

Supplementary methods S3

edgeR analysis

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Introduction

The edgeR analysis is performed 6 times, one for each tissue/statistical approach. Here, as an example, we show how it is performed with leaf samples against reference. For the other analysis, we follow the same pipeline, adjusting the variables to each wanted condition.

Pipeline - Leaf samples (against reference)

Data imput:

We load the summarized experiment obtained previously (supplementary file 1)

```
#1-. Load the SE objects:  
load("/home/albert/Desktop/Lolium/P226/se_P226.RData")  
#2-. Extract leaf samples:  
se.f <- se_P226[, se_P226$Tissue == "Leaf"]
```

Extracting the count matrices and the phenotype:

```
#1-. We extract the phenotype contained in the columns  
pheno <- colData(se.f)  
#2-. We set 35% as our first level:  
pheno$Soil_Humidity <- relevel(pheno$Soil_Humidity, "35%")  
#3-. We extract the count matrices  
counts <- assay(se.f)
```

```
#4-. We rename the count matrices according to the phenotype
colnames(counts) <- pheno$SampleName
```

Independent Filtering:

```
#1-. We establish our independent filtering threshold
keep <- rowSums(counts)>0
#2-. We apply the threshold to our counting matrices
counts.f <- counts[keep,]
```

Data normalization:

```
#1-. We create a dge object:
dge <- DGEList(counts=counts.f, group=pheno$Soil_Humidity)
#2-. We normalize our data using the TMM method:
dge.n <- calcNormFactors(dge, method = "TMM")
```

Model Building:

```
#1-. We set the design for our model:
design <- model.matrix(~0+dge.n$samples$group)
#2-. We change the rownames of our design:
rownames(design) <- colnames(dge.n)
#3-. We change the colnames of our design:
colnames(design) <- c("Leaf_35%", "Leaf_01%", "Leaf_05%", "Leaf_15%")
#4-. We estimate the dispersion parameters of our data:
dge.p <- estimateDisp(dge.n, design, robust=TRUE)
#5-. We fit our model:
fit <- glmQLFit(dge.p, design, robust=TRUE)
```

Data testing:

35% vs 15% analysis:

```
#1-. We contrast the conditions we want to assess:
qlf.1 <- glmQLFTest(fit, contrast=c(-1,0,0,1))
#2-. We obtain our genes results:
res_35vs15 <- topTags(qlf.1, n= nrow(qlf.1$table))
#3-. We transform our data into a data.frame:
res.df.1 <- as.data.frame(res_35vs15)
#4-. We get rid off those genes with NA values:
res.df.1 <- res.df.1[complete.cases(res.df.1),]
#5-. We order the results by
res.ord.1 <- res.df.1[order(res.df.1$FDR, decreasing = TRUE), ]
#6-. We save the results in a .csv file:
write.csv(file="P226_edgeR_Results_Leaf_AR_1.csv", x=res.ord.1)
```

```
#7-. We save the results in an excel file:
write.xlsx(res.ord.1, file="P226_edgeR_Results_Leaf_AR_1.xlsx", row.names = TRUE)
```

35% vs 5% analysis:

```
#1-. We contrast the conditions we want to assess:
qlf.2 <- glmQLFTest(fit, contrast=c(-1,0,1,0))
#2-. We obtain our genes results:
res_35vs05 <- topTags(qlf.2, n= nrow(qlf.2$table))
#3-. We transform our data into a data.frame:
res.df.2 <- as.data.frame(res_35vs05)
#4-. We get rid off those genes with NA values:
res.df.2 <- res.df.2[complete.cases(res.df.2),]
#5-. We order the results by
res.ord.2 <- res.df.2[order(res.df.2$FDR, decreasing = TRUE), ]
#6-. We save the results in a .csv file:
write.csv(file="P226_edgeR_Results_Leaf_AR_2.csv", x=res.ord.2)
#7-. We save the results in an excel file:
write.xlsx(res.ord.2, file="P226_edgeR_Results_Leaf_AR_2.xlsx", row.names = TRUE)
```

35% vs 1% analysis:

```
#1-. We contrast the conditions we want to assess:
qlf.3 <- glmQLFTest(fit, contrast=c(-1,1,0,0))
#2-. We obtain our genes results:
res_35vs01 <- topTags(qlf.3, n= nrow(qlf.3$table))
#3-. We transform our data into a data.frame:
res.df.3 <- as.data.frame(res_35vs01)
#4-. We get rid off those genes with NA values:
res.df.3 <- res.df.3[complete.cases(res.df.3),]
#5-. We order the results by
res.ord.3 <- res.df.3[order(res.df.3$FDR, decreasing = TRUE), ]
# 6-. We save the results in a .csv file:
write.csv(file="P226_edgeR_Results_Leaf_AR_3.csv", x=res.ord.3)
#7-. We save the results in an excel file:
write.xlsx(res.ord.3, file="P226_edgeR_Results_Leaf_AR_3.xlsx", row.names = TRUE)
```

Data saving:

```
#1-. We create an .RData object containing all data generated:
save.image(file= "P226_edgeR_Leaf_AR.RData")
```