

Electronic Supplementary Materials

Biomarker panels associated with progression of renal disease in type 1 diabetes

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ESM Methods

Use of hierarchical shrinkage priors

We adopted a Bayesian modelling approach based on hierarchical shrinkage priors [1], in which the clinical covariates used to control for confounding in the models were assigned a Gaussian prior (which induces some shrinkage), while the biomarkers were penalised through the horseshoe prior to promote sparsity [2], [3]. Standard methods for penalized regression include ridge regression (equivalent to a Gaussian prior on all coefficients) or LASSO regression (equivalent to a double exponential prior). Hierarchical shrinkage priors (such as the horseshoe) are more flexible than either of these, allowing the model for instance to learn a spike-and-slab distribution of effect sizes (in which a few variables have large effects but most have near zero effects) if that is what the data support. Developments in Bayesian computation, implemented in programs such as Stan and PyMC3, have made it possible to use these more flexible priors, even when some variables have high posterior correlations.

Note on the expected information for discrimination

For logistic regression models besides reporting the area under the ROC curve (AUC), we also presented the expected information for discrimination Λ expressed in bits [4]. The expected information for discrimination is the average weight of evidence (log-likelihood ratio) favouring true (case or control) over false (control or case) status, where the average is taken over the sampling distributions of the weight of evidence in cases and controls. A key advantage of using the information for discrimination is that the contributions of independent predictors are additive. Therefore, this is a better measure of the incremental contribution of biomarkers to the predictive performance, as it captures the amount of additional information that they contain over and beyond the initial set of clinical covariates. Computations were done with the R package `wevid` (version 0.6: <https://CRAN.R-project.org/package=wevid>).

Note on the projection predictive method

To recover a sparse model, we applied a projection approach according to which the high-dimensional posterior draws of the model containing all biomarkers (full model) are projected to lower-dimensional subspaces [5], [6]. The advantages of the projection predictive method for variable selection have been discussed by Piironen and Vehtari [7]. The method compares the predictive distribution of each candidate model based on a subset of variables to the predictive distribution of the full model, using the Kullback-Leibler (KL) divergence to evaluate the discrepancy between the candidate model and the full model. By fitting the candidate model to the predictive distribution rather than to the target variable, overfitting is reduced.

ESM References

- [1] Peltola T, Havulinna AS, Salomaa V, Vehtari A. Hierarchical Bayesian survival analysis and projective covariate selection in cardiovascular event risk prediction. In: Laskey KB, Jones J, Almond R, editors. Proceedings of eleventh UAI Bayesian Modeling Applications Workshop (BMAW 2014), CEUR workshop proceedings, vol. 1218, 2014, pp. 79–88.
- [2] Carvalho C, Polson N, Scott J. The horseshoe estimator for sparse signals. *Biometrika* 2010;97:465–80.
- [3] Piironen J, Vehtari A. On the hyperprior choice for the global shrinkage parameter in the horseshoe prior. In: Proceedings of the 20th international conference on artificial intelligence and statistics (AISTATS), PMLR, vol. 54, 2017, pp. 905–13.
- [4] McKeigue P. Quantifying performance of a diagnostic test as the expected information for discrimination: Relation to the C-statistic. *Statistical Methods in Medical Research* 2018.
- [5] Goutis C, Robert CP. Model choice in generalised linear models: A Bayesian approach via Kullback-Leibler projections. *Biometrika* 1998;85:29–37.
- [6] Dupuis J, Robert CP. Variable selection in qualitative models via an entropic explanatory power. *Journal of Statistical Planning and Inference* 2003;111:77–94.
- [7] Piironen J, Vehtari A. Comparison of Bayesian predictive methods for model selection. *Statistics and Computing* 2017;27:711–35.

ESM Table 1: Participant characteristics at study day stratified by rapid progression status

Covariate	SDRNT1BIO		FimDiane	
	Stable (N = 665)	Rapid progressor (N = 194)	Stable (N = 188)	Rapid progressor (N = 127)
Main characteristics				
Age (years)	55.7 (46.2, 64.5) 58.2%	54.5 (45.9, 63.8) 52.1%	46.8 (37.8, 53.7) 47.9%	43.6 (33.8, 50.7) 40.2%
Sex (Female)	26.4 (16.9, 37.9)	26.7 (18.9, 36.5)	33.4 (26.9, 39.1)	29.9 (24.3, 34.8)
Diabetes duration (years)				
Kidney function				
eGFR (ml/min/1.73m ²)	72.5 (62.8, 83.7)	70.2 (57.3, 85.4)	61.1 (47.5, 68.4)	54.3 (43.6, 64.0)
Achieved eGFR (ml/min/1.73m ²)	76.6 (64.2, 88.3)	53.3 (32.8, 74.1)	51.8 (30.9, 64.3)	10.0 (10.0, 10.0)
Weighted average of historical eGFR (ml/min/1.73m ²)	81.5 (70.6, 91.3)	80.2 (70.7, 91.9)	68.2 (58.6, 78.0)	58.7 (45.8, 71.9)
Albumin/creatinine ratio (mg/mmol)	0.5 (0.3, 1.2)	1.1 (0.4, 10.2)	11.8 (4.7, 43.3)	56.3 (20.5, 129.5)
ACR category (Normo/Micro/Macro)	87.9%/10.0%/2.1%	67.7%/ ^a 13.4%/ ^b 18.8%	0%/ ^a 37.8%/ ^b 62.2%	0%/ ^a 11.8%/ ^b 88.2%
Prospective eGFR slope (ml/min/1.73m ² /year)	-0.1 (-1.4, 1.1)	-5.2 (-6.7, -4.0)	-1.3 (-2.1, -0.3)	-5.0 (-7.1, -3.9)
Other covariates				
Length of follow-up (years)	5.2 (4.5, 5.7)	5.1 (4.4, 5.7)	10.3 (7.2, 13.3)	6.5 (4.8, 9.8)
HbA1c (mmol/mol)	67 (60, 77)	73 (64, 88)	68 (58, 76)	79 (69, 88)
HbA1c (%)	8.3 (7.6, 9.2)	8.8 (8.0, 10.2)	8.4 (7.5, 9.1)	9.4 (8.5, 10.2)
Body mass index (kg/m ²)	27.1 (24.6, 30.3)	27.4 (24.6, 30.7)	26.0 (23.2, 28.0)	25.6 (23.4, 28.7)
HDL-C (mmol/l)	1.6 (1.3, 1.9)	1.5 (1.2, 1.9)	0.9 (0.7, 1.1)	0.8 (0.6, 1.0)
Total cholesterol (mmol/l)	4.5 (3.9, 5.1)	4.5 (4.0, 5.3)	5.0 (4.4, 5.6)	5.3 (4.7, 6.0)
Systolic blood pressure (mmHg)	133 (121, 144)	137 (126, 151)	141 (130, 154)	150 (132, 159)
Diastolic blood pressure (mmHg)	74 (68, 80)	75 (68, 82)	80 (71, 87)	84 (78, 90)
Ever smoker	64.2%	65.5%	47.9%	58.3%
On any anti-hypertensive treatment	59.2%	70.1%	93.6%	95.3%
On ACE or ARB	53.8%	65.5%	85.6%	86.6%

We report median and interquartile range (IQR) for continuous variables, and frequency for categorical variables

Stable: prospective eGFR slope $\geq -3 \text{ ml min}^{-1} [1.73\text{m}]^{-2}$ Rapid progressor: prospective eGFR slope $< -3 \text{ ml min}^{-1} [1.73\text{m}]^{-2}$

ESM Table 2: Summary characteristics of all biomarkers measured in the study

Biomarker	Units	SDRNTIBIO			FinDiane		
		Median (IQR)	Range	Below	Median (IQR)	Range	Below
Luminex proteins							
Alpha-1-Microglobulin	µg/mL	17.0 (14.0, 22.0)	4.3 – 77.5	31.0 (23.0, 40.0)	6.6 – 119.0		
Beta-2-Microglobulin	µg/mL	2.3 (1.8, 2.9)	0.9 – 8.3	3.9 (3.0, 5.2)	1.0 – 13.0		
Calbindin	ng/mL	Intraclass correlation < 0.4	0.4 – 510.0	51.2	Affected by storage conditions	43.1	
CD27 antigen	U/mL	42.0 (29.0, 69.0)	0.1	183.8 (112.0, 301.5)	19.0 – 867.0		
Clusterin	µg/mL	Intraclass correlation < 0.4	333.0 – 2100.0	Affected by storage conditions			
Cystatin-C	ng/mL	758.0 (632.5, 948.0)	51.0 – 6510.0	111.0 (900.5, 1370.0)	365.0 – 4480.0		
Eotaxin-2	pg/mL	884.0 (569.5, 1420.0)	4.6 (4.6, 4.6)	Intraclass correlation < 0.4			
Fatty Acid-Binding Protein, heart	ng/mL	0.1 (0.1, 0.3)	0.1 – 2.8	0.3 (0.2, 0.5)	0.1 – 5.3		
Fibroblast Growth Factor 21	ng/mL	0.1 (0.1, 0.1)	0.0 – 2.7	0.1 (0.0, 0.1)	0.0 – 0.8		
Fibroblast Growth Factor 23	ng/mL	123.0 (92.0, 173.0)	13.0 – 968.0	128.0 (90.5, 189.0)	21.0 – 653.0		
Growth-Regulated alpha protein	pg/mL	43.0 (36.0, 49.0)	9.3 – 90.0	53.0 (45.0, 64.0)	6.6 – 166.5		
Insulin-like Growth Factor-Binding Protein 7	ng/mL	1250.0 (1060.0, 1460.0)	216.0 – 2840.0	Affected by storage conditions			
Interleukin-1 receptor type 1	pg/mL	8.6 (7.2, 10.0)	1.6 – 19.0	Affected by storage conditions			
Interleukin-1 receptor type 2	pg/mL	2620.0 (2100.0, 3340.0)	955.0 – 12600.0	Affected by storage conditions			
Interleukin-2 receptor alpha	pg/mL	0.0 (0.0, 0.1)	0.0 – 1.3	3250.0 (2475.0, 4165.0)	1130.0 – 12100.0		
Kidney Injury Molecule-1	ng/mL	11.0 (8.6, 12.8)	0.6 – 20.0	0.1 (0.0, 0.2)	0.0 – 2.5		
Latency-Associated-Peptide	ng/mL	0.8 (0.3, 1.3)	0.3 – 7.1	9.2 (6.8, 12.0)	0.8 – 23.0		
Lectin-Like Oxidized LDL Receptor 1	ng/mL	381.0 (282.0, 519.0)	58.0 – 1410.0	0.3 (0.3, 1.1)	0.3 – 6.6		
Neutrophil Gelatinate-Associated Lipocalin	ng/mL	289.0 (136.0, 632.5)	7.5 – 10650.0	Affected by storage conditions			
N-terminal prohormone of brain natriuretic peptide	pg/mL	12.0 (8.5, 17.0)	0.9 – 65.0	39.0 (7.5, 182.5)	7.5 – 7690.0		
Osteopontin	ng/mL	6.8 (5.5, 8.4)	0.6 – 19.0	9.1 (6.4, 13.0)	0.9 – 54.0		
Osteoprotegerin	pM	11.0 (7.9, 16.0)	2.4 – 2190.0	6.6 (5.4, 8.1)	1.8 – 16.0		
Progranulin	ng/mL	0.1 (0.0, 0.1)	0.0 – 0.1	14.0 (11.0, 21.0)	2.4 – 276.0		
Tamm-Horsfall Urinary Glycoprotein	ng/mL	5.0 (4.2, 6.2)	1.8 – 19.0	0.1 (0.0, 0.1)	0.0 – 0.1		
Thrombomodulin	pg/mL	275.0 (154.0, 457.0)	39.0 – 4040.0	7.7 (5.7, 10.0)	2.3 – 22.0		
Thymus-Expressed Chemokine	ng/mL	141.0 (123.0, 163.0)	65.0 – 340.0	277.0 (180.5, 437.0)	47.5 – 2670.0		
Tissue Inhibitor of Metalloproteinases 1	pg/mL	0.1 (0.1, 0.2)	0.0 – 3.0	168.0 (142.0, 193.0)	70.0 – 323.0		
Tumor Necrosis Factor Receptor 1	pg/mL	1760.0 (1340.0, 2385.0)	99.0 – 8340.0	Affected by storage conditions			
Vascular Endothelial Growth Factor	pg/mL	256.0 (167.0, 388.0)	21.0 – 1060.0	270.0 (190.5, 3620.0)	242.0 – 11300.0		
			0.4	276.0 (155.0, 446.0)	21.0 – 2030.0		
LC-MS/MS metabolites							
1-Methyl-L-histidine	nM/L	5.1 (2.2, 11.8)	0.2 – 79.0	3.6	5.6 (2.9, 12.5)	0.2 – 150.0	0.6
3-hydroxykynurenine	nM/L	Excessive left-censoring		99.9	Excessive left-censoring	98.2	
3-hydroxytryptamine HCl [conf]	nM/L	0.1 (0.1, 0.1)	0.1 – 9.6	97.7	Excessive left-censoring	98.5	
3-hydroxytyramine HCl	nM/L	0.1 (0.1, 0.1)	0.1 – 7.0	97.7	Excessive left-censoring	98.2	
3-Methyl-histidine	nM/L	4.5 (3.4, 6.0)	0.7 – 24.2	7.1 (5.1, 9.8)	Intraclass correlation < 0.4	97.9	
3OMD	nM/L	189.0 (161.0, 219.0)	69.0 – 3788.0	Excessive missingness at random			
5-HtAA	nM/L	Excessive left-censoring		99.7	Excessive missingness at random	98.2	
5-hydroxytryptophan	nM/L	Excessive left-censoring		99.8	Excessive left-censoring	98.5	
8OHdG	nM/L	Excessive left-censoring		99.7	Excessive left-censoring	97.9	
Adenine [conf]	nM/L	Excessive missingness at random		99.7	Excessive missingness at random	98.5	
Adenine	nM/L	Excessive missingness at random		99.1	Excessive left-censoring	98.2	
Adenosine	nM/L	Excessive left-censoring		478.0 (431.0, 537.0)	220.0 – 769.0		
ADMA	nM/L	468.5 (411.6, 523.0)	270.0 – 958.0	Affected by storage conditions			
Alanine	nM/L	452.5 (383.9, 525.9)	152.0 – 1329.3	250.0 – 250.0	250.0 – 1043.0		
Anthranilic acid	nM/L	Excessive left-censoring		52.0 – 341.3	Affected by storage conditions	91.2	
Arginine	nM/L	116.9 (102.1, 134.9)	99.9	Excessive left-censoring			
Arginosuccinic acid	nM/L	Excessive left-censoring		99.9	Excessive left-censoring	98.5	
Arginosuccinic anhydride	nM/L	24.5 (20.4, 29.6)	11.1 – 49.9	99.9	Excessive left-censoring	98.5	
Aspartic acid	nM/L				Affected by storage conditions		

ESM Table 2: Summary characteristics of all biomarkers measured in the study (continued)

Biomarker	Units	SDRNtBIO			FinDiane		
		Median (IQR)	Range	Below	Median (IQR)	Range	Below
Beta-Alanine	µM/L	9.8 (8.4, 11.8)	4.3 – 83.9		9.5 (8.3, 11.3)	5.2 – 46.5	
C10:1 Carnitine	nM/L	99.0 (69.0, 141.0)	5.0 – 525.0	0.6	80.5 (43.0, 125.0)	5.0 – 403.0	1.8
C10 Carnitine	nM/L	157.0 (100.0, 252.1)	15.0 – 1472.0	1.5	149.5 (88.5, 252.8)	15.0 – 1490.0	2.1
C12 Carnitine	nM/L	5.0 (5.0, 10.0)	5.0 – 58.5	73.7	8.0 (2.5, 17.8)	2.5 – 81.0	32.3
C14:1 Carnitine	nM/L	399.5 (320.0, 519.0)	60.0 – 1443.0	0.3	410.5 (308.2, 557.5)	60.0 – 3433.0	9.1
C14 Carnitine	nM/L	125.0 (92.2, 163.0)	25.0 – 461.0	1.6	122.0 (98.0, 161.0)	25.0 – 489.0	1.8
C16:0H Carnitine	nM/L	20.0 (10.0, 29.0)	10.0 – 100.0	48.1	10.0 (10.0, 26.0)	10.0 – 63.0	52.2
C16 Carnitine	nM/L	309.0 (245.2, 396.0)	82.0 – 1027.0		271.0 (202.0, 350.8)	11.7.0 – 814.0	
C16OH Carnitine	nM/L	10.0 (10.0, 10.0)	10.0 – 65.0	89.9	10.0 (10.0, 10.0)	10.0 – 43.0	93.3
C18 Carnitine	nM/L	157.0 (116.0, 206.0)	25.0 – 514.0	2.8	140.0 (92.8, 200.0)	25.0 – 1354.0	6.2
C18OH Carnitine	nM/L	10.0 (10.0, 10.0)	10.0 – 100.0	82.1	Intraclass correlation < 0.4 Affected by storage conditions		85.6
C2 Carnitine	µM/L	7.0 (5.4, 9.3)	1.9 – 23.9				93.0
C3 Carnitine	nM/L	421.5 (323.0, 551.0)	101.0 – 5230.0		133.0 (85.1, 187.0)	25.0 – 798.0	7.9
C3DC Malonyl/3OHB	nM/L	137.0 (99.0, 216.8)	25.0 – 2927.0	1.4	131.0 (89.0, 206.8)	25.0 – 1416.0	2.9
C4 Carnitine	nM/L	212.0 (155.2, 282.0)	53.0 – 1394.0	0.1	Affected by storage conditions		2.9
C4DC Methylmalonyl/C5OH	nM/L	125.0 (100.0, 158.0)	50.0 – 707.0		166.5 (132.2, 222.0)	64.0 – 662.0	
C4DC Methylmaloyl	nM/L	Intraclass correlation < 0.4		66.9	13.0 (10.0, 26.0)	10.0 – 85.0	48.7
C5 Carnitine	nM/L	114.0 (89.0, 146.8)	25.0 – 1000.0	1.6	109.2 (83.2, 147.0)	25.0 – 1030.0	3.2
C5DC (Glutaryl) Carnitine [conf]	nM/L	105.0 (86.0, 133.0)	25.0 – 413.0	1.8	124.0 (95.2, 163.5)	25.0 – 482.0	1.2
C5DC (Glutaryl) Carnitine	nM/L	135.0 (109.0, 165.8)	25.0 – 489.0	0.2	163.5 (127.0, 230.4)	25.0 – 718.0	0.3
C5OH specific	nM/L	25.0 (25.0, 25.0)	25.0 – 140.0	76.1	25.0 (25.0, 58.0)	25.0 – 157.0	68.0
C6 Carnitine	nM/L	52.0 (36.0, 74.0)	11.0 – 276.0		22.0 (14.0, 37.0)	5.0 – 113.0	12.0
C6DC (methylglutaryl) Carnitine	nM/L	82.0 (58.0, 112.0)	25.0 – 662.0	17.2	92.0 (62.0, 135.0)	25.0 – 684.0	15.0
C8 Carnitine	nM/L	108.0 (70.0, 158.8)	5.0 – 958.0	0.1	115.0 (79.0, 177.8)	5.0 – 673.0	0.3
Carnosine	nM/L	Intraclass correlation < 0.4		83.5	Affected by storage conditions		1.2
Citrate [conf]	µM/L	Excessive missingness at random			Excessive missingness at random		1.8
Citrate	µM/L	Excessive missingness at random			Excessive missingness at random		0.3
Citrulline	µM/L	41.0 (34.1, 50.1)	8.3 – 108.3		43.8 (36.8, 51.6)	14.5 – 128.0	
Creatine	µM/L	20.5 (14.0, 32.3)	4.4 – 576.0		19.6 (14.1, 28.2)	7.1 – 110.7	
Cystathione [conf]	nM/L	130.0 (85.0, 202.8)	12.5 – 3589.0	9.2	Affected by storage conditions		32.6
Cystathione	nM/L	134.0 (88.0, 204.0)	7.5 – 3682.0	3.9	Affected by storage conditions		18.8
Dihydrothymine	µM/L	Excessive left-censoring		99.9	Excessive left-censoring		98.2
Dihydouracil	µM/L	Excessive left-censoring		99.9	Excessive left-censoring		98.2
Free Carnitine	µM/L	34.2 (28.9, 40.8)	9.3 – 77.1		Affected by storage conditions		14.4
Free Cysteine	µM/L	41.4 (33.3, 51.5)	12.9 – 113.6		Affected by storage conditions		97.4
Free homocystine	µM/L	Excessive left-censoring		99.0	Excessive left-censoring		98.2
Free homocysteine	µM/L	Excessive left-censoring		99.1	Excessive left-censoring		
Free Sialic acid	nM/L	793.0 (650.5, 1014.0)	325.0 – 6369.0		983.0 (789.2, 1300.8)	336.0 – 3362.0	
Galactose-1-P Phosphate [conf]	µM/L	Excessive missingness at random			Excessive missingness at random		
Galactose-1-Phosphate	µM/L	0.8 (0.4, 1.4)	0.1 – 9.4	11.1	0.7 (0.1, 1.6)	0.1 – 15.6	34.3
Glutamate	µM/L	61.6 (51.0, 73.2)	25.3 – 192.5		Affected by storage conditions		
Glutamate/Glutamine ratio	µM/L	0.1 (0.1, 0.1)	0.1 – 0.4		Affected by storage conditions		
Glycine	µM/L	562.4 (503.9, 631.0)	284.2 – 1239.0		Affected by storage conditions		
Glycine/Glycine/Valine	µM/L	281.4 (237.5, 342.1)	128.0 – 791.3		Affected by storage conditions		
Guamidinoacetic acid	µM/L	1.2 (0.9, 1.6)	0.3 – 7.7		312.1 (260.8, 365.5)	157.7 – 802.2	
Hexitol	µM/L	2.4 (1.9, 2.9)	0.8 – 7.2		1.3 (1.1, 1.7)	0.5 – 5.5	
Histidine	µM/L	10.4 (7.4, 16.2)	3.4 – 599.2		2.3 (1.8, 2.7)	0.8 – 6.0	
HMMA	µM/L	84.9 (77.0, 95.5)	37.7 – 341.8		13.8 (9.0, 21.8)	1.1 – 228.9	0.3
Homovanillic acid	nM/L	718.0 (476.0, 1032.5)	125.0 – 416201.0	4.5	81.3 (70.5, 91.1)	38.0 – 142.0	97.9
Hydroxyproline	µM/L	12.2 (8.9, 16.8)	3.4 – 69.8		Excessive left-censoring		1.2
Hypoxanthine	µM/L	7.8 (6.1, 10.2)	0.5 – 45.2	0.2	1057.0 (643.5, 1562.0)	125.0 – 4552.0	0.5 – 24.0

ESM Table 2: Summary characteristics of all biomarkers measured in the study (continued)

Biomarker	Units	SDRNtBIO			FinDiane		
		Median (IQR)	Range	Below	Median (IQR)	Range	Below
Inosine	nM/L	1137.0 (530.2, 2711.1)	25.0 – 15287.0	0.3	1339.0 (587.0, 3068.0)	25.0 – 15287.0	3.5
Isoleucine	nM/L	72.2 (57.9, 90.5)	21.1 – 247.6	0.4	84.8 (70.5, 100.7)	32.8 – 177.9	
Kynurenic acid	nM/L	61.0 (48.0, 78.0)	10.0 – 262.0	0.4	82.5 (62.0, 109.0)	28.0 – 404.0	
Kynurene	nM/L	2.9 (2.4, 3.6)	1.3 – 8.7	0.3 (2.4, 3.7)	1.0 – 9.5		
Leucine/Alanine ratio		0.3 (0.2, 0.3)	0.1 – 1.1	0.3 (0.3, 0.4)	0.2 – 0.7		
Leucine	μM/L	125.5 (102.0, 154.6)	34.1 – 174.9	151.8 (126.4, 175.9)	61.3 – 306.9		
Lysine	μM/L	197.3 (170.4, 223.0)	88.6 – 474.3	201.3 (180.9, 230.8)	99.3 – 368.7		
Malate	nM/L	26.5 (22.2, 31.5)	5.8 – 71.0	55.1	134.8 (50.0, 202.5)	50.0 – 1025.0	32.3
Methionine	μM/L	1211.5 (723.2, 2010.5)	125.0 – 23028.0	5.0	Affected by storage conditions	0.3	
Methylcitrate	nM/L	304.0 (234.0, 415.0)	25.0 – 76024.0	0.2	291.5 (225.0, 373.0)	25.0 – 1465.0	2.3
Methylmalonic acid	nM/L	1045.5 (869.5, 1258.0)	432.0 – 1947.0	620.5 (524.0, 757.0)	190.0 – 1554.0	0.6	
N-acetyl-aspartate [conf]	nM/L	679.0 (574.0, 808.8)	310.0 – 1757.0	576.0 (485.0, 750.0)	125.0 – 1554.0	0.9	
N-acetyl-aspartate [conf]	nM/L	Excessive missingness at random		446.5 (175.0, 583.2)	175.0 – 1146.0	29.0	
N-acetylglutamine [conf]	nM/L	392.5 (175.0, 516.5)	175.0 – 978.0	43.5	Excessive missingness at random		
N-acetylglutamine	μmol/L/min	30.0 (25.3, 35.5)	12.3 – 71.2	25.0 (21.2, 30.9)	11.0 – 64.8		
NAG	nM/L	7.5 (7.5, 7.5)	7.5 – 32.0	91.2	7.5 (7.5, 7.5)	7.5 – 53.5	76.0
Neopterin	nM/L	Excessive left-censoring		99.9	Excessive left-censoring		98.2
Nitrotyrosine	μM/L	100.8 (86.6, 121.8)	44.4 – 272.3	91.5 (79.8, 107.0)	36.1 – 183.4		
Ornithine	nM/L	Intraclass correlation < 0.4			Intraclass correlation < 0.4		
Orotic	nM/L	75.8 (67.7, 85.5)	34.1 – 222.5	82.4 (71.5, 93.0)	34.4 – 144.8		
Phenylalanine	μM/L	1.0 (0.9, 1.2)	0.5 – 3.2	1.2 (1.0, 1.4)	0.6 – 2.8		
Phenylalanine/Tyrosine Phosphotransferase	μM/L	Excessive left-censoring		99.9	Excessive left-censoring		98.5
Phosphoserine [conf]	nM/L	Excessive missingness at random		99.9	Excessive missingness at random		
Phosphoserine	μM/L	7.3 (5.5, 11.1)	1.3 – 25.8	99.9	Intraclass correlation < 0.4		0.6
Proline	μM/L	212.9 (173.9, 257.7)	79.5 – 778.5	97.4	Affected by storage conditions		
Pyridoxal 5prime-phosphate	nM/L	Excessive left-censoring		97.4	Excessive left-censoring		
Pyridoxal 5prime-phosphate	nM/L	Excessive left-censoring		97.4	Excessive left-censoring		
Quinolonic acid	nM/L	25.0 (25.0, 25.0)	25.0 – 98.5	97.4	Affected by storage conditions		
Quinolonic acid	nM/L	Intraclass correlation < 0.4		97.4	1437.0 (1107.2, 1959.5)	410.0 – 10322.0	
Sarcosine	nM/L	6.1 (2.5, 8.4)	2.5 – 69.5	42.1	5.7 (2.5, 7.7)	2.5 – 46.1	38.4
SDMA/ADMA ratio	nM/L	1.2 (1.1, 1.4)	0.7 – 3.7	686.0 (563.0, 842.0)	1.4 (1.2, 1.7)	0.8 – 3.7	
SDMA	nM/L	546.0 (478.0, 651.8)	293.0 – 1881.0	181.0 (152.0, 201.0)	356.0 – 1766.0		
Serine	μM/L	142.1 (123.5, 165.0)	79.0 – 290.3	79.0 – 290.3	81.0 – 359.0		
Succinate	nM/L	17.4 (14.6, 20.8)	5.6 – 63.0	99.9	Intraclass correlation < 0.4		
Succinylacetone	nM/L	Excessive left-censoring		0.3	Affected by storage conditions		
Sulphocysteine	nM/L	716.5 (590.0, 868.0)	100.0 – 2539.0	99.9	Excessive left-censoring		
Taurine	μM/L	111.7 (86.0, 143.1)	32.4 – 511.1	99.9	Intraclass correlation < 0.4		
Theanine	nM/L	118.4 (95.7, 140.5)	30.2 – 562.8	0.3	Affected by storage conditions		
Thymine	nM/L	Excessive missingness at random		0.3	18.5 (14.9, 23.7)	4.7 – 52.6	
TMAO	μM/L	5.0 (3.0, 8.1)	0.3 – 67.4	36.4	6.7 (4.5, 10.1)	1.7 – 156.8	
Tryptophan	μM/L	58.9 (50.8, 67.4)	28.4 – 137.2	6.7 (4.5, 10.1)			
Tryptophan/Kynurenic acid ratio	nM/L	20.4 (16.2, 24.6)	5.3 – 48.1	30.0 – 204.8	18.5 (14.9, 23.7)	4.7 – 52.6	
Tryptophan/Kynurenic acid ratio	nM/L	75.1 (63.1, 90.5)	30.2 – 204.8	69.0 (57.5, 81.2)	37.6 – 137.3		
Tyrosine	μM/L	970.0 (768.2, 1206.0)	264.0 – 2677.0	99.9	Intraclass correlation < 0.4		1.2
Uracil	nM/L	Excessive left-censoring		99.9	Excessive left-censoring		98.5
Ureidopropionate	nM/L	363.0 (295.0, 459.0)	79.0 – 858.0	231.0 (188.0, 279.0)	15.0 – 601.0		
Uric acid	μM/L	232.1 (197.7, 268.6)	69.5 – 596.5	232.8 (201.0, 272.2)	101.9 – 436.6		
Valine	nM/L	2.0 (1.7, 2.5)	0.2 – 14.2	0.2	1.9 (1.5, 2.4)	0.2 – 12.6	0.3
Xanthine	nM/L	25.0 (25.0, 119.0)	25.0 – 579.0	50.9	77.0 (25.0, 128.4)	25.0 – 353.0	28.7
LC-MSMS tryptic peptides							
ACTIN (758.9 / 765.4)		*					
Intraclass correlation < 0.4		0.8					

ESM Table 2: Summary characteristics of all biomarkers measured in the study (continued)

Biomarker	Units	SDRNtBIO			FinDiane		
		Median (IQR)	Range	Below	Median (IQR)	Range	Below
Acyloxyacyl hydrolase	(703.9 / 931.3)	*	193.8 (151.5, 245.2)	5.0 – 633.6	0.5	194.6 (123.7, 269.7)	19.2 – 735.6
Afamin	(416.8 / 572.4)	*	Intraclass correlation < 0.4	18.2	Intraclass correlation < 0.4	0.3	38.4
Afamin	(563.8 / 825.4)	*	Intraclass correlation < 0.4	17.6	Intraclass correlation < 0.4	0.3	23.8
AGR2 (lung cancer biomarker)	(407.7 / 701.4)	*	Intraclass correlation < 0.4	1.0	Intraclass correlation < 0.4	0.3	1.0
Albumin T31	(337.3 / 416.3)	*	Intraclass correlation < 0.4	0.4	1897.4 (1677.8, 2174.7)	417.0 – 3948.7	
Albumin T34	(441.0 / 680.5)	*	1192.0 (1051.5, 1353.5)	5.0 – 2520.9	1016.5 (898.7, 1860.2)	290.9 – 1860.2	
Albumin T6	(575.4 / 937.4)	*	27296.9 (24671.0, 30189.0)	5.0 – 47576.5	23164.3 (20356.6, 26048.2)	9566.2 – 47832.8	
Albumin T70	(501.2 / 587.5)	*	4303.3 (3687.7, 4970.2)	5.0 – 10234.8	3181.2 (2453.8, 3845.9)	344.5 – 7405.8	
Alpha-1-acid-glycoprotein 1	(723.3 / 937.4)	*	106.4 (85.9, 131.9)	5.0 – 276.8	90.2 (75.5, 115.7)	29.0 – 234.3	
Alpha-1-antichymotrypsin isol.	(608.4 / 775.4)	*	236.7 (192.7, 286.9)	5.0 – 980.6	204.4 (172.4, 241.5)	83.5 – 981.2	
Alpha-1-antitrypsin	(444.8 / 718.4)	*	78.0 (61.9, 100.0)	5.0 – 205.0	74.5 (58.2, 92.3)	18.4 – 201.1	
Alpha-1-antitrypsin	(536.4 / 797.7)	*	45.2 (34.2, 58.3)	5.0 – 205.5	39.9 (30.0, 53.3)	5.0 – 114.9	
Alpha-1-antitrypsin	(631.3 / 889.5)	*	62.5 (44.5, 84.1)	5.0 – 193.1	47.2 (29.9, 63.8)	5.0 – 129.2	
Alpha-1B-glycoprotein	(619.3 / 894.5)	*	84.1 (68.2, 104.0)	5.0 – 197.2	78.3 (62.4, 95.6)	5.0 – 173.0	
Alpha-1B-glycoprotein	(687.4 / 960.6)	*	Intraclass correlation < 0.4	0.4	Intraclass correlation < 0.4	0.3	
Alpha-2-macroglobulin	(697.8 / 737.4)	*	131.5 (99.2, 164.4)	5.0 – 350.6	164.5 (127.3, 211.5)	2.9	
Alpha-fetoprotein	(490.7 / 833.3)	*	47.0 (37.6, 57.2)	5.0 – 120.6	36.4 (28.8, 44.6)	54.9 – 460.3	
Aminopeptidase B	(547.8 / 680.4)	*	Intraclass correlation < 0.4	2.6	5.0 – 95.2	3.5	
Angiotensin II	(349.8 / 136.1)	*	1104.0 (967.5, 1369.5)	5.0 – 2393.0	0.4	0.3	
Angiotensinogen	(634.9 / 956.6)	*	34.6 (25.7, 45.1)	5.0 – 203.9	6.8	970.5 (830.0, 1168.4)	
ApoA1	(700.8 / 1023.5)	*	419.0 (337.9, 510.4)	5.0 – 907.8	0.4	317.6 (249.3, 399.0)	
ApoA-II-pre	(486.8 / 546.4)	*	80.2 (65.2, 97.3)	5.0 – 196.6	0.4	44.8 – 755.4	
ApoA-Q1	(626.8 / 422.2)	*	127.9 (104.5, 156.8)	5.0 – 315.3	0.4	109.1 (87.8, 133.7)	
ApoB-100	(655.1 / 975.4)	*	50.6 (38.2, 68.7)	5.0 – 294.9	2.2	109.1 (87.8, 133.7)	
ApoB-Q1	(640.8 / 838.4)	*	93.6 (73.3, 117.2)	5.0 – 289.3	0.5	94.4 (70.6, 116.8)	
ApoC-I	(516.8 / 466.2)	*	135.9 (102.3, 178.8)	5.0 – 906.0	0.4	94.5 (67.7, 140.2)	
ApoC-II	(745.1 / 1149.7)	*	Intraclass correlation < 0.4	63.3	Intraclass correlation < 0.4	15.7 – 533.9	
ApoC-III	(598.8 / 854.4)	*	297.4 (231.7, 381.6)	5.0 – 1258.3	0.4	60.1	
ApoE-Q	(484.8 / 588.3)	*	72.1 (58.4, 91.5)	5.0 – 250.1	0.6	220.7 (170.9, 284.5)	
Apoll	(637.8 / 932.5)	*	24.5 (5.0, 36.3)	5.0 – 90.7	26.8	48.2 – 1034.2	
Beta-2-glycoprotein I	(511.8 / 751.4)	*	46.4 (37.0, 58.3)	5.0 – 132.5	1.7	50.0 – 430.0	
Beta-2-macroglobulin	(575.0 / 920.3)	*	847.0 (569.5, 1162.8)	5.0 – 2247.0	0.4	94.4 (70.6, 116.8)	
Bone morphogenetic protein 5	(723.5 / 577.3)	*	204.5 (174.8, 241.9)	5.0 – 618.1	0.4	5.0 – 242.2	
BPI fold-containing family A member 2	(481.3 / 500.4)	*	2213.5 (1982.9, 2475.4)	5.0 – 4531.0	0.4	0.3	
ClQTNF1	(547.7 / 707.4)	*	Intraclass correlation < 0.4	0.5	162.7 (110.7, 196.1)	24.0 – 411.8	
Caeruloplasmin	(602.3 / 695.3)	*	29.8 (21.4, 41.4)	5.0 – 99.8	9.5	9.7	
Caeruloplasmin	(760.1 / 1059.8)	*	Intraclass correlation < 0.4	8.1	Intraclass correlation < 0.4	6.2	
Carboxypeptidase A4	(492.9 / 547.7)	*	72.8 (54.0, 92.5)	5.0 – 250.2	0.6	119.9 (88.0, 166.9)	
Carboxypeptidase A4	(493.5 / 510.3)	*	168.3 (135.5, 210.3)	5.0 – 558.0	0.4	5.0 – 343.2	
Carboxypeptidase M	(501.8 / 874.5)	*	49.0 (39.5, 58.4)	5.0 – 101.2	0.9	2.1	
CART std	(576.2 / 705.3)	*	63.7 (49.9, 77.5)	5.0 – 151.3	0.6	52.4 (39.2, 64.8)	
Cathepsin S	(359.2 / 717.4)	*	33.3 (23.9, 49.2)	5.0 – 135.0	5.5	Intraclass correlation < 0.4	
C-C motif chemokine 14	(476.2 / 666.3)	*	267.1 (215.2, 326.4)	5.0 – 815.5	0.4	150.0	
Cellular repressor of E1A-stimulated genes 1	(575.8 / 704.4)	*	186.9 (164.3, 211.4)	5.0 – 341.3	0.4	159.8 (141.1, 180.4)	
Chromogranin A	(488.2 / 775.4)	*	36.9 (25.6, 46.9)	5.0 – 123.5	10.7	69.4 – 278.5	
Chromogranin B	(496.2 / 764.3)	*	55.4 (42.1, 69.9)	5.0 – 215.9	2.2	34.9 (25.1, 48.8)	
Chromogranin B	(579.3 / 815.4)	*	70.1 (55.4, 85.0)	5.0 – 169.3	0.6	50.5 (40.0, 63.2)	
Chromogranin B	(1073.5 / 822.4)	*	Intraclass correlation < 0.4	0.8	Intraclass correlation < 0.4	0.3	
Clusterin	(697.5 / 922.4)	*	74.2 (59.2, 91.8)	5.0 – 287.0	0.5	6.2	
Clusterin isoform I	(559.3 / 903.5)	*	73.2 (58.5, 89.7)	5.0 – 188.5	17.6	Intraclass correlation < 0.4	
Coagulation factor IX pre	(626.3 / 792.4)	*	Intraclass correlation < 0.4	5.0 – 778.0	5.0	19.3 – 162.0	
Coagulation factor V	(555.8 / 895.8)	*	329.0 (215.0, 432.5)	5.0 – 297.9 (209.0, 359.8)	5.0	5.0 – 675.1	

ESM Table 2: Summary characteristics of all biomarkers measured in the study (continued)

Biomarker	Units	SDRNtBIO			FinDiane		
			Median (IQR)	Range	Below	Median (IQR)	Range
Coagulation factor V (659.0 / 925.5)	*	Intraclass correlation < 0.4	63.5	Affected by storage conditions	68.0		
Coagulation factor XIIa HC (442.3 / 685.4)	*	123.2 (95.6, 154.9)	5.0 – 270.8	0.4	108.8 (83.6, 133.1)	5.0 – 278.9	0.3
Complement C1q subcomponent subunit A (420.0 / 244.0)	*	224.6 (178.2, 271.4)	5.0 – 782.7	0.4	203.1 (160.3, 253.4)	63.5 – 547.0	0.3
Complement C3 (501.8 / 731.4)	*	64.4 (52.8, 78.2)	5.0 – 154.1	0.8	Intraclass correlation < 0.4		
Complement C3 (673.4 / 646.4)	*	130.5 (100.1, 155.2)	5.0 – 295.8	0.4	113.0 (89.9, 135.4)	36.8 – 241.7	0.3
Complement C4 beta (557.8 / 629.4)	*	109.4 (88.0, 138.2)	5.0 – 276.5	0.4	100.3 (84.9, 206.0)	29.8 – 218.0	
Complement C6 (612.8 / 758.4)	*	30.8 (50.4, 47.5)	5.0 – 105.2	32.6	Intraclass correlation < 0.4	22.0	
Complement C7 (550.7 / 577.3)	*	Intraclass correlation < 0.4	76.2	Intraclass correlation < 0.4	58.4		
Complement C8 alpha (501.7 / 726.3)	*	Intraclass correlation < 0.4	13.4	Intraclass correlation < 0.4	11.7		
Complement C8 beta (694.9 / 942.6)	*	453.0 (360.8, 541.4)	5.0 – 1128.7	0.4	303.0 (249.7, 380.3)	5.0 – 106.7	
Complement C8 delta (809.9 / 1091.5)	*	59.2 (38.7, 85.5)	5.0 – 228.9	7.0	349.5 (23.4, 73.5)	131.4 – 809.6	
Complement C8 gamma (810.9 / 836.4)	*	Intraclass correlation < 0.4	43.4	Intraclass correlation < 0.4	5.0 – 190.1		
Complement factor B (578.3 / 671.4)	*	235.1 (187.8, 292.0)	5.0 – 610.8	0.4	219.8 (173.8, 292.2)	74.8 – 557.0	
Complement Factor I (1007.5 / 1023.5)	*	171.5 (95.0, 386.5)	5.0 – 741.5	0.5	Intraclass correlation < 0.4	0.3	
Corticosteroid-binding globulin (539.1 / 517.3)	*	Intraclass correlation < 0.4	1.0	Intraclass correlation < 0.4	0.3		
Corticotropin-releasing factor-binding protein (617.8 / 674.3)	*	Intraclass correlation < 0.4	0.8	Intraclass correlation < 0.4	0.3		
C-reactive protein (564.8 / 609.4)	*	93.0 (74.9, 116.1)	5.0 – 234.1	0.8	Intraclass correlation < 0.4	1.8	
C-reactive protein (569.7 / 829.3)	*	Intraclass correlation < 0.4	28.1	Intraclass correlation < 0.4	24.9		
C-reactive protein iso1 (696.9 / 1035.6)	*	Intraclass correlation < 0.4	19.8	Intraclass correlation < 0.4	14.1		
Cystatin C (685.0 / 412.1)	*	Intraclass correlation < 0.4	0.6	Intraclass correlation < 0.4	0.3		
E-selectin (613.8 / 982.5)	*	143.4 (117.5, 178.3)	5.0 – 385.5	0.4	127.0 (102.7, 154.0)	58.3 – 381.4	0.9
Extracellular glycoprotein lacitin (481.3 / 501.3)	*	204.0 (1845.5, 2283.0)	5.0 – 3990.0	0.4	1797.4 (1562.7, 2034.6)	400.9 – 3403.0	
Fas (TNFRSF6) associated factor 1 (667.8 / 786.3)	*	340.0 (242.1, 451.3)	5.0 – 1898.5	1.4	306.5 (227.3, 405.6)	5.0 – 1925.4	1.2
Ferritin HC (673.3 / 1035.5)	*	Intraclass correlation < 0.4	0.4	Intraclass correlation < 0.4	14.1		
Fetuin-B (498.4 / 467.3)	*	Intraclass correlation < 0.4	27.5	Intraclass correlation < 0.4	19.4		
Fibronectin isol (536.3 / 680.4)	*	66.4 (45.0, 101.5)	5.0 – 980.4	1.2	39.8 (24.6, 63.5)	5.0 – 408.3	6.2
Fractalkine (612.8 / 977.5)	*	49.5 (34.9, 67.1)	2.5 – 157.2	8.1	44.7 (31.0, 59.7)	5.0 – 168.0	7.9
Haptoglobin (490.5 / 562.6)	*	157.1 (110.6, 216.2)	5.0 – 711.9	0.5	138.9 (95.8, 190.7)	25.8 – 590.8	
Haptoglobin (559.4 / 658.4)	*	145.8 (110.7, 184.4)	5.0 – 598.0	0.9	120.1 (78.5, 170.0)	5.0 – 537.6	1.2
Haptoglobin (602.8 / 803.7)	*	123.4 (94.7, 161.4)	5.0 – 514.9	1.2	105.7 (80.7, 135.0)	11.6 – 372.9	
Haptoglobin beta chain (490.8 / 562.3)	*	Intraclass correlation < 0.4	0.5	789.3 (537.7, 1034.9)	114.5 – 3028.0		
Heat shock protein 60kDa (456.8 / 515.4)	*	72.6 (56.3, 94.2)	5.0 – 209.5	1.1	65.4 (53.1, 80.5)	5.0 – 178.3	1.2
Heat shock protein 90alpha isol (675.4 / 921.5)	*	5.0 (5.0, 5.0)	5.0 – 60.2	73.4	Intraclass correlation < 0.4	72.7	
Hemopexin (571.5 / 650.4)	*	204.0 (168.0, 252.0)	5.0 – 505.0	0.4	176.3 (146.6, 212.2)	83.9 – 305.2	
Heparin cofactor II (514.8 / 814.4)	*	929.0 (808.5, 1054.5)	5.0 – 1913.0	0.4	802.0 (651.7, 933.8)	354.0 – 1559.6	
Hyaluronan-binding protein 2 (575.2 / 901.5)	*	81.1 (67.0, 98.0)	5.0 – 191.5	0.4	76.0 (61.4, 90.2)	33.9 – 203.4	
Ig gamma-1 chain C region (594.0 / 698.7)	*	197.0 (169.7, 226.7)	5.0 – 335.4	0.4	172.2 (145.4, 195.1)	69.7 – 321.7	
Immunoglobulin J chain (695.5 / 971.4)	*	9.5 (65.2, 157.2)	5.0 – 355.5	0.4	Intraclass correlation < 0.4		
Immunoglobulin lambda-like polypeptide 5 (421.3 / 429.2)	*	40.7 (25.4, 58.6)	5.0 – 141.1	16.4	40.7 (25.4, 58.6)	5.0 – 134.8	8.2
Inhibin beta B chain (594.8 / 419.3)	*	97.5 (79.4, 117.5)	5.0 – 239.0	0.4	91.1 (74.0, 110.4)	5.0 – 221.6	0.3
Insulin-like growth factor-binding protein2 (530.6 / 785.4)	*	159.6 (130.3, 202.4)	5.0 – 625.4	0.4	129.2 (96.2, 163.6)	31.4 – 1012.4	
Inter-alpha-trypsin inhibitor HC (579.3 / 902.5)	*	20.4 (13.8, 27.2)	5.0 – 77.1	15.6	Intraclass correlation < 0.4	16.4	
Interlenkin-26 (575.2 / 512.3)	*	143.3 (119.6, 166.2)	5.0 – 275.1	0.4	111.7 (90.1, 134.8)	44.1 – 231.5	
Kallistatin (643.4 / 971.6)	*	104.2 (88.1, 122.2)	5.0 – 224.1	0.4	85.7 (73.2, 103.4)	42.4 – 189.0	
Kininogen-1 (626.3 / 173.1)	*	340.2 (289.4, 407.1)	5.0 – 857.7	0.4	Affected by storage conditions	48.4	
Leucine-rich alpha-2-glycoprotein (590.5 / 725.4)	*	Intraclass correlation < 0.4	0.4	Intraclass correlation < 0.4			
MPO (575.3 / 939.4)	*	141.6 (115.9, 173.6)	5.0 (5.0, 34.6)	0.4	128.1 (102.5, 159.6)	43.7 – 2840.0	33.4
Osteopontin isoA (694.3 / 853.4)	*	241.2 (191.0, 286.7)	5.0 – 556.3	0.4	Intraclass correlation < 0.4		
Pericentriolar material 1 (536.3 / 825.5)	*	49.7 (39.0, 64.4)	5.0 – 167.2	2.1	48.2 (37.0, 61.1)	5.0 – 117.3	2.1
Peroxidases (492.6 / 703.3)	*	30.3 (21.3, 42.9)	5.0 – 93.5	9.8	26.8 (18.0, 38.1)	5.0 – 100.3	16.1
Peroxiredoxin 2 (489.8 / 735.4)	*	140.0 (107.2, 186.0)	5.0 – 617.5	0.6	115.8 (86.0, 160.1)	5.0 – 344.8	0.3

ESM Table 2: Summary characteristics of all biomarkers measured in the study (continued)

Biomarker	Units	SDRNtBIO			FinDiane			
		Median (IQR)	Range	Below	Median (IQR)	Range	Below	
Phosphatidylcholine-sterol acyltransferase	(693.7 / 941.5)	*	28.3 (17.5, 44.4)	5.0 – 291.7	18.6	26.7 (12.6, 53.3)	5.0 – 146.5	20.5
Plasma glutamate carboxypeptidase	(577.3 / 743.3)	*	Intraclass correlation < 0.4	48.4	Intraclass correlation < 0.4	41.9		
Plasma protease C1 inhibitor	(633.0 / 1049.6)	*	56.5 (44.9, 71.5)	5.0 – 288.6	1.0	Intraclass correlation < 0.4	0.3	
Plasminogen	(438.3 / 502.3)	*	28.6 (20.7, 36.4)	5.0 – 85.2	11.5	Intraclass correlation < 0.4	8.2	
Plasminogen	(438.3 / 615.3)	*	31.0 (261.5, 379.5)	5.0 – 2073.0	0.4	294.1 (247.7, 347.9)	118.7 – 1190.0	
Plasminogen	(570.8 / 691.4)	*	92.0 (75.0, 111.0)	5.0 – 225.1	0.5	84.6 (72.6, 103.9)	32.9 – 195.2	
Polymerase (RNA) II polypeptide D	(425.2 / 635.3)	*	36.9 (26.6, 48.0)	5.0 – 137.0	5.1	33.0 (22.9, 45.3)	5.0 – 131.6	7.0
PON1 Serum paraoxonase/arylesterase 2	(942.5 / 472.3)	*	234.0 (174.0, 317.0)	5.0 – 869.0	0.4	250.0 (174.6, 372.9)	52.7 – 666.8	2.1
Prothrombin	(626.3 / 679.4)	*	45.4 (34.9, 57.6)	5.0 – 117.2	1.9	Intraclass correlation < 0.4	2.6	
Prothrombin	(626.3 / 879.5)	*	39.0 (30.0, 50.2)	5.0 – 108.7	3.6	Intraclass correlation < 0.4	2.6	
P-selectin	(500.2 / 492.3)	*	18.1 (5.0, 33.2)	5.0 – 456.5	31.9	17.4 (5.0, 134.6)	5.0 – 366.2	32.8
P-selectin	(712.4 / 876.5)	*	Intraclass correlation < 0.4	15.9	Intraclass correlation < 0.4	7.9		
RBP1	(367.2 / 505.3)	*	415.6 (3666.6, 4584.2)	5.0 – 6884.7	55.2	5.0 (5.0, 10.6)	5.0 – 30.9	70.4
Retinal binding protein 2	(575.8 / 695.3)	*	Intraclass correlation < 0.4	0.4	3571.3 (3086.9, 3986.7)	1725.1 – 6580.2		
RBP4	(481.2 / 833.4)	*	Intraclass correlation < 0.4	0.4	136.9 (101.4, 183.0)	29.1 – 333.8		
RBP	(583.3 / 669.5)	*	59.3 (44.2, 78.6)	5.0 – 219.6	0.9	67.4 (53.2, 91.8)	12.6 – 230.4	
Renin isoform1	(520.8 / 502.3)	*	86.1 (59.4, 120.2)	5.0 – 321.5	0.6	67.2 (34.5, 105.1)	5.0 – 224.7	0.9
Renin isoform1	(610.3 / 959.4)	*	41.7 (24.3, 98.0)	5.0 – 474.3	7.1	Intraclass correlation < 0.4	5.3	
Reticulon-4 isoform1	(618.8 / 894.5)	*	13.6 (5.0, 21.5)	2.5 – 86.9	34.5	Intraclass correlation < 0.4	30.5	
sCD40L	(471.2 / 837.4)	*	122.8 (97.7, 152.2)	5.0 – 351.5	0.4	Intraclass correlation < 0.4		
Serotransferrin	(489.9 / 735.4)	*	164.0 (129.8, 215.1)	5.0 – 602.5	0.4	133.3 (100.5, 174.7)	47.2 – 366.0	
Serotransferrin	(625.5 / 675.4)	*	330.4 (255.4, 419.8)	5.0 – 866.0	0.5	265.6 (210.1, 335.8)	72.6 – 696.5	
Serum Amyloid A	(739.4 / 1151.6)	*	Intraclass correlation < 0.4	21.1	Intraclass correlation < 0.4	21.7		
Serum amyloid P-component	(578.8 / 508.3)	*	Intraclass correlation < 0.4	0.9	Intraclass correlation < 0.4	0.3		
Somatostatin receptor type 2-P30874	(575.8 / 674.4)	*	97.0 (81.4, 114.5)	5.0 – 207.9	0.4	81.5 (68.7, 98.2)	31.5 – 154.5	
TAO kinase 1	(645.3 / 804.4)	*	78.8 (58.8, 98.2)	5.0 – 200.8	0.6	60.9 (48.6, 77.8)	5.0 – 160.5	0.6
Thyroxine binding globulin	(550.8 / 244.1)	*	51.0 (28.1, 114.4)	5.0 – 527.5	7.9	Intraclass correlation < 0.4	6.7	
Thyroxine-binding globulin	(801.1 / 993.9)	*	77.2 (56.3, 99.5)	5.0 – 210.8	1.4	Intraclass correlation < 0.4	34.9	
TNF alpha	(691.9 / 1067.7)	*	67.1 (54.6, 83.2)	5.0 – 204.1	1.2	Intraclass correlation < 0.4	0.9	
TNFRSF9	(409.7 / 519.3)	*	572.8 (421.8, 715.1)	5.0 – 489.0	0.4	508.7 (350.3, 820.6)	101.4 – 3230.4	
Transferrin	(489.7 / 735.4)	*	138.9 (106.1, 179.4)	5.0 – 619.0	0.5	109.9 (84.7, 157.2)	29.5 – 409.3	
Transferrin	(598.3 / 993.5)	*	72.3 (56.2, 88.8)	5.0 – 180.4	1.3	58.5 (46.2, 73.8)	5.0 – 183.7	0.9
Transferrin	(739.9 / 1184.6)	*	Intraclass correlation < 0.4	0.4	271.4 (168.1, 361.2)	37.1 – 832.9		
Triglyceride	(581.8 / 935.4)	*	Intraclass correlation < 0.4	0.8	91.5 (70.8, 113.7)	25.3 – 251.3		
Ubiquinol-cytochrome c reductase core protein II	(547.3 / 639.3)	*	Intraclass correlation < 0.4	0.4	1069.0 (658.1, 1377.4)	162.5 – 2569.7		
Vitamin K-dependent protein C	(516.3 / 603.3)	*	62.3 (49.9, 74.9)	5.0 – 167.3	0.8	Intraclass correlation < 0.4	30.8	
Vit D binding	(628.2 / 681.3)	*	70.0 (52.9, 87.8)	5.0 – 209.0	1.2	Intraclass correlation < 0.4	53.1	
Vitronectin	(711.8 / 647.3)	*	98.7 (82.6, 119.5)	5.0 – 320.4	0.4	86.9 (74.7, 104.5)	38.2 – 200.5	
Von Willebrand factor	(500.8 / 788.4)	*	57.8 (46.4, 70.0)	5.0 – 120.9	1.5	67.9 (57.1, 81.2)	24.6 – 138.5	
WD repeat domain 67-isol1	(487.8 / 604.3)	*	44.4 (33.9, 55.6)	5.0 – 130.0	4.7	44.4 (33.3, 55.5)	5.0 – 119.6	3.2
Zinc-alpha-2-glycoprotein	(697.8 / 789.4)	*	51.4 (34.0, 68.3)	5.0 – 172.6	11.5	Zinc-alpha-2-glycoprotein	8.5	

Note:

Biomarkers that were not analyzed report the reason for their exclusion from the analysis

An * indicates that the value is a ratio of the biomarker to a stable isotope of albumin T6

Metabolites with “[conf]” in their name represent confidence transitions

“Below” reports the percentage of observations with levels below the detection threshold

ESM Table 3: Associations of each biomarker (considered separately) with achieved eGFR from linear regression models adjusted for age, sex, duration of diabetes, study day eGFR, length of follow-up and ACR category

Biomarker	SDRNT1BIO		FinnDiane	
	Coefficient (95% CI)	p	Coefficient (95% CI)	p
Luminex proteins				
CD27 antigen	-0.22 (-0.27, -0.17)	2E-16	-0.34 (-0.46, -0.23)	9E-09
Kidney Injury Molecule-1	-0.16 (-0.21, -0.11)	6E-10	-0.27 (-0.37, -0.18)	6E-08
Alpha-1-Microglobulin	-0.18 (-0.24, -0.13)	2E-11	-0.26 (-0.36, -0.16)	4E-07
Beta-2-Microglobulin	-0.22 (-0.27, -0.17)	1E-15	-0.26 (-0.37, -0.14)	1E-05
Cystatin-C	-0.23 (-0.29, -0.18)	3E-15	-0.20 (-0.31, -0.09)	3E-04
Thrombomodulin	-0.19 (-0.24, -0.14)	3E-12	-0.22 (-0.32, -0.11)	5E-05
Tumor Necrosis Factor Receptor 1	-0.17 (-0.22, -0.12)	1E-10	-0.20 (-0.30, -0.09)	3E-04
Osteopontin	-0.12 (-0.17, -0.07)	1E-06	-0.18 (-0.28, -0.08)	7E-04
Interleukin-2 receptor alpha	-0.16 (-0.20, -0.11)	2E-10	-0.11 (-0.21, -0.01)	3E-02
Fibroblast Growth Factor 21	-0.11 (-0.15, -0.06)	5E-06	-0.15 (-0.25, -0.06)	1E-03
N-terminal prohormone of brain natriuretic peptide	-0.12 (-0.17, -0.07)	6E-06	0.00 (-0.10, 0.09)	9E-01
Tamm-Horsfall Urinary Glycoprotein	0.11 (0.06, 0.15)	3E-05	0.09 (-0.01, 0.18)	8E-02
Trefoil Factor 3	-0.11 (-0.16, -0.06)	2E-05	<i>Not tested</i>	<i>Not tested</i>
LC-MS/MS metabolites				
Free Sialic acid	-0.20 (-0.26, -0.15)	2E-12	-0.21 (-0.33, -0.08)	1E-03
SDMA	-0.15 (-0.21, -0.09)	2E-07	-0.21 (-0.33, -0.09)	6E-04
Tryptophan/Kynurenine	0.17 (0.12, 0.22)	3E-10	0.08 (-0.03, 0.18)	2E-01
C5DC (Glutaryl) Carnitine	-0.11 (-0.16, -0.06)	4E-05	-0.07 (-0.18, 0.04)	2E-01
Free Cystine	-0.11 (-0.16, -0.05)	7E-05	<i>Not tested</i>	<i>Not tested</i>
Tryptophan	0.11 (0.07, 0.16)	2E-06	<i>Not tested</i>	<i>Not tested</i>
Methionine	0.10 (0.05, 0.14)	3E-05	<i>Not tested</i>	<i>Not tested</i>
LC-MS/MS tryptic peptides				
Albumin T70 (501.2 / 587.5)	0.10 (0.05, 0.14)	3E-05	-0.01 (-0.10, 0.08)	8E-01
Chromogranin A (488.2 / 775.4)	-0.10 (-0.14, -0.05)	3E-05	0.00 (-0.09, 0.09)	1E+00

Regression coefficients are per unit of standard deviation of gaussianised biomarker

We report median and interquartile range (IQR) for continuous variables, and frequency for categorical variables

ESM Table 4: Associations of each biomarker (considered separately) with achieved eGFR from linear regression models adjusted for full clinical covariates

	Biomarker	SDRNT1BIO		FinnDiane	
		Coefficient (95% CI)	p	Coefficient (95% CI)	p
Luminex proteins					
CD27 antigen		-0.17 (-0.22, -0.12)	9E-12	-0.32 (-0.43, -0.21)	4E-08
Beta-2-Microglobulin		-0.16 (-0.21, -0.11)	2E-09	-0.26 (-0.37, -0.15)	9E-06
Alpha-1-Microglobulin		-0.14 (-0.19, -0.09)	1E-07	-0.23 (-0.32, -0.13)	5E-06
Kidney Injury Molecule-1		-0.14 (-0.19, -0.10)	6E-09	-0.23 (-0.33, -0.13)	6E-06
Cystatin-C		-0.17 (-0.22, -0.11)	2E-09	-0.22 (-0.33, -0.12)	5E-05
Osteopontin		-0.09 (-0.14, -0.05)	9E-05	-0.19 (-0.29, -0.09)	2E-04
Thrombonodulin		-0.14 (-0.18, -0.09)	1E-07	-0.19 (-0.29, -0.09)	2E-04
Tumor Necrosis Factor Receptor 1		-0.12 (-0.17, -0.08)	2E-07	-0.19 (-0.29, -0.09)	3E-04
Interleukin-2 receptor alpha		-0.11 (-0.16, -0.07)	1E-06	-0.11 (-0.21, -0.02)	2E-02
LC-MS/MS metabolites					
SDMA		-0.11 (-0.16, -0.06)	3E-05	-0.21 (-0.33, -0.10)	3E-04
Free Sialic acid		-0.14 (-0.19, -0.08)	6E-07	-0.18 (-0.30, -0.05)	6E-03
Tryptophan/Kynurenone		0.14 (0.09, 0.18)	6E-08	0.11 (0.00, 0.21)	5E-02
Tryptophan		0.09 (0.04, 0.13)	6E-05	<i>Not tested</i>	

Regression coefficients are per unit of standard deviation of gaussianised biomarker

We report median and interquartile range (IQR) for continuous variables, and frequency for categorical variables
 Full clinical covariates: age, sex, diabetes duration, study day eGFR, length of follow-up, categorical ACR,
 BMI, DBP, SBP, HbA1c, HDL-cholesterol, total cholesterol, smoking status, weighted average of historical eGFR

ESM Table 5: Ranking of the top biomarkers for prediction of achieved eGFR produced by the projective selection approach from linear regression models adjusted for age, sex, duration of diabetes, study day eGFR and length of follow-up

#	SDRNT1BIO	FinnDiane
Luminex		
1	CD27 antigen	CD27 antigen
2	Kidney Injury Molecule-1	Kidney Injury Molecule-1
3	Cystatin-C	Tamm-Horsfall Urinary Glycoprotein
4	Tamm-Horsfall Urinary Glycoprotein	Alpha-1-Microglobulin
5	Tumor Necrosis Factor Receptor 1	Fibroblast Growth Factor 23
6	Thrombomodulin	Osteopontin
7	N-terminal prohormone of brain natriuretic peptide	Tissue Inhibitor of Metalloproteinases 1
8	Alpha-1-Microglobulin	Latency-Associated-Peptide
9	Osteopontin	Thrombomodulin
10	Lectin-Like Oxidized LDL Receptor 1	Growth-Regulated alpha protein
LC-MS/MS		
1	Free Sialic acid	Free Sialic acid
2	Albumin T70 (501.2 / 587.5)	Beta-2-microglobulin (575.0 / 920.3)
3	Tryptophan/Kynurenine	Heat shock protein 60kDa (456.8 / 515.4)
4	Chromogranin A (488.2 / 775.4)	SDMA
5	Hydroxyproline	Retinal binding protein 2 (575.8 / 695.3)
6	Osteopontin isoA (694.3 / 853.4)	Xanthine
7	Extracellular glycoprotein lacritin (481.3 / 501.3)	Alpha-2-macroglobulin (697.8 / 737.4)
8	SDMA	Hexitol
9	ApoC-III (598.8 / 854.4)	Glycine/Valine
10	Tyrosine	Alpha-fetoprotein (490.7 / 833.3)
Both platforms		
1	CD27 antigen	CD27 antigen
2	Kidney Injury Molecule-1	Kidney Injury Molecule-1
3	Free Sialic acid	Beta-2-microglobulin (575.0 / 920.3)
4	Angiotensin II (349.8 / 136.1)	SDMA
5	Thrombomodulin	Histidine
6	Tryptophan/Kynurenine	Alpha-1-Microglobulin
7	3-Methyl-histidine	Serotransferrin (625.5 / 675.4)
8	Histidine	Fibronectin iso1 (536.3 / 680.4)
9	Citrulline	Free Sialic acid
10	Heparin cofactor II (514.8 / 814.4)	Thrombomodulin

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