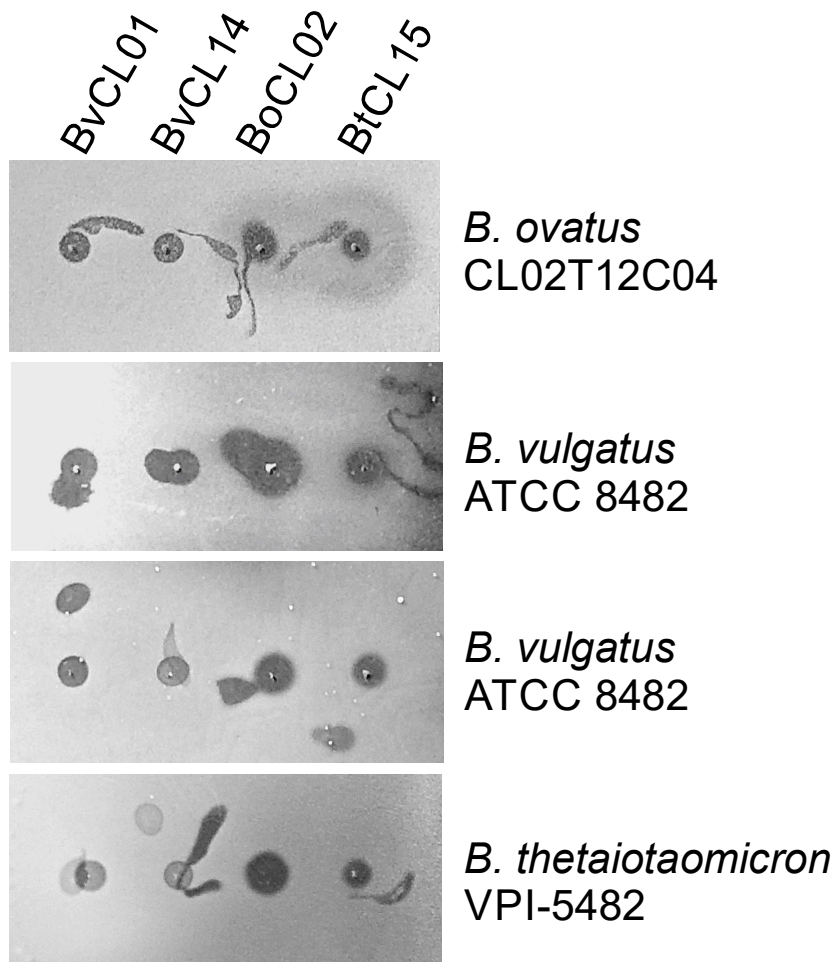


Supplementary Information

A family of anti-Bacteroidales peptide toxins wide-spread in the human gut microbiota

Coyne, Béchon, *et al.*



Supplementary Figure 1. Agar spot overlay assays demonstrating the movement of the toxin on the agar surface due to physical application of the overlay. The four toxin-producing strains are shown at the top and were cultured on the plate overnight, removed, then the plate was overlaid with top agar containing the strains shown on the left. Representative overlays are shown to illustrate the effect.

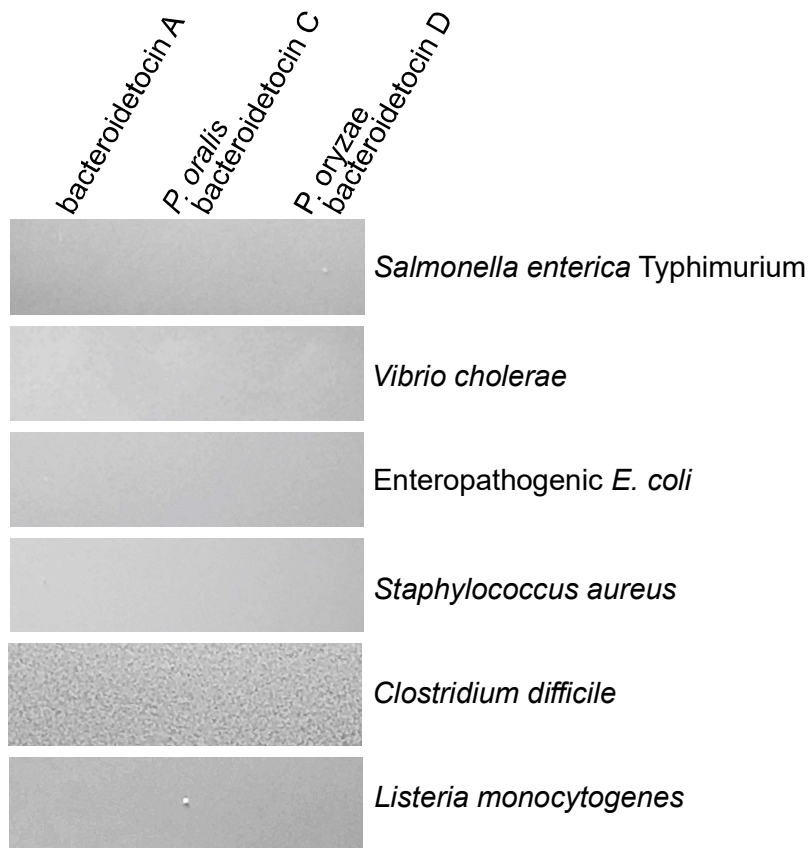
EH214_03825 *B. vulgatus* CL01T12C17 (bacteroidetocin A)

	no. times seen
M K K I S N E T L S Q V T G G K F W G S R V E C R V Q A N G H C M C R K V Y Y R F G I R S Y G G W W P A N P S Q C	2
K F W G S	55
K F W G S R	1
K F W G S R V E	12
F W G S R	1
F W G S R V E	11
V Q A N G H	170
K V Y Y R	89
V Y Y R	1
V Y Y R F	1
V Y Y R F G I R S Y G G W	1
Y R F G I R	2
R F G I R	1
F G I R S Y G G W W	1
S Y G G W W P A	3
S Y G G W W P A N	2
S Y G G W W P A N P	2
S Y G G W W P A N P S	57
S Y G G W W P A N P S Q C	9
Y G G W W P A N P S	1
Y G G W W P A N P S Q	1
G G W W P A N P S Q	1
G W W P A N P S Q	1

EH213_01844 *B. thetaiotomicron* CL15T12C11 (bacteroidetocin B)

	no. times seen
M K K V S N K V L S R A F G G K M F G S K T E C E R W E G G C R C R Q V Y Y T F W V K S Y G A W N P A A D W Q C N	11
K M F G S K	9
K M F G S K T E C E R	1
Q V Y Y T F	21
Q V Y Y T F W V K	2
Y T F W V K	1
T F W V K	1
S Y G A W N P A	2
S Y G A W N P A A D	4
S Y G A W N P A A D W Q	6
S Y G A W N P A A D W Q C N	1
G A W N P A A D W Q C N	1

Supplementary Figure 2. Fragments of bacteroidetocin A and B identified by LC-MS/MS analysis. Yellow highlight indicate residues not detected. Blue highlights are completely or partially digested tryptic fragments. Numbers on the right indicate the number of times the fragment was detected.



Supplementary Figure 3. Toxin dot assays of synthesized peptides bacteroidetocin A (left), the *Prevotella oralis* ATCC 33269 HMPREF0663_ORF3 peptide (bacteroidetocin C) (middle), and the *Prevotella oryzae* DSM 17970 XylorDRAFT_0090 peptide (bacteroidetocin D) (right) against bacterial pathogens of non-Bacteroidetes phyla. 1 µg of peptide was applied in a 5 µl spot on top of an agar overlay containing the indicated strains.

Supplementary Table 1. Strains used in this study.

Genome name	media liquid/solid	source
<i>Bacteroides vulgatus</i> CL01T12C17	basal/BHIS	Comstock Lab
<i>Bacteroides vulgatus</i> CL14T03C19	basal/BHIS	Comstock Lab
<i>Bacteroides ovatus</i> CL02T12C04	basal/BHIS	Comstock Lab
<i>Bacteroides ovatus</i> CL02T12C04 tn mut 206	basal/BHIS	Comstock Lab
<i>Bacteroides ovatus</i> CL02T12C04 tn mut 512	basal/BHIS	Comstock Lab
<i>Bacteroides ovatus</i> CL02T12C04 tn mut 363	basal/BHIS	Comstock Lab
<i>Bacteroides ovatus</i> CL02T12C04 tn mut 724	basal/BHIS	Comstock Lab
<i>Bacteroides thetaiotaomicron</i> CL15T12C11	basal/BHIS	Comstock Lab
<i>Bacteroides thetaiotaomicron</i> CL15T12C11 Δ 1843	basal/BHIS	Comstock Lab
<i>Bacteroides thetaiotaomicron</i> CL15T12C11 Δ 1844	basal/BHIS	Comstock Lab
<i>Bacteroides thetaiotaomicron</i> CL15T12C11 Δ 1845	basal/BHIS	Comstock Lab
<i>Bacteroides thetaiotaomicron</i> VPI 5482	basal/BHIS	ATCC
<i>Bacteroides vulgatus</i> ATCC 8482	basal/BHIS	ATCC
<i>Bacteroides fragilis</i> 638R	basal/BHIS	lab collection
<i>Parabacteroides merdae</i> CL03T12C32	basal/BHIS	Comstock Lab
<i>Parabacteroides johnsonii</i> CL02T12C29	basal/BHIS	Comstock Lab
<i>Parabacteroides goldsteinii</i> CL02T12C30	basal/BHIS	Comstock Lab
<i>Prevotella melaninogenica</i> D18	TSB-A/TSA-A	BEI
<i>Prevotella denticola</i> F0289, Oral Taxon 291	TSB-A/TSA-A	BEI
<i>Prevotella nigrescens</i> F0103	TSB-A/TSA-A	BEI
<i>Prevotella histicola</i> F0411	TSB-A/TSA-A	BEI
<i>Prevotella oralis</i> HGA0225	TSB-A/TSA-A	BEI
<i>Prevotella bivia</i> DNF00188	TSB-A/TSA-A	BEI
<i>Prevotella veroralis</i> F0319	TSB-A/TSA-A	BEI
<i>Prevotella intermedia</i> str. 17	TSB-A/TSA-A	lab collection
<i>Prevotella copri</i> DSM 18205	TSB-A/TSA-A	DSM
<i>Porphyromonas gingivalis</i> W83	TSB-A/TSA-A	lab collection
<i>Sphingobacterium spiritovorum</i> ATCC 33861	BHIS	ATCC
<i>Sphingobacterium multivorum</i> ATCC 35656	BHIS	ATCC
<i>Flavobacterium johnsoniae</i> ATCC 17061	BHIS	ATCC
<i>Elizabethkingia meningoseptica</i> ATCC 13253	BHIS	ATCC
<i>Escherichia coli</i> HS	BHIS	lab collection
<i>Lactobacillus rhamnosus</i> LMS2-1	BHIS	BEI
<i>Bifidobacterium breve</i> EX336960VC18	BHIS	BEI
<i>Enterococcus faecium</i> 503	basal/BHIS	BEI
<i>Salmonella enterica</i> Typhimurium ATCC 14028	BHIS	ATCC
Enteropathogenic <i>E. coli</i> 2348/69	BHIS	lab collection
<i>Vibrio cholerae</i> N16961	BHIS	lab collection
<i>Staphylococcus aureus</i> 502A	TSA-A	J. Lee
<i>Clostridium difficile</i> NAP07	BHIS	BEI
<i>Listeria monocytogenes</i> 104035	BHIS	M. Waldor
<i>Escherichia coli</i> DH5 α	LB	lab collection
<i>Escherichia coli</i> BL21/DE3	LB	lab collection
<i>Escherichia coli</i> S17 λ pir	LB	E. Martens

Supplementary Table 2. Primers used in this study.

Purpose	Primer Sequence ^a	Orientation
Sequence out of pYT646 junction	GACTTGGATACCTCACGCC	
Cloning of EH213_01845	AACGGGATCCATAGCAAAACTTCATGCGTAGCAA	Forward
	TTCAGGATCCCGGGACAAAACCTTTGTTGCTTACTT	Reverse
Cloning of EH213_01844	TTTTGGATCCGGAATAATAACAACCTTTATTGGTTTT	Forward
	GTCAGGATCCATTTACCTACTGGTTAGTTGCACTG	Reverse
Cloning of EH213_01843	TTATGGATCC GTAAAGCCAATTGATGGTATATCTCTTTTATT	Forward
	TCGCGGATCCTGTTGCGTAAATACTTTCTTCGCT	Reverse
Deletion of EH213_01845	CATAGGATCCGATATTCGCCGCTTTTCATTCTAC	left flank forward
	TGGTACGCGTTCAAATATTAAGTTTTCCAGTATGGT	left flank reverse
	TTTTACGCGTTCATACGGTATTGGTTTTGTGTTTT	right flank forward
	AGTTGGATCCTTTATGGATGCTTGAAGAATTTGA	right flank reverse
Deletion of EH213_01844	AGAGGGATCCATACAGGGAAATACGGATTTGAGA	left flank forward
	TTCAACGCGTGGGACAAAACCTTTGTTGCTTACTT	left flank reverse
	AATAACGCGTCAGTGCAACTAACCAGTAGGTAATAA	right flank forward
	AATAGGATCCACCGATCCGACCAAATATAGATAA	right flank reverse
Deletion of EH213_01843	AGTGGATCCCCCGGGCTGCACCATACTGGGAACTTAATATTTG	left flank forward
	TTAGTTGTTTTGTTGCAATACAATATTTTTTTCTATTC	left flank reverse
	TATTGCAACAAAACAACATAAAAAGTAATATGTAATGATATTAATATC	right flank forward
	CTTGATATCGAATTCCTGCAGACAACCAAATAAGCGAAG	right flank reverse
Cloning 1845-1842 into into pET16b	ACTTTAAGAAGGAGATATAC	Forward
	ATGCGTAGCAAGAAATATAAAAAATATATTTTG	
	CTGTGATGATGATGATGATGATGATGATGATGGCC AATCGTCATAGCAACCAAC	Reverse

^a Restriction sites used for cloning are underlined