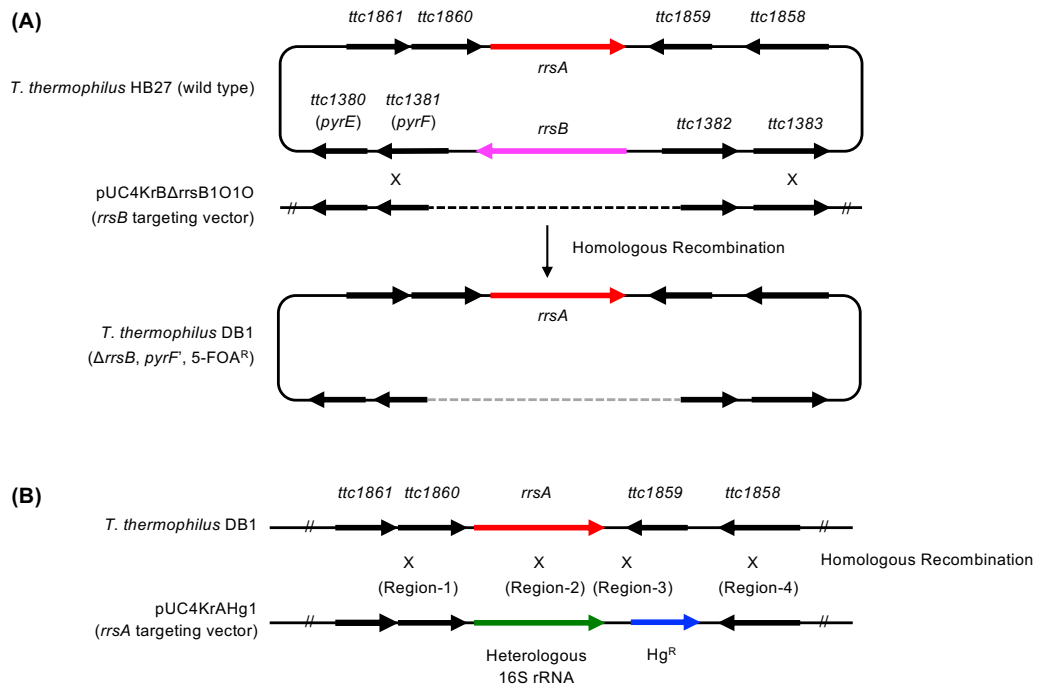


Supplementary Information

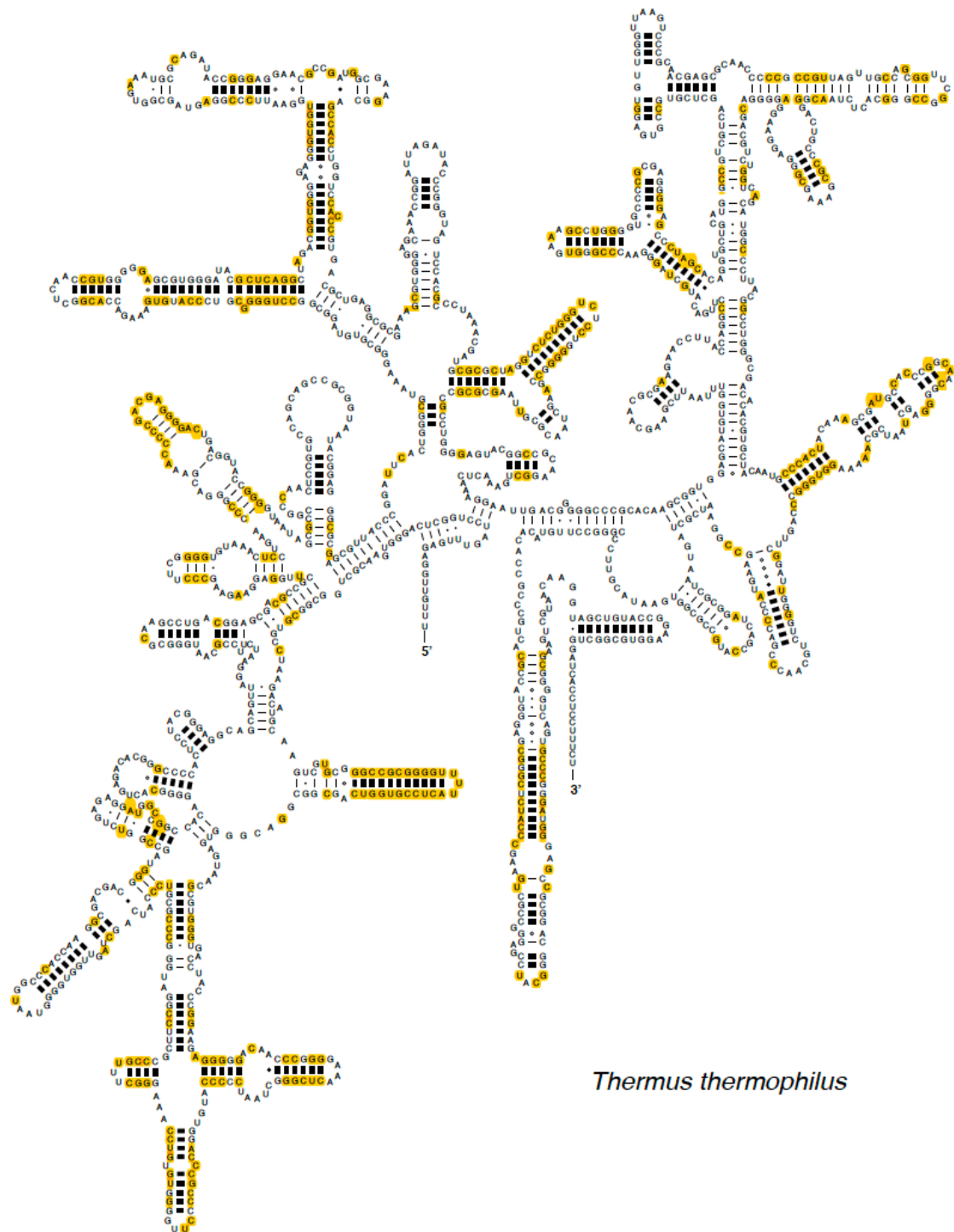
Occurrence of randomly recombined functional 16S rRNA genes in *Thermus thermophilus* suggests genetic interoperability and promiscuity of bacterial 16S rRNAs

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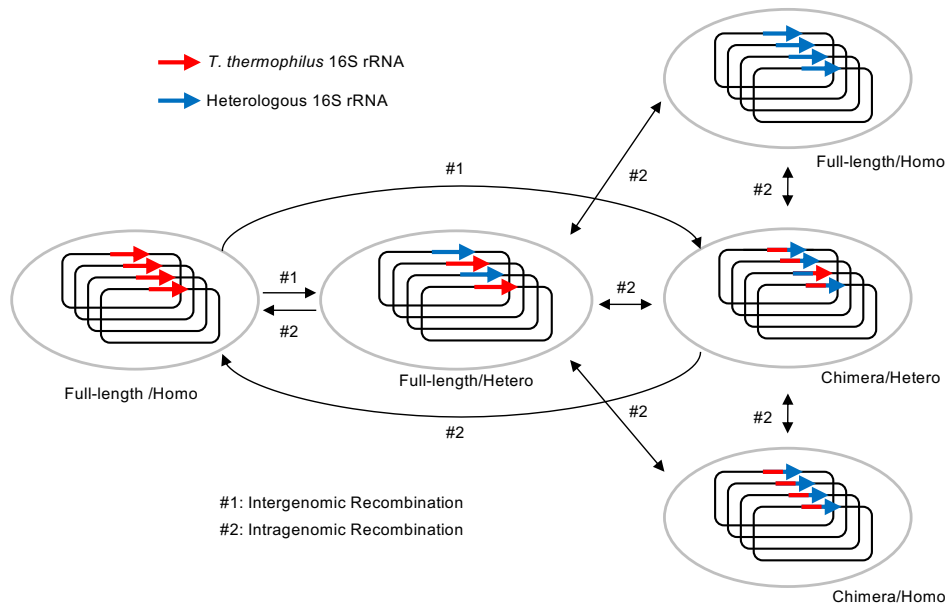
Supplementary Fig. S1. Experimental horizontal gene transfer of 16S rRNA in *T. thermophilus*.

(A) Creation of 16S rRNA gene knockout mutant of *T. thermophilus* HB27. A targeting vector (pUC4KrB $\Delta rrsB$ 1O10) was used to transform *T. thermophilus* HB27. Through a double crossover mechanism, a mutant strain designated DB1 was obtained. Given that the flanking *ttc1381* codes *pyrF*, by partially deleting the gene, the *rrsB*-deletion mutant was obtained as a 5-FOA resistant. **(B)** Interspecies exchange of 16S rRNA gene. *T. thermophilus rrsA* gene was replaced by a heterologous gene. *ttc1859*, which is nonessential for growth, was replaced by a thermostable Hg^R gene for efficient screening of recombinants.



Thermus thermophilus

Supplementary Fig. S2. Different nucleotides in *M. ruber* and *D. radiodurans* 16S rRNAs mapped on the secondary structure of *T. thermophilus* 16S rRNA. Variable sites are shown in yellow. Large portions of the different nucleotides localized at the stem regions.

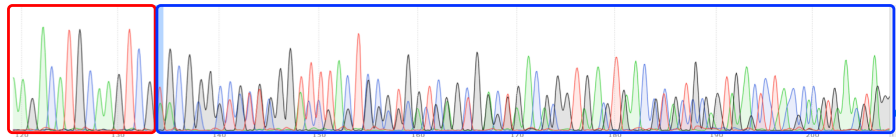


Supplementary Fig. S4. Mechanism for generation of chimera in *T. thermophilus*. *T. thermophilus* is a polyploid. Intergenomic followed by iterative intragenomic recombination promotes chimerization of 16S rRNA genes.

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T. thermophilus AGACATGCAA GTTCGTGCGGGCCGCGGGGTTTTACTCCGTGGTCAGCGGGGACGGGTGAGTAACGCGTGG 120
D. radiodurans AGACATGCAA GTCGAACGCGGT-----CTT---CGGACCGAGTGGCGCACGGGTGAGTAACACGTAA 109
***** ** * ** * ** ** ** ** ** ** ** ** ** ** ** ** **

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Supplementary Fig. S5. An example of mixed sequencing chromatograms. *T. thermophilus* is a polyploid and in some cases, allopolyploid status was observed as a mixed sequencing chromatogram in a heterogeneous region. In this figure, *T. thermophilus* and *D. radiodurans* sequences share identical sites but the different sites show superimposed sequences that may reflect the presence of insertions/deletions.

Supplementary Table S1. List of *T. thermophilus* DB2 mutants

Donors Mutants	Chimera (Type) ¹ or Full-length	Boundaries ²	Database ID
<i>T. kowarayensis</i>			LC436704
TY2	Chimera (HD)	1115	LC436735
TY5	Chimera (HD)	1424	LC436727
TY15	Chimera (HD)	1075	LC436739
TY16	Full-length	N.A. ³	LC436729
<i>T. aquaticus</i>			LC440032
TY10	Chimera (DHDHDH)	748/846/1006/1251/1297	LC436726
TY23	Chimera (DHD)	846/1006	LC436717
<i>T. Brockianus</i>			LC436697
TY1	Chimera (DHDHD)	262/265/1251/1424	LC436725
TY11	Chimera (HD)	1424	LC436732
TY12	Full-length	N.A. ³	LC436719
<i>T. scotoductus</i>			LC436703
TY3	Full-length	N.A. ³	LC436721
<i>M. ruber</i>			LC436693
TY9	Chimera (HD)	974	LC436709
TY19	Chimera (HDHD)	543/822/1244	LC436695
TY20	Chimera (HD)	1409	LC436700
TY25	Full-length	N.A. ³	LC436711
TY40	Chimera (HDH)	1075/1474	LC436714
Mru2	Chimera (HD)	382	LC440033
<i>D. radiodurans</i>			LC436701
TY32	Chimera (DHD)	989/1410	LC436728
TY34	Full-length	N.A. ³	LC436723
TY38	Chimera (HDH)	264/286	LC436715
S52	Chimera (HD)	1357	LC436736
S66	Chimera (HD)	1242	LC436699
<i>R. marinus</i>			LC436722
42	Full-length	N.A. ³	LC436737
48	Chimera (DHD)	955/1356	LC436702
53	Chimera (DHD)	812/904	LC436696
S40	Chimera (HD)	881	LC436731
S41	Chimera (HD)	1409	LC436734
S44	Chimera (HD)	1356	LC436740

S86	Chimera (DHD)	955/1326	LC436738
<i>R. xylanophilus</i>			LC436710
61	Full-length	N.A. ³	LC436724
63	Chimera (DH)	1410	LC436720
89	Chimera (HD)	906	LC436712
Rxy3	Chimera (DHD)	907/1343	LC440343
<i>T. maritima</i>			LC436716
S1	Chimera (HDHD)	1116/1152/1232	LC436698
S2	Chimera (HD)	1414	LC436713
S7	Chimera (HD)	904	LC436694
S17	Chimera (HD)	949	LC436707
<i>A. aeolicus</i>			LC436706
S4	Chimera (HD)	1393	LC436705
S8	Chimera (HD)	1414	LC436730
S10	Chimera (HD)	1383	LC436718
S29	Chimera (HD)	1510	LC436708

¹ Donor sequence is denoted by D and host *T. thermophilus* sequence is denoted by H.

² Boundary is defined by the first nucleotide site that distinguishes two sequences. For example, the TY2 chimeric mutant from 5' end to nucleotide position 1114 derives from *T. thermophilus* (H) and from 1115 to 3' end from *T. kowarayensis* (D). *E. coli* numbering.

³ N.A., not applicable.

Supplementary Table S2. Nucleotide composition of *T. thermophilus*, *M. ruber*, and *D. radiodurans* 16S rRNA genes

		A	T	G	C	%GC	Total
Full-length	<i>T. thermophilus</i>	311	237	546	427	0.64	1521
	<i>M. ruber</i>	347	288	488	368	0.574	1491
	<i>D. radiodurans</i>	367	304	483	350	0.554	1504
Consensus		260	188	391	267	0.595	1106
Variable	<i>T. thermophilus</i>	51	49	155	160	0.759	415
	<i>M. ruber</i>	87	100	97	101	0.514	385
	<i>D. radiodurans</i>	107	116	92	83	0.44	398

Supplementary Table S3. Molecular mass of ribosomal proteins in 30S subunit ¹

Proteins	<i>T. thermophilus</i>	<i>A. aeolicus</i>	<i>T. maritima</i>
S1	59851.6	60769.2	61356.7
S2	29228.3	31429.0	29887.9
S3	26672.6	24137.2	24057.8
S4	24306.0	24796.6	24702.3
S5	17525.2	23192.7	18461.4
S6	11954.6	13455.3	15351.3
S7	17997.7	18606.5	13439.3
S8	15791.3	19439.6	15270.7
S9	14369.4	17046.5	15179.4
S10	11911.8	12217.2	11618.6
S11	13694.6	13508.5	14074.2
S12	15119.7	14188.4	14042.4
S13	14312.6	14321.6	14413.6
S14	7121.6	7485.0	7324.9
S15	10536.2	10888.7	10789.7
S16	10766.2	13020.2	11235.9
S17	12278.4	12817.8	12549.4
S18	10241.1	8694.2	8997.9
S19	10563.3	21624.1	10880.8
S20	11684.9	13249.3	11274.1
Total	345927.0	374887.5	344908.4

¹ Genomic sequences used for calculation were as follows: AE017221.1 (*T. thermophilus*); NC_000918.1 (*A. aeolicus*); NC_000853.1 (*T. maritima*).

Supplementary Table S4. Natural occurrence of HGT in 16S rRNAs

Phylogeny	Type of HGT	References
<i>Haloarcula marismortui</i> (Euryarchaeota)	Intragenomic heterogeneity	Mylvaganam and Dennis, 1992 45
<i>Aeromonas</i> (Proteobacteria)	Chimeragenesis	Sneath et al., 1993 46
<i>Rhizobium</i> (Proteobacteria) <i>Aeromonas</i> (Proteobacteria)	Chimeragenesis	Eardly et al., 1996 47
<i>Escherichia</i> and <i>Salmonella</i> (Proteobacteria)	Intragenomic heterogeneity Chimeragenesis	Cilia et al., 1996 48
<i>Thermobispora bispora</i> (Actinobacteria)	Intragenomic heterogeneity	Wang et al., 1997 49
<i>Vibrio cholerae</i> (Proteobacteria)	Intragenomic heterogeneity	Lan and Reeves, 1998 50
<i>Streptomyces</i> (Actinobacteria)	Chimeragenesis	Ueda et al., 1999 51
<i>Thermomonospora chromogena</i> (Actinobacteria)	Intragenomic heterogeneity Chimeragenesis	Yap et al., 1999 52
<i>Actinomycetes</i> (Actinobacteria)	Chimeragenesis	Wang and Zhang, 2000 53
<i>Bradyrhizobium</i> (Proteobacteria)	Chimeragenesis	Parker, 2001 54
<i>Streptococcus anginosus</i> (Firmicutes)	Chimeragenesis	Schouls et al., 2003 55
Rhizobia (Proteobacteria)	Chimeragenesis	van Berkum et al. 56
<i>Thermoanaerobacter tengcongensis</i> (Firmicutes)	Intragenomic heterogeneity	Acinas et al., 2004 57
<i>Desulfotomaculum kuznestovii</i> (Firmicutes)	Intragenomic heterogeneity	Tuorova, et al., 2001 58
<i>Halosimplex carlsbadense</i> (Euryarchaeota)	Intragenomic heterogeneity	Boucher et al., 2004 59
<i>Natrinema</i> sp. (Euryarchaeota)		

<i>Rhizobium galegae</i> (Proteobacteria)	Chimeragenesis	Eardly et al., 2005 60
<i>Helicobacter</i> (Proteobacteria)	Chimeragenesis	Dewhirst et al., 2005 61
<i>Scytonema hyalinum</i> (Cyanobacteria)	Intragenomic heterogeneity	Johansen et al., 2017 62

Supplementary Table S5. Oligonucleotide primers used in this study

Oligonucleotides	Sequence (5' to 3')
TTC1380Rev	CTAGACCTCCTCCAAGGGCACC
TTC1383Rev	CTACACGTCTAGGATGACCTGGGC
TTC1381_100OUT	ACACCCTCGAGCTCCTCGAGGCCCTGGCCC
TTC1382_100OUT	TCACCCTGACCCGGGGGAAGCCGGGAGAA
TTC1858Fwd	ATGAGCATCCCCGAGGAGCTTTGGCGCCGC
TTC1860DownFwd	TAGCATGGCTTTTGCTGCGCGGGCG
TTC1861Fwd	ATGTCACGGGACCTTCTGGGCCTCGAGGGG
TTC1859UpRev	ACCCCTCCTTACGGCAACAAAAA
TTC1859Fwd300	CTCATGCGCCACCTGGGGG
Thermus_1F	TTGTTGGAGAGTTTGATCCTGGCTCAGGG
Thermus_1521R	AGAAAGGAGGTGATCCAGCCGCACC
Thermus_1R	CCCTGAGCCAGGATCAAACCTCTCCAACAA
Thermus_1521F	GGTGCGGCTGGATCACCTCCTTTCT
Thermus_1F(AT)	TTTGTGGAGAGTTTGATCCTGGCT
Thermus_1499R(AT)	GGAGGTGATCCAGCCGCA
Thermus_1R(AT)	AGCCAGGATCAAACCTCTCCAACAAA
Thermus_1499F(AT)	TGCGGCTGGATCACCTCC
HgFwd	ATGAAAAGCCTGAACTCACCGTG
HgRev	TTAGGCTGCCAGGTCGACGGAT

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