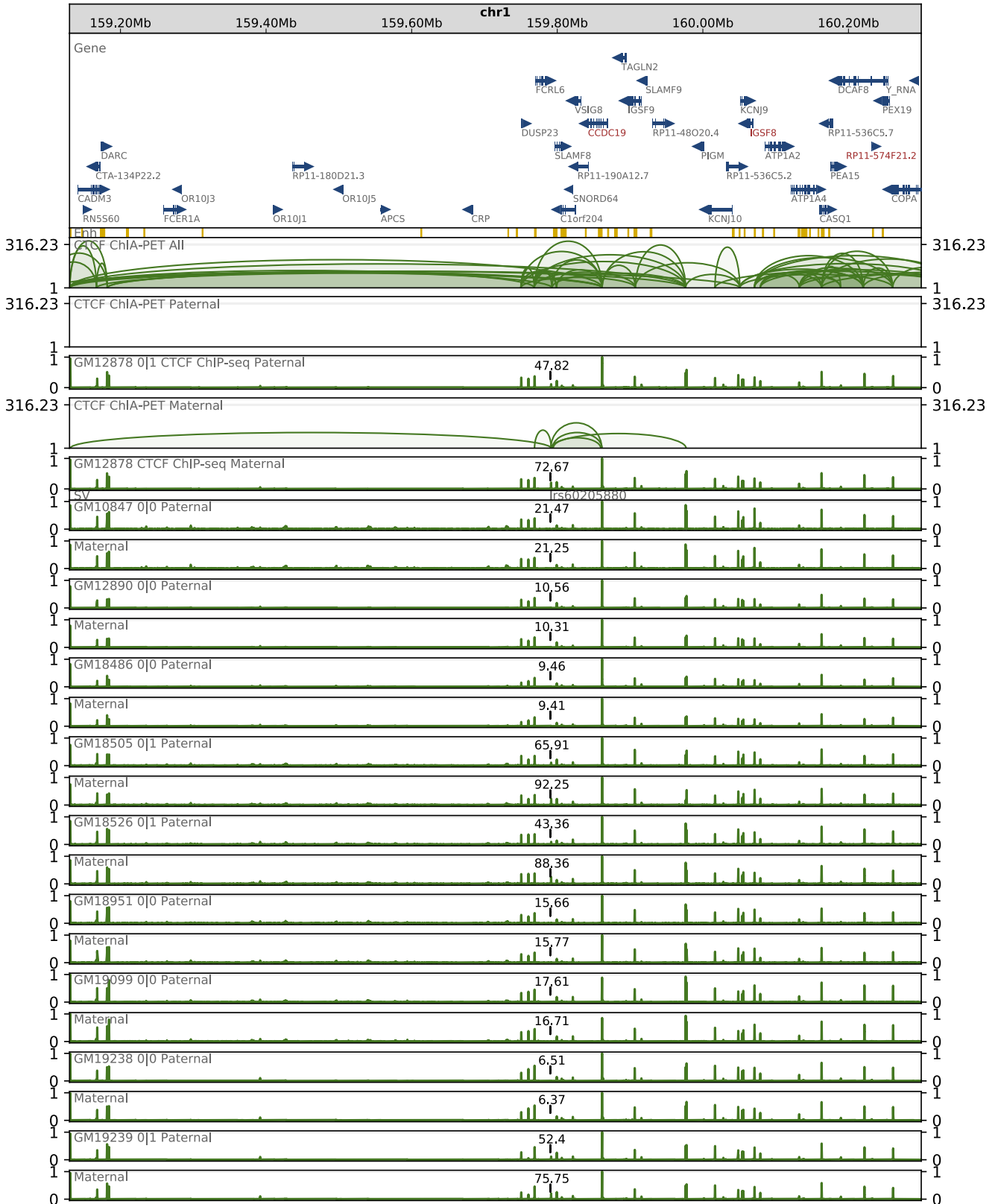
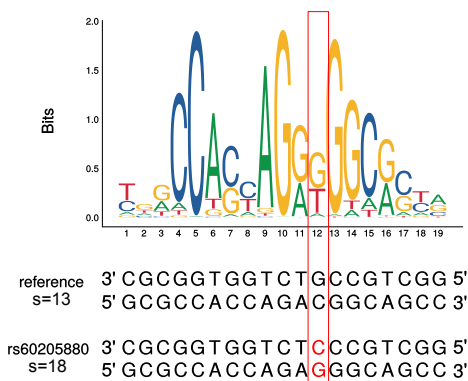


Fig. S1: Distributions of CTCF and cohesin signals around genomic structural elements. Aggregation plots showing CTCF, SMC3 and RAD21 binding distribution profiles around CCDs, CCD boundaries and CTCF anchors. CCDs were taken with $\pm 10\%$ extension proportional to the genomic regions enclosed by the domains. In case of CCD boundaries and CTCF anchors, 2 kb genomic regions centered at CTCF binding motifs indicating the boundaries and around the anchor centers were taken respectively. Each of the extended genomic segment corresponding to a CCD was split into 1,000 equal-sized bins and RPM value of binding signal along each bin was extracted. Similarly, 2 kb genomic region around each CCD boundary or CTCF anchor center was split into 200 equal-sized bins. The values across corresponding bins from all instances of a certain genomic structural element were averaged and plotted as a curve. Even though CTCF anchors are highly enriched with CTCF, SMC3 and RAD21 binding signals, those signals are significantly higher at CCD borders. In plots for whole CCDs, high peaks at boundaries can be observed as positions of CTCF anchors inside domains vary across CCDs and the signal from them averages out.

A



B



C

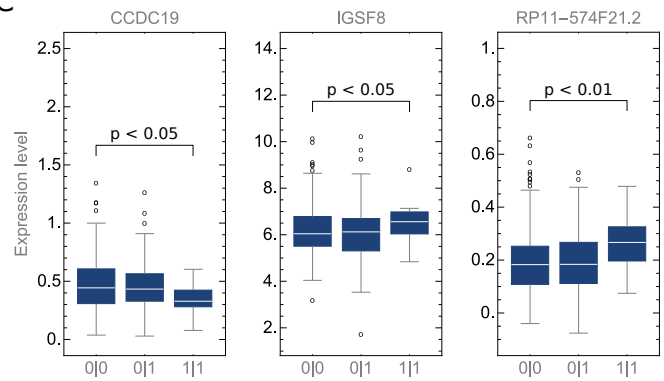


Fig. S2: SNP in a CTCF motif in region chr1:159130000-160300000. **a** Browser view of a 1 Mb genomic segment with SNP rs60205880 identified in a part of the human population. SNP rs60205880 alters sequence of a CTCF motif residing in an interaction anchor. Haplotype-specific CTCF ChIP-seq signals from 10 lymphoblastoid cell lines genotyped for this SNP are presented for comparison along with genes, enhancers and haplotype-specific (GM12878) CTCF ChIA-PET interactions located in this genomic segment. ChIP-seq signal from each sample is measured in RPMs and divided by the maximal value of the signal in the visualized region (separately for each homologous chromosome from the pair). In each individual signal track, value of the signal summed over genomic region occupied by the altered CTCF anchor is additionally marked. **b** Comparison of sequences and scores of CTCF motifs with reference and alternative alleles defined by rs60205880. CTCF sequence logo demonstrates the importance of particular nucleotide positions in the motif. **c** Differences in gene transcription rates between genotypes set for rs60205880. Only genes exhibiting differences in transcription which pass Mann-Whitney test with p-value < 0.05 were reported.

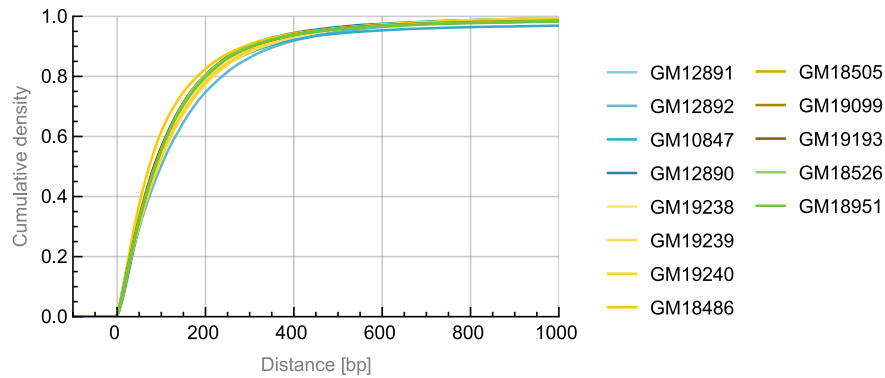


Fig. S3: Cumulative density plot of distances between CTCF ChIP-seq peaks located in GM12878 interaction anchors and closest CTCF ChIP-seq peaks from different lymphoblastoid cells.

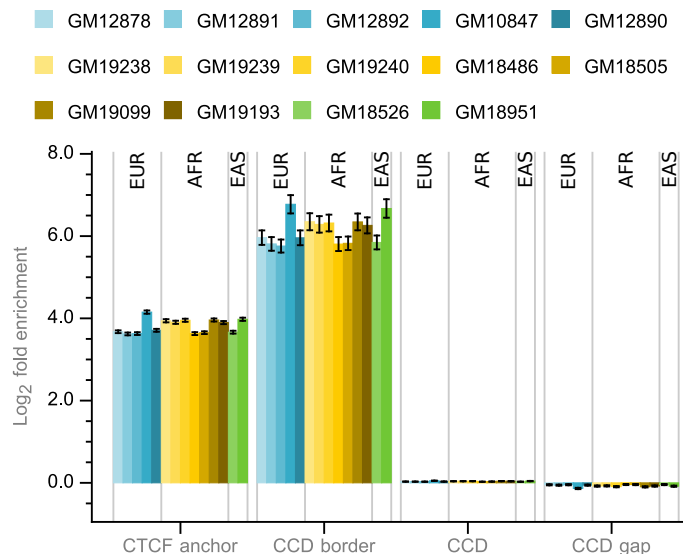


Fig. S4: Enrichment/depletion of structural elements identified in the GM12878 cell line with CTCF ChIP-seq peaks from different lymphoblastoid cells.

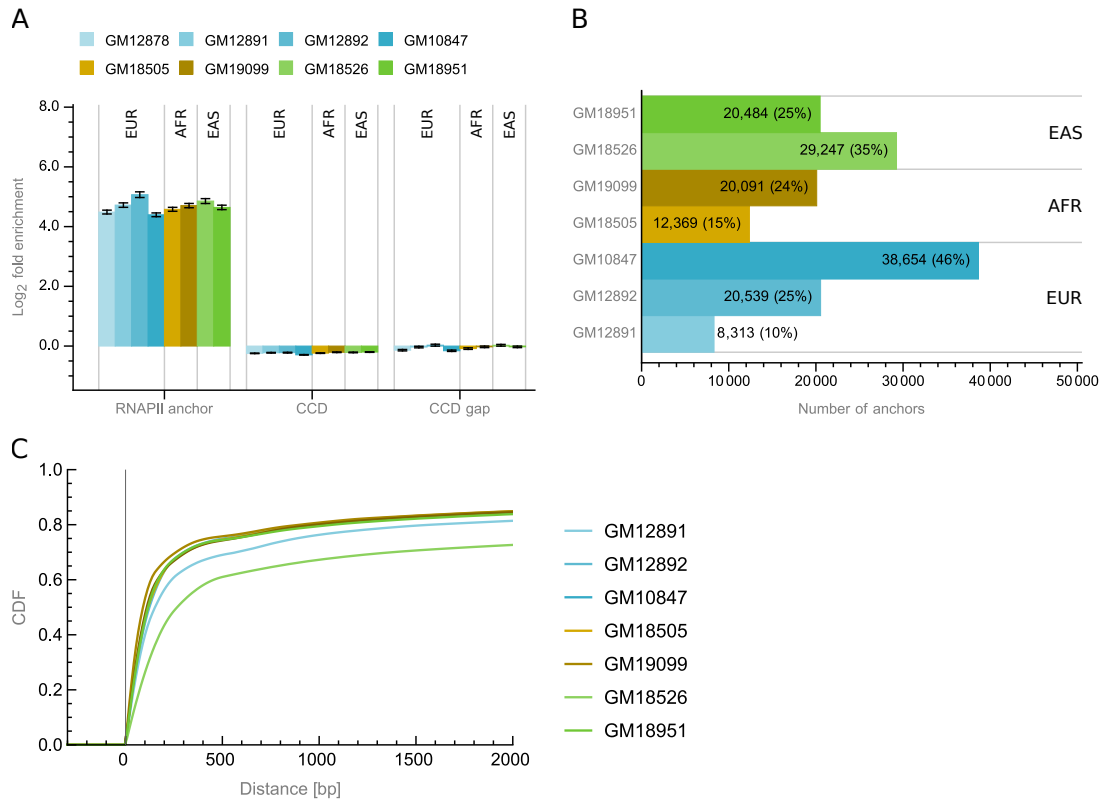


Fig. S5: RNAPII ChIP-seq peaks in interaction segments. **a** Enrichment/depletion of structural elements identified in the GM12878 cell line with RNAPII ChIP-seq peaks from different lymphoblastoid cells. **b** RNAPII anchors from the GM12878 not intersected with RNAPII ChIP-seq peaks identified in different lymphoblastoid cells. **c** Cumulative density plot of distances between RNAPII ChIP-seq peaks located in GM12878 interaction anchors and closest RNAPII ChIP-seq peaks from different lymphoblastoid cells.

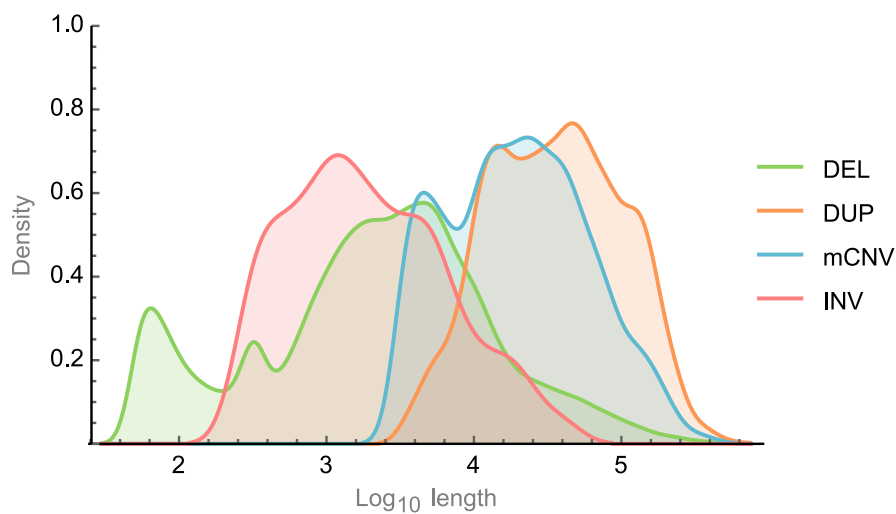


Fig. S6: Density plot showing genomic span distribution of SVs by type.

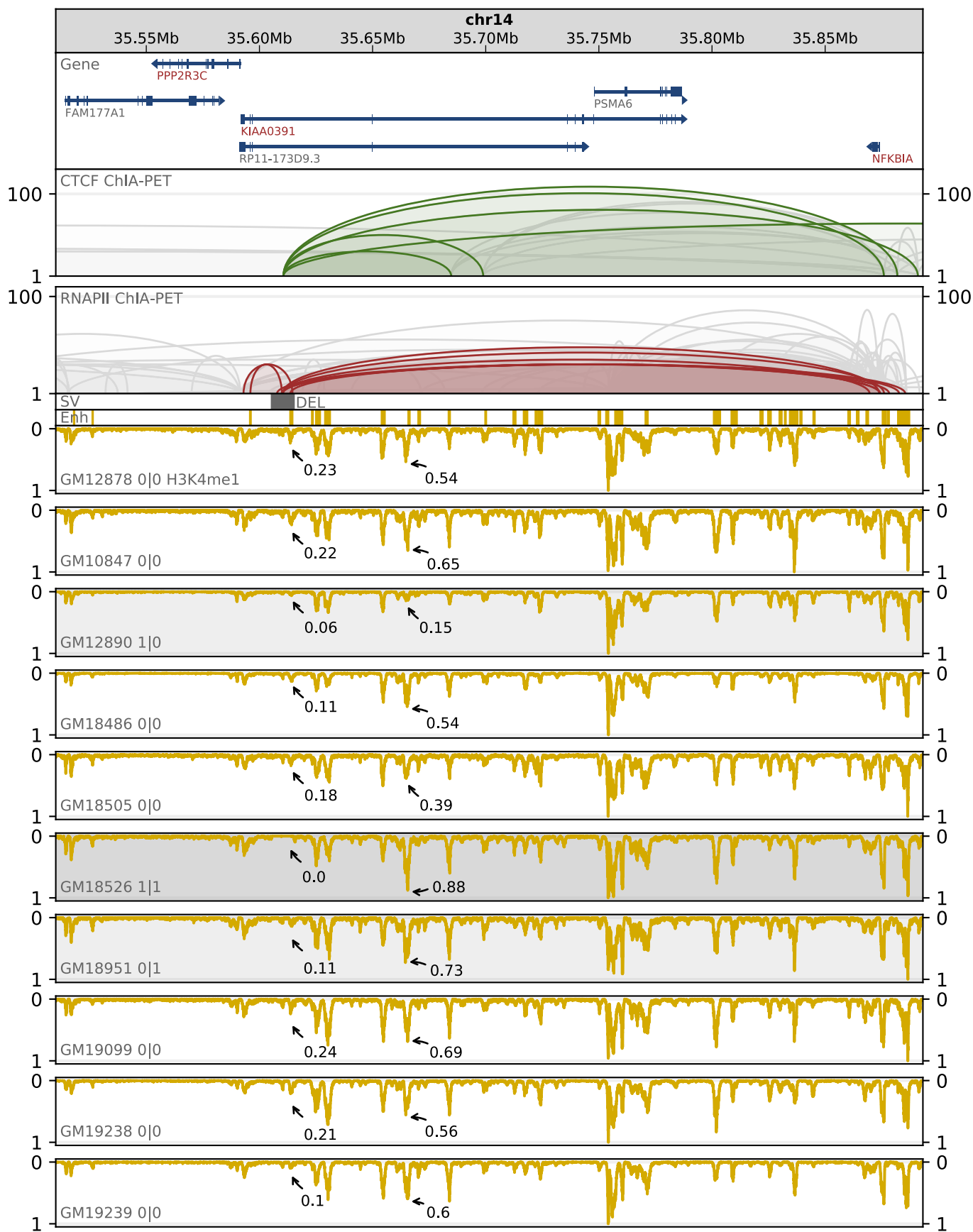


Fig. S7: Deletion of a CTCF interaction anchor in region chr14:35510000-35893204. ChIA-PET interactions at the deletion site displayed above the ChIP-seq profiles of H3K4me1 (primarily associated with active enhancers) for a number of lymphoblastoid cell lines exhibiting different genotypes. The deletion removes a CTCF-mediated interaction anchor possibly involved in formation of insulated neighborhoods separating the PPP2R3C gene (upstream) from a group of enhancers (downstream) and promotes higher PPP2R3C transcription. H3K4me1 signal is notably stronger in deletion-affected homozygous GM18526 than in non-affected homozygous GM12878 in the enhancer region of interest. The signal seems comparable among samples with the reference genotype. It is higher for heterozygous GM18951, but not for heterozygous GM12890. The distinctive profile of the latter shows that other factors may also be involved in enhancer activity of this region in this cell. Compare with Fig. 4b.

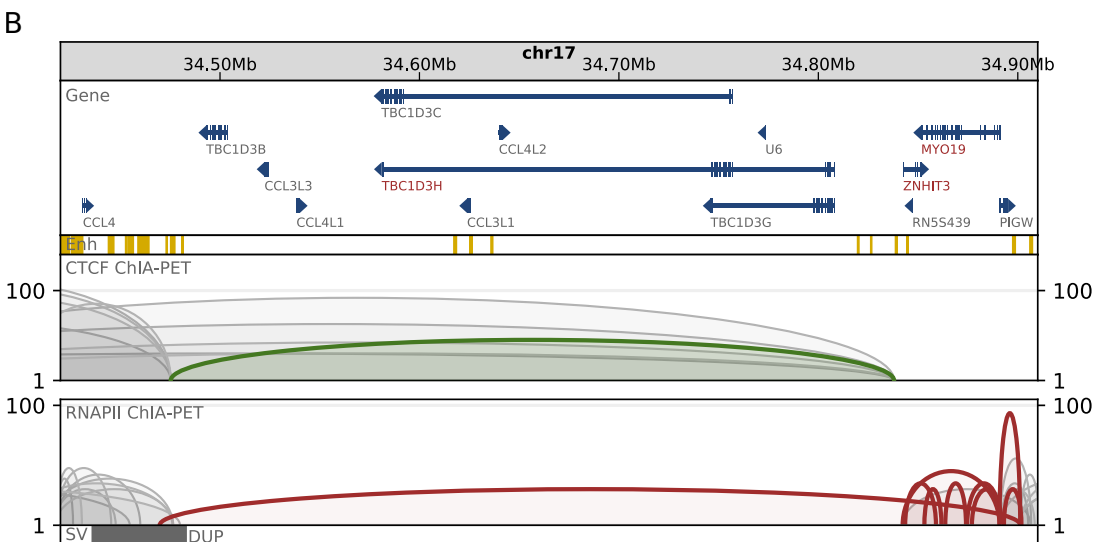
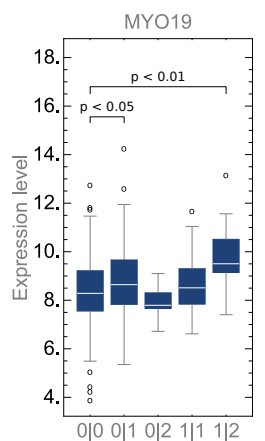
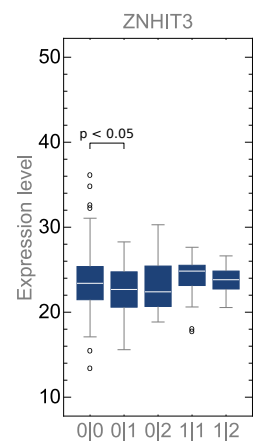
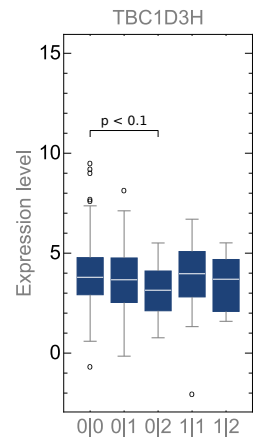
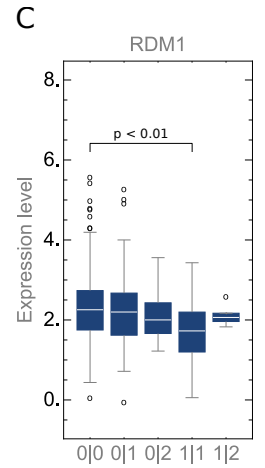
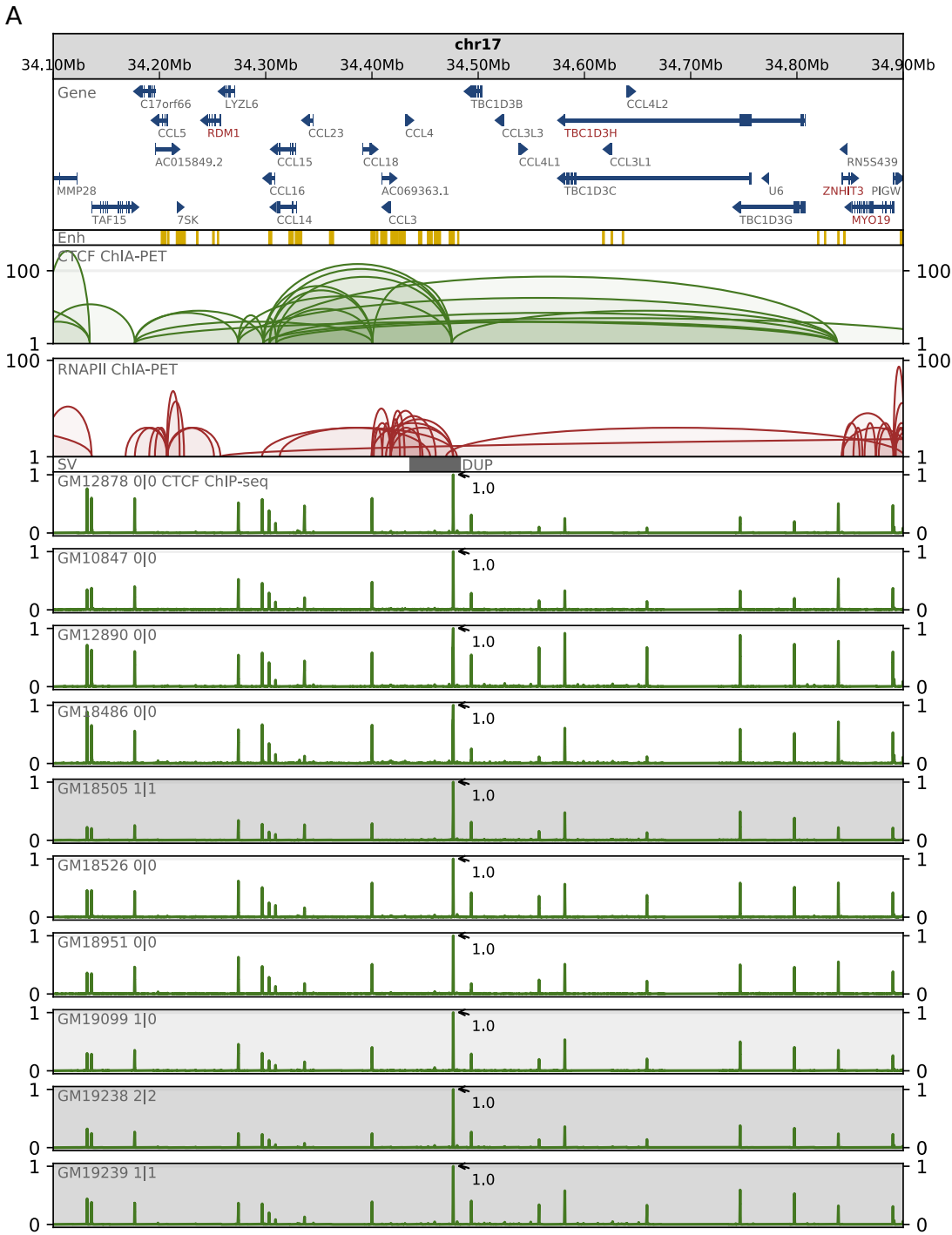
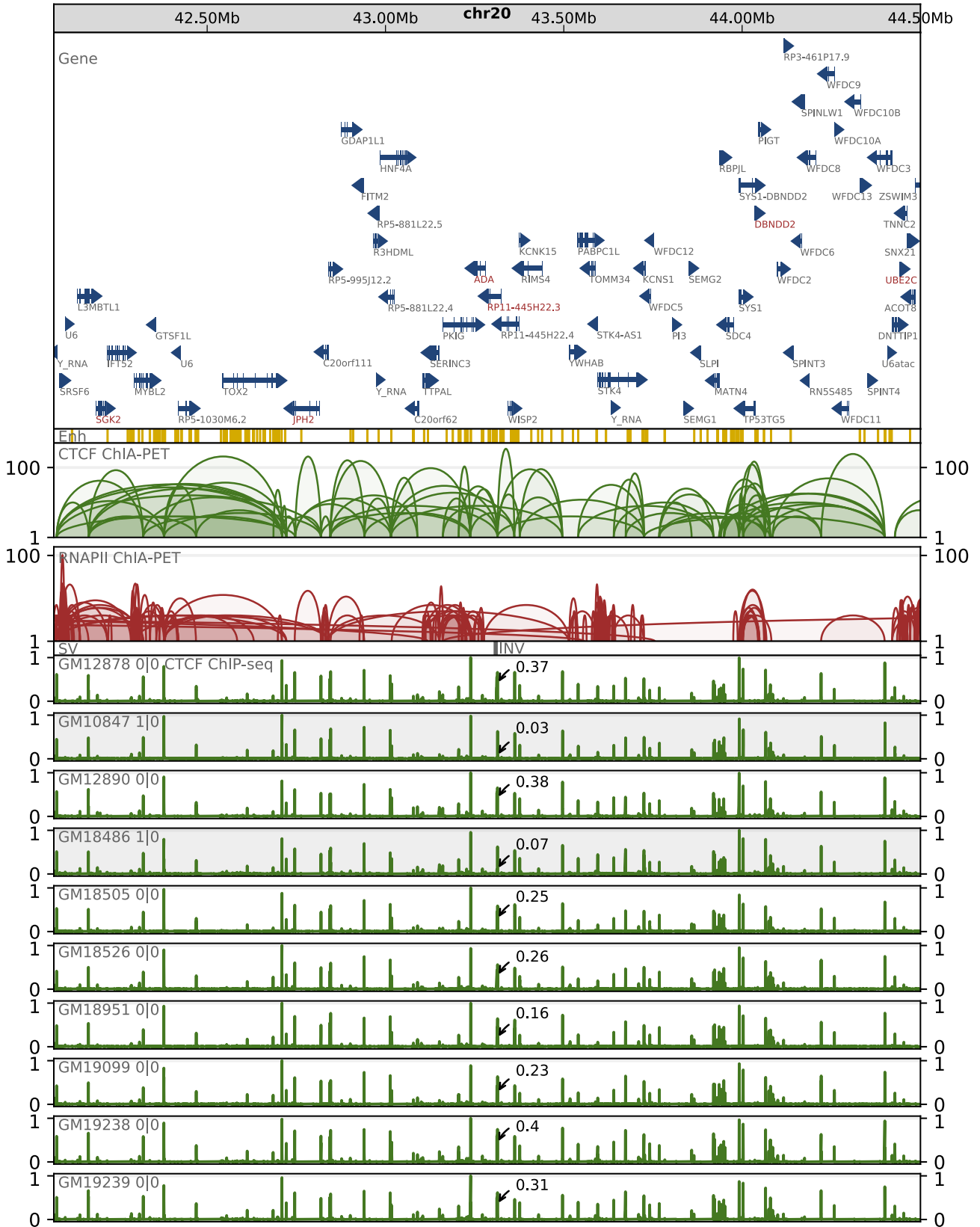


Fig. S8: Duplication of a CTCF interaction anchor in region chr17:34100000-34900000. **a** Browser view of a 0.8 Mb genomic segment with duplication chr17:34436099-34482872 identified in a part of the human population, which duplicates a CTCF anchor containing enhancers. CTCF ChIP-seq signals from 10 lymphoblastoid cell lines genotyped for this duplication are presented for comparison along with genes, enhancers and CTCF and RNAPII ChIA-PET interactions located in this genomic segment. In each ChIP-seq signal track, the value of the highest signal peak in genomic region targeted by the SV is marked. ChIP-seq signal from each sample is measured in RPMs and divided by the value of the peak neighboring the annotated peak. **b** Close-up on the CTCF and RNAPII ChIA-PET interactions affected by duplication chr17:34436099-34482872. **c** Differences in gene transcription rates between genotypes set for duplication chr17:34436099-34482872. Only genes exhibiting differences in transcription which pass Mann-Whitney test with p-value < 0.1 were reported.

A



B

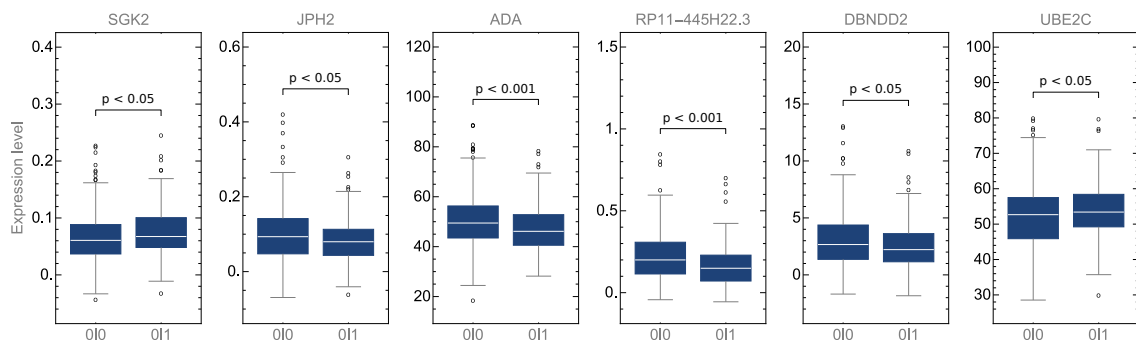


Fig. S9: Inversion in a CTCF interaction anchor in region chr20:42070000-44500000. **a** Browser view of a 2.5 Mb genomic segment with inversion chr20:43305746-43312261 identified in a part of the human population, which inverts a CTCF motif in an interaction anchor containing an enhancer. CTCF ChIP-seq signals from 10 lymphoblastoid cell lines genotyped for this inversion are presented for comparison along with genes, enhancers and CTCF and RNAPII ChIA-PET interactions located in this genomic segment. ChIP-seq signal from each sample is measured in RPMs and divided by the maximal value of the signal in the visualized region. In each individual signal track, value of the highest signal peak in genomic region targeted by the SV is additionally marked. **b** Differences in gene transcription rates between genotypes set for inversion chr20:43305746-43312261. Only genes exhibiting differences in transcription which pass Mann-Whitney test with p -value < 0.05 were reported.

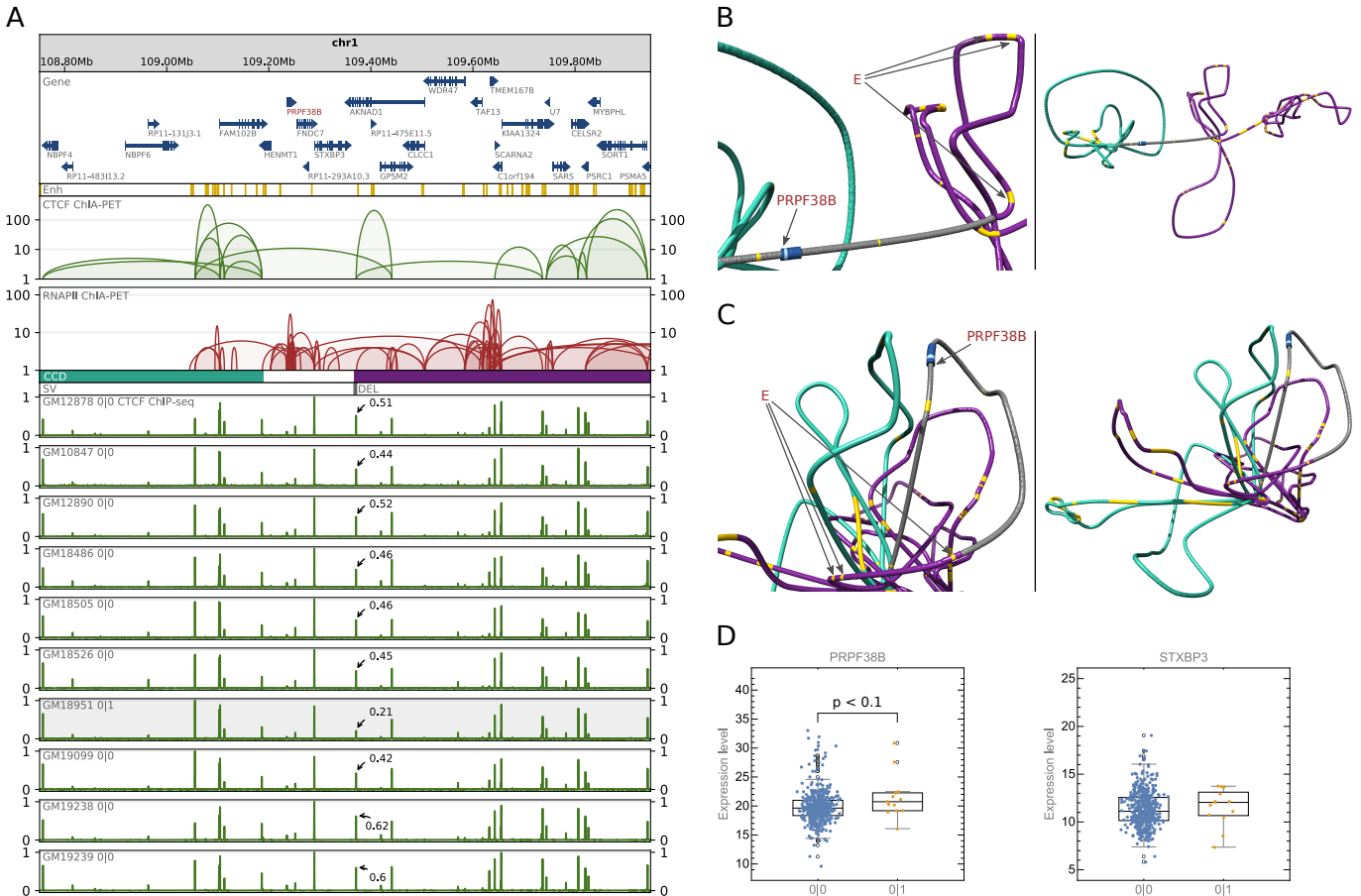


Fig. S10: Deletion of a CCD border in region chr1:108751000-109948000. **a** ChIA-PET interactions at the deletion site chr1:109366972-109372000 displayed above the CTCF ChIP-seq profiles for a number of lymphoblastoid cells exhibiting specific genotypes. The deletion removes a topological domain boundary. ChIP-seq signal from each sample is measured in RPMs and divided by the maximal value of the signal in the visualized region. In each individual signal track, value of the highest signal peak in genomic region targeted by the SV is additionally marked. **b** Models showing PRPF38B gene and enhancers relative arrangement in 3D (left) and arrangement of all chromatin loops in the genomic region shown in (a) (right). In genomes of the reference genotype, the PRPF38B gene is located at the genomic segment linking two topological domains and insulated from the enhancers located in those domains. **c** Similar to (b), but models represent topology of genomes having the deletion. The PRPF38B gene is fused with a bigger domain and becomes a part of a chromatin loop including a set of enhancers. The gene may spatially interact with those enhancers, which could be the cause of increased transcription rates (d). There is one another gene at the gap for which transcription rates are at our disposal (STXBP3), but its expression does not show significant differences (d). The reason for this could be sparsely represented genotype 0|1, but other mechanisms could also be involved. Experiments should be performed for comprehensive explanation. **d** Transcription of genes located at the gap neighboring the domain boundary excised by the deletion. The difference in PRPF38B transcription between genotypes is statistically significant (Mann-Whitney test).

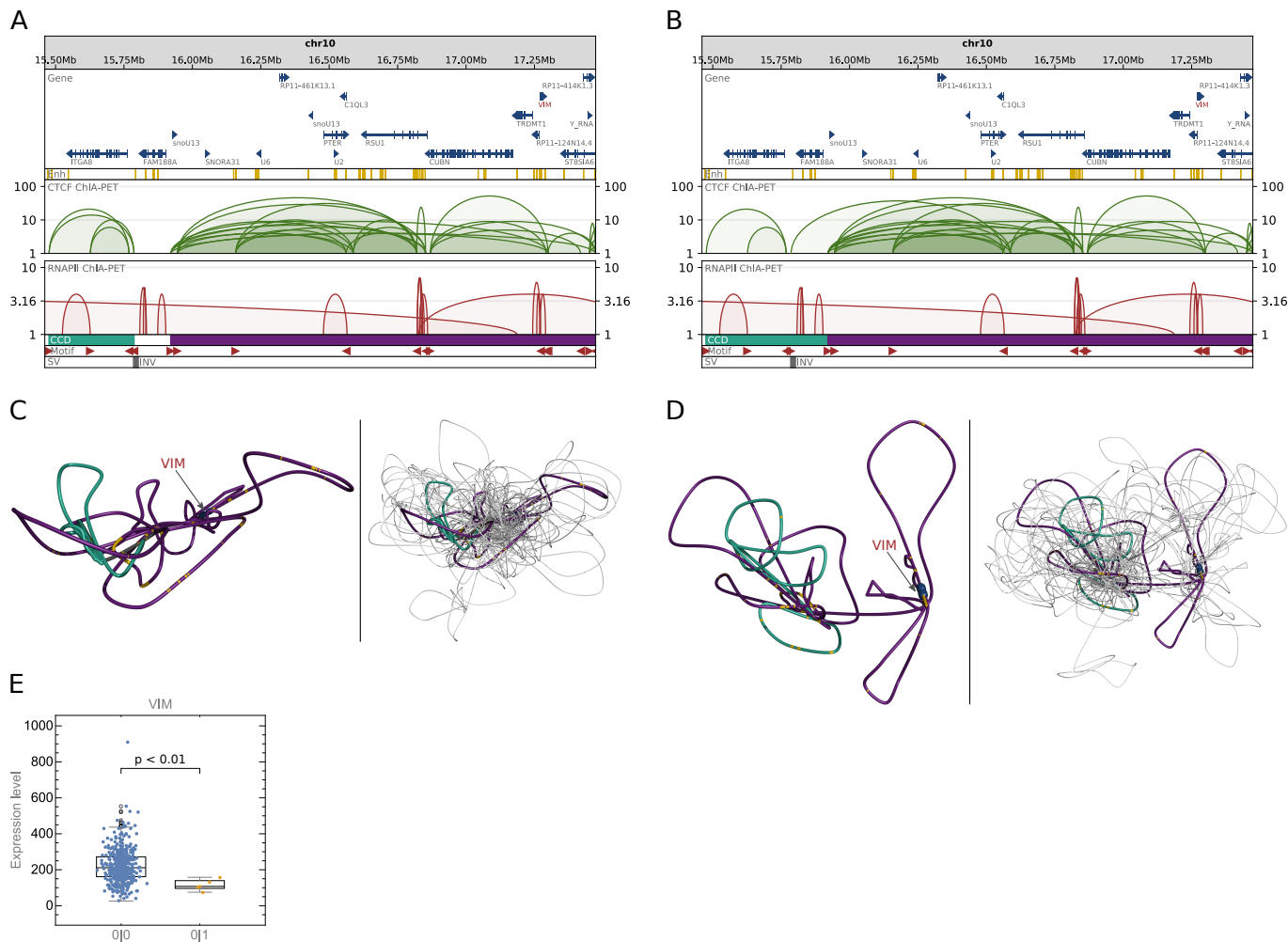


Fig. S11: Inversion of a CCD border in region chr10:15460694-17473625. **a** ChIA-PET interactions at the inversion chr10:15784798-15802449 site in genomes not carrying the inversion. Orientation of CTCF motifs at the interaction anchors are marked with red arrows. **b** Predicted ChIA-PET interactions at the inversion site in genomes carrying the inversion. **c** Model showing spatial arrangement of chromatin loops in the genomic region shown in (a) (left) and an ensemble of such models superimposed (right). **d** Similar to (c), but interaction network shown in (b) is modeled. According to our model, CTCF anchor after the inversion loses the old contacts and is linked to the closest interaction anchor that can form a convergent loop with it. Since the anchor affected by the inversion forms convergent loop encompassing the domain, the domain boundary is lost after the inversion. Interestingly, it seems that the motif inversion leads to a topology with two domains, but different than those before inversion. The smaller of the two borders contains the VIM gene, which transcription drops with the inversion. **e** The VIM gene which transcription correlates with occurrence of the inversion (p -value from the Mann-Whitney test).

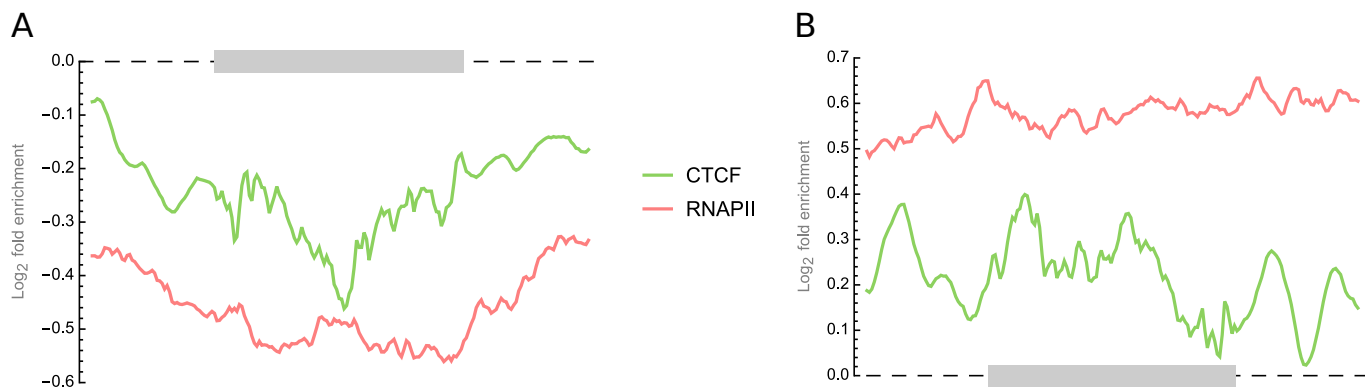


Fig. S12: Aggregate analysis of ChIP-seq signals in altered interaction anchors. **a** Enrichment/depletion of the CTCF and RNAPII ChIP-seq signal in anchors hit by deletions. **b** Enrichment/depletion of the CTCF and RNAPII ChIP-seq signal in anchors hit by duplications.

Fig. S13: Deletions introducing architectural changes at olfactory receptor family genes locus.

a ChIA-PET interactions at the deletion site chr1:248560000-249105000 displayed above the haplotype-specific CTCF ChIP-seq profiles for a number of lymphoblastoid cells. For each track, ChIP-seq signal values (originally in RPMs) were divided by the maximal value of the signal in the visualized region. In each signal track, value of the highest signal peak in genomic region targeted by a SV is additionally marked. **b** Similar to (a), but H3K4me1 ChIP-seq signals are presented. **c** ChIA-PET interactions forming the topological domain around the deletion site. Predicted chromatin loop disrupted in GM12878 cell by DEL2 is represented by the black dotted line. **d** Models showing enhancers and selected genes relative arrangement in 3D within topological domain shown in (c). Topology of genomes without the deletions (left) and with the deletions (right). DEL2 removes a potential insulation boundary between a group of olfactory receptor family genes (light green region) and a group of enhancers (magenta region). There is a strong CTCF motif at the positive strand of DNA fragment covered by the deletion. The loss of the insulation site may promote higher transcription rates of those genes (we did not obtain transcription rates for OR2T11, OR2T27 and Y_RNA). DEL1 (in a very strong LD with DEL2) intersects the OR2T10 gene, which may be the reason why OR2T10 is the only gene which transcription drops with occurrence of the deletions. DEL3 excises a strong CTCF binding motif in a interaction anchor. However it is not reflected in the CTCF signal and requires experiments to gain more certainty about the details of the chromatin topology in the region. Interestingly, it seems that there are no RNAPII chromatin loops within the CTCF loop containing the genes, which may suggest that potential spatial interactions between those genes and enhancers are regulated by CTCF. **e** Genes which transcription correlates with occurrence of DEL2 (p-values from the Mann-Whitney test). **f** Linkage disequilibrium (measured as r^2 value in the CEU population) between the deletions shown in (a), (b) and (c).

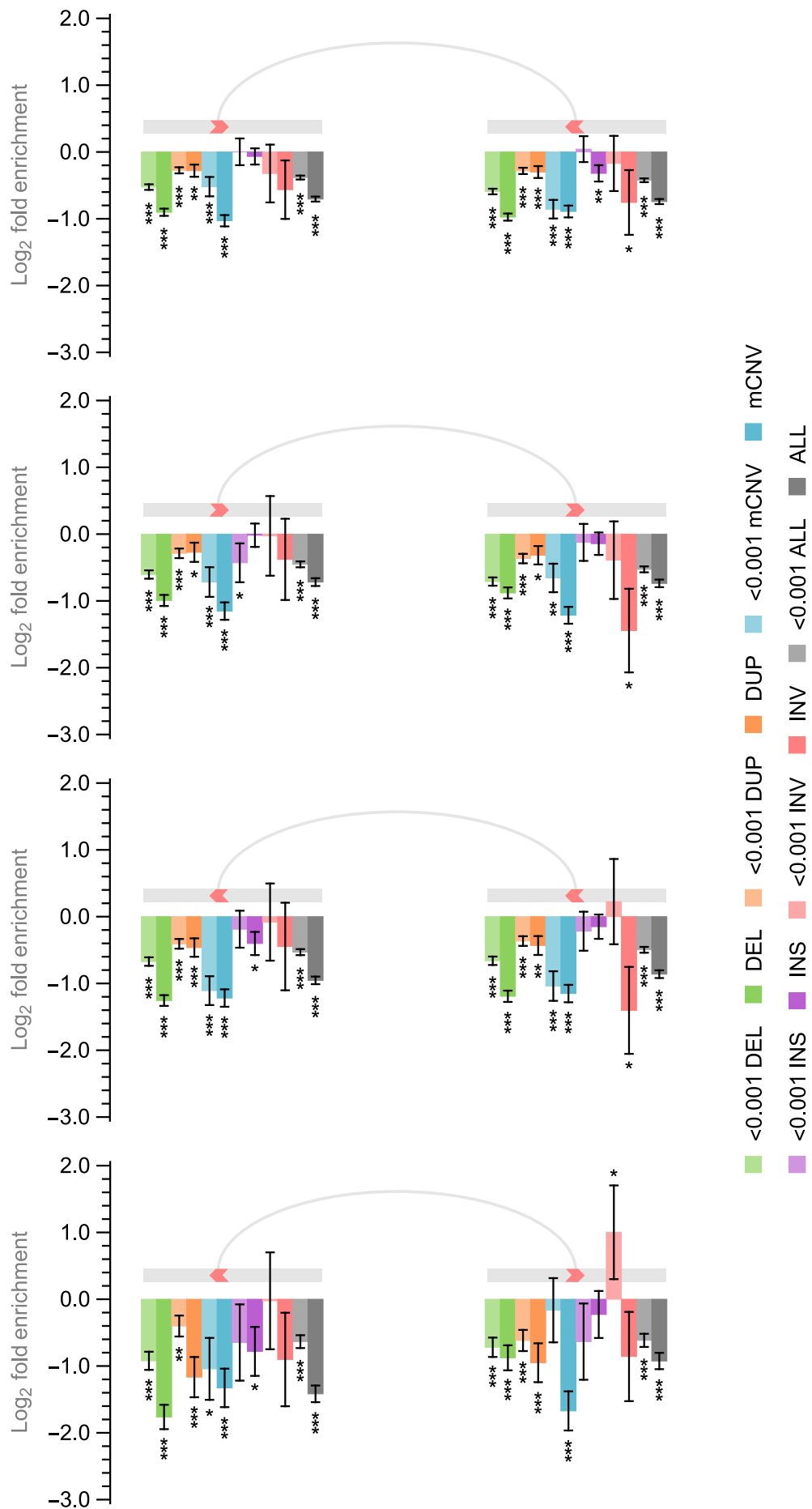
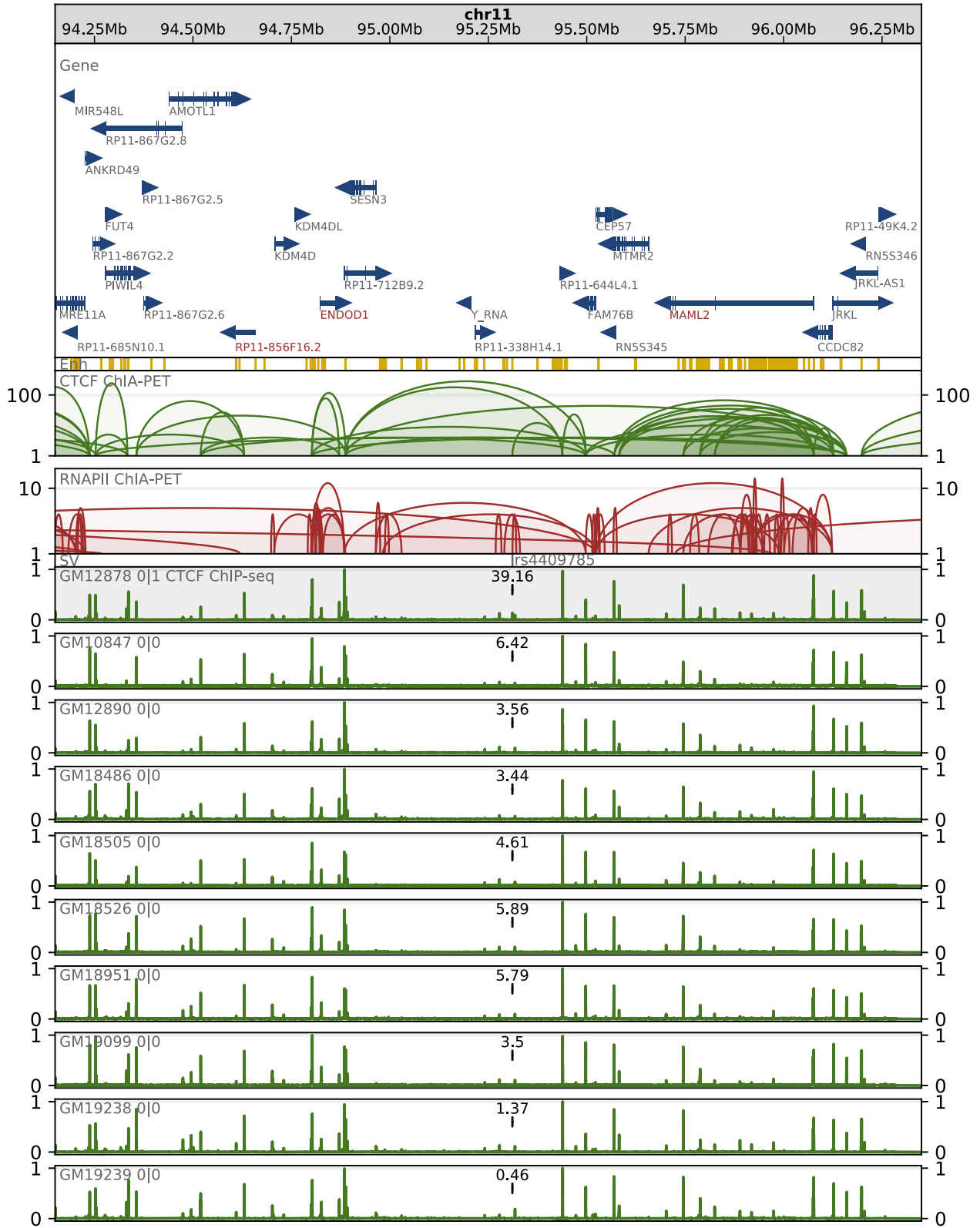
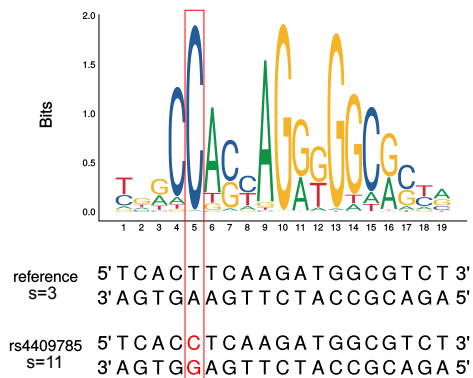


Fig. S14: Impact of structural variants on CTCF ChIA-PET interactions in GM12878 cells. Enrichment/depletion of anchors of CTCF PET clusters with SVs of different types and of different VAF (VAF < 0.001 and VAF \geq 0.001). Calculations for convergent, tandem right, tandem left and divergent loops are plotted separately.

A



B



C

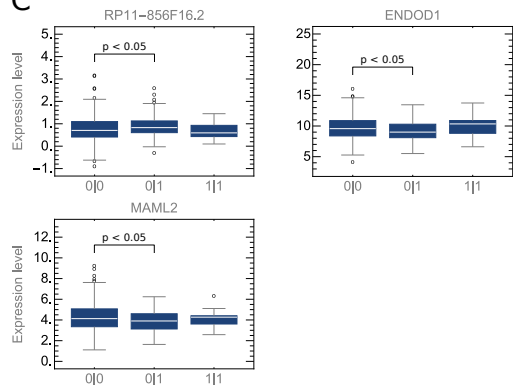
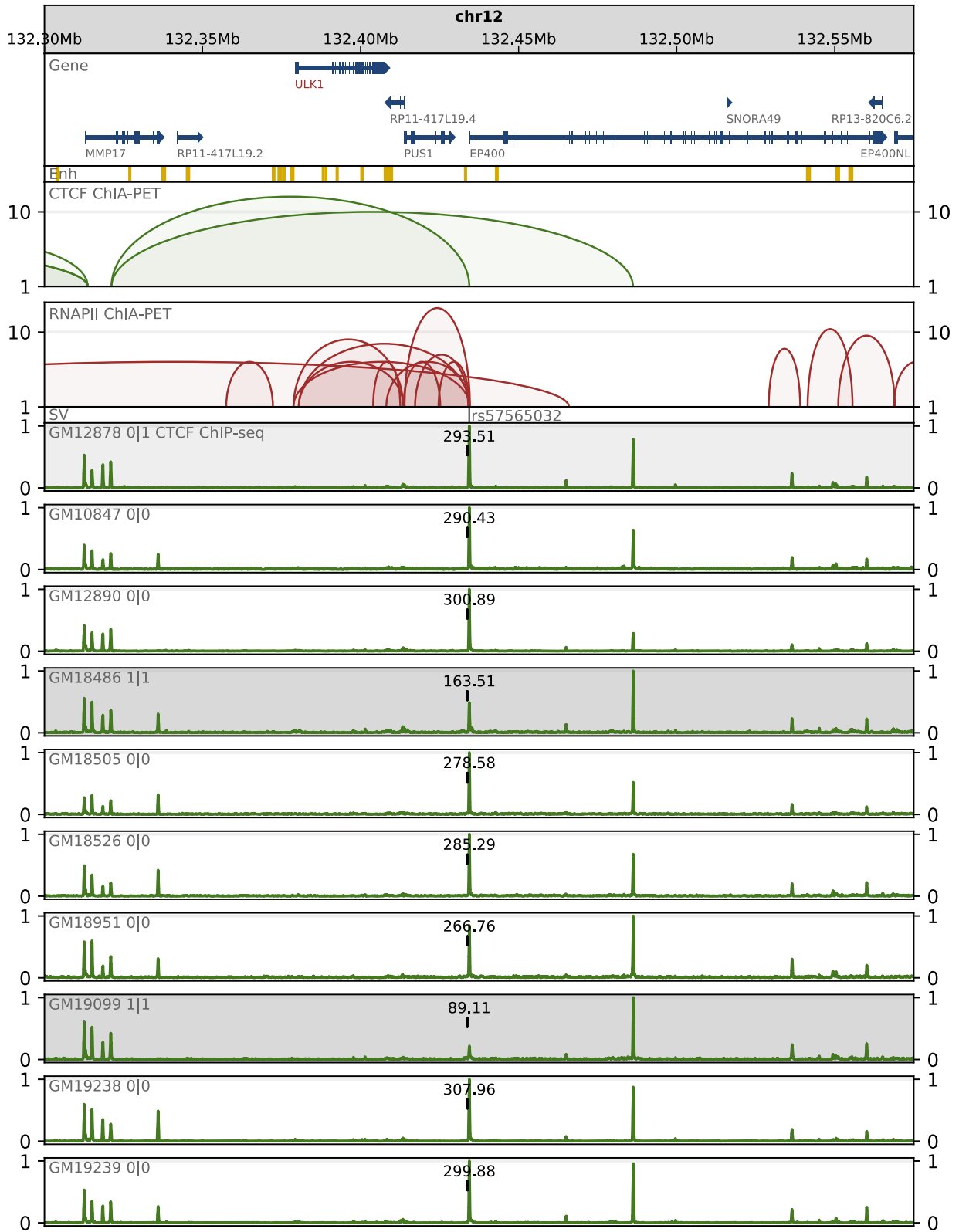
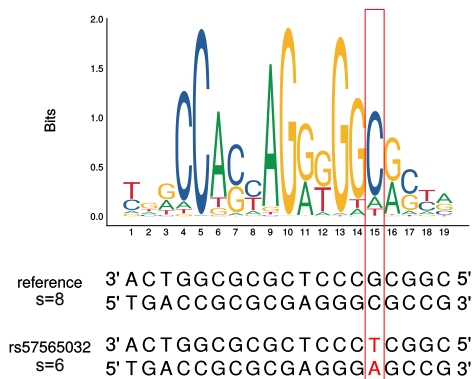


Fig. S15: GWAS SNP in a CTCF motif in region chr11:94150000-96350000. **a** Browser view of a 2 Mb genomic segment with SNP rs4409785 identified in a part of the human population and associated with rheumatoid arthritis and vitiligo. SNP rs4409785 alters sequence of a CTCF motif residing in an interaction anchor. CTCF ChIP-seq signals from 10 lymphoblastoid cell lines genotyped for this SNP are presented for comparison along with genes, enhancers and CTCF and RNAPII ChIA-PET interactions located in this genomic segment. ChIP-seq signal from each sample is measured in RPMs and divided by the maximal value of the signal in the visualized region. In each individual signal track, value of the signal summed over genomic region occupied by the altered CTCF anchor is additionally marked. **b** Comparison of sequences and scores of CTCF motifs with reference and alternative alleles defined by rs4409785. CTCF sequence logo demonstrates the importance of particular nucleotide positions in the motif. **c** Differences in gene transcription rates between genotypes set for rs4409785. Only genes exhibiting differences in transcription which pass Mann-Whitney test with p-value < 0.05 were reported.

A



B



C

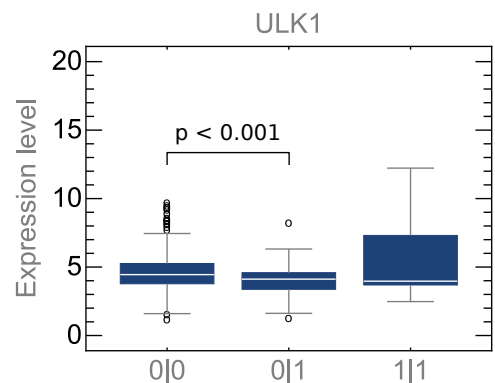


Fig. S16: GWAS SNP in a CTCF motif in region chr12:132300000-132575000. **a** Browser view of a 0.3 Mb genomic segment with SNP rs57565032 identified in a part of the human population and associated with red blood cell distribution width. SNP rs57565032 alters sequence of a CTCF motif residing in an interaction anchor. CTCF ChIP-seq signals from 10 lymphoblastoid cell lines genotyped for this SNP are presented for comparison along with genes, enhancers and CTCF and RNAPII ChIA-PET interactions located in this genomic segment. ChIP-seq signal from each sample is measured in RPMs and divided by the maximal value of the signal in the visualized region. In each individual signal track, value of the signal summed over genomic region occupied by the altered CTCF anchor is additionally marked. **b** Comparison of sequences and scores of CTCF motifs with reference and alternative alleles defined by rs57565032. CTCF sequence logo demonstrates the importance of particular nucleotide positions in the motif. **c** Transcription of gene ULK1 by genotypes set for rs57565032.

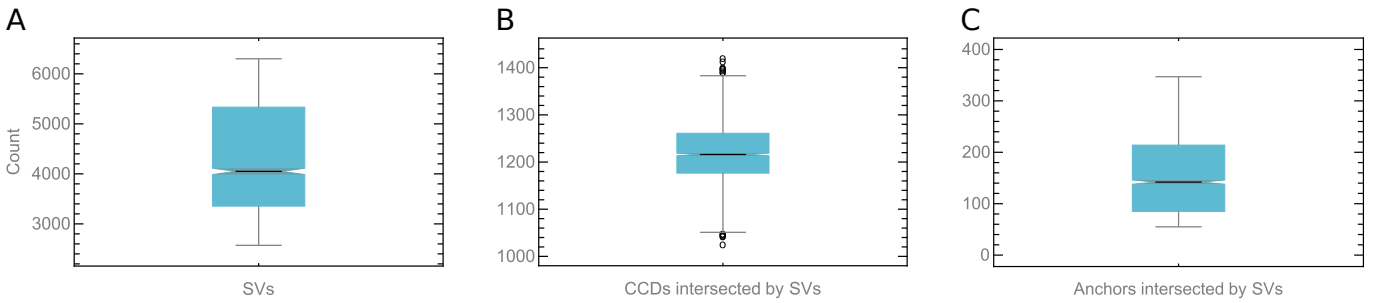


Fig. S17: Quantification of genomic elements in individual genomes. **a** Number of SVs (deletions, duplications, inversions and insertions adopted from the 1000 Genomes Project) per sample. **b** Number of the reference CCDs intersected with SVs in other lymphoblastoid cells. **c** Number of the reference CTCF-mediated interaction anchors intersected with SVs in other lymphoblastoid cells.

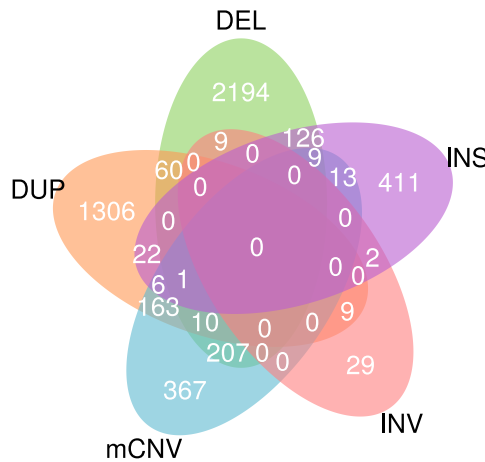


Fig. S18: Division of domain variability patterns by SV classes they contain.

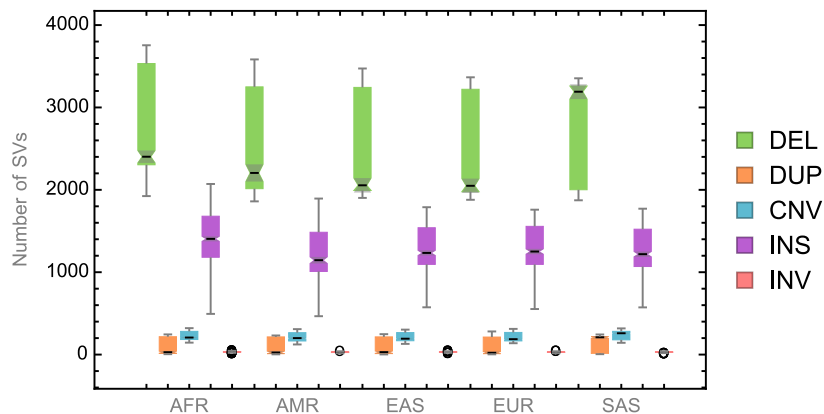


Fig. S19: Number of SVs per individual genome, grouped by SV class and population.

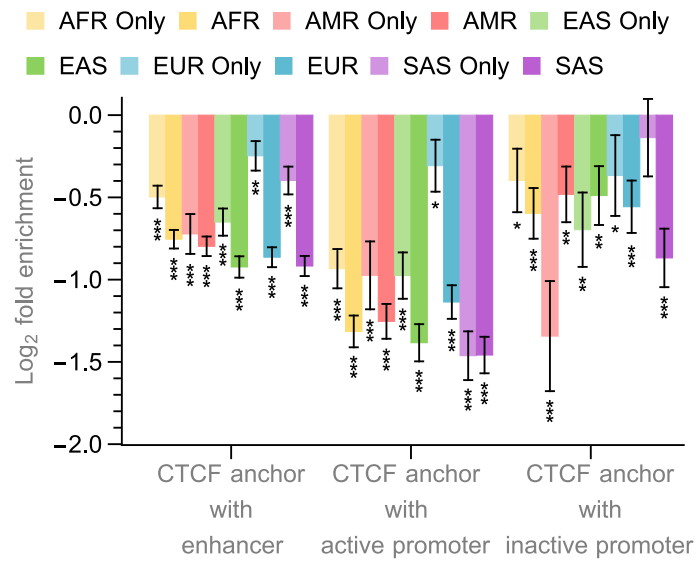


Fig. S20: Enrichment/depletion of CTCF anchors intersected with genomic functional elements identified in GM12878 cells with SVs divided by continental groups.

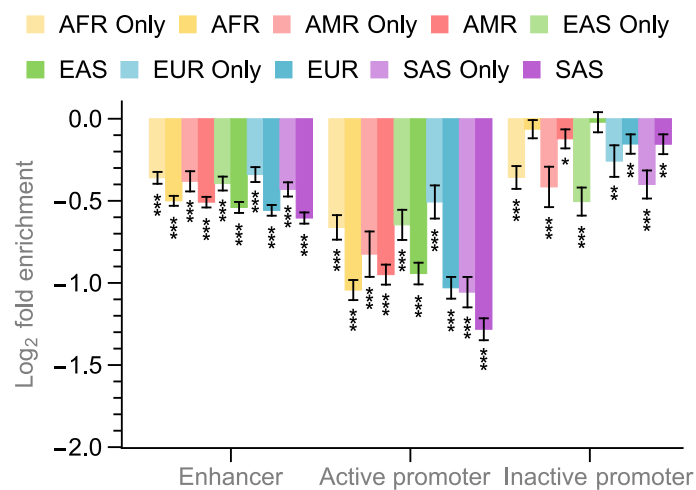


Fig. S21: Enrichment/depletion of genomic functional elements identified in GM12878 cells with SVs divided by continental groups.

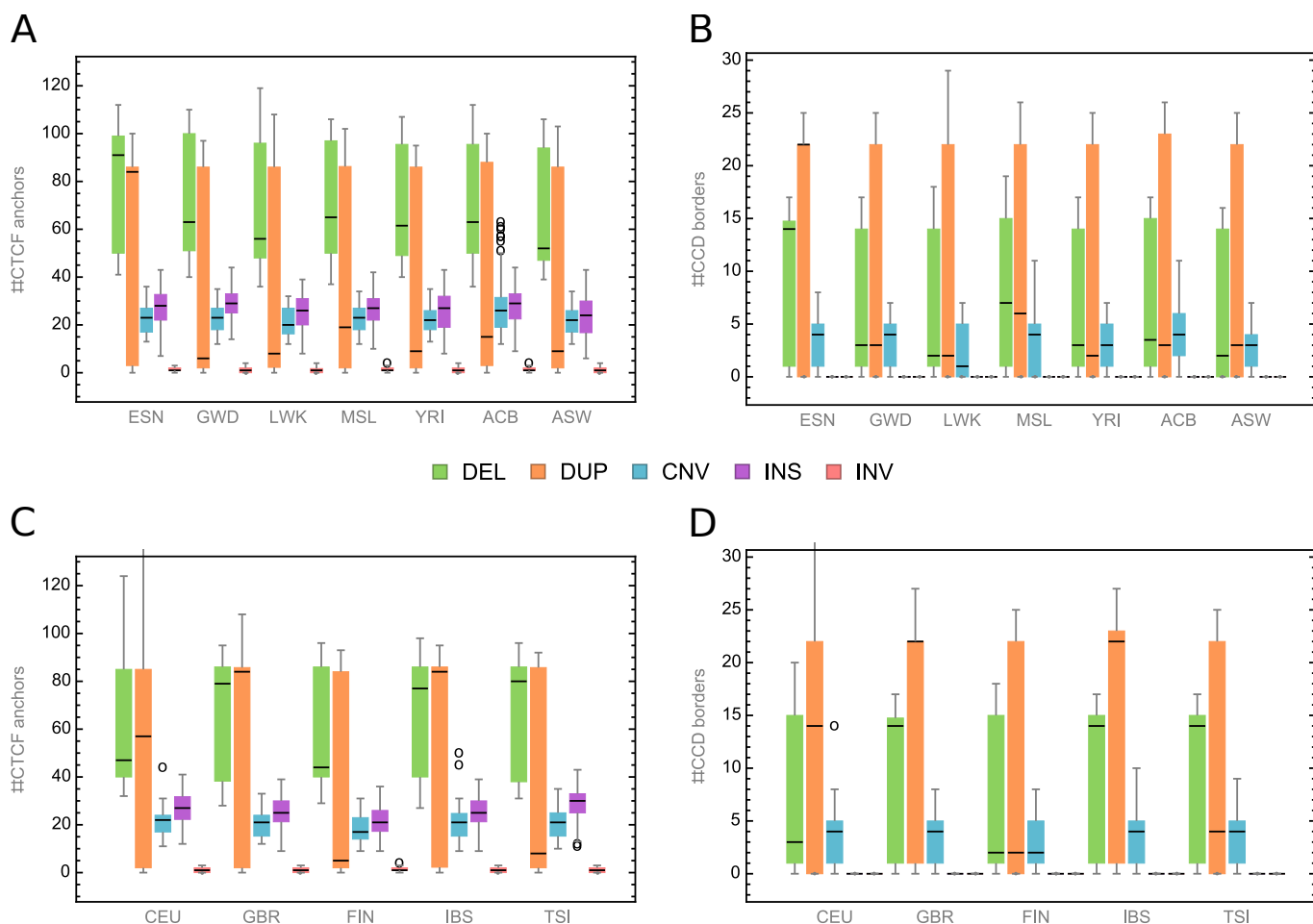
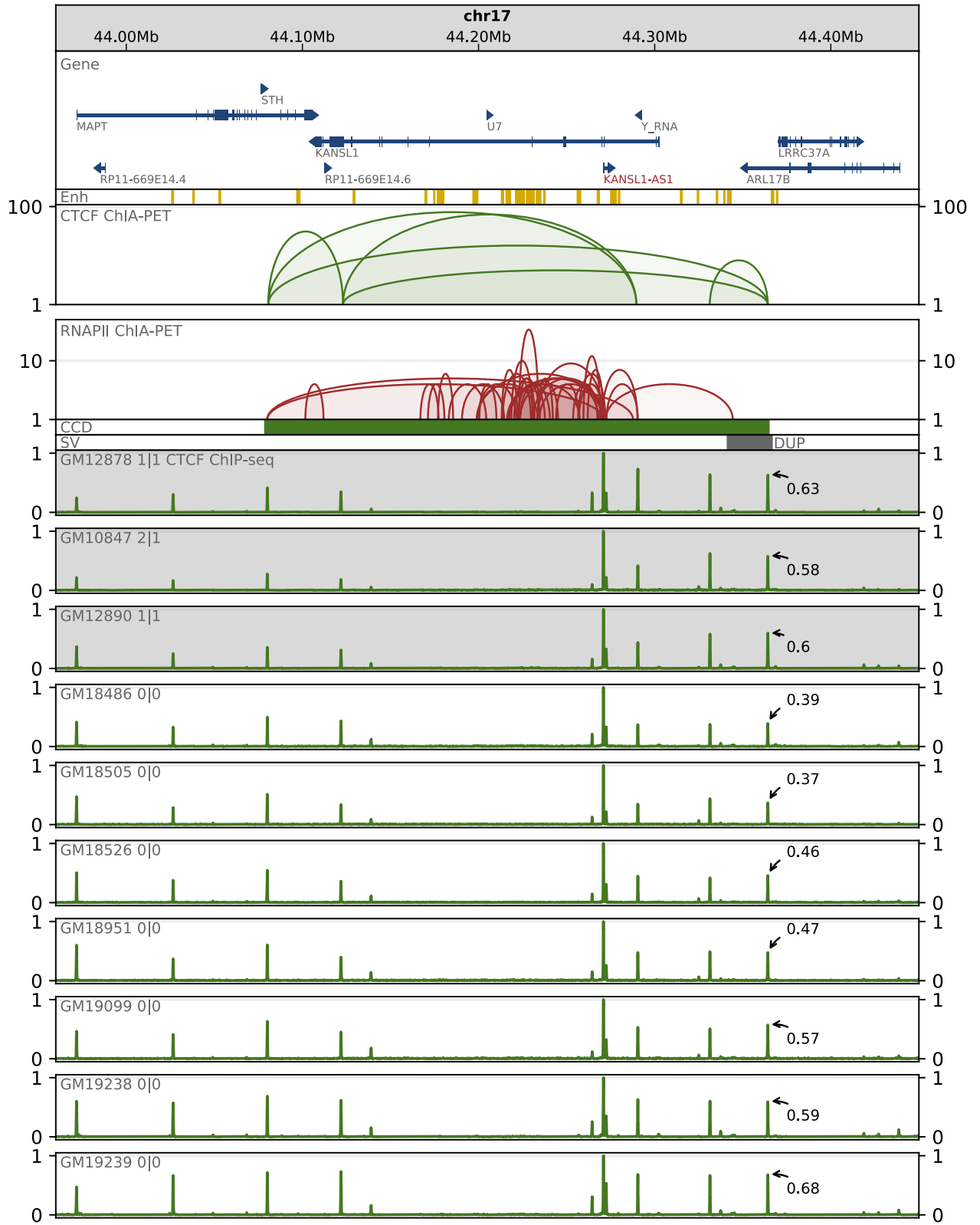


Fig. S22: Individual structural variation in African and European continental groups. **a** Number of CTCF anchors intersected by SVs of a given type identified in individuals from African continental group. **b** Number of domain borders fully overlapped by SVs of a given type identified in individuals from African continental group. **c** Similar to (a), but for European continental group. **d** Similar to (b), but for European continental group.

A



B

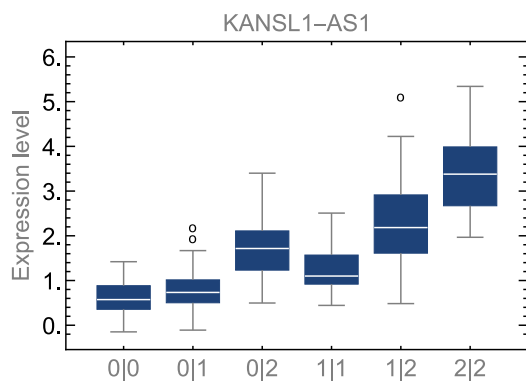


Fig. S23: Duplication of a CCD border in region chr17:43960000-44450000. **a** Browser view of a 0.5 Mb genomic segment with duplication chr17:44341412-44366497 identified in a part of the human population, which duplicates border of a topological domain and is an eQTL for the KANSL1-AS1 gene. CTCF ChIP-seq signals from 10 lymphoblastoid cell lines genotyped for this duplication are presented for comparison along with genes, enhancers and CTCF and RNAPII ChIA-PET interactions located in this genomic segment. ChIP-seq signal from each sample is measured in RPMs and divided by the maximal value of the signal in the visualized region. In each individual signal track, value of the highest signal peak in genomic region targeted by the SV is additionally marked. **b** Transcription of the KANSL1-AS1 gene by genotypes.

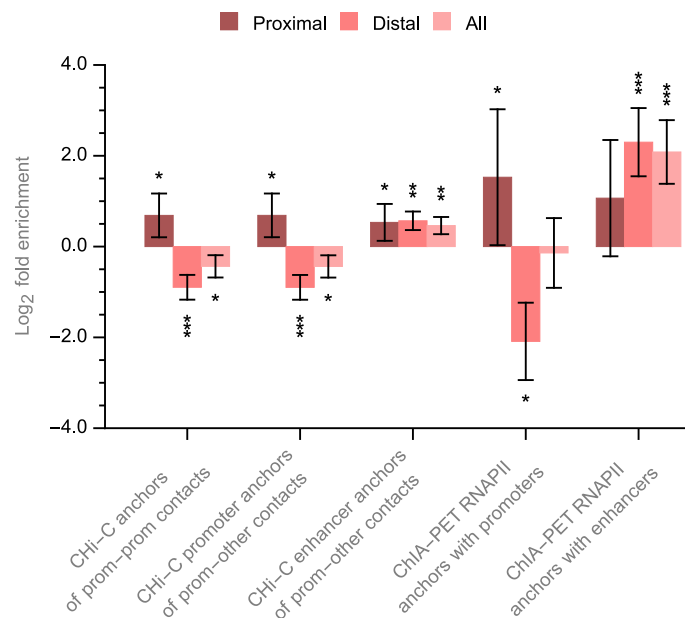
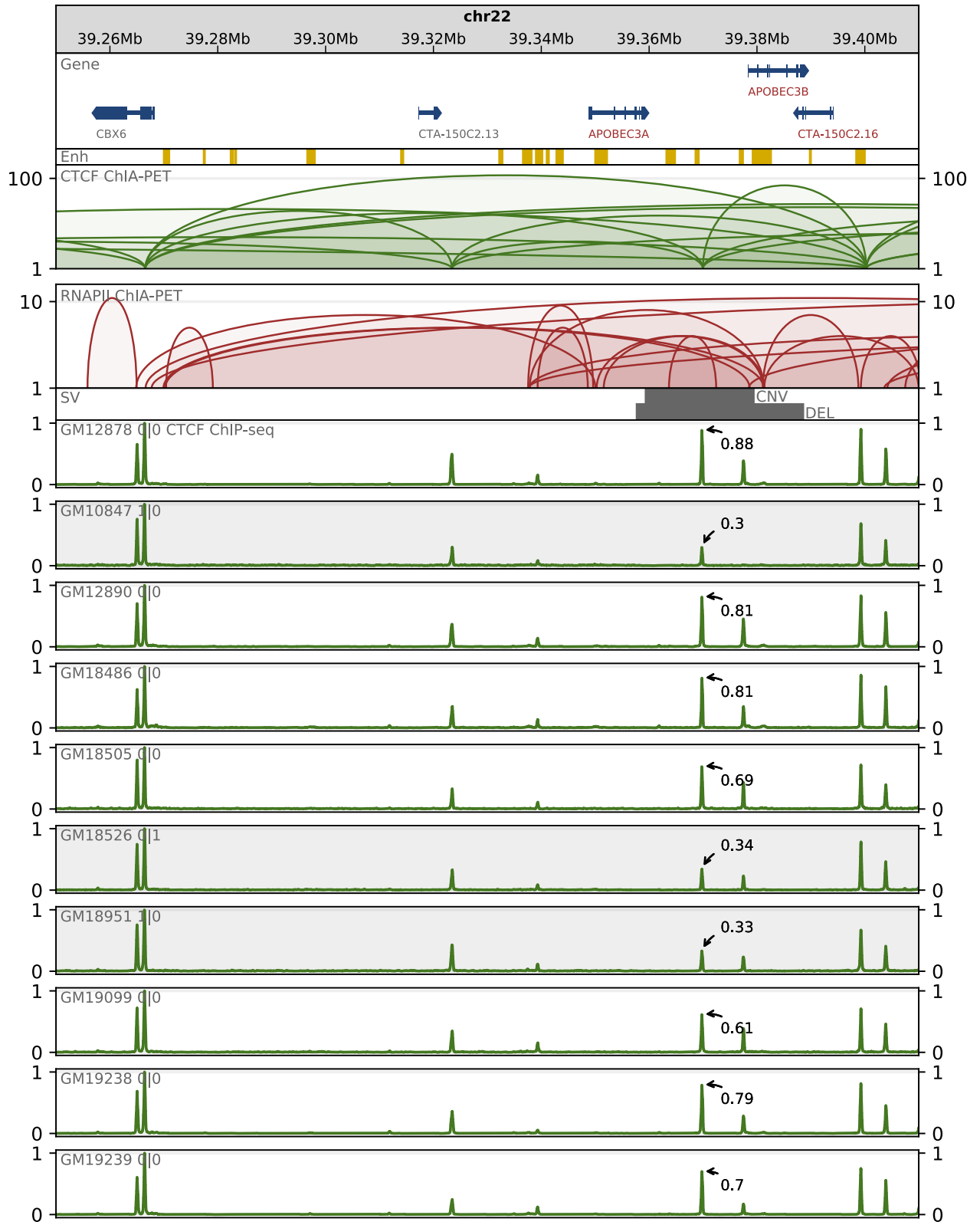


Fig. S24: eQTLs versus promoter interactions. Enrichment/depletion of ChIA-PET and ChI-C anchors containing promoters or enhancers with proximal and distal eQTLs. ChI-C anchors were divided into 3 groups: anchors of promoter-promoter interactions, promoter anchors of promoter-other region interactions and anchors intersected by enhancers of promoter-other region interactions.

A



B

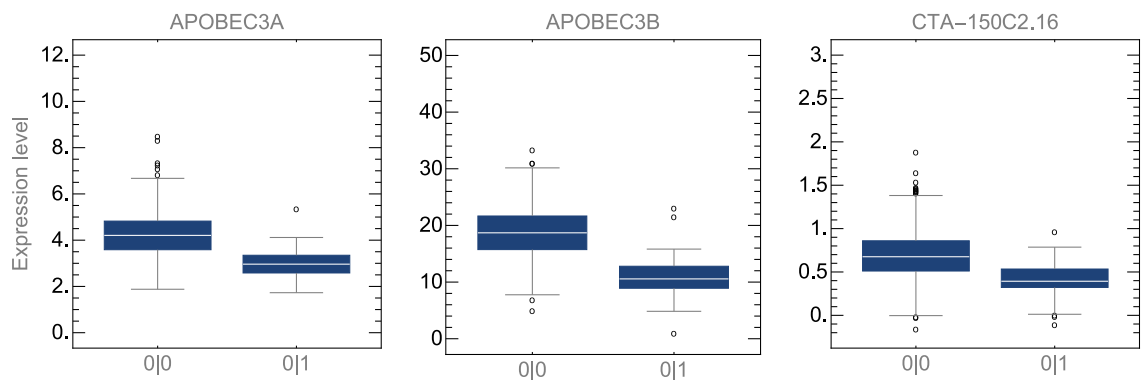
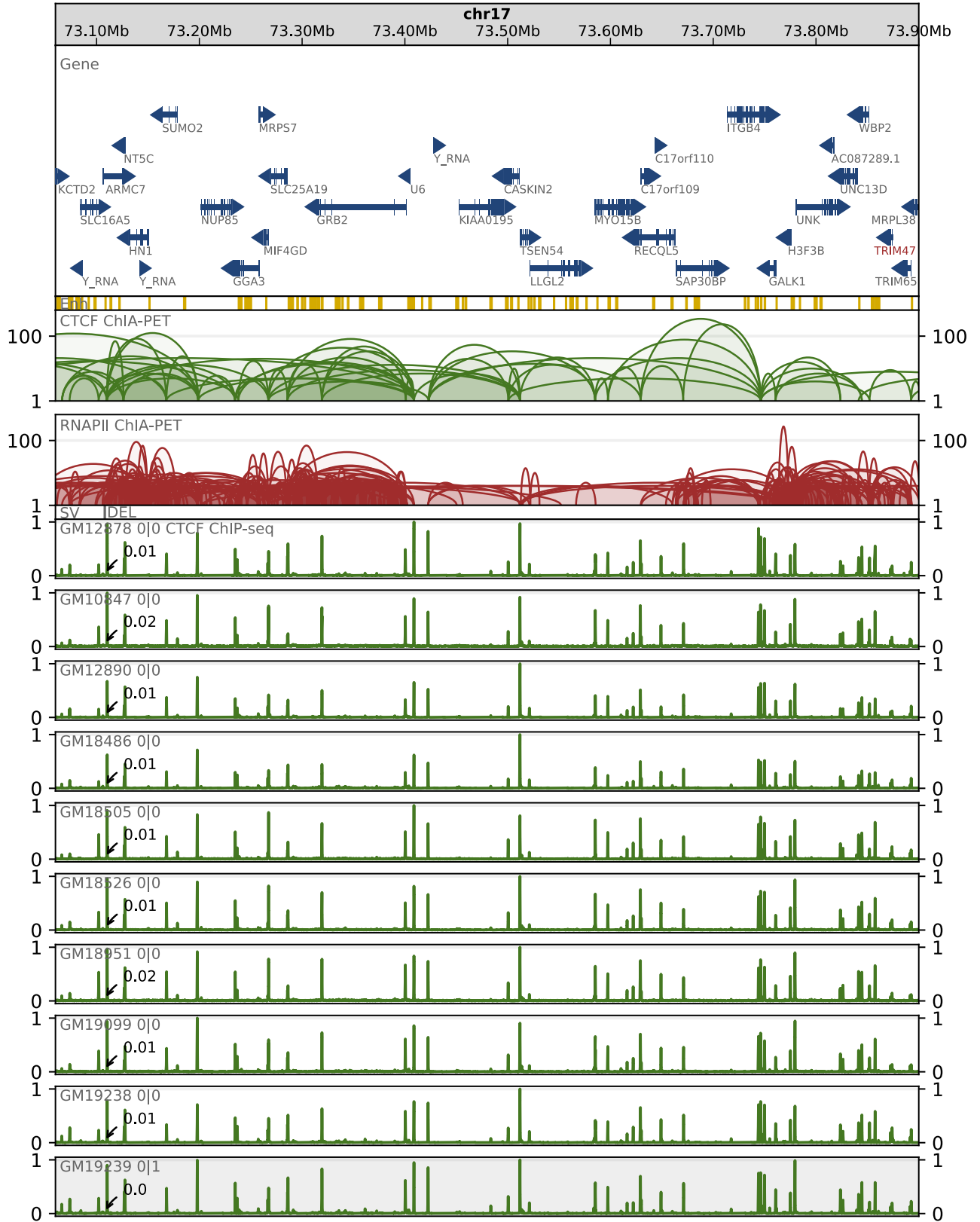


Fig. S25: Deletion of a CTCF interaction anchor in region chr22:39250000-39410000. **a** Browser view of a 0.2 Mb genomic segment with deletion chr22:39357694-39388574/CNV chr22:39359355-39379392 identified in a part of the human population, which removes/removes or multiplies CTCF anchor and is an eQTL for 3 neighboring genes (signed with the red font). APOBEC3A and APOBEC3B are immune-related genes. CTCF ChIP-seq signals from 10 lymphoblastoid cell lines are presented for comparison along with genes, enhancers and CTCF and RNAPII ChIA-PET interactions located in this genomic segment. All the presented cell lines with non-reference genotypes carry deletion chr22:39357694-39388574. ChIP-seq signal from each sample is measured in RPMs and divided by the maximal value of the signal in the visualized region. In each individual signal track, value of the highest signal peak in genomic region targeted by the SV is additionally marked. **b** Genes which transcription is correlated with deletion chr22:39357694-39388574 and CNV chr22:39359355-39379392 (p-value much smaller than 0.001).

A



B

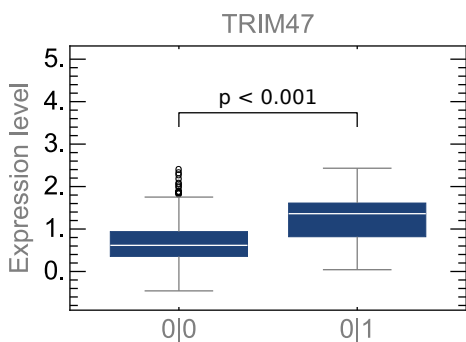


Fig. S26: Deletion in a CTCF interaction anchor in region chr17:73060000-73900000. **a** Browser view of a 0.8 Mb genomic segment with deletion chr17:73107713-73108273 identified in a part of the human population, which removes part of a CTCF anchor and is an eQTL for immune-related gene TRIM47 (signed with the red font). CTCF ChIP-seq signals from 10 lymphoblastoid cell lines genotyped for this deletion are presented for comparison along with genes, enhancers and CTCF and RNAPII ChIA-PET interactions located in this genomic segment. ChIP-seq signal from each sample is measured in RPMs and divided by the maximal value of the signal in the visualized region. In each individual signal track, value of the highest signal peak in genomic region targeted by the SV is additionally marked. **b** Immune-related gene TRIM47 which transcription is correlated with deletion chr17:73107713-73108273 (p-value much smaller than 0.001).

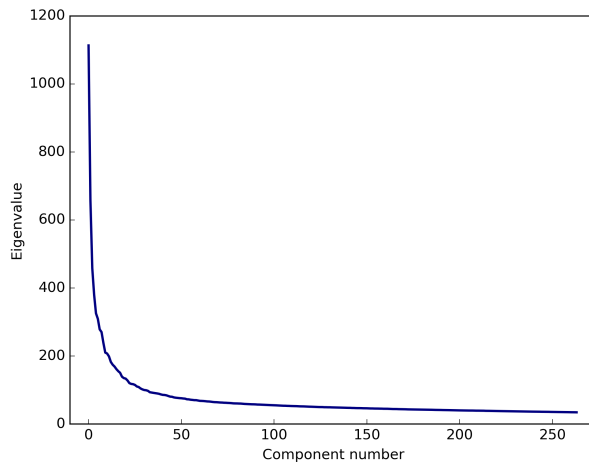


Fig. S27: Scree Plot. Showing sorted eigenvalues for PCA applied to the space of gene expression rates.

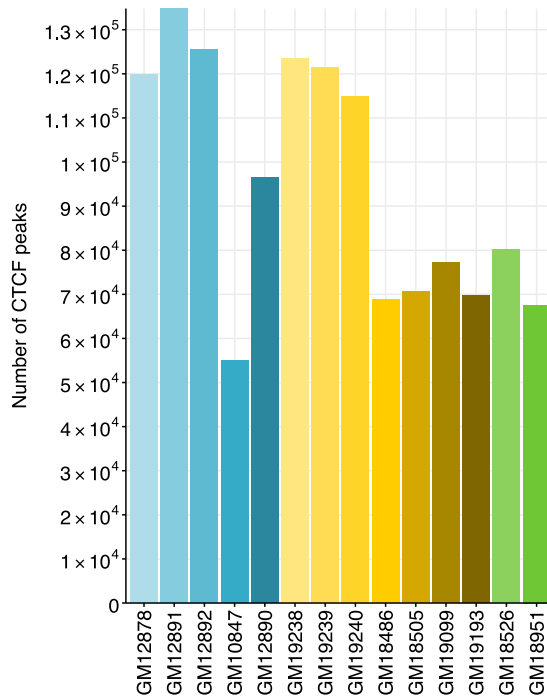


Fig. S28: Number of CTCF ChIP-seq peaks called in different human lymphoblastoid cell lines.

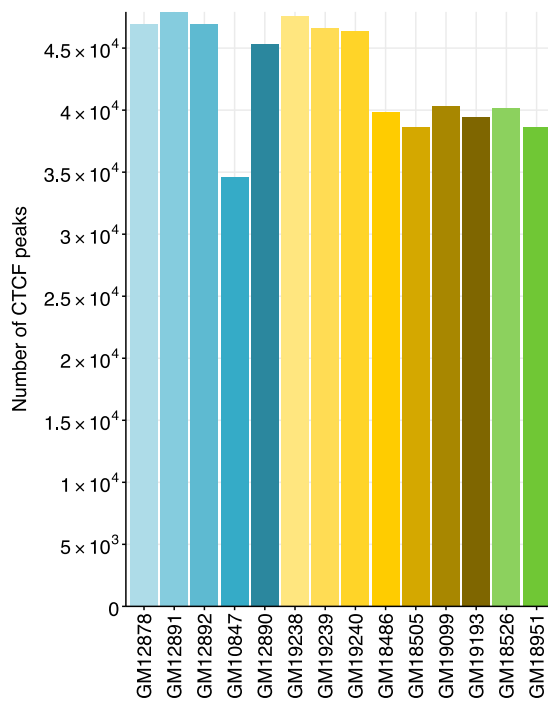


Fig. S29: Number of CTCF ChIP-seq peaks called in different human lymphoblastoid cell lines after filtering by conserved CTCF binding sites.

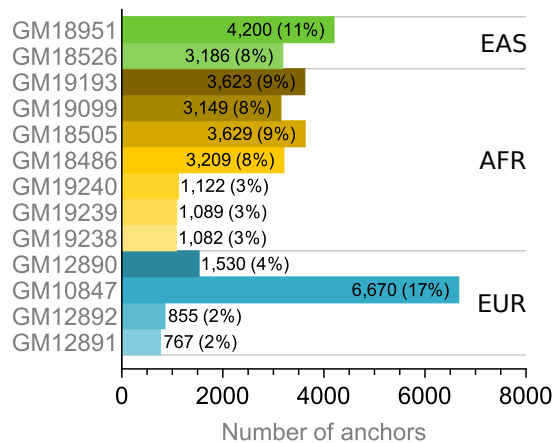


Fig. S30: CTCF anchors from GM12878 not intersected with CTCF ChIP-seq peaks identified in different lymphoblastoid cells.