

Supplemental Information

Deciphering the essentiality and function of the anti- σ^M factors in *Bacillus subtilis*

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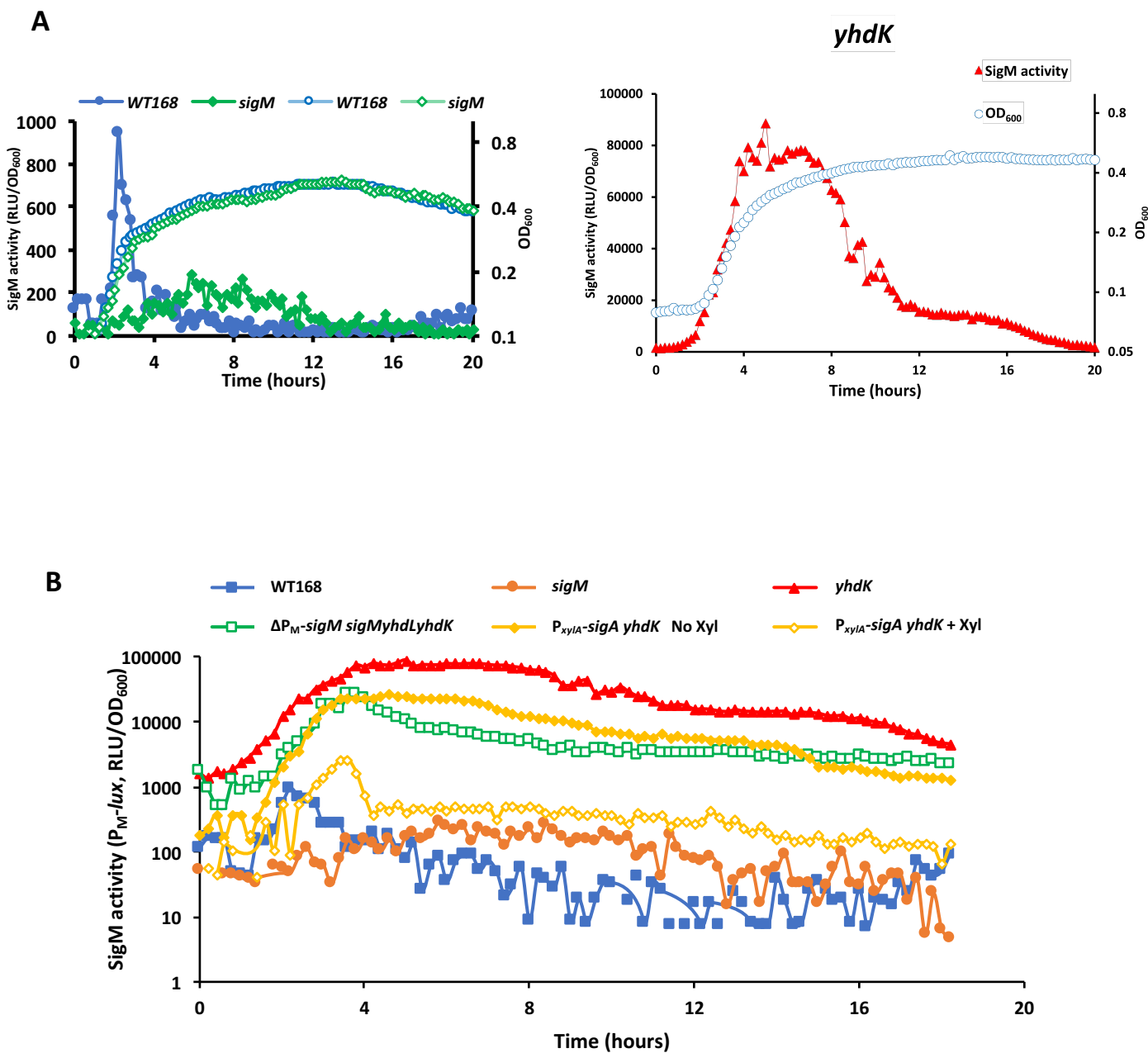
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Table S1. Primers and gBlock used in this study

Number	Name	Sequence
6578	xyIR-F-BamHI	ATCGGGATCCCCGGTGGATAAACAAAGGAGTGG
6579	xyIR-R-NotI	ATCGCGCGCCCGGGAGCTCCTAACTTATAGGGGT
6580	sigM-F-NotI	ATCGCGCGCCCGCCTATCTTTTGGCGCCAT
6581	sigM-R-HindIII	ATCGAAGCTTTGGTGCCTCATTTCGCCATT
6582	yhdL-cln-up-F	GCCGTTTTTCGTGCGAGAAT
6583	yhdL-cln-up-R	CGCCGACATTGCTGATTTTTCTGGTTCGCTCATTTC
6584	yhdL-cln-mid-F	GGGAAATGAGCGACCAGGAAAATCAGCGAATGTCGGCG
6585	yhdL-cln-mid-R	CCTATCACCTCAAATGGTTCGCTGCCGAAAACCGGTATAACGAAA
6586	yhdL-cln-down-F	CGAGCGCCTACGAGGAAATTTGTATCGAGATACGAAATTTACAGTTTGGCT
6587	yhdL-cln-down-R	ACGAATCGGGCAATCATGTG
6588	chr-sigM-seq-F	CCATTGTGCCACTCCTTCAC
6589	chr-sigM-seq-R	TGCAGTCATTTCTGGTTCGC
6590	pAX01-check-F	GGGGGAAATGACAAATGGTCC
6591	pAX01-check-R	ACGAAAGGGCCTCGTGATAC
6599	Pxyl-yhdL-F-BamHI	ATCGGGATCCTAGAGGGGAGAAAAGGCAATGATGAATGAAGAAATTTAAAAAGC
6600	Pxyl-yhdL-R-SacII	ATCGCCGCGGTCCAGCCGAATACATTGTG
6693	pAX01-erm-cm-up-F	GCCGCACTCTTCTTTTTCAA
6694	pAX01-erm-cm-up-R	CTTGATAATAAGGGTAACATAATTCCTTTGGTTGAGTACTTTTTCACTCG
6695	pAX01-erm-cm-down-F	GGGTAACTAGCCTCGCCGGTCCACGCTGGGGGAGGAAATAATCTATGAGTCCG
6696	pAX01-erm-cm-down-R	TCCGCATTTTTGCATGGAGC
6759	yhdL-check-F	ACGCTGGGAAAGCTACCTCTA
6760	yhdL-check-R	TCTGCTTTTGCAGTCTGTTG
6808	PsigM-F-EcoRI	AGCTGAATTCGCCGTTTGCATGTAATGTG
6809	PsigM-R-PstI	AGCTCTGCAGCAGTAAGTCTTCAGCAAGATGC
6814	pBs1ClacZ(lux)-check-F	AAAGGATTTGAGCGTAGCGA
6815	pBs1ClacZ-check-R	TTGGGTAACGCCAGGGTTTT
6816	pBs3Clux-check-R	GAGAGTCTCTGTCGACCT
7249	yhdL-in-check-F	GAAAACACAGCACCGGCAAT
7250	yhdL-in-check-R	AATTCACCTCGCCGACATT
7324	rpoB-gRNA-F	TACGGACTGTTAAGTCTGATGACG
7325	rpoB-gRNA-R	AAACCGTCATCAGACTTAACAGTC
7326	rpoB-repair-up-F	AAGGCCAACGAGCCTCGCCTTCGCAATCTTGATG
7327	rpoB-repair-up-R	CTTGATGTTTTACACGCTCCAACAACATCATTAGACTTAACAGTCAGAAATTTCTGA
7328	rpoB-repair-down-F	TCAAAGAAATTCGACTGTAAAGTCTAATGATGTTGTTGGACGTGTGAAAACATACGAAAG
7329	rpoB-repair-down-R	AAGGCCATTTAGCCCTACGGCGTTGTCTTGTGAA
7340	rpoC10034-gRNA-F	TACGGGTAACGCTGTCGTAATCT
7341	rpoC10034-gRNA-R	AAACGAGTAATCGACAGCTTACC
7344	rpoC1004-repair-up-R	CAACAACGATTACAGAAGCTCCTGAATAGTCCACATGTTACCAAGAAGGTTTTGACGGA
7345	rpoC1004-repair-down-F	TCCGTCAAAACCTTCTTGGTAAACATGTGGACTATTCAGGACGTTCTGTAATCGTTGTTG
7348	pJOE8999-check-F	CCTTTTTGCGTGTGATGCGA
7349	pJOE8999-check-R	GTCAGCTAGGAGGTGACTGA
7405	SigA-R-SacII	ATCGCCGCGGACAAAATTTGAATAGAAACATGCCT
7406	SigA-F-SpeI	ATCGACTAGTTTGGCGAGGAGCAATAGAT
7625	NFLAG-YhdK-Up-F	AAGGCCAACGAGGCGGGAAATGAGCGACCAGGAA
7628	NFLAG-YhdK-Down-R	AAGGCCTATTGGCCGACATCGAGCAAGCTGGAGA
7635	N-FLAG-YhdK-gRNA-F	TACGGCGAGGTGGAATTAAGGAAAC
7636	N-FLAG-YhdK-gRNA-R	AAACGTTCCATAATTCACCTCCG
7656	PfosB-XbaI-F	CTAGACTGTATGAAACTTTCTTATGAAAAAAGTCGTATATGTTGGATGATCAGCTTCTGCA
7657	PfosB-PstI-R	GAACTGATCATCCACATATACGACTTTTTTATAAGAAAGTTTCATACAGT
7738	gBLOCK-CL-NK-up-R	TTGGCCCTGTAAACGACCGAAACCGTATACCTTTGACACCGTTTTTCTCTACA
7739	gBLOCK-CL-NK-down-F	TTGCCATGGCTGCCGTTCCGTTCCGGCGTTGTTGAACTCTT
7545	PsigW-XbaI-F	ATCGCTAGAACTTTGACTCCGTCATCGCT
7827	sigM-int-F	CCGCTGTCTGGCAGGAAATA
7828	sigM-int-R	TTTTCAAGTGGCGCGAAACG
8110	PspoIIM-XbaI-F	ATCGCTAGACATACAGCAGTTGATGATAAGG
8111	PspoIIM-PstI-R	ATCGCTGACGCGCTCTAGTGTATTTGATTTAATA
8112	PspoVFA-XbaI-F	ATCGCTAGAGCGGGCTCTAAGAAAAACAT
8113	PspoVFA-PstI-R	ATCGCTGACGCTGGATCTCTAGTTGTTAAGC
8114	sigH-check-F	ATTTTCGGACAGGGGGCATT
8115	sigH-check-R	GGTTTCCGCATGTCTTGCAAT
8116	sigE-check-F	CAGGGGAGTTGGTACGACAAA
8117	sigE-check-R	ACCCCGCAGATTTTCGACTTT
8118	spoIIIC-check-F	TGCCTGCAACTTGGACTGAT
8119	spoIIIC-check-R	AGCTTTTAGAACGTCGCGCT
8122	spoIVCB-check-F	ACAGGCCTGCCATCCATTTT
8123	spoIVCB-check-R	GGCCGCAAGCGTCTTTTAAC
8132	PspoVG42-up-F	ATCGCTAGAAATAAGAAAAGTGATTTCTGGGAGA
8133	PspoVG42-up-R	TCCTGCTGTTTTTAAAAATTTTTTAAAAAATAGGATATAGTTACACAATTAGGT
8134	PspoVG42-down-F	ACTATATCCTATTTTTTAAAAAATTTTTTAAAAACGAGCAGGA
8135	PspoVG42-down-R	ATCGCTGCAGTCCCTATATAAAAGCATTAGTGTATCA
DR194	SigM operon Fwd	GGAAAAGGTGGTGAACACTACTAGCCGATTCTGGAGGCTTC
DR197	SigM auto rev	CTATGTTATACTTTTATAAGAAAGGTTTAAAGTTGCAC
DR196	sigM auto Fwd	TTTCTTATGAAAGTATAACATAGAGGG
DR198	sigM rev	CATGGCCTGCCCGGTTATTACTGGTTCGCTCATTTCCTCA TGTAGAGAAAACCGGTGCTCAATGTATACGGTTCGGTCTTACAGGGCCAACCAAGGAAATCCAGCGCATGCTGAAAAATAAATCTGTAATAATCAGCGA ATGTCGGAGAAAGTGGAACTTTGGAATGGGGATCAGGCCAAGGACCGGGAAAGCGGACAAAGGCCCTGATTACAAAAGATGACGATGACAAAGGCGAGGT GGAATATGGAATAGTAAGAGATTATAAAGATGATGATGATAAAGGCTCAGGCCAAGGACCTGGATCAGGCCAAGGCCCTATGGAACCTGTTAGAATT TTTAAAGAACACAAATGTTTCGGCTGGATCTGTCGGGACAGCGGTTCTGCCCTGCTGTTGCTGAATTTGGCCATTATCAGCAACGTCACGTTTTTAT CCTATCAAATGCTTCCGTTCCGCTAGGCTGCCGTTCCGTTCCGGCTGTTGAACTCTT

gBlock sequence for C-FLAG-yhdL N-FLAG-yhdK

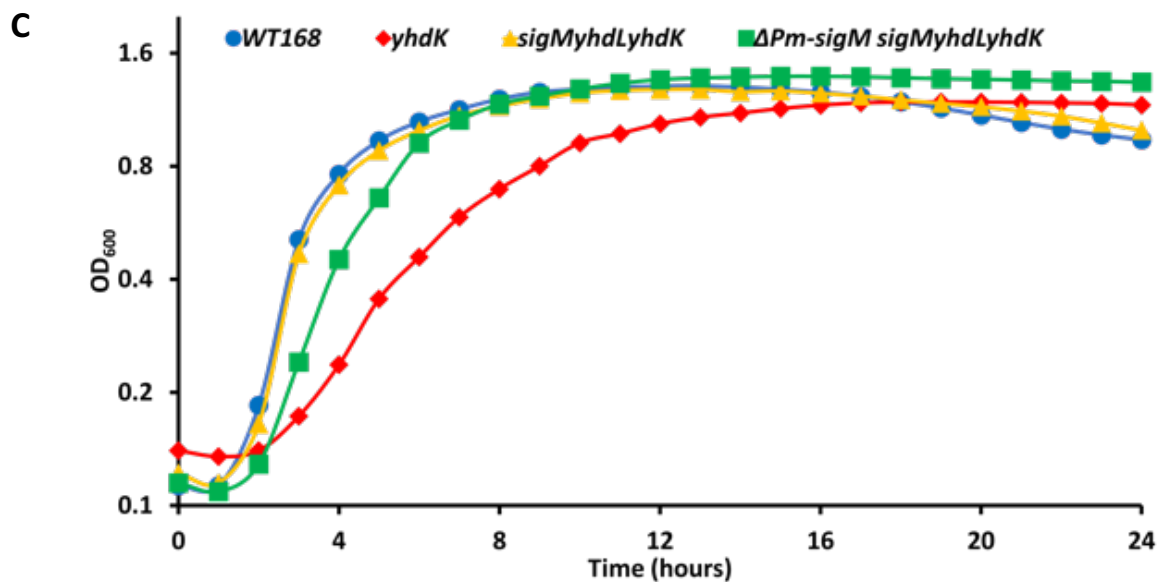
Figure S1. Growth and SigM activity in the absence of each anti-sigM factor



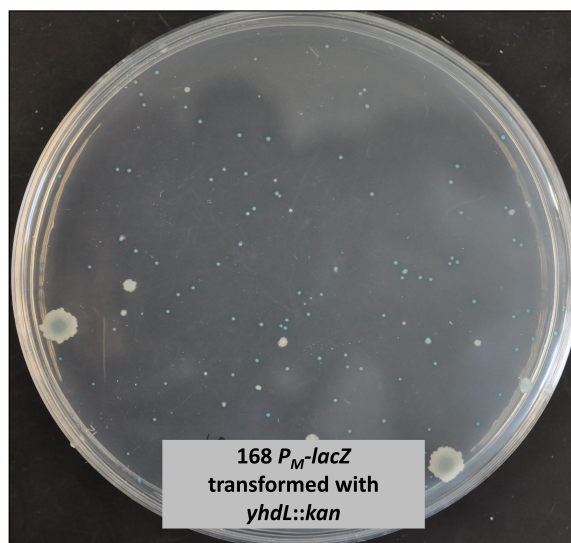
A. [Left panel] SigM activity of WT (168) strain (HB17325) and the *sigM* null mutant (HB17494) during growth in LB medium at 37°C as measured using a P_M -*lux* reporter (promoter activity is in closed symbols and was calculated by dividing relative light unit (RLU) by OD_{600} , in open symbols, with measurements every 12 min). [Right panel] SigM activity of the *yhdK* null mutant during growth in LB medium. Note that OD_{600} was measured with a culture volume of 100 μ l per well in a 96-well plate.

B. SigM activity of different strains as measured in panel A. A representative measurement is shown.

Figure S1. Growth and SigM activity in the absence of each anti-sigM factor



D

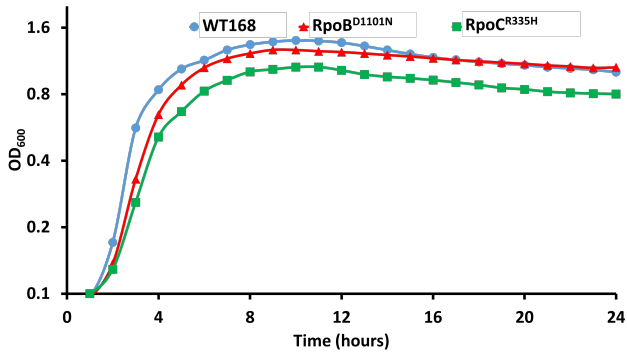


C. Representative growth curve of WT and mutants in liquid LB medium. OD₆₀₀ was measured with a culture volume of 200 μ l per well in a Bioscreen plate, so final OD₆₀₀ values are twice those shown in Figure S1A and correspond roughly to those normally detected using a 1 cm path length in a spectrophotometer.

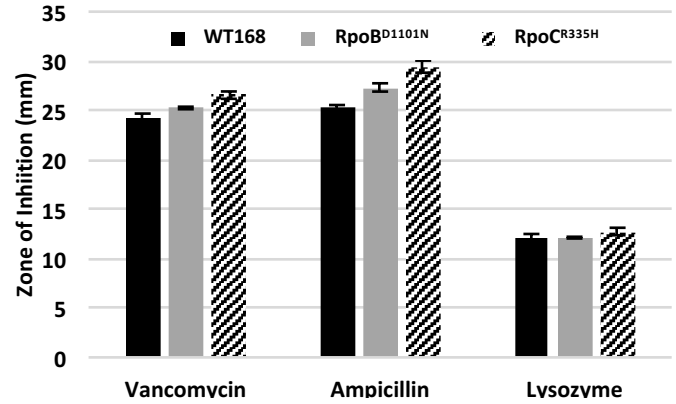
D. Transformation plate using WT 168 strain with a P_M -lacZ reporter as recipient, and chromosomal DNA of a *yhdL* deletion strain (HB17264) as donor. Transformed cells were plated on a LB plate containing kanamycin and X-gal and incubated at 37° C for 48 hours

Figure S2. Effect of *rpoB/C* mutations on cell growth, antibiotic sensitivity, morphology and alternative sigma factor activity.

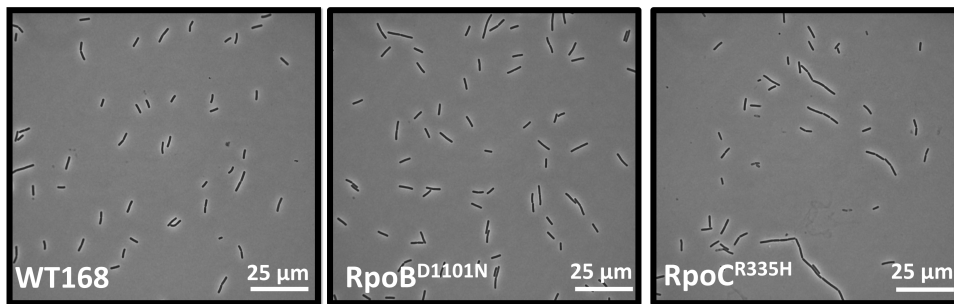
A



B



C



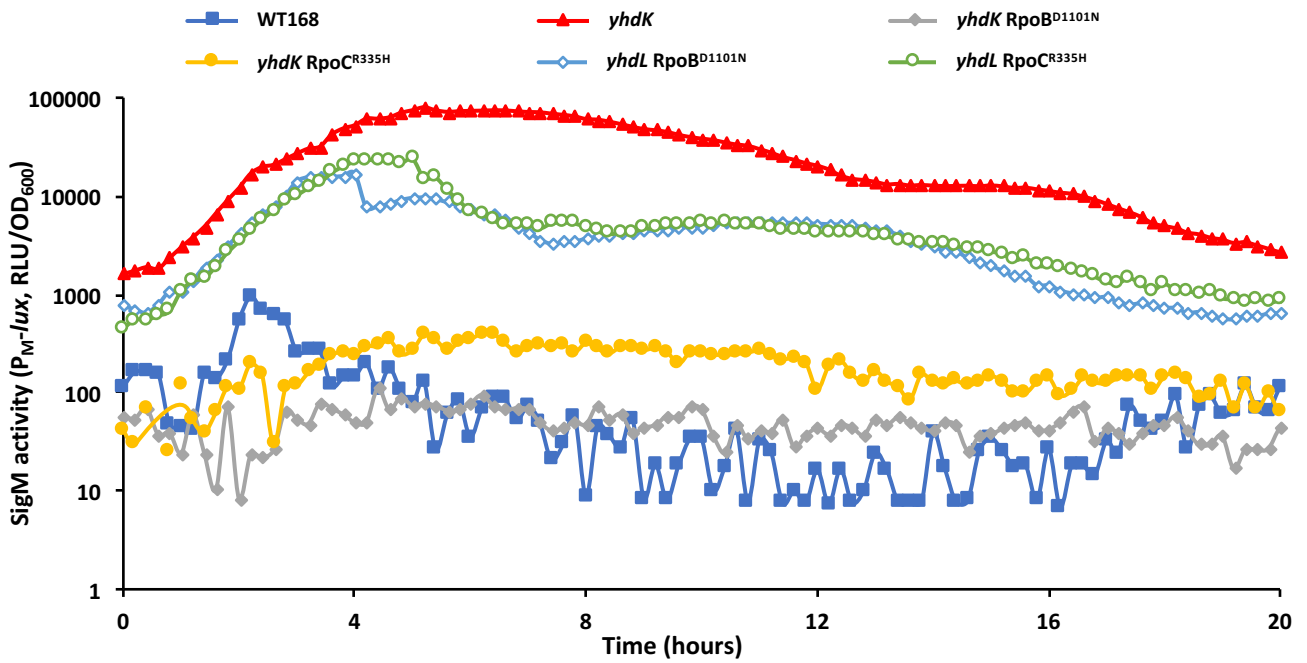
A. Growth curve of WT and *rpoB/C* mutants in LB medium at 37 °C in Bioscreen.

B. Zone of inhibition assay showing sensitivity of WT and *rpoB/C* mutants against vancomycin, ampicillin, and lysozyme. Data was presented as mean ± SEM, n=3.

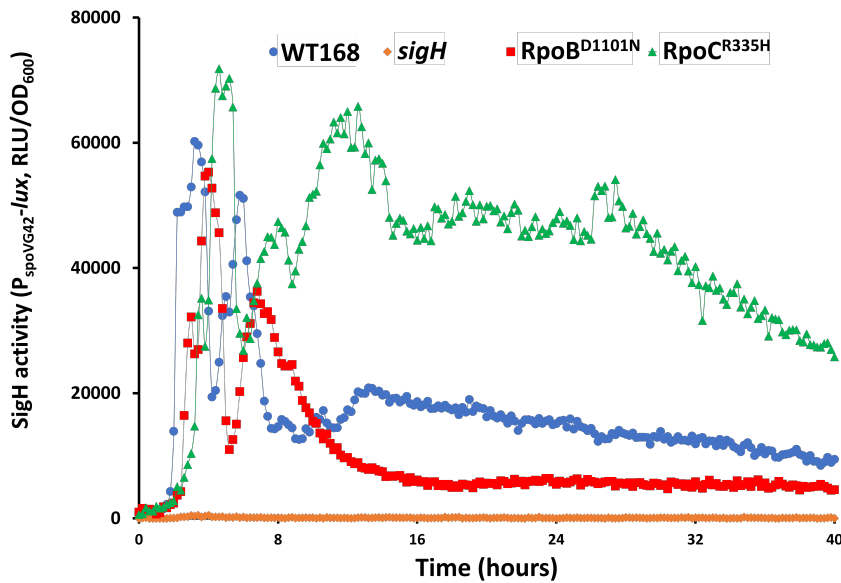
C. Representative phase contrast microscopic images of WT and mutants grown in LB medium at exponential phase.

Figure S2. Effect of *rpoB/C* mutations on cell growth, antibiotic sensitivity, morphology and alternative sigma factor activity.

D



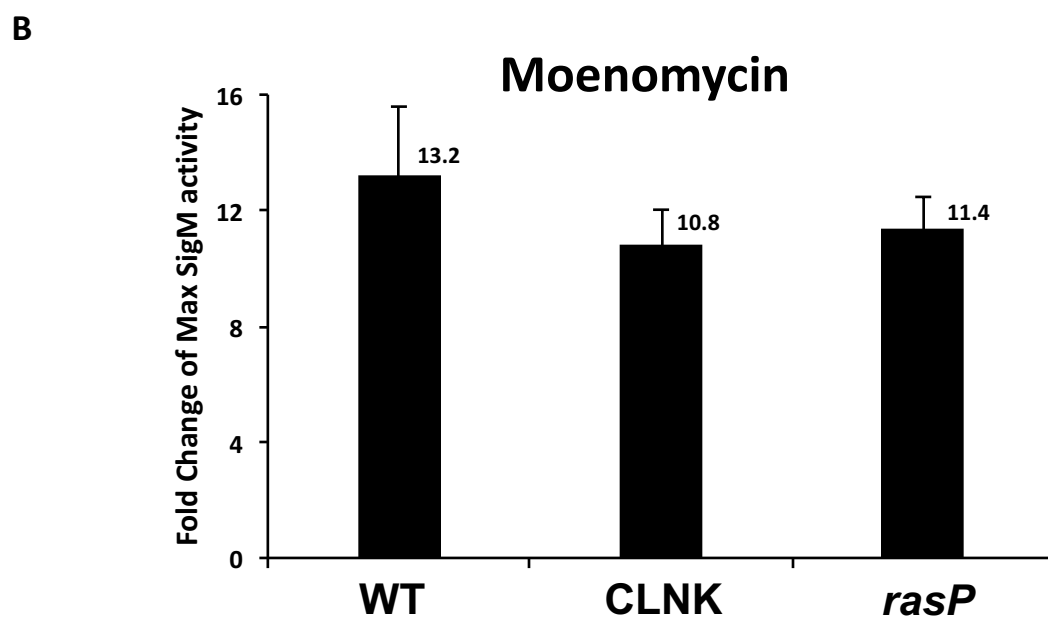
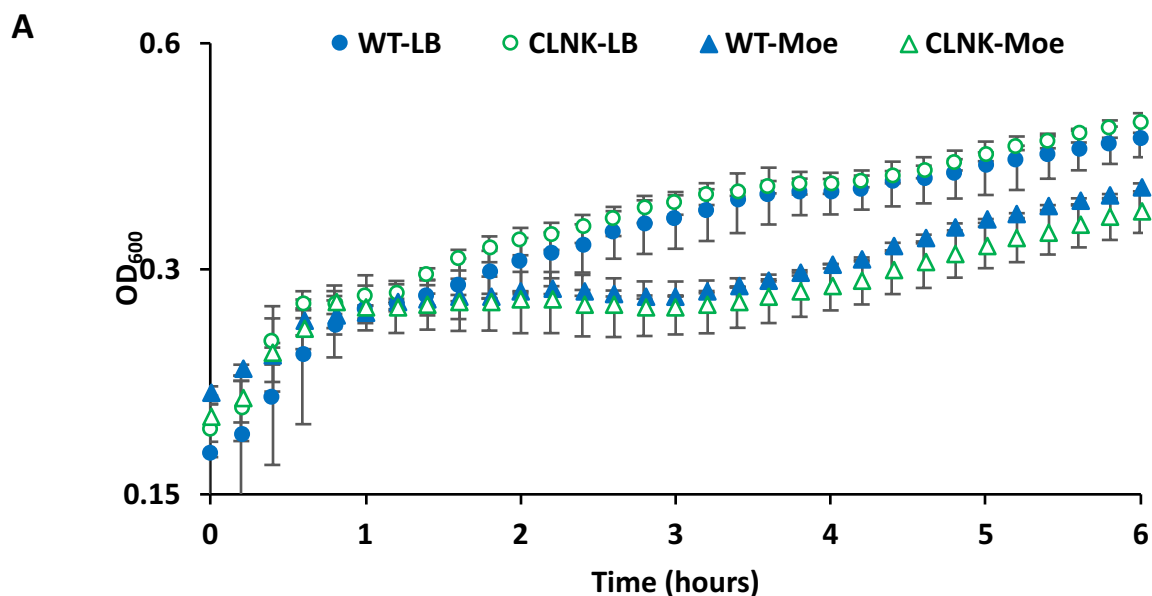
E



D. SigM activity of *yhdK* and *yhdL* mutants with or without single amino acid substitution in RpoB or RpoC.

E. SigH activity of WT168 strain and mutants during growth in LB medium. SigH activity was measured using a $P_{spoVG42-lux}$ reporter, with relative light unit (RLU) and OD₆₀₀ measured every 12 minutes. The promoter activity is calculated by dividing RLU with OD₆₀₀.

Figure S3. C-FLAG-YhdL and N-FLAG-YhdK exhibit function similar to WT



A. Curves of OD₆₀₀ of WT and epitope tag labelled C-FLAG-YhdL N-FLAG-YhdK (CLNK) in LB or LB supplemented with 10 $\mu\text{g ml}^{-1}$ moenomycin. Measurement was performed every 6 minutes and every other measurement was shown for simplicity. Data was presented as mean \pm SD, n=3.

B. Fold change of maximum SigM activity of different strains after treatment of final concentration of 10 $\mu\text{g ml}^{-1}$ moenomycin. SigM activity was measured every 6 minutes after treatment using the P_M-*lux* reporter and the maximum value was used to calculate fold changes between samples with or without treatment.