

## *Supplementary Material*

# **Toll-like receptor 2 release by macrophages: an anti-inflammatory program induced by glucocorticoids and lipopolysaccharide**

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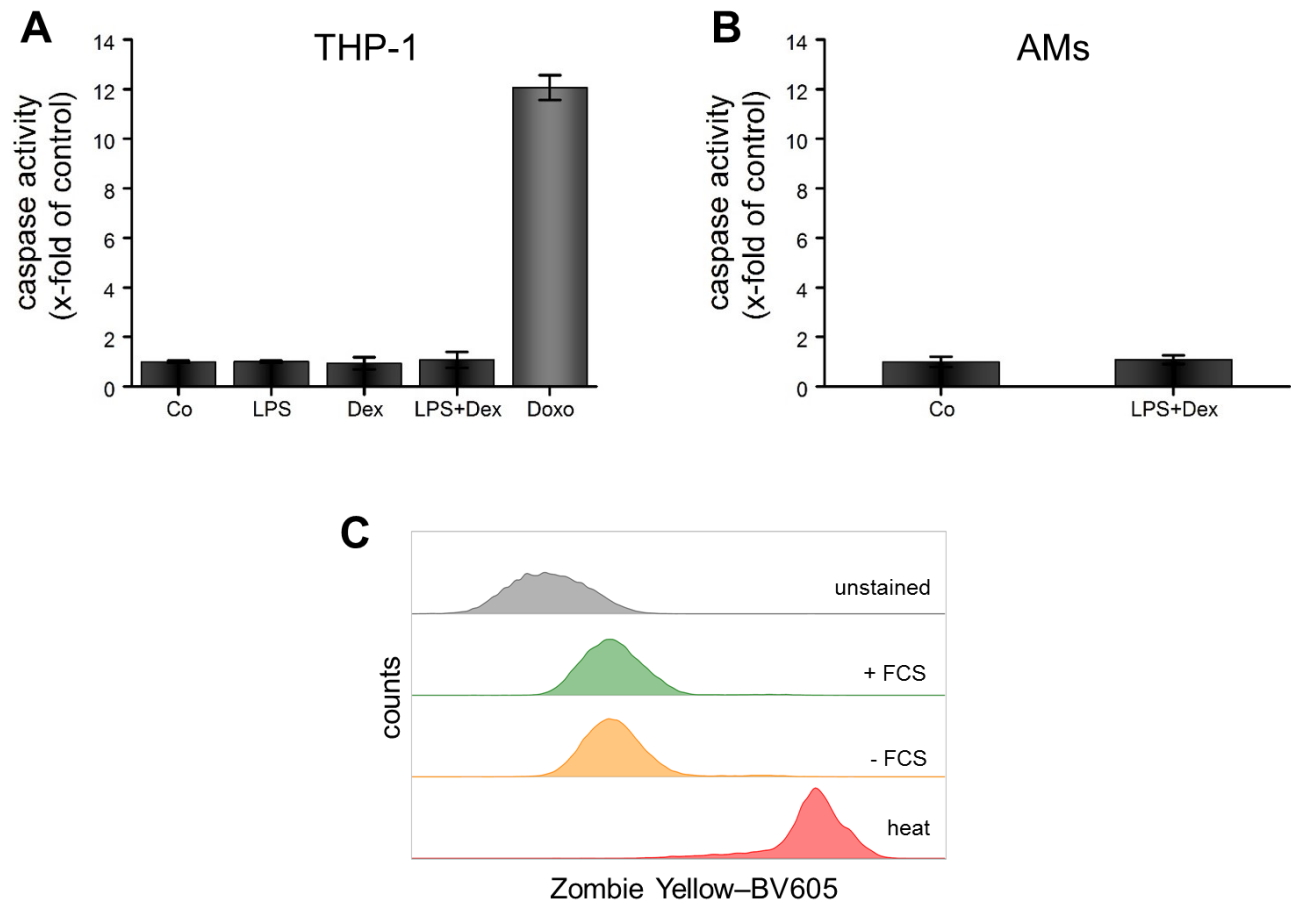
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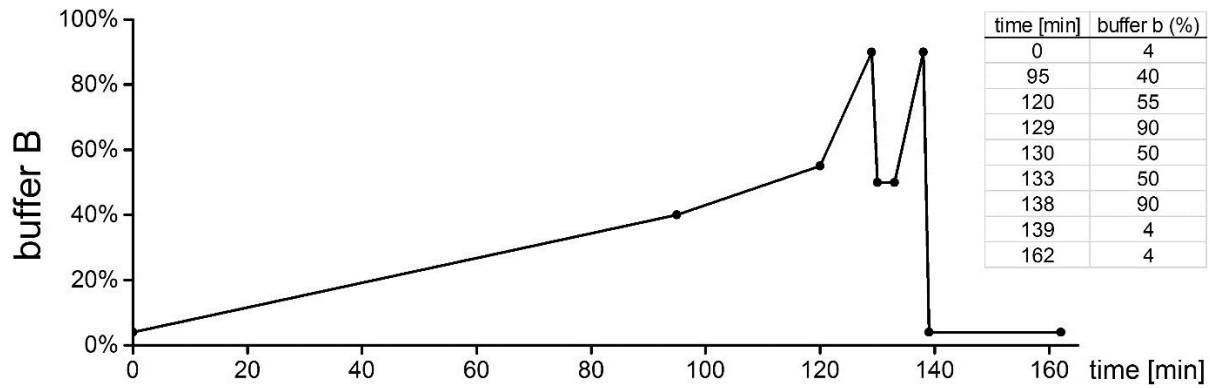
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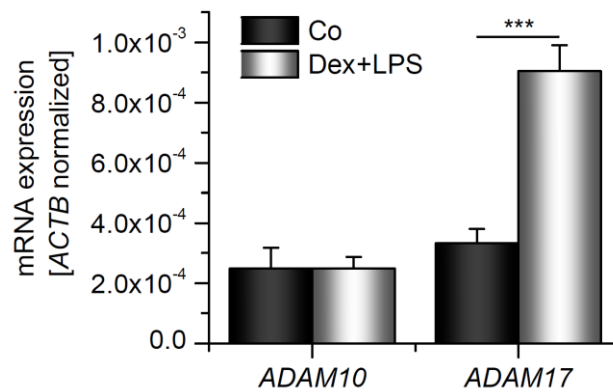
## Supplementary Figures



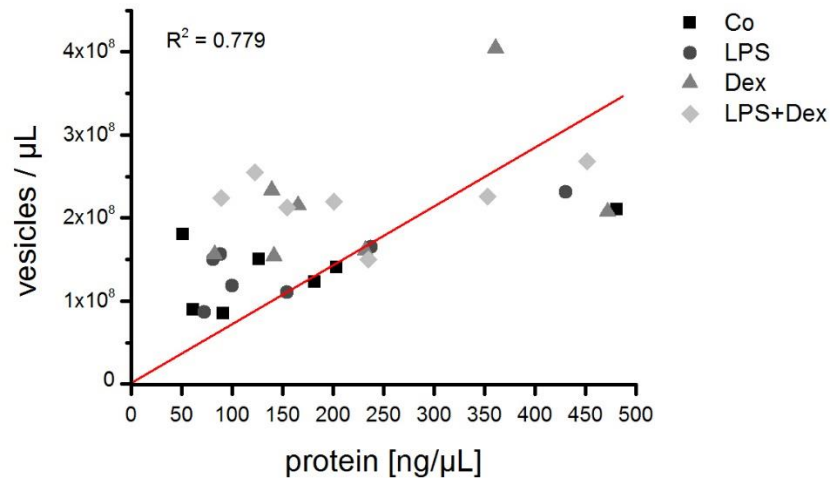
**Supplementary Figure 1.** *Treatment schemes and serum deprivation do not impair cell viability.* A, B: Caspase-3-like activity assay. Cells were treated with the vehicle control (0.1% DMSO, Co), LPS (100 ng/mL), Dex (1  $\mu$ M), or LPS + Dex for 3 d in serum-free medium. Treatment with 10  $\mu$ M doxorubicin (Doxo) for 24 h served as a positive control. An increase in apoptosis-associated caspase-3-like activity was neither detected in THP-1 cells (A) nor in AMs (B) (n = 3). C: Zombie yellow viability staining. Differentiated THP-1 cells were cultured in in serum-containing (+ FCS) or serum-free (- FCS) medium for 3 d. Cells incubated at 60°C for 1 h (heat) served as a positive control. Cell viability was not affected by serum-free medium (n = 2, triplicates).



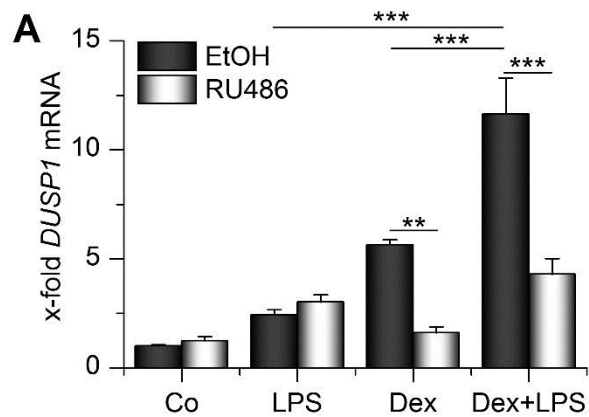
**Supplementary Figure 2.** Gradient for nano-liquid chromatography. Time points for changes in buffer B concentrations are given in the table.



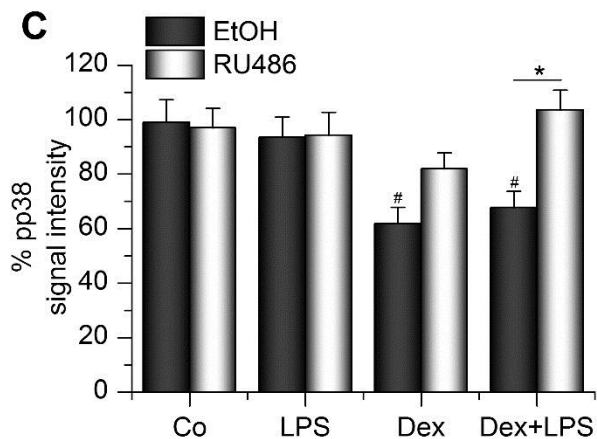
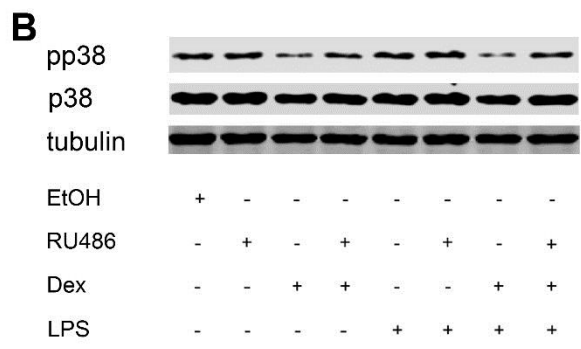
**Supplementary Figure 3:** ADAM10 and ADAM17 expression in AMs. AMs were treated with vehicle (Co) or LPS (100 ng/ml) + Dex (1  $\mu$ M) for 24 h. ADAM10 and ADAM17 expression was determined by qPCR, and values were normalized to the housekeeping gene *ACTB*. The means of 3 experiments performed in triplicate + SEM are shown. \*\*\* $p < 0.001$ .  $p$ -values were generated by ANOVA with Bonferroni's post-hoc test.



**Supplementary Figure 4:** Correlation between EV concentration and protein amount. EVs were isolated from THP-1 macrophages treated with the vehicle control (0.1% DMSO, Co), LPS (100 ng/mL), Dex (1  $\mu\text{M}$ ) or LPS + Dex for 3 d in serum-free medium. Values for seven individual isolations are given. ■ = Co, ● = LPS, ▲ = Dex and ◆ = LPS+Dex.



**Supplementary Figure 5: Influence of dexamethasone and LPS on DUSP1 expression and p38 MAPK activation.** AMs were preincubated with the GR inhibitor RU486 (10  $\mu$ M) or solvent control (0.1% EtOH) and treated with LPS (100 ng/mL), Dex (1  $\mu$ M), or both for 24 h. A: *DUSP1* expression was measured by qPCR. B, C: Phospho-p38 and total p38 were analyzed by Western Blot. Tubulin served as a loading control. Data from at least three independent experiments performed in duplicate with cells from different donors are presented as means + SEM and A, C: \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ . C: # $p < 0.05$  vs. vehicle-treated Co. p-values were generated by ANOVA with Bonferroni's post-hoc test.



## Supplementary Tables

Supplementary Table 1: PCR conditions.

Gene	Sequence (5'→3') forward primer	Sequence (5'→3') reverse primer	Probe sequence (5'FAM→3'BHQ)	Probe [nM]	MgCl <sub>2</sub> [mM]	Annealing [°C]
<i>ACTB</i>	TGCGTGACA TTAAGGAGA AG	GTCAGGCAG CTCGTAGCT CT	CACGGCTGCTTC CAGCTCCTC	60	4	60
<i>ADAMI10</i>	TGCCCAGAT ATCCAGTCA TGTT	TCACCATGA AACTGATGT TACGG	no probe	N/A	N/A	60
<i>ADAMI17</i>	AGAGAACCA CCTGAAGAG CTTG	TCCCCTCTG CCCATGTAT CT	no probe	N/A	N/A	60
<i>CCL2</i>	TTGATGTTT TAAGTTTAT CTTTCATGG	CAGGGGTAG AACTGTGGT TCA	no probe	N/A	N/A	60
<i>CXCL10</i>	GAGCCTACA GCAGAGGAA CC	AAGGCAGCA AATCAGAAT CG	TCCAGTCTCAGC ACCATGAATCAAA	60	4	60
<i>DUSP1</i>	CAGCTGCTG CAGTTTGAG TC	AGGTAGCTC AGCGCACTG TT	no probe	N/A	N/A	64
<i>FPR2</i>	GCATCCTCA GGAAAATGC ACC	GCATCCTCA GGAAAATGC ACC	no probe	N/A	N/A	60
<i>ICAM</i>	GAAGTGGCC CTCCATAGA CA	TCAAGGGTT GGGGTCAGT AG	no probe	N/A	N/A	60
<i>IL10</i>	CAACAGAAG CTTCCATTC CA	AGCAGTTAG GAAGCCCCA AG	AGCCTGACCACG CTTCTAGCTGTTGA G	100	4	60

<b><i>MMP9</i></b>	CTTTGAGTC CGGTGGACG AT	TCGCCAGTA CTTCCCATC CT	no probe	N/A	N/A	60
<b><i>SELE</i></b>	AGCCCAGAG CCTTCAGTG TA	CCCTGCATG TCACAGCTT TA	no probe	N/A	N/A	60
<b><i>TNF</i></b>	CTCCACCCA TGTGCTCCT CA	CTCTGGCAG GGGCTCTTG AT	CACCATCAGCCG CATCGCCGTCTC	100	3	60
<b><i>TLR1</i></b>	AGCAAAGAA ATAGATTAC ACATCA	TTACCTACA TCATACACT CAAAT	ATTCCTCCTGTT GATATTGCTTTTG	60	5	57
<b><i>TLR2</i></b>	GCAAGCTGC GGAAGATAA TG	CGCAGCTCT CAGATTTAC CC	ATGGACGAGGCT CAGCGGGAAG	60	3	60
<b><i>TLR4</i></b>	ATGAAATGA GTTGCAGCA GA	AGCCATCTG TGTCTCCCT AA	AAGTGATGTTTG ATGGACCTCTGA ATCT	60	4	58
<b><i>TLR6</i></b>	TTTACTTGG ATGATGGTG AATAGT	AGTTCCCCA GATGAAACA TT	GTCGTAAGTAAC TGCTGGAGGTGC	100	5	57
<b><i>VCAM</i></b>	CGAGACCAC CCCAGAATC TA	CTGTGGTGC TGCAAGTCA AT	no probe	N/A	N/A	60

**Supplementary Table 2: Antibodies and TLR2 ligands for flow cytometry.**

<b>antibody / ligand</b>	<b>amount per sample</b>	<b>order no.</b>	<b>supplier</b>
<b>FITC anti-human CD9, Mouse IgG1, kappa [HI9a 25]</b>	0.5 µg	BLD-312103	Biozol
<b>FITC anti-human CD63, Mouse IgG1, [H5C6]</b>	1 µg	BLD-353005	Biozol
<b>FITC Mouse IgG1, kappa Isotype Ctrl (FC) [MOPC-21]</b>	1 µg	BLD-400109	Biozol
<b>APC anti-human TLR2 (CD282) Mouse IgG2a [TL2.1]</b>	2 µg	17-9922-41	ThermoFisher
<b>APC Mouse IgG2a kappa Isotype Control [eBM2a]</b>	2 µg	17-4724-81	ThermoFisher
<b>Rhodamine-conjugated Pam<sub>3</sub>CSK<sub>4</sub></b>	0.5 µg	tlrl-rpms	Invivogen