Supplementary Information

Maternal control of suspensor programmed cell death via gibberellin signaling

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Supplementary Fig. 1: Biochemical characters of NtCRF.

a The interaction of NtCRF2 with three AATTT motifs (M1-M5) of the *NtCYS* promoter tested by yeast one-hybrid assay. **b** A schematic diagram showing the structure of NtCRF. **c** EMSA of His-tag with NtCRF1 used as a negative control. **d** EMSA of NtCRF1 with M3, M4 and M5, respectively. **e** and **f** EMSA of NtCRF2 with M3 motif of the *NtCYS* promoter. **g** NtCRF1 and NtCRF2 could form homo- or hetero-dimer by yeast two-hybrid test [domains for test is marked in (**b**)]. The source data of the uncropped immunoblots are provided in the Source Data file.



Supplementary Fig. 2: The expression pattern of NtCRF1.

a Semi-quantitative PCR analysis of *NtCRF1* expression in stem, anther, leaf, root and 6-DAP seeds. *GAPDH* was used as a control. **b** Expression of *pNtCRF1::H2B-GFP* in root, ovule, leaf and pollen, respectively. **c** Location of NtCRF1-GFP in the hypocotyl of *pNtCRF1::NtCRF1-GFP* line. **d** Expression of NtCRF1-GFP during ovule development in *pNtCRF1::NtCRF1-GFP* plant. Scale bars: 100 μ m (**b**), 20 μ m (**c**) and 50 μ m (**d**). The source data of the uncropped gels are provided in the Source Data file.



Supplementary Fig. 3: *Ntcrf2* mutants have no effects on *NtCYS* expression.

a Localization of NtCRF2-GFP during early embryogenesis in *pNtCRF2::NtCRF2-GFP* plants. Scale bars: 10µm. **b** Alignment of the sequences of WT and *crf2* mutants showing the insertion or deletion sites (red); numbers, translated amino acids (aa). **c** aborted seeds in WT and *crf2* mutant lines. Scale bars: 500µm. **d** The frequency of aborted seeds in WT and *crf2* mutant lines. Data represent the mean \pm SE from five independent experiments, with 200 - 300 seeds per line in each experiment. **e** The relative expression of *NtCYS* in *crf2* mutants measured by RT-qPCR. The expression level of *NtCYS* in the WT is set to 1. Data represent the mean \pm SE from three independent experiments (ns, *P* > 0.05, Sudeten's t-test). The source data of the graphs are provided in the Source Data file.



Supplementary Fig. 4: Downregulation of *NtCRF1* deceases the *NtCYS* expression and induces precocious PCD in the basal cell lineage.

a A schematic diagram of *NtCRF1* gene showing the RNAi target region. **b** Promoter activity of *NtCRF1 (pNtCRF1::H2B-GFP)* during embryogenesis. **c** Quantitative analysis of *NtCRF1* expression in 5DAP seeds of independent *NtCRF1* RNAi (Ri) lines. Quantitative analysis of *NtCYS* expression in 5DAP seeds of corresponding *NtCRF1-Ri* lines. The levels of gene expression are normalized to *GAPDH* and *UBI* expression. The level of *NtCYS* expression in wild-type anthers is set to 1. **d** Nuclear DNA fragmentation in two-celled proembryos of WT and *NtCRF1-Ri* lines stained with TUNEL. **e**, The frequency of two-celled proembryos with TUNEL-positive basal cells in WT and *NtCRF1 Ri* lines (line 59 and 67) (n = 97 - 124). **f** The abnormal suspensors at early stages of *NtCRF1* Ri embryo showed by modified pseudo-Schiffpropidium iodide staining. Stars indicated suspensor. **g** The frequency of aborted seeds in WT and *NtCRF1-Ri* lines. (n=1,000-1,500) **h** The relative frequency of abnormal or absent suspensors in (**f**) at stage 7 (n=150). Data are the means ± SE of 3 independent experiments in (**c**, **e**) and 5 in (**g**); Scale bars: 10 µm. (Student's t-test, ns, *P* > 0.05, **P* < 0.05, **P* < 0.01, ****P* < 0.001, and *****P* < 10⁻⁴). The source data of the graphs are provided in the Source Data file.



Supplementary Fig. 5: Overexpressing *NtCRF1-GFP* has no effect on suspensor PCD at stage 4.

a Relative expression of *NtCRF1* and *NtCYS* at stage 4, as measured by qRT-PCR. The expression level in WT was set to 1. **b** The Nuclear DNA fragmentation in the suspensor SC of *OE-NtCRF1* at stage 4 stained with TUNEL. **c** The frequency of embryos with TUNEL-positive basal cells in *OE-NtCRF1* line at stage 4. Data represent the mean \pm SE from 3 independent experiments (n = 90) (Student's t-test, ns, P > 0.05). Scale bar: 10 µm. The source data of the graphs are provided in the Source Data file.



Supplementary Fig. 6: Phylogenetic analysis and sequence alignment and of NtCRF and DELLAs.

a Phylogenetic analysis of NtCRFs and DELLA homologs in plants. The scale bar indicates the number of amino acid substitutions per site. **b** Alignment of NtCRFs with the known DELLAs, including tomato DELLA (PROCERA), rice DELLA (OsSLR1) and Arabidopsis (AtRGA, AtGAI and AtRGL1, AtRGL2, AtRGL3).



Supplementary Fig. 7: NtCRF1 interacts with NtGID1s depending on bioactive GA.

a Phylogenetic analysis of NtGID1s [NtGID1B (XP_016477389.1), NtGID1B2 (XP_016514746.1), NtGID1B4 (XP_016476806.1), and NtGID1C (XP_016452257.1)] and its three homologs in Arabidopsis. The scale bar indicates the number of amino acid substitutions per site. **b** Semi-quantitative analysis of *NtGID1* in two-celled proembryo (stage 1). **c** NtCRF1 interacts with NtGID1B3, NtGID1B5 and NtGID1C2 depending on GA₁ (10 μ M) or GA₄ (10 μ M), confirmed by yeast two-hybrid, respectively. The control is 0.01% ethyl alcohol. **d** qRT-PCR analysis of *NtGID1B3*, *NtGID1B5* and *NtGID1C2* in embryo proper and suspensor of 32-celled embryos (stage 4). The *GAPDH* was used as an internal control.

Supplementary Fig. 8: NtCRF1 shows the properties of known DELLAs.

a Relative expression level of *p35S::NtCRF1-GFP* lines. Values are means \pm SE of three biological replicates. **b** and **c** Representative images of hypocotyl and the lengths of 6-d-old WT, *Ntcrf1.1* and *p35S::NtCRF1-GFP* (L8) seedlings (n = 90). **d** and **e** The lengths of 3 cells in the hypocotyl (n = 90). The sizes of the 3 cells are indicated by the red regions. **f** The cell number of the hypocotyl (n = 90). **g** Hypocotyl lengths of the seedlings grown under increasing concentrations of GA₃ (0, 0.5, 1, 2 and 5 μ M). **h** The relative response to GA treatment. **i** Hypocotyl lengths of the seedlings grown in the presence of increasing concentrations of the seedlings grown in the presence of increasing concentrations of the GA biosynthesis inhibitor PAC (0, 0.1, 0.2, 0.4 and 1 μ M). **j** The relative response to PAC treatment. **k** Effect of GA₃ treatment on fluorescence of transgenic BY-2 cells expressing NtCRF1-GFP and (*crf1*- Δ 17)-GFP fusion protein, respectively. 0.05% ethyl alcohol was used as control. Scale bar: 100 μ m. (Student's t-test, ns, *P* > 0.05, **P* < 0.05, ***P* < 0.01, ****P* < 0.001, and *****P* < 10⁻⁴). The source data of the graphs are provided in the Source Data file.

Supplementary Fig. 9: Expression of the genes related to GA biosynthesis and deactivation during embryogenesis.

a Quantification of GA₅₃, GA₄₄, GA₁₉, GA₂₀ and GA₁ in WT seeds at stage 1, stage 3 and stage 5, respectively. Plotted data are the means of three replicates and presented as pg per 1,000 seeds, \pm SE. nd, not detected. **b** RT-PCR analysis of GA biosynthesis and deactivation gene in embryo proper and suspensor of 32-celled embryo (stage 4). The *GAPDH* was used as an internal control. **c** Expression of *NtGA3ox2* during embryogenesis. **d** Expression of *NtGA3ox1* in the developing seed coat. Asterisks, micropylar ends; Scale bars: 10 µm (**c**), 100 µm (**d**). The source data of the graphs are provided in the Source Data file.

Supplementary Fig. 10: Inhibition of GA biosynthesis delayed suspensor PCD and induced extra suspensor division.

a The Nuclear DNA fragmentation in embryo at stage 4 with PAC or control (0.05% ethyl alcohol) treatment. Arrows indicate extra divisions. Scale bars: 10 μ m. **b** The frequency of the embryos with TUNEL-positive basal cells at stage 4 after PAC or control treatment. Data represent the mean \pm SE from 3 independent experiments (n = 105 - 120) (Student's t-test, ****P* < 0.001). The source data of the graphs are provided in the Source Data file.

Supplementary Fig. 11: Phylogenetic analysis and sequence alignment of NtGA3ox proteins.

a Phylogenetic analysis of NtGA3ox and homologs proteins in *Nicotiana tabacum*. **b** A bootstrap consensus of phylogenetic tree representing similarities of NtGA3ox protein sequence.

Supplementary Fig. 12: Integument-, endosperm- and embryo-specific expression of *NtGA3ox1-GFP* did not influence *NtCYS* expression.

a Promoter specificity of *NtWCP28* (*pNtWCP28::H2B-GFP* expression), *NtCYS8* (*pNtCYS8::H2B-GFP* expression), *NtTPE8* (*pNtTPE8::H2B-GFP* expression) and *NtDRP* (*pNtDRP::H2B-GFP* expression) in the seeds (left) and in the fertilized embryo sac with a proembryo (arrow) and endosperm cells (right). **b** Relative expression of *NtGA3ox1* and *NtCYS* in WT and *pNtWCP28::NtGA3ox1-GFP* lines. **c** Relative expression of *NtGA3ox1* and *NtCYS* in WT and *pNtCYS8::NtGA3ox1-GFP* lines. **d** *NtGA3ox1-GFP* expressed in proembryo and relative expression of *NtCYS* in WT and *pNtCYS8::NtGA3ox1-GFP* lines. **d** *NtGA3ox1-GFP* lines. The expression level in WT was set to 1. Data represent the mean \pm SE from three independent experiments. (Student's t-test, ns, P > 0.05, *P < 0.05, and **P < 0.01). Scale bars: 100 µm (**a**), 10 µm (**d**). The source data of the graphs are provided in the Source Data file.

Supplementary Fig. 13: The phenotypes in *global-della* and *ga1-3* mutants in Arabidopsis. **a** Abortive seeds and reduced fertility in Arabidopsis *global-della* mutants. Arrow indicates abortive seeds. Stars indicate unfertilized ovules. **b** The proembryo of *global-della* mutant at the same stage. Arrows indicate extra cell divisions in an abnormal suspensor. **c** Arabidopsis *ga1-3* mutants show abnormal inflorescences and pistil. **d** Ovules of 48HAP WT (L*er*) and *ga1-3* plants. Arrow indicates arrested ovule with aborted female gametophyte development (98%, n=350). Scale Bar: 20µm (**b**) and (**d**) bottom; 0.5mm (**d**) top; 1mm (**c**).

Supplementary Fig. 14: The work model of suspensor PCD.

Maternal-embryo communication via gibberellin signaling precisely controls the duration of maternal nursing of proembryos.