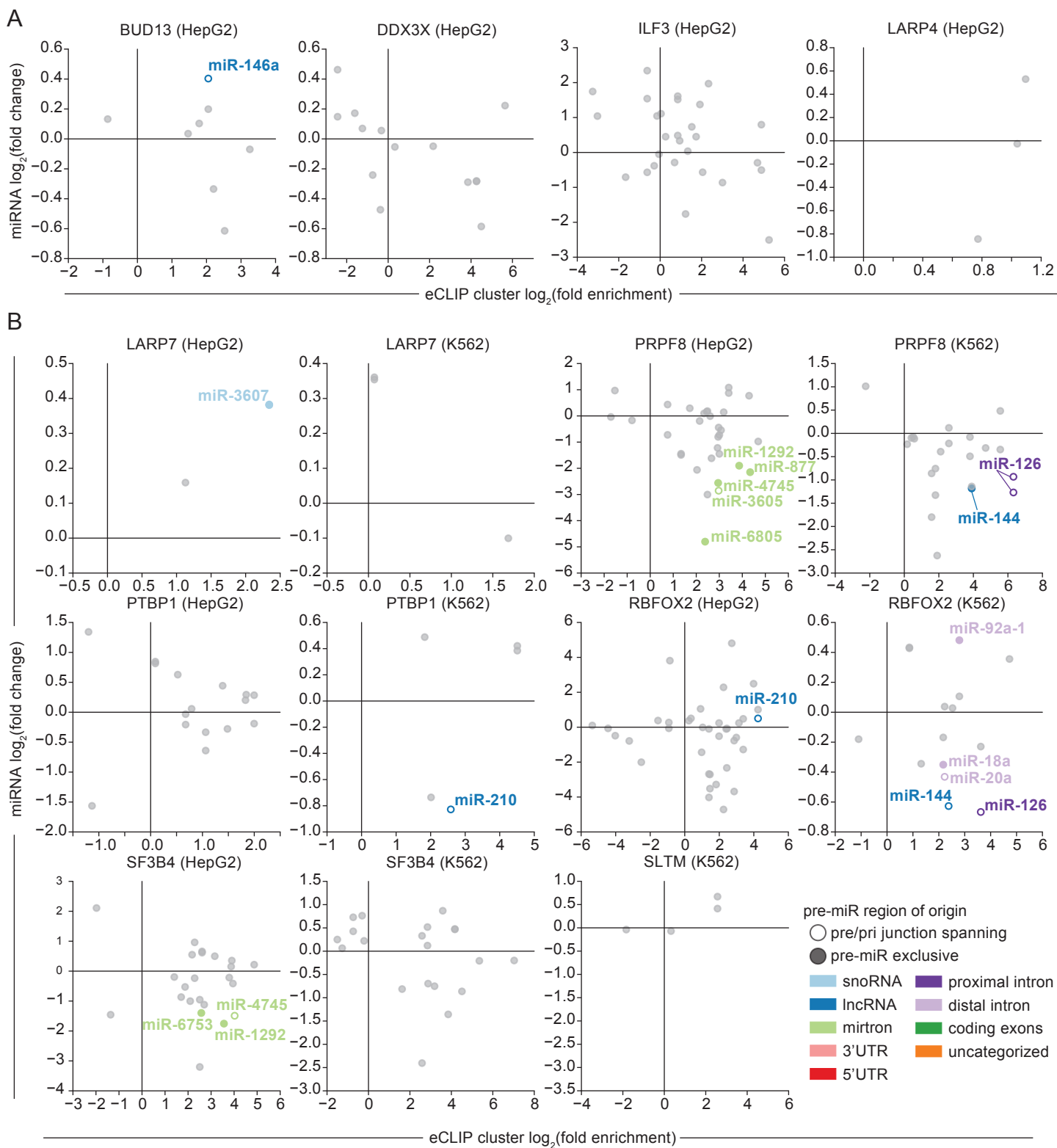


Figure S5. Comparison of findings with Treiber et al 2017. Related to Figure 3B.

(A) Comparison of RBPs identified as putative binders of pre-miRs/miRNA loci. The 180 RBPs identified by Trieber *et al.* (red) were intersected with the RBPs we identified as having at least n eCLIP cluster at a miRNA locus exceeding the fold enrichment and p-adjusted cutoffs in at least 1 cell line tested (yellow) as well as the RBPs for which <n clusters exceeded the fold enrichment and p-adjusted cutoffs in the cell line(s) tested (blue).

Figure S5

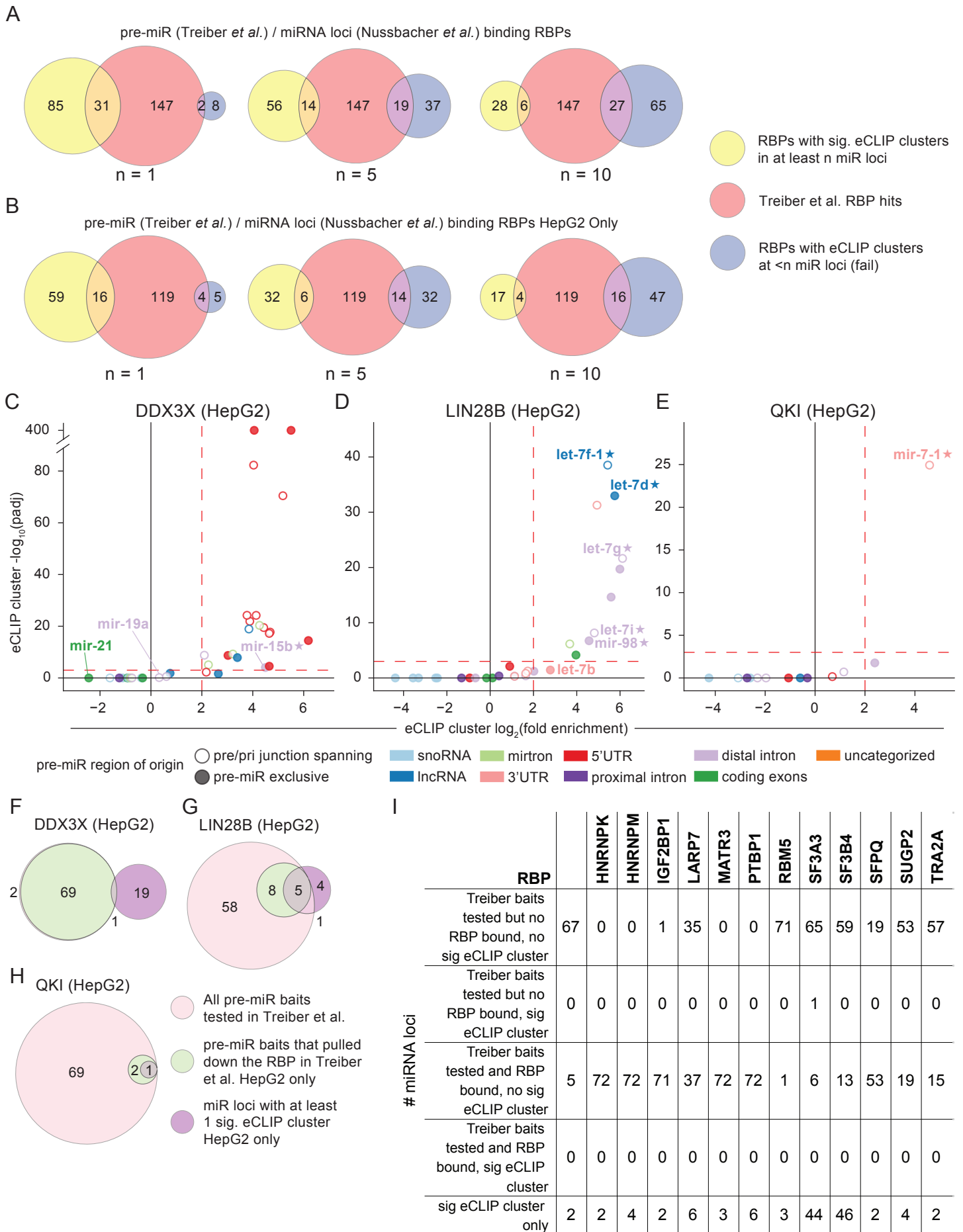


Table S1: shRNAs for RBPs. Related to STAR Methods

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Oligonucleotides		
shRNA targeting BUD13	Sigma-Aldrich	TRCN0000074896
shRNA targeting DDX3X	Sigma-Aldrich	TRCN0000000003
shRNA targeting DKC1	Sigma-Aldrich	TRCN0000039738
shRNA targeting ILF3	Sigma-Aldrich	TRCN0000329787
shRNA targeting LARP4	Sigma-Aldrich	TRCN0000161048
shRNA targeting LARP7	Sigma-Aldrich	TRCN0000122544
shRNA targeting LIN28B	Sigma-Aldrich	TRCN0000144508
shRNA targeting PRPF8	Sigma-Aldrich	TRCN0000075112
shRNA targeting PTBP1	Sigma-Aldrich	TRCN0000001062
shRNA targeting RBFOX2	Sigma-Aldrich	TRCN0000074544
shRNA targeting SF3B4	Sigma-Aldrich	TRCN0000000039
shRNA targeting SLTM	Sigma-Aldrich	TRCN0000135106