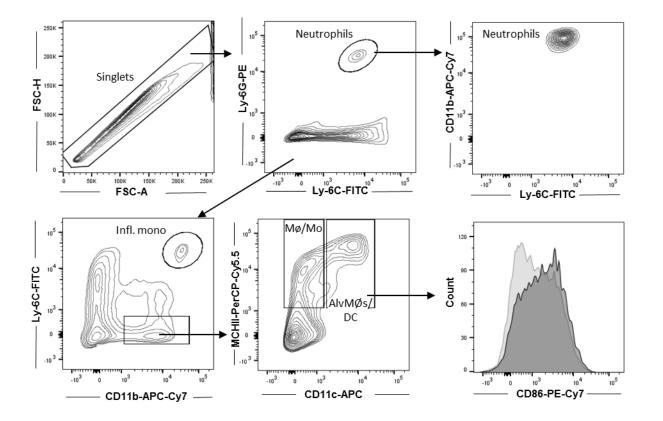
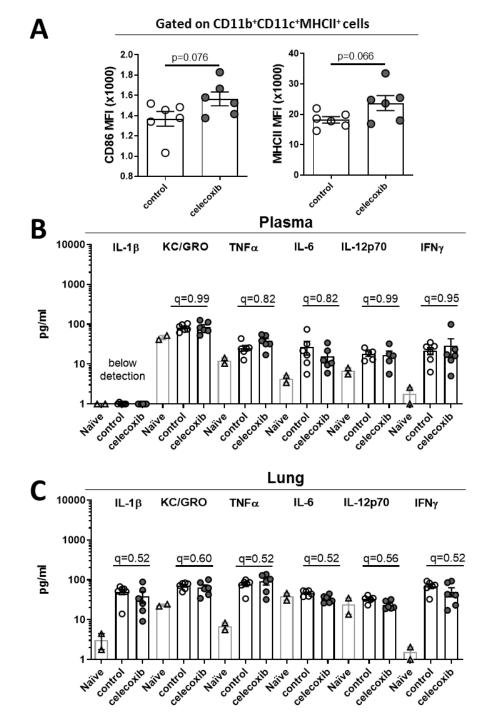
Supplementary Figure 1.

Example of gating strategy to identify innate cell populations in lungs of Mtb infected animals. In the "Singlets" gate, neutrophils were defined as CD11b⁺/Ly-6C^{dim}/Ly-6G⁺ and with in the "not neutrophils" gate, inflammatory monocytes were identified as CD11b⁺/Ly-6C^{high}. Gating on Ly-6C⁻/Ly-6G⁻ CD11b⁺cells, macrophages/monocytes (Mø/Mo) were defined as CD11b⁺/MHCII⁺/CD11c⁻ and a mixed population of alveolar macrophages/dendritic cells (AlvMø/DCs) was defined as CD11b⁺/MHCII⁺/CD11c⁺. Expression of MHCII and CD86 (example shown) in AlvMØ/DCs was assessed by calculating the mean fluoresence intensity (MFI).



Supplementary Figure 2.

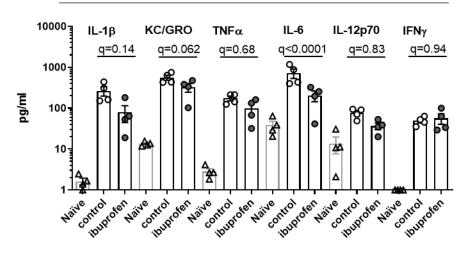
(A) Female CB6F1 mice of five to eight weeks of age were infected with ~25-50 CFU Mtb Erdman by the aerosol route. Four weeks into the infection, mice were fed with normal chow or chow containing 500ppm celecoxib. Two weeks after treatment (week 6 of the infection), the activation status of CD11b⁺CD11c^{+MHCII+} cells in the lung was assessed by staining for CD86 and MHCII (n=6). (**B**,**C**) Two weeks after treatment (week 6 of the infection) soluble proinflammatory cytokines were measured in plasma (B) or lung homogenates (C) by multiplex analysis (n=6). Bars indicate means ± SEM. p values were calculated with a students t-test and q values (adjusted p values) were calculated with an ANOVA followed by the Benjamini-Hochberg method to control the false discovery rate during multiple comparisons.



Supplementary Figure 3.

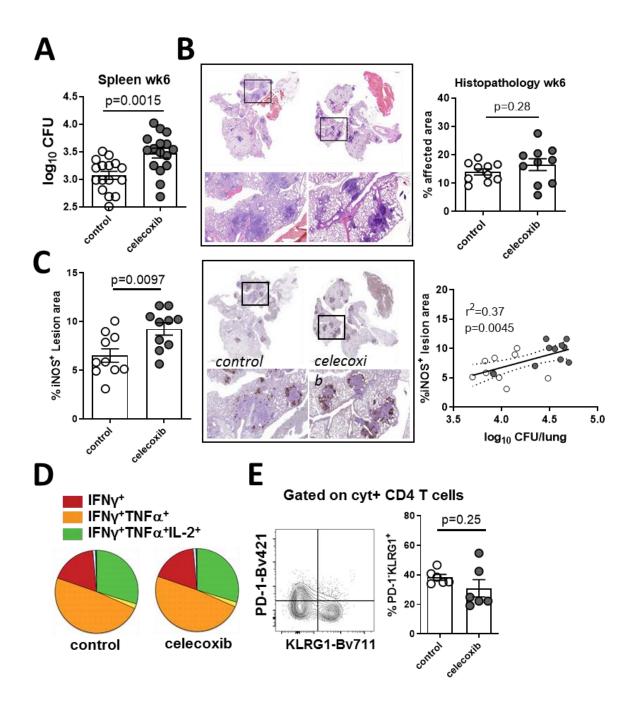
Soluble proinflammatory cytokines measured in lung homogenates of C3HeB/FeJ (n=4) by multiplex analysis one week of ibuprofen treatment (week four after i.v. infection). Bars indicate means \pm SEM. q values (adjusted p values) were calculated with an ANOVA followed by the Benjamini-Hochberg method to control the false discovery rate during multiple comparisons.

Lung (i.v. infection C3HeB/FeJ)



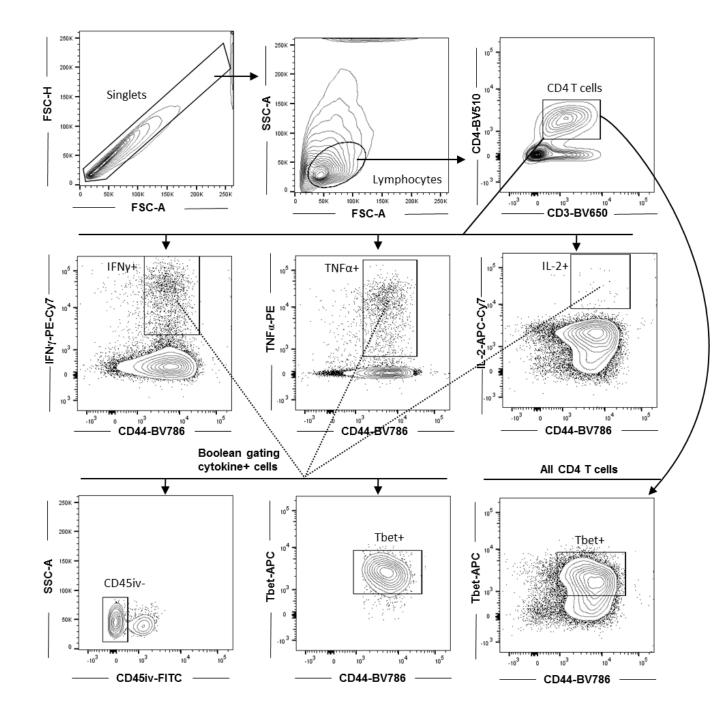
Supplementary Figure 4.

(A) CB6F1 mice were infected with ~25-50 CFU Mtb Erdman by the aerosol route, treated with celecoxib for eight weeks as previously and cleared with antibiotic treatment before Mtb reinfection (50-100 CFUs). Mtb bacilli were enumerated in the spleen by plating spleen homogenates at week six of the re-infection (n=16). (**B**) Lungs were harvested at week six (n=10) of the re-infection to investigate whether the initial celecoxib treatment induced changes in histopathology. (left) Hematoxylin and eosin stain of representative tissue sections of formalin fixed lungs. (right) percentage of lesion area (granulation lesion area/lung tissue area*100) for individual animals in each group. (C) Analysis of iNOS expression by immunohistochemistry (n=10). (left) Percentage of the iNOS⁺ area within the total lesion area for individual animals in each group, (middle) iNOS staining of representative tissue sections of formalin fixed lungs, (right) correlation between lung bacterial burden and iNOS staining showing that higher lung CFU is associated with more iNOS staining in the lesions. (D) CB6F1 mice (n=6) were infected with ~25-50 CFU Mtb Erdman by the aerosol route and treated with celecoxib after four weeks. Two weeks into the treatment, the cytokine expression profile of lung CD4 T cells stimulated by overlapping ESAT-6 peptides was assed by ICS followed by combinatorial Boolean gating (see Supplementary Figure 5). Pies indicate the proportion of cell subsets expressing different combinations of cytokines: IFN γ^+ single positive (red), IFN γ^+ TNF⁺ double positive (orange) and IFN γ^+ TNF⁺IL-2⁺ triple positive CD4 T cells (green). (E) Analysis of PD-1⁻KLRG1⁺ CD4 T cells producing any cytokine combination after stimulation with overlapping ESAT-6 peptides (n=6). Bars indicate means \pm SEM. p values were calculated with a students t-test. In the correlation plot in Supplementary Figure 3C (right) a linear regression model was used to calculate the p value and r^2 coefficient.



Supplementary Figure 5.

Example of gating strategy for intracellular cytokine staining (ICS) of stimulated CD4 T cells from Mtb infected lungs. In the "lymphocytes" gate, CD4 T cells were defined as CD3⁺CD4⁺. CD44^{hi} Cytokine expressing cells, expressing any combination of IFN γ , TNF α or IL-2, were identified via Boolean gating. Parenchymal-localized T cells were identified by gating on cells that were <u>not</u> stained by the intravenously injected CD45-FITC antibody. Tbet expression was assessed in all CD4 T cells as wells as in cytokine producing CD4 T cells.



Supplementary Figure 6.

(A) Example of gating strategy to identify $ESAT_{4-17}$ tetramer positive CD4 T cells in pooled lymph nodes and spleens from Mtb infected animals. (B) Staining for FoxP3, ROR γ t and GATA-3 in ESAT-6₄₋₁₇ positive CD4 T cells purified from pooled spleen and LNs harvested from infected WT C57BL/6 mice four weeks into celecoxib treatment.

