

Electronic Supporting Information for:

Unnatural spirocyclic oxindole alkaloids biosynthesis in *Uncaria guianensis*

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natural oxindole alkaloids

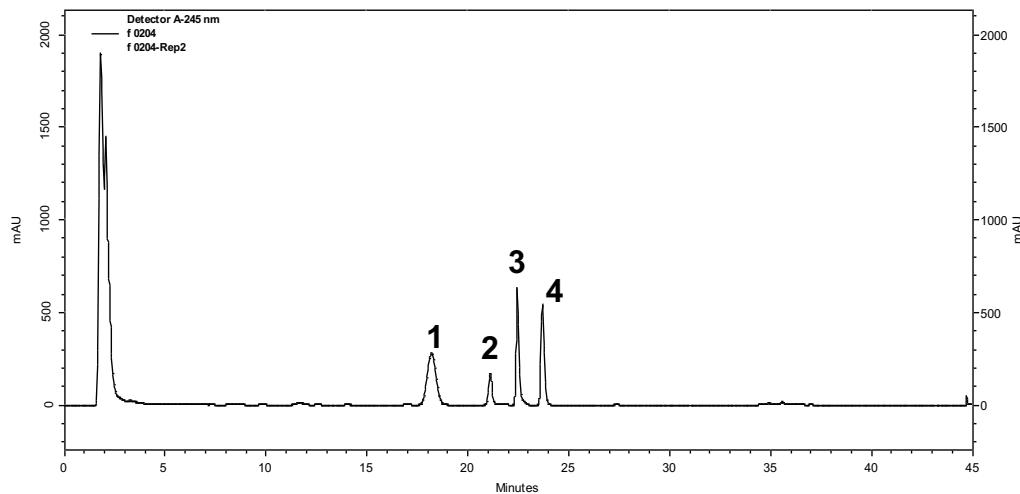
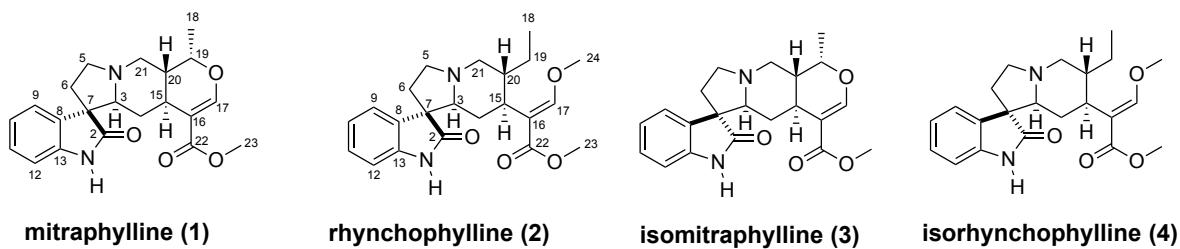


Figure S1. Chromatogram of methanolic extract from aerial parts of *U. guianensis* using HPLC conditions described in methods section.

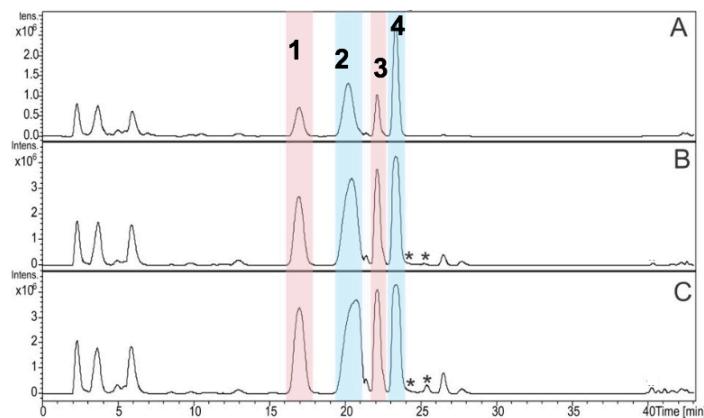


Figure S2. Chromatogram of control extract (A) and extracts obtained after supplementation with **a**-5-methyl-tryptamine (B) and **b**-7-methyl-tryptamine (C), an asterisk is used to represent unnatural oxindole alkaloids biosynthesis.

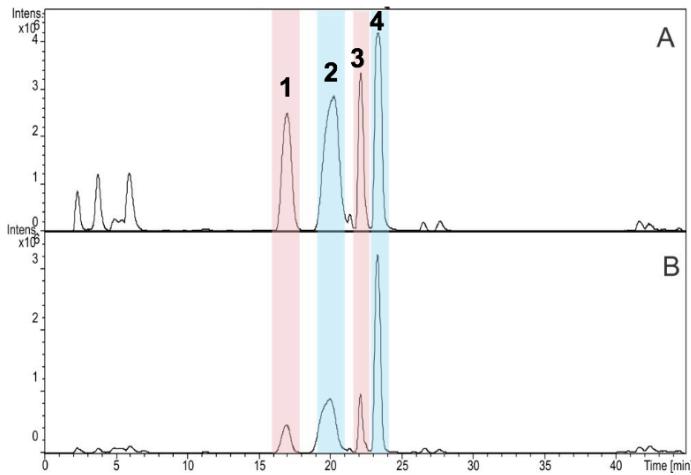


Figure S3. Chromatogram of control extract (A) and extracts obtained after supplementation with *c*-6-fluoro-tryptamine (B), the unnatural oxindole alkaloids co-eluted with natural oxindole alkaloids (**1-4**).

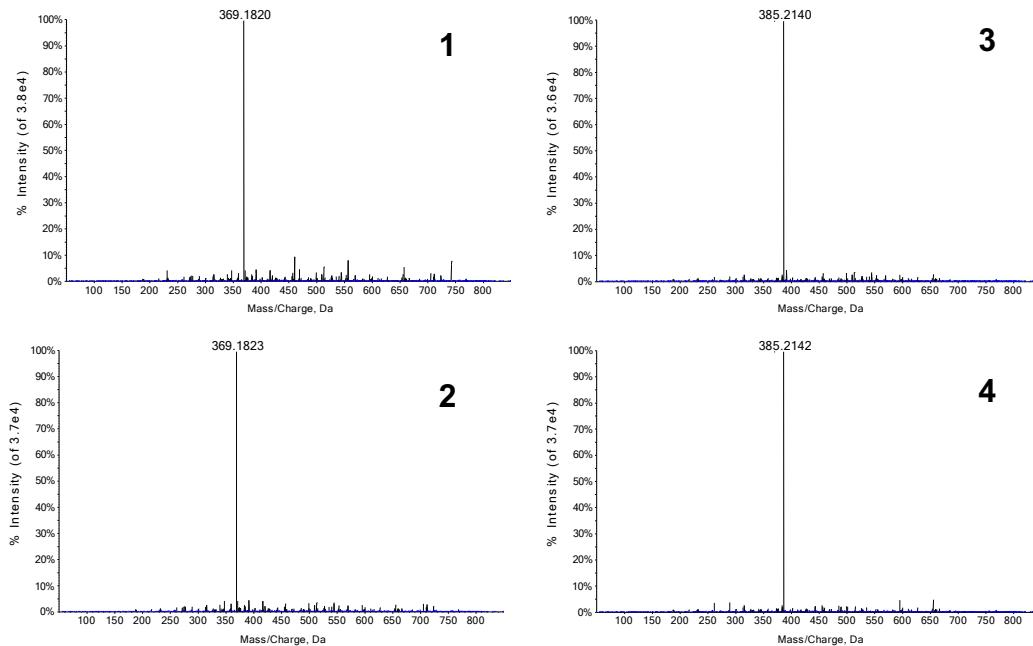


Figure S4. ESI-QTOF-MS/MS of natural oxindole alkaloids (**1-4**).

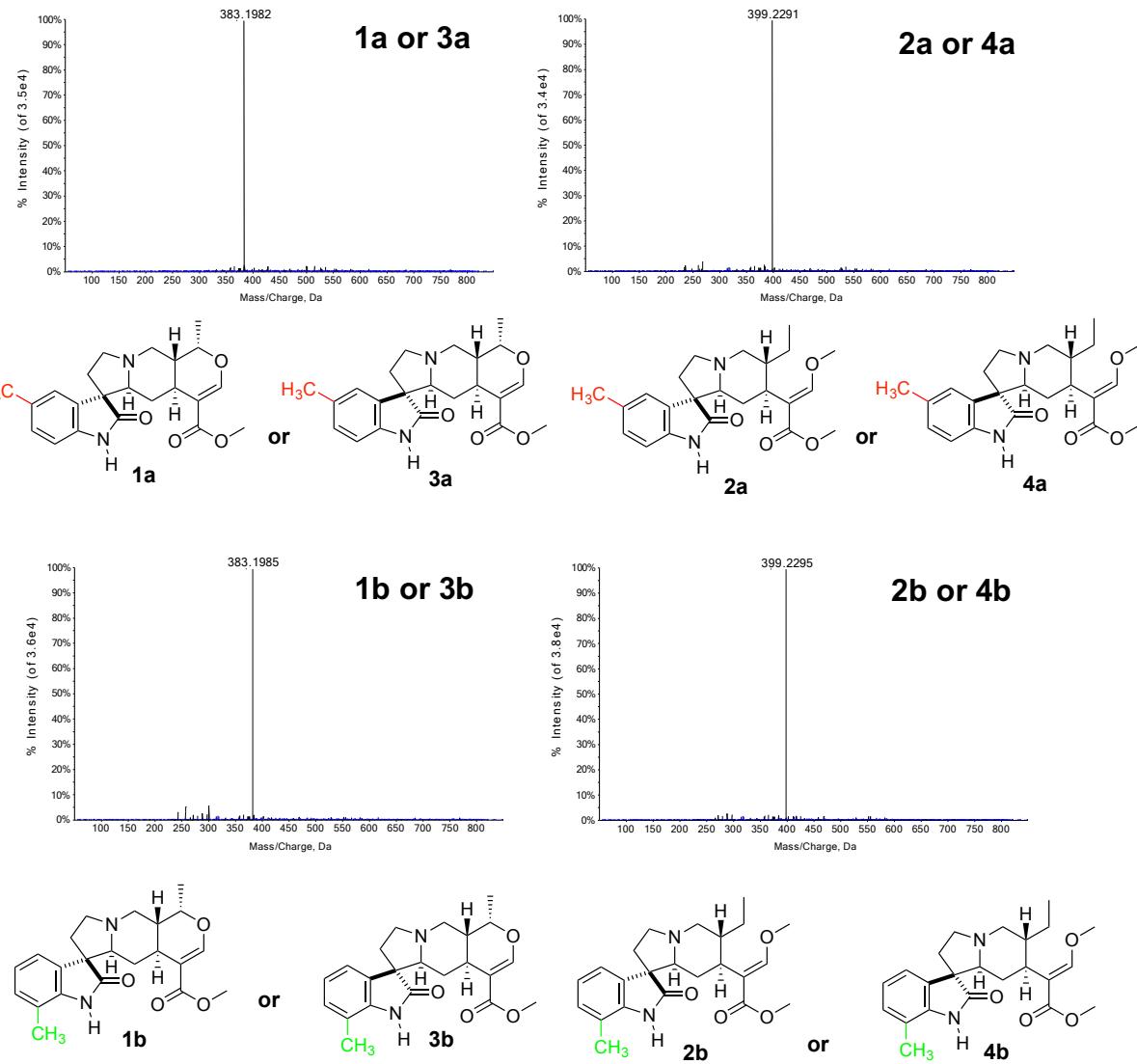


Figure S5. ESI-QTOF-MS/MS of methylated oxindole alkaloids produced by *U. guianensis* plantlets.

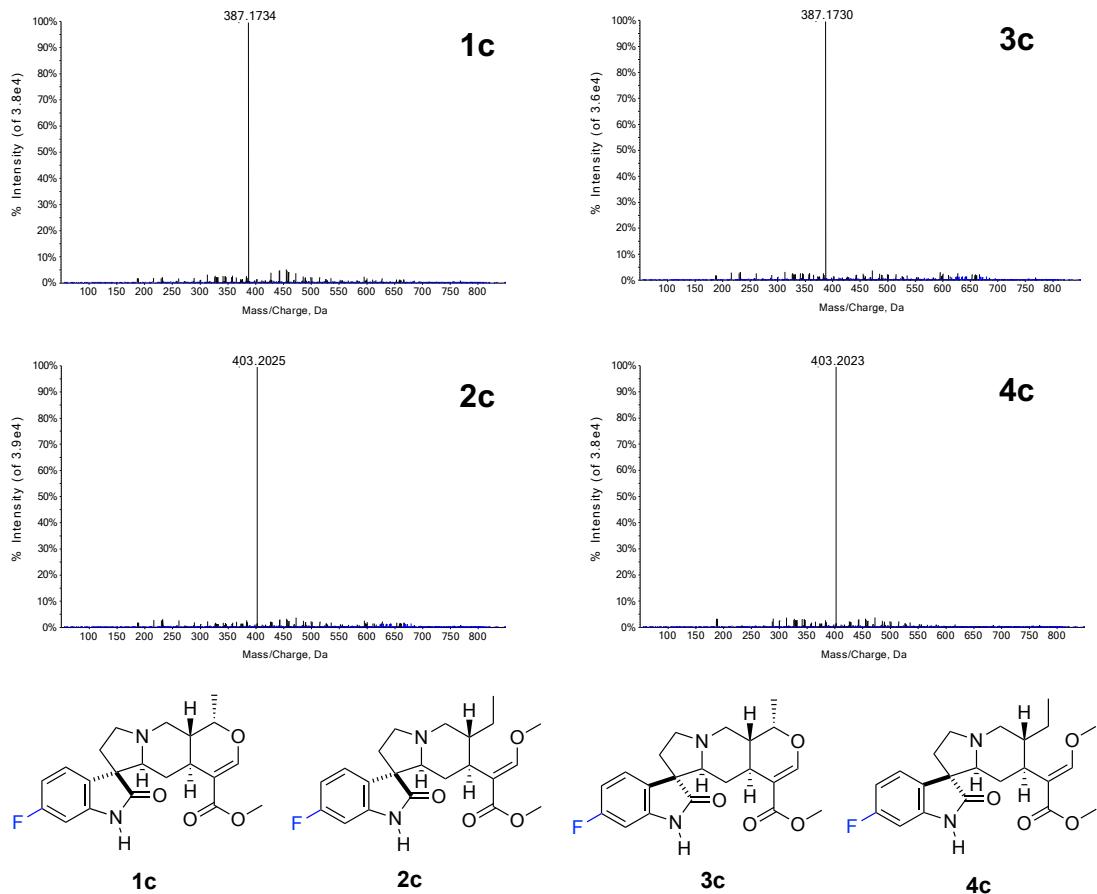


Figure S6. ESI-QTOF-MS/MS of fluorated oxindole alkaloids produced by *U. guianensis* plantlets.

Table S1. ESI-QTof-MS data of compounds.

| Compounds | Observed <i>m/z</i> | Calculated <i>m/z</i> | error (ppm) | Molecular Formula |
|---|------------------------|--------------------------|----------------|--|
| mitraphylline (1) | 369.1820 | 369.1814 | 1.6 | [C ₂₁ H ₂₄ N ₂ O ₄ +H] ⁺ |
| isomitraphylline (3) | 369.1823 | 369.1814 | 2.4 | [C ₂₁ H ₂₄ N ₂ O ₄ +H] ⁺ |
| rinchophylline (2) | 385.2140 | 385.2127 | 3.4 | [C ₂₂ H ₂₃ N ₂ O ₄ +H] ⁺ |
| isorinchophylline (4) | 385.2142 | 385.2127 | 3.9 | [C ₂₂ H ₂₃ N ₂ O ₄ +H] ⁺ |
| 5-methyl-mitraphylline/5-methyl-isomitraphylline (1a/3a) | 383.1982 | 383.1970 | 3.1 | [C ₂₂ H ₂₆ N ₂ O ₄ +H] ⁺ |
| 5-methyl-rinchophylline/5-methyl-isorinchophylline (2a/3a) | 399.2291 | 399.2284 | 1.8 | [C ₂₃ H ₃₀ N ₂ O ₄ +H] ⁺ |
| 7-methyl-mitraphylline/7-methyl-isomitraphylline (1b/3b) | 383.1985 | 383.1970 | 3.9 | [C ₂₂ H ₂₆ N ₂ O ₄ +H] ⁺ |
| 7-methyl-rinchophylline/7-methyl-isorinchophylline (2b/3b) | 399.2295 | 399.2284 | 2.8 | [C ₂₃ H ₃₀ N ₂ O ₄ +H] ⁺ |
| 6-fluor-mitraphylline (1c) | 387.1734 | 387.1720 | 3.6 | [C ₂₁ H ₂₃ FN ₂ O ₄ +H] ⁺ |
| 6-fluor-isomitraphylline (3c) | 387.1730 | 387.1720 | 2.6 | [C ₂₁ H ₂₃ FN ₂ O ₄ +H] ⁺ |
| 6-fluor-rinchophylline (2c) | 403.2025 | 403.2033 | -2.0 | [C ₂₂ H ₂₇ FN ₂ O ₄ +H] ⁺ |
| 6-fluor-isorinchophylline (4c) | 403.2023 | 403.2033 | -2.5 | [C ₂₂ H ₂₇ FN ₂ O ₄ +H] ⁺ |

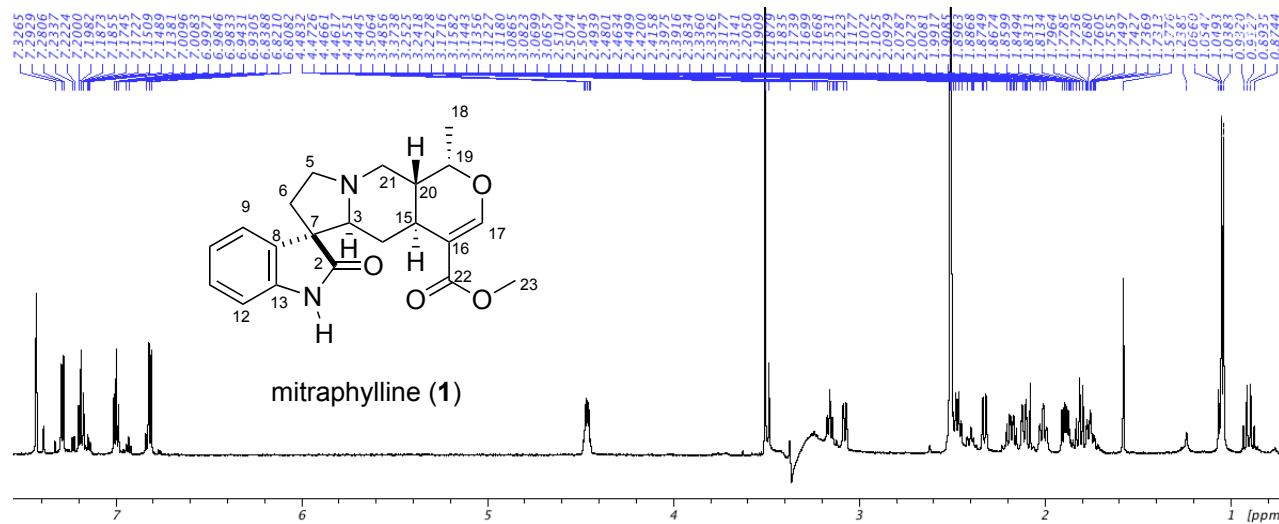


Figure S7. ^1H -NMR spectrum of **1** (600 MHz, DMSO- d_6).

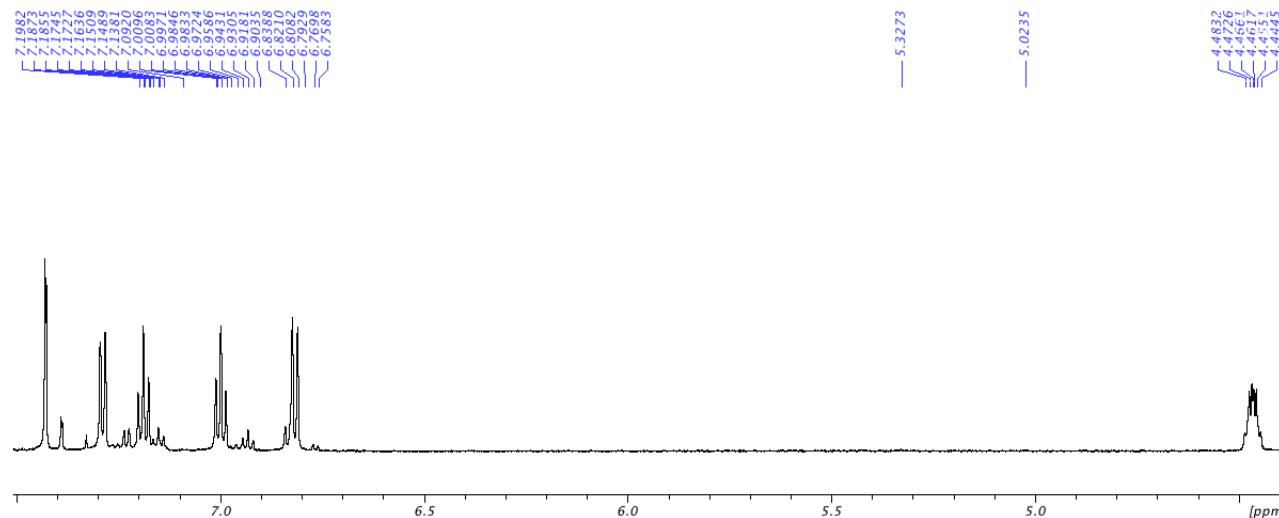


Figure S8. ^1H -NMR spectrum (expanded view) of **1** (600 MHz, DMSO- d_6).

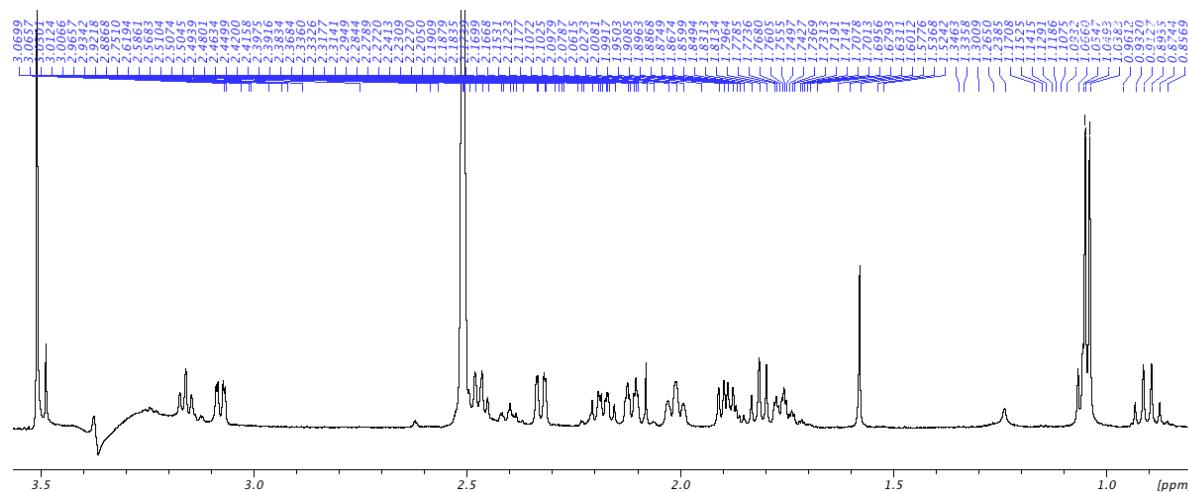


Figure S9. ^1H -NMR spectrum (expanded view) of **1** (600 MHz, DMSO- d_6).

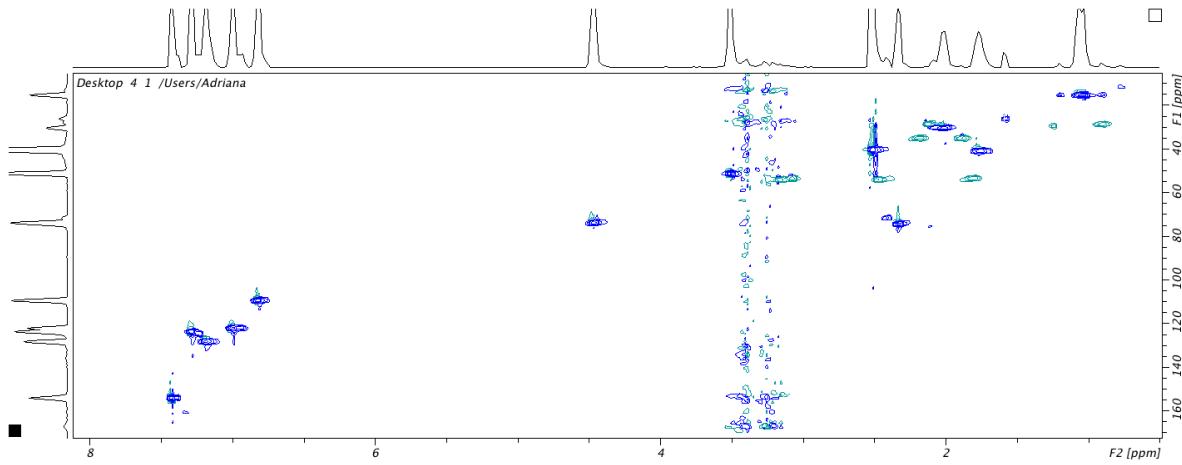


Figure S10. HSQC spectrum of **1** in DMSO-d₆ (600 MHz).

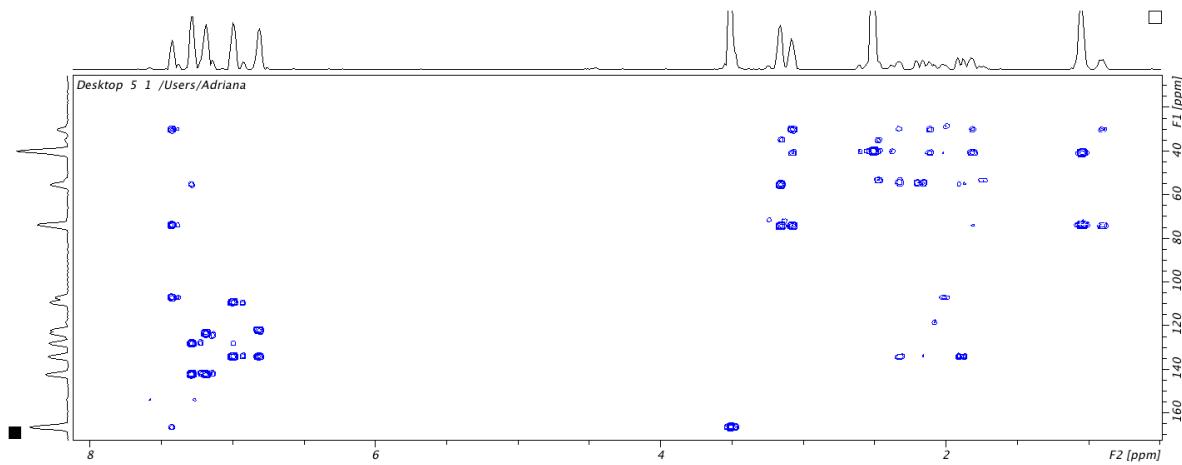


Figure S11. HMBC spectrum of **1** in DMSO-d₆ (600 MHz).

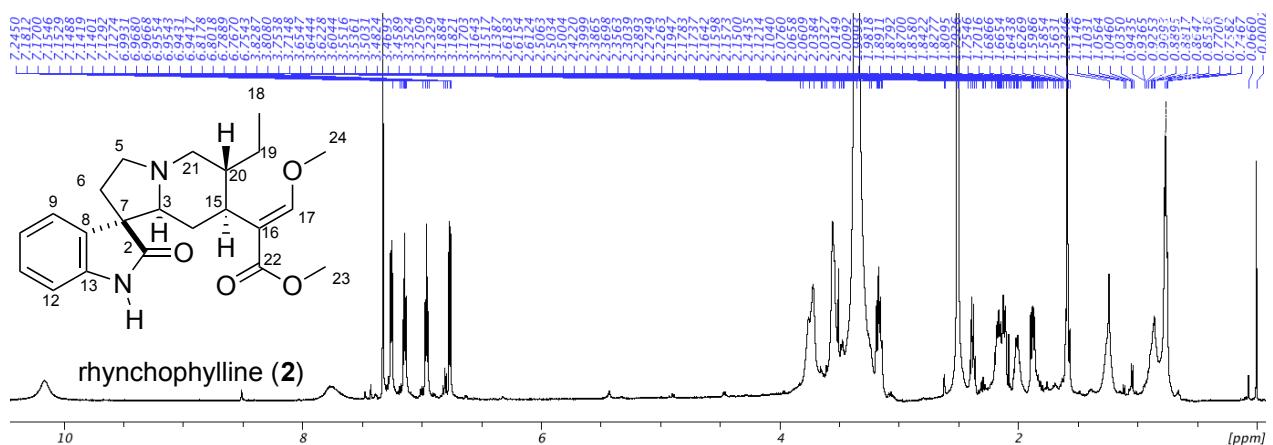


Figure S12. ¹H-NMR spectrum of **2** (600 MHz, DMSO-d₆).

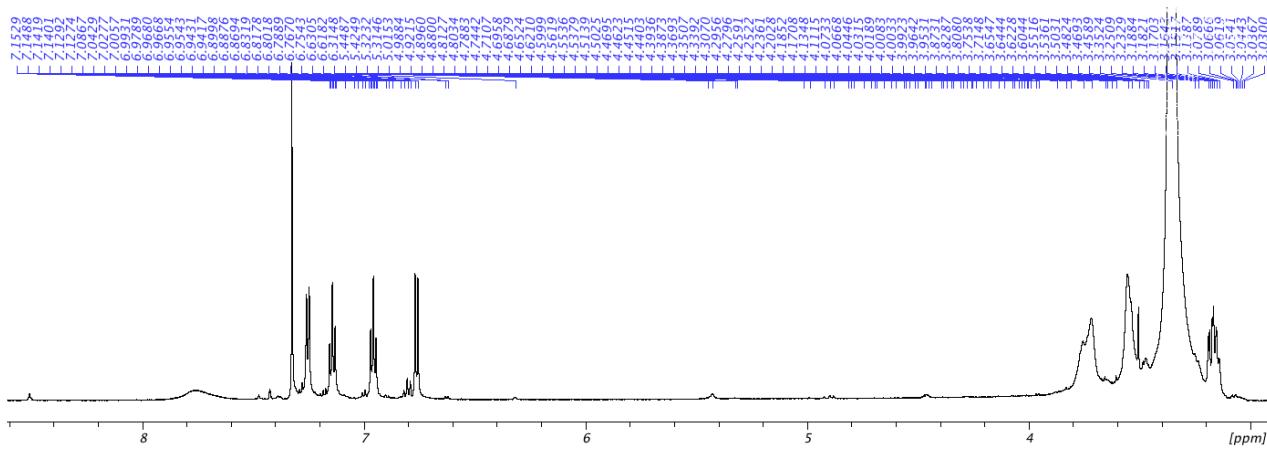


Figure S13. ^1H -NMR spectrum (expanded view) of **2** (600 MHz, DMSO-d_6).

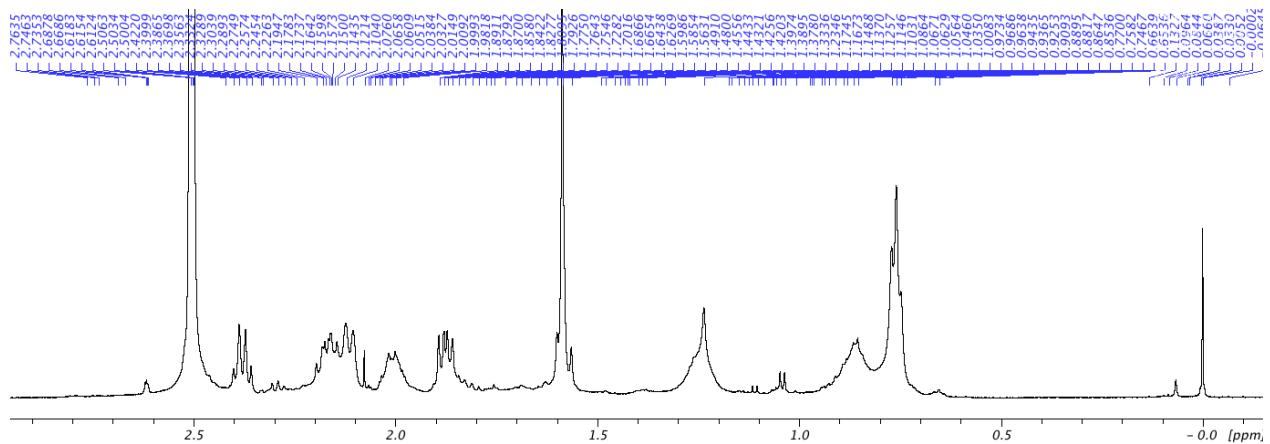


Figure S14. ^1H -NMR spectrum (expanded view) of **2** (600 MHz, DMSO-d_6).

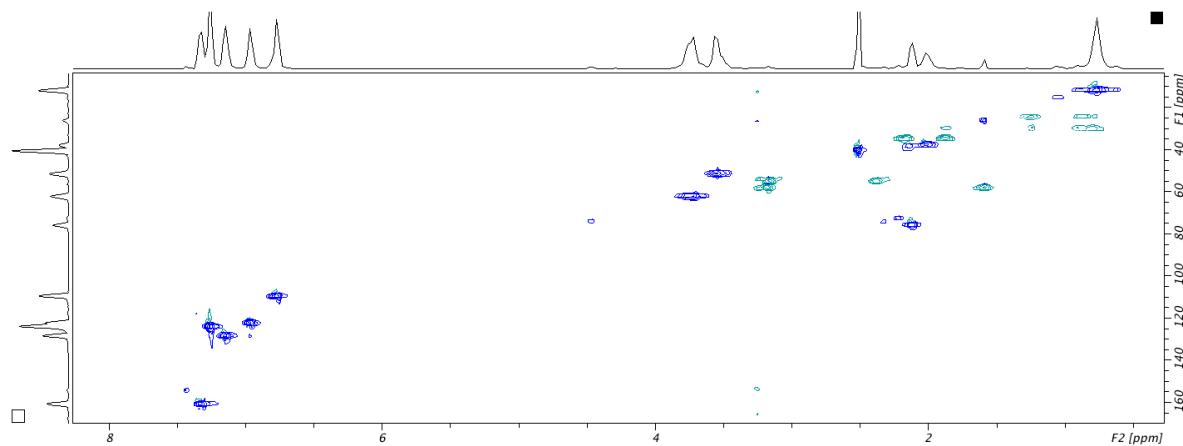


Figure S15. HSQC spectrum of **2** in DMSO-d_6 (600 MHz).

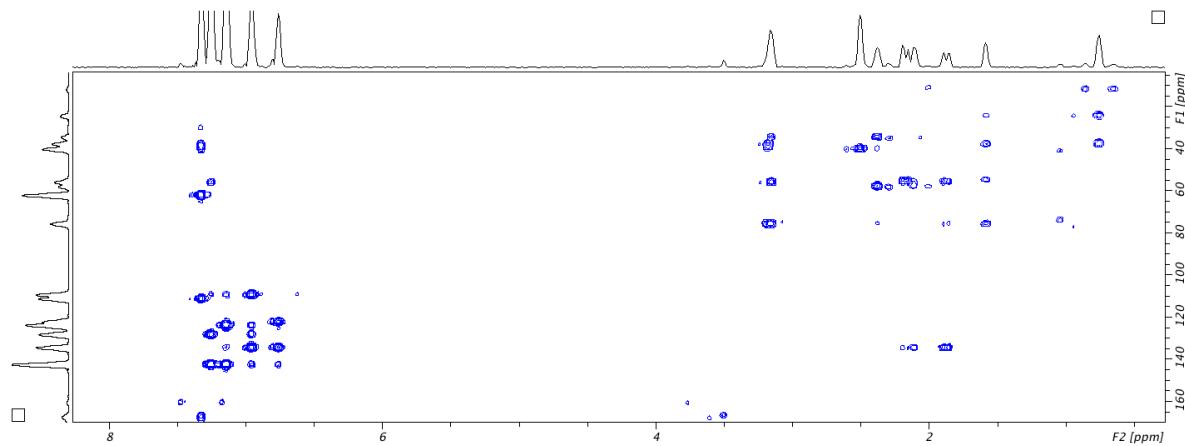


Figure S16. HMBC spectrum of **2** in DMSO-d₆ (600 MHz).

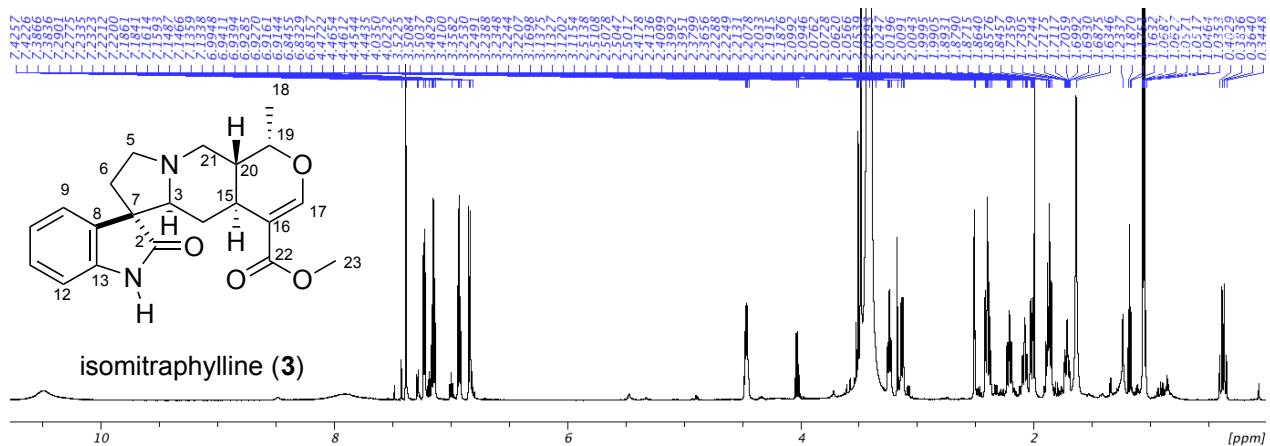


Figure S17. ¹H-NMR spectrum of **3** (600 MHz, DMSO-d₆).

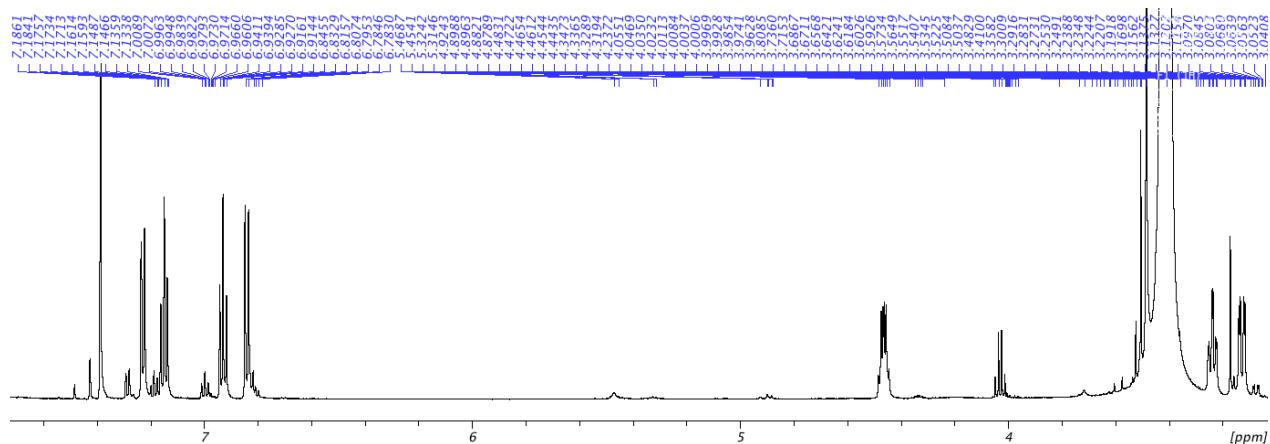


Figure S18. ¹H-NMR spectrum (expanded view) of **3** (600 MHz, DMSO-d₆).

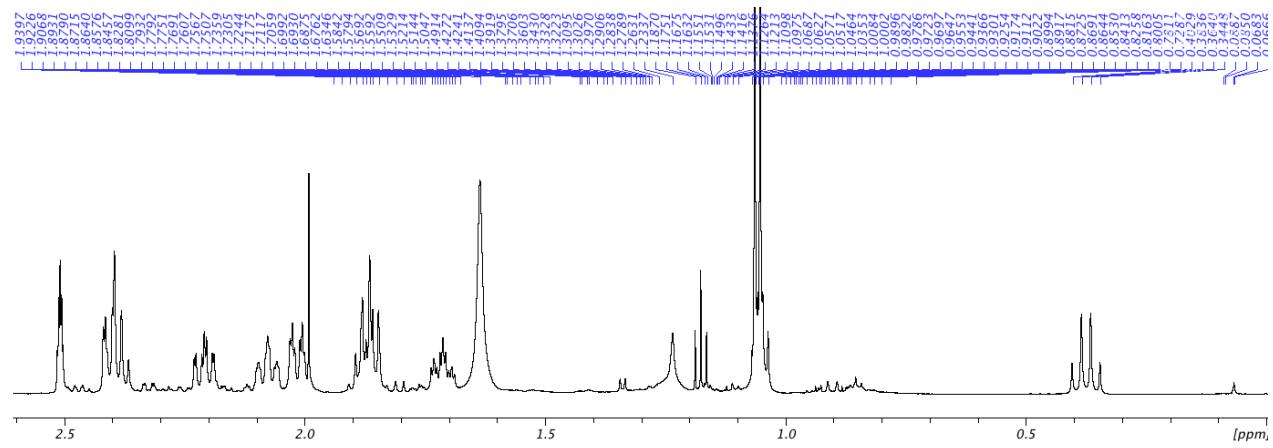


Figure S19. ^1H -NMR spectrum (expanded view) of **3** (600 MHz, DMSO-d_6).

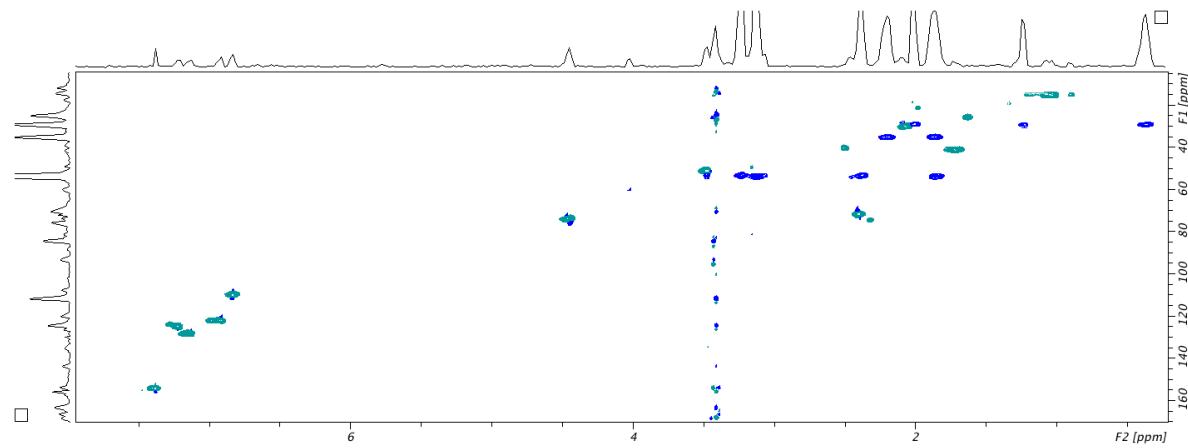


Figure S20. HSQC spectrum of **3** in DMSO-d_6 (600 MHz).

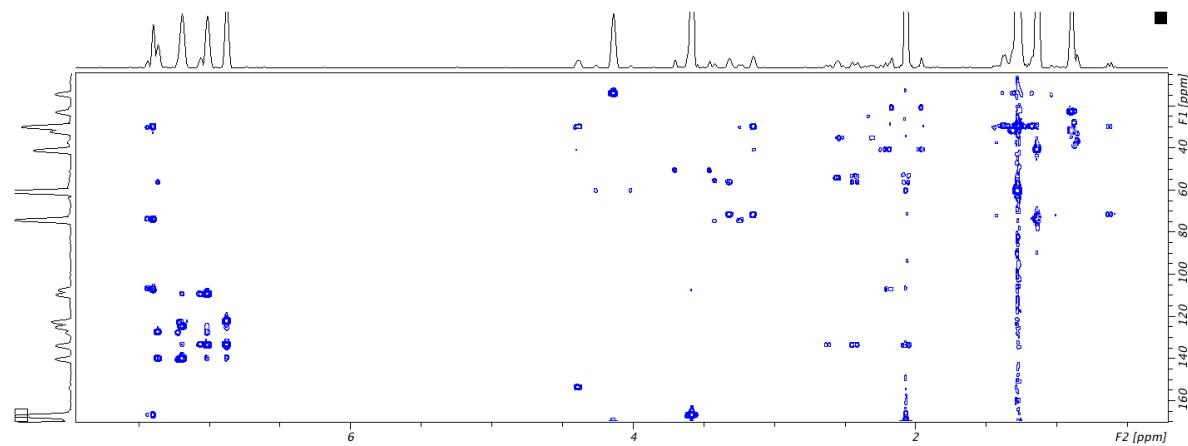


Figure S21. HMBC spectrum of **3** in DMSO-d_6 (600 MHz).

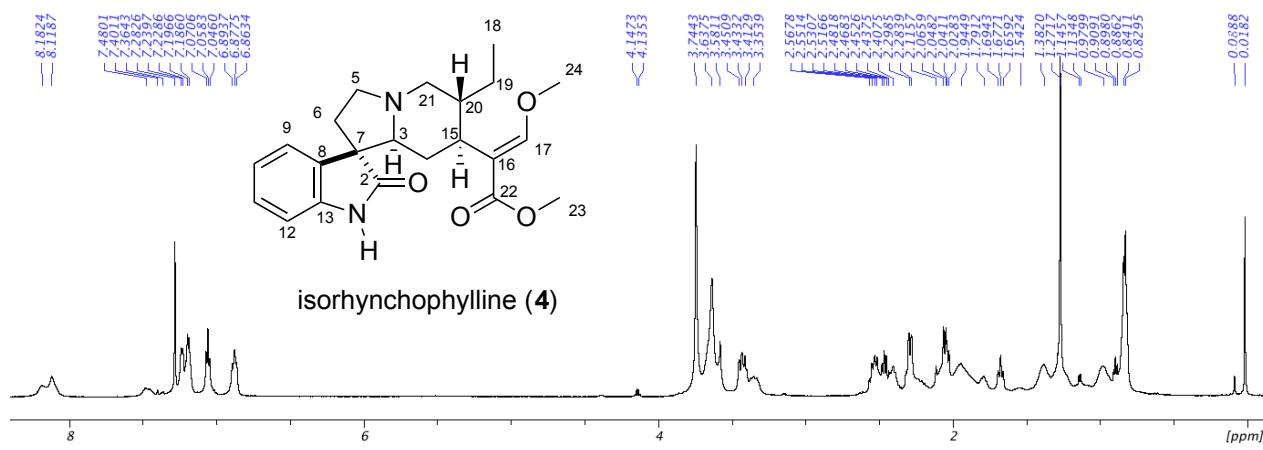


Figure S22. ^1H -NMR spectrum of **4** (600 MHz, DMSO- d_6).

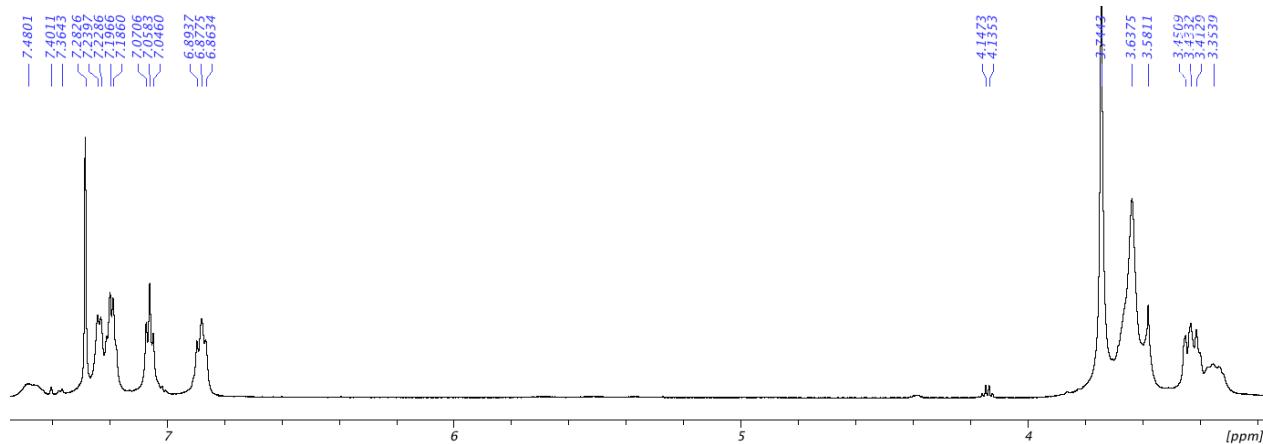


Figure S23. ^1H -NMR spectrum (expanded view) of 4 (600 MHz, DMSO- d_6).

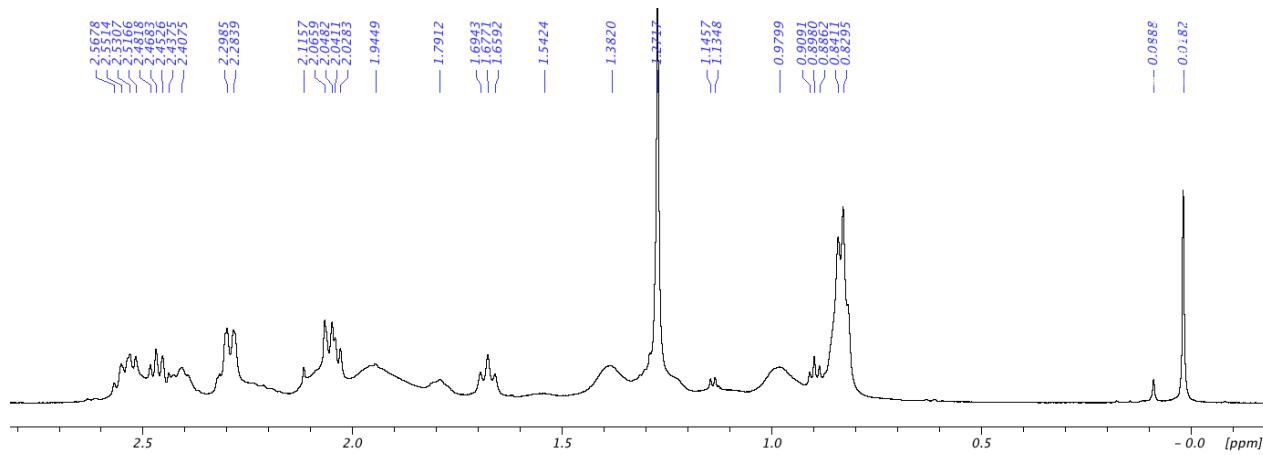


Figure S24. ^1H -NMR spectrum (expanded view) of **4** (600 MHz, DMSO- d_6).

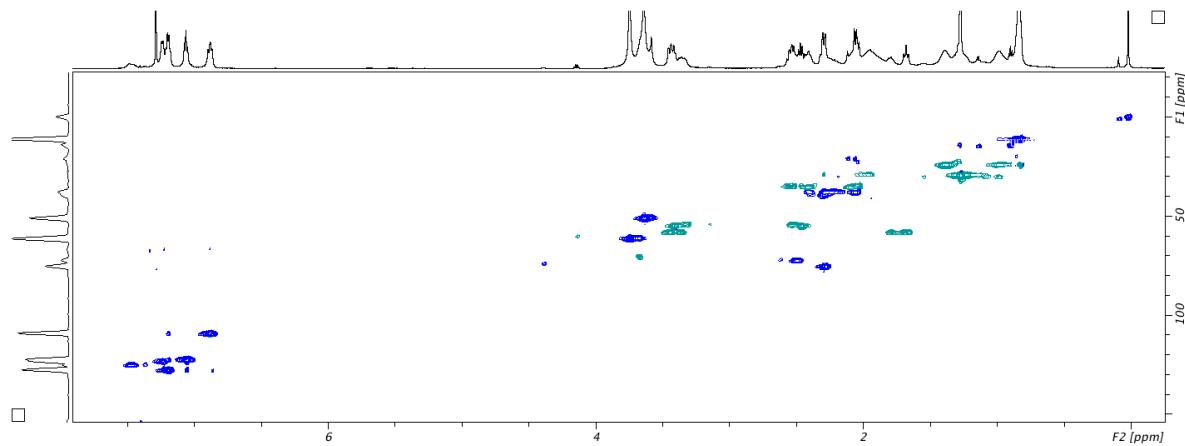


Figure S25. HSQC spectrum of **4** in DMSO-d₆ (600 MHz).

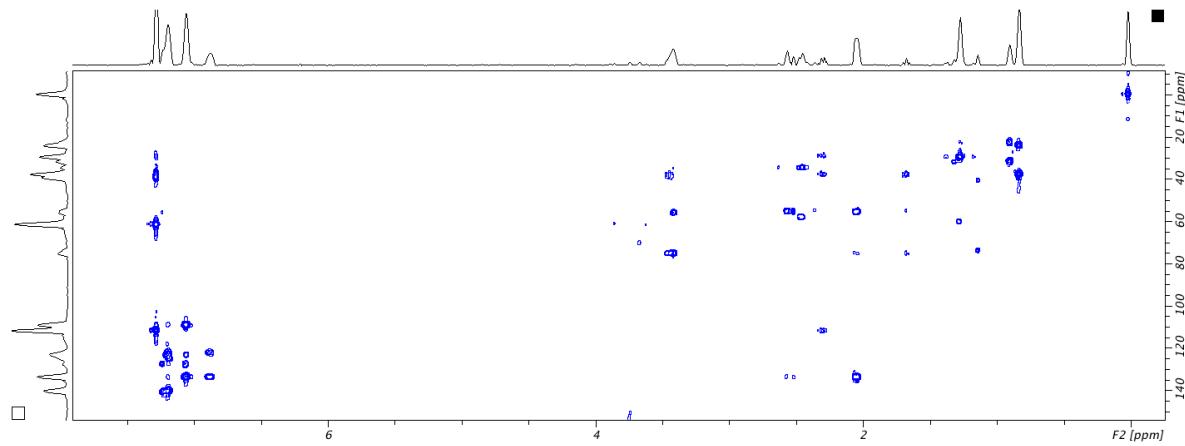


Figure S26. HMBC spectrum of **4** in DMSO-d₆ (600 MHz).

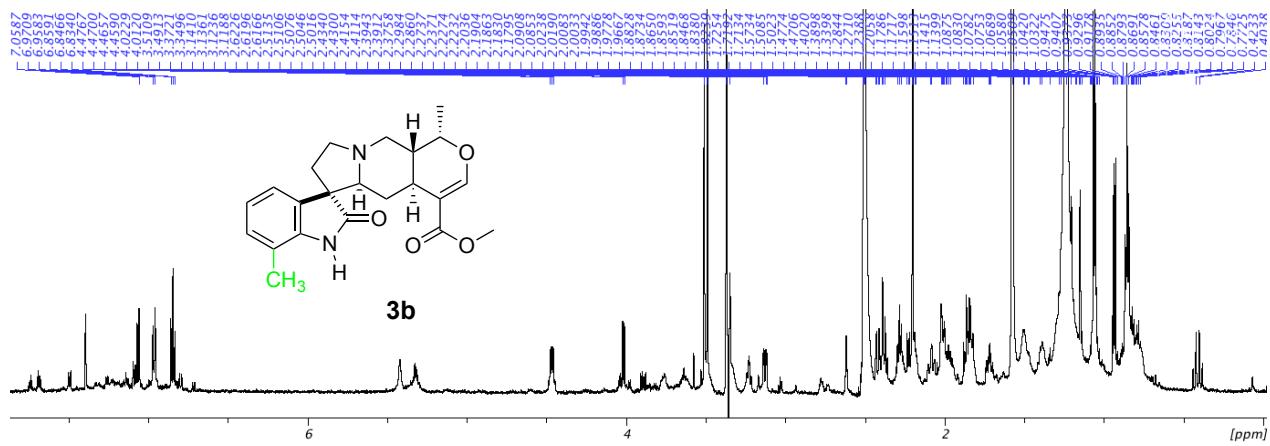


Figure S27. ^1H -NMR spectrum of **3b** (600 MHz, DMSO-d_6).

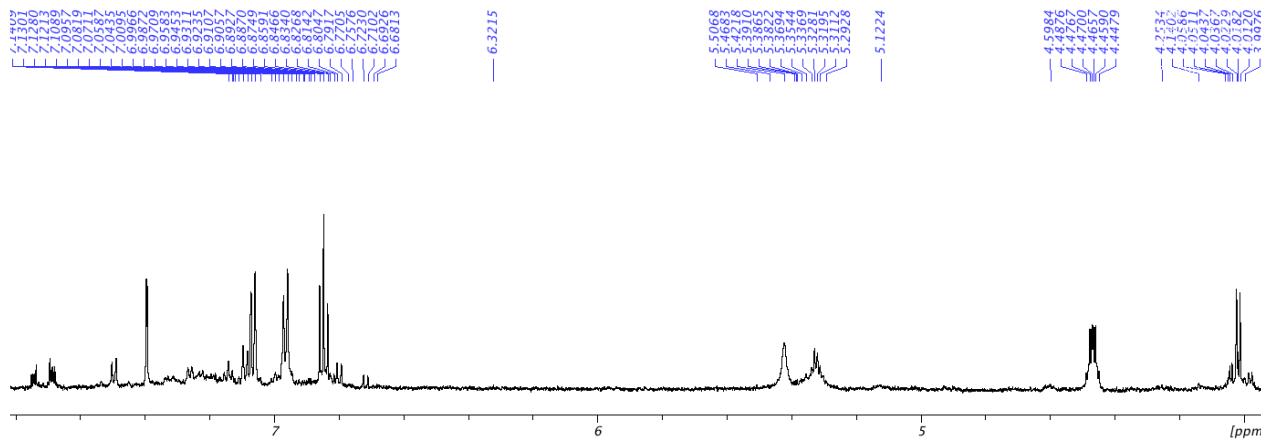


Figure S28. ^1H -NMR spectrum (expanded view) of **3b** (600 MHz, DMSO-d_6).

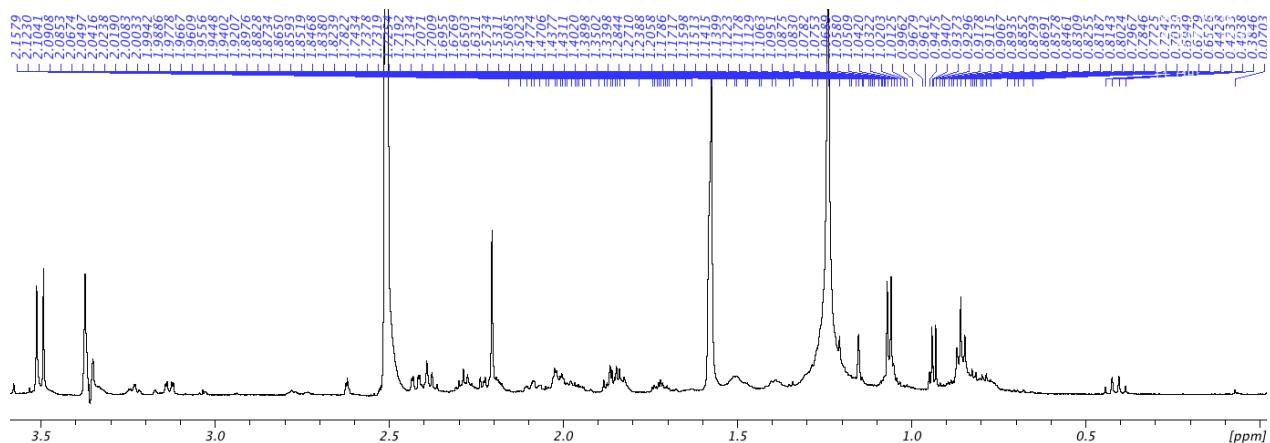


Figure S29. ^1H -NMR spectrum (expanded view) of **3b** (600 MHz, DMSO-d_6).

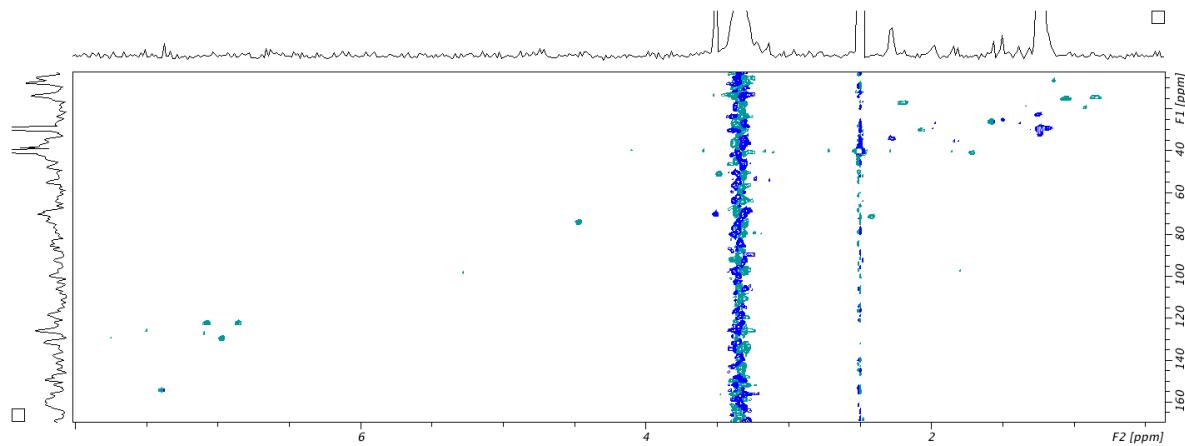


Figure S30. HSQC spectrum of **3b** in DMSO-d₆ (600 MHz).

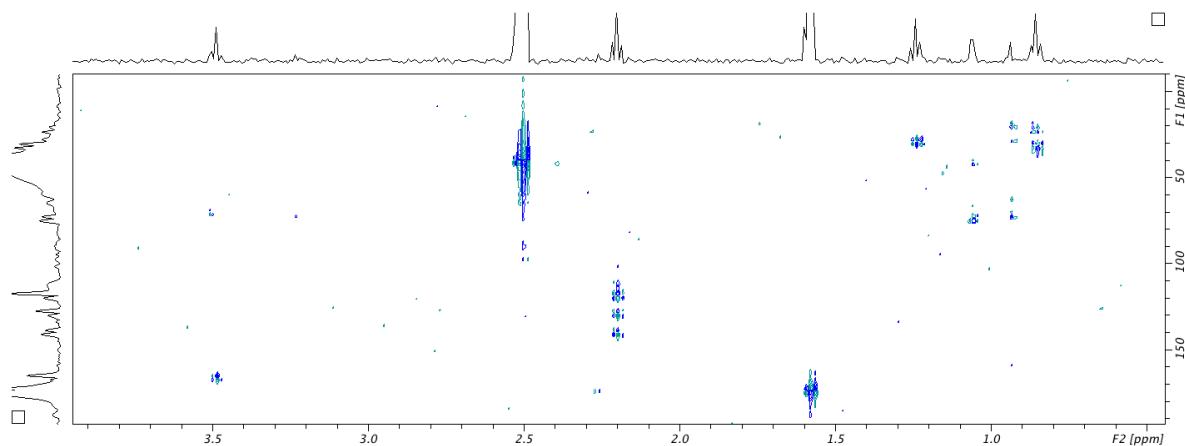


Figure S31. HMBC spectrum of **3b** in DMSO-d₆ (600 MHz).

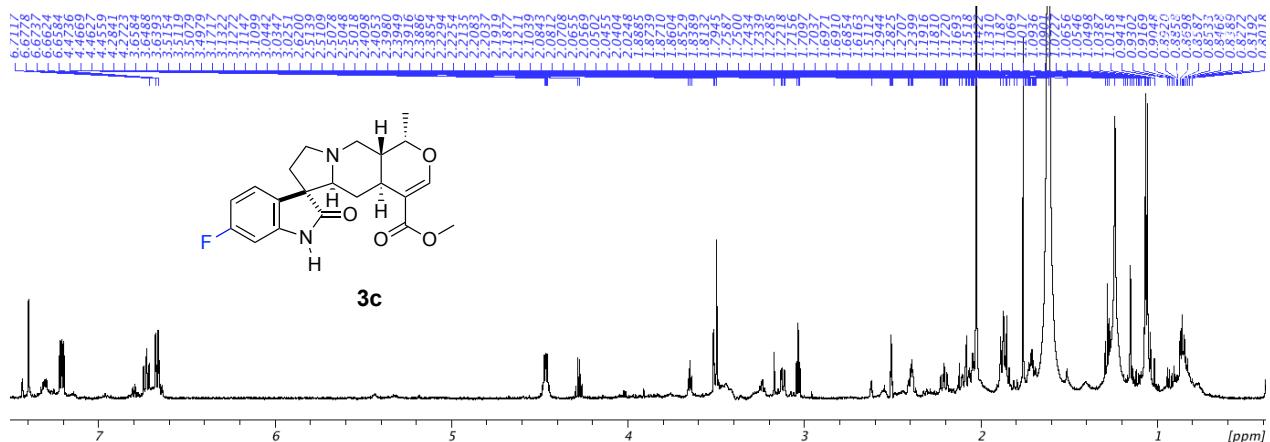


Figure S32. ¹H-NMR spectrum of **3c** (600 MHz, DMSO-d₆).

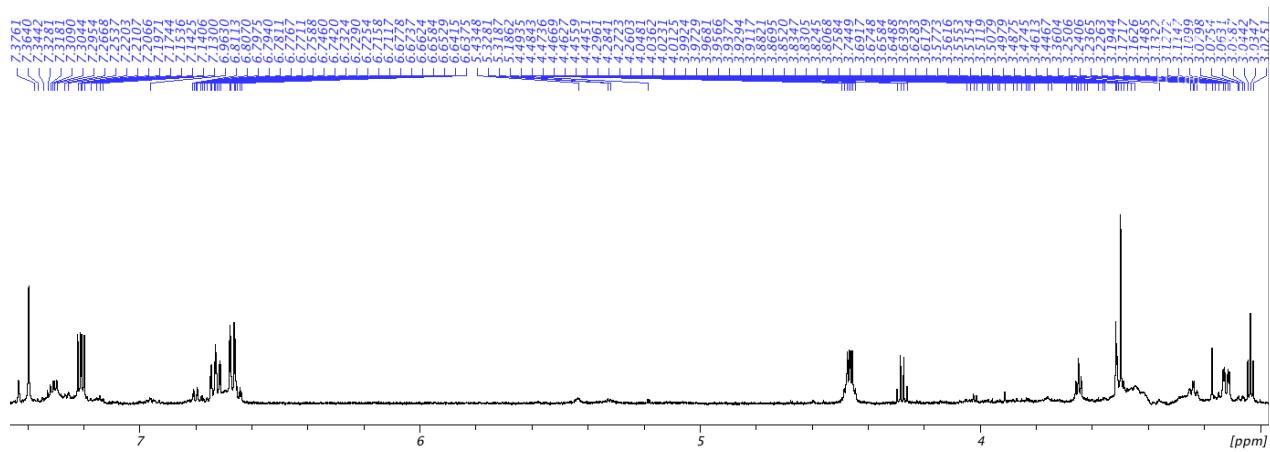


Figure S33. ^1H -NMR spectrum (expanded view) of **3c** (600 MHz, DMSO-d_6).

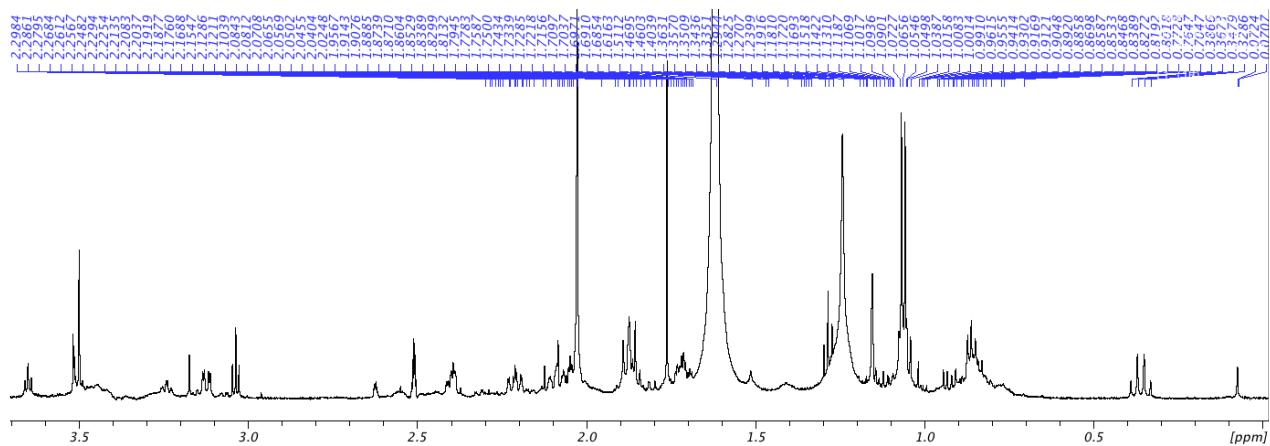


Figure S34. ^1H -NMR spectrum (expanded view) of **3c** (600 MHz, DMSO-d_6).

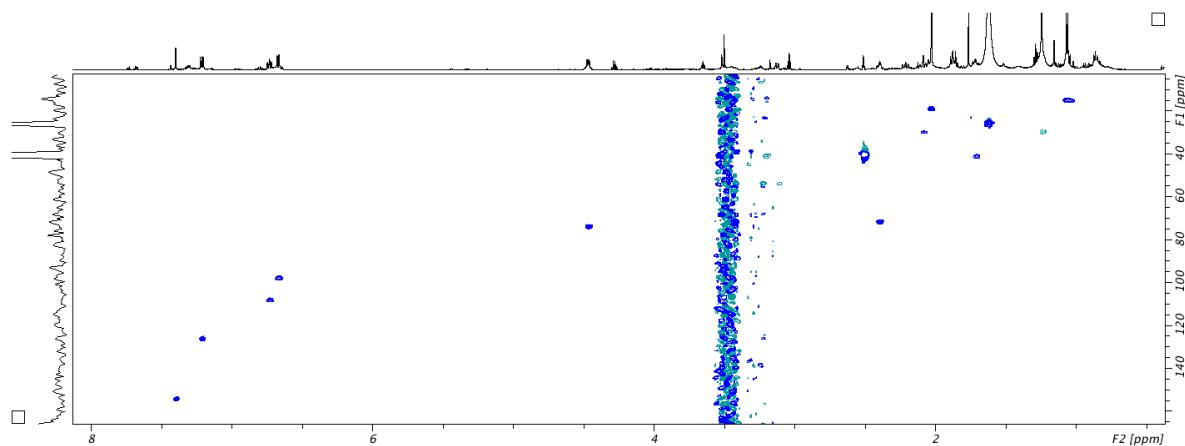


Figure S35. HSQC spectrum of **3c** in DMSO-d_6 (600 MHz).

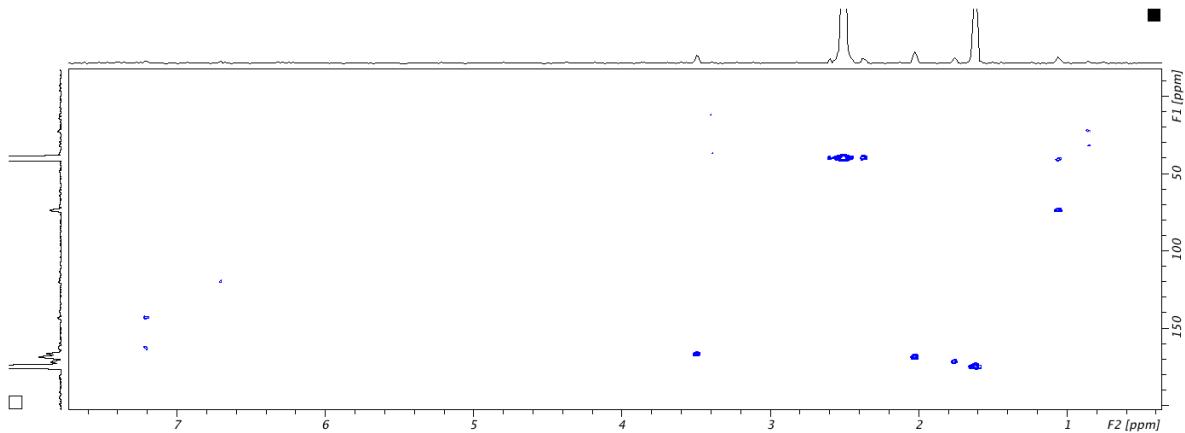


Figure S36. HMBC spectrum of **3c** in DMSO-d₆ (600 MHz).

Table S2. ^1H and ^{13}C NMR data for isomitraphylline (**3**), 7-methyl-isomitraphylline (**3b**) and 6-fluoro-isomitraphylline (**3c**) (600 MHz; DMSO-d₆)

| | 3¹ | | 3b | | 3c | |
|-----------|--|--|---------------------------------------|----------------------|---|----------------------|
| | RMN de ^1H | RMN de ^{13}C | RMN de ^1H | <i>g</i>HMQC* | RMN de ^1H | <i>g</i>HMQC* |
| 2 | - | 181.6 | - | <i>no</i> | - | <i>no</i> |
| 3 | 2.50 m | 71.6 | 2.41 m | 71.8 | 2.39 m (1H) | 71.8 |
| 5 | 2.50 m; 3.19 t (J=6.8 Hz) | 53.3 | 3.2 m (2H) | 53.5 | <i>no</i> | <i>no</i> |
| 6 | 2.0 m; 2.3 m | 35.3 | 2.28 m; 1.57 m (2H) | 34.1 | 1.85 m | 35.3 |
| 7 | - | 56.4 | - | <i>no</i> | - | 58.0 |
| 8 | - | 133.7 | - | <i>no</i> | - | 129.3 |
| 9 | 7.27 d (J=7.5 Hz) | 124.7 | 7.06 d (J= 7.5 Hz; 1H) | 122.1 | 7.21 dd (J= 8.4; 6.0 Hz; 1H) | 126.4 |
| 10 | 6.90 ddd (J= 7.5; 7.6; 0.9 Hz) | 122.2 | 6.8 t (J= 7.5 Hz; 1H) | 122.1 | 6.72 ddd (J= 10.0 ; 7.8; 2.4 Hz; 1H) | 107.8 |
| 11 | 7.09 ddd (J= 7.6; 7.7; 1.3 Hz) | 127.4 | 6.96 d (J= 7.5 Hz; 1H) | 129.3 | - | 162.6 |
| 12 | 6.81 d (J= 7.7 Hz) | 109.6 | - | 119.0 | 6.68 dd (J= 9.0 ; 2.4 Hz; 1H) | 97.9 |
| 13 | - | 140.3 | - | 140.0 | - | 142.0 |
| 14 | 0.54 q (J=11.6 Hz), 2.5 m | 29.1 | 0.93 m | 19.4 | 0.36 q (J=11.6 Hz; 1H), 2.03 m | 29.2 |
| 15 | 1.80 m; 2.10 m | 29.9 | 2.07 m (1H) | 30.7 | 2.07 m (1H) | 30.0 |
| 16 | - | 107.2 | - | <i>no</i> | - | 108.0 |
| 17 | 7.31 d (J=1.4 Hz) | 153.7 | 7.39 d (J= 1.5 Hz, 1H) | 154.2 | 7.39 d (J= 1.8 Hz, 1H) | 154.1 |
| 18 | 1.05 d (J=6.6 Hz) | 14.8 | 1.05 d (J= 6.6 Hz, 3H) | 15.1 | 1.05 d (J= 6.6 Hz, 3H) | 15.0 |
| 19 | 4.30 m | 73.8 | 4.47 m (1H) | 74.2 | 4.46 m (1H) | 73.8 |
| 20 | 1.85 m | 40.8 | 1.71 m (1H) | 41.08 | 1.71 m | 41.2 |
| 21 | 1.8 t (J=10.4 Hz); 3.10 dd (J= 10.4; 2.0 Hz) | 54.2 | 3.13 m; 1.8 m (2H) | 54.4 | 3.12 dd (J= 10.0; 2.0 Hz, 1H); 1.85 m | 53.7 |
| 22 | - | 167.0 | - | <i>no</i> | - | 166.6 |
| 23 | 3.49 s | 50.7 | 3.48 s | 51.0 | 3.49 s | <i>no</i> |
| 24 | - | - | 2.20 s (3H) | 17.2 | - | - |

no- “non-observed”

* ^{13}C data based in *g*HMQC

The coupling constant in bold is related to H-F.

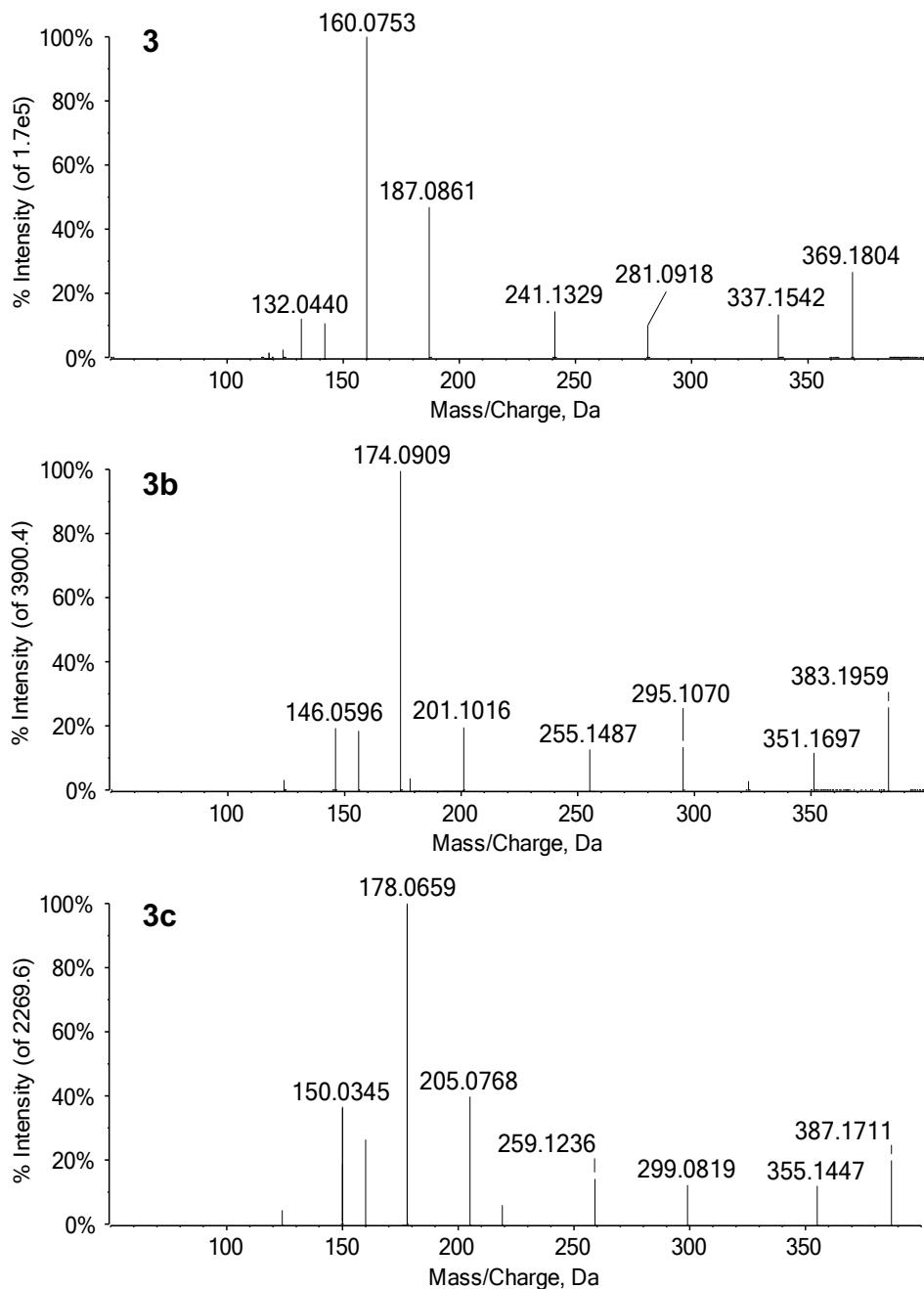


Figure S37. ESI-MS/MS spectra of **3**, **3b** and **3c** obtained on an ESI-QTOF.

Table S3. ESI-QTOF-MS/MS data of oxindole alkaloids **3**, **3b** and **3c**.

| Observed <i>m/z</i> ^{a,b} | Calculated <i>m/z</i> | Error (ppm) | Formula of the ion | Assignment |
|------------------------------------|-----------------------|-------------|---|---|
| 3 | | | | |
| 369.1804 (25) | 369.1809 | +1.4 | C ₂₁ H ₂₅ N ₂ O ₄ ⁺ | A |
| 337.1542 (13) | 337.1547 | +1.5 | C ₂₀ H ₂₁ N ₂ O ₃ ⁺ | B (A -CH ₃ OH) |
| 281.0918 (10) | 281.0921 | +1.1 | C ₁₆ H ₁₃ N ₂ O ₃ ⁺ | C (B -C ₄ H ₈) |
| 241.1329 (15) | 241.1335 | +2.5 | C ₁₅ H ₁₇ N ₂ O ⁺ | D (A -C ₆ H ₈ O ₃) |
| 187.0861 (48) | 187.0866 | +2.7 | C ₁₁ H ₁₁ N ₂ O ⁺ | E (D -C ₄ H ₆) |
| 160.0753 (100) | 160.0757 | +2.5 | C ₁₀ H ₁₀ NO ⁺ | F (E -HCN) |
| 142.0646 (11) | 142.0651 | +3.5 | C ₁₀ H ₈ N ⁺ | G (F -H ₂ O) |
| 132.0440 (13) | 132.0444 | +3.0 | C ₈ H ₆ NO ⁺ | H (E -C ₂ H ₄ -HCN) |
| 3b | | | | |
| 383.1959 (26) | 383.1965 | +1.6 | C ₂₂ H ₂₇ N ₂ O ₄ ⁺ | A |
| 351.1697 (11) | 351.1703 | +1.7 | C ₂₁ H ₂₃ N ₂ O ₃ ⁺ | B (A -CH ₃ OH) |
| 295.1070 (14) | 295.1077 | +2.4 | C ₁₇ H ₁₅ N ₂ O ₃ ⁺ | C (B -C ₄ H ₈) |
| 255.1487 (13) | 255.1492 | +2.0 | C ₁₆ H ₁₉ N ₂ O ⁺ | D (A -C ₆ H ₈ O ₃) |
| 201.1016 (20) | 201.1022 | +3.0 | C ₁₂ H ₁₃ N ₂ O ⁺ | E (D -C ₄ H ₆) |
| 174.0909 (100) | 174.0913 | +2.3 | C ₁₁ H ₁₂ NO ⁺ | F (E -HCN) |
| 156.0803 (18) | 156.0808 | +3.2 | C ₁₁ H ₁₀ N ⁺ | G (F -H ₂ O) |
| 146.0596 (20) | 146.0600 | +2.7 | C ₉ H ₈ NO ⁺ | H (E -C ₂ H ₄ -HCN) |
| 3c | | | | |
| 387.1711 (20) | 387.1715 | +1.0 | C ₂₁ H ₂₄ FN ₂ O ₄ ⁺ | A |
| 355.1447 (13) | 355.1452 | +1.4 | C ₂₀ H ₂₀ FN ₂ O ₃ ⁺ | B (A -CH ₃ OH) |
| 299.0819 (14) | 299.0826 | +2.3 | C ₁₆ H ₁₂ FN ₂ O ₃ ⁺ | C (B -C ₄ H ₈) |
| 259.1236 (15) | 259.1241 | +1.9 | C ₁₅ H ₁₆ FN ₂ O ⁺ | D (A -C ₆ H ₈ O ₃) |
| 205.0768 (41) | 205.0772 | +2.0 | C ₁₁ H ₁₀ FN ₂ O ⁺ | E (D -C ₄ H ₆) |
| 178.0659 (100) | 178.0663 | +2.2 | C ₁₀ H ₉ FNO ⁺ | F (E -HCN) |
| 160.0553 (26) | 160.0557 | +2.5 | C ₁₀ H ₇ FN ⁺ | G (F -H ₂ O) |
| 150.0345 (38) | 150.0350 | +3.3 | C ₈ H ₅ FNO ⁺ | H (E -C ₂ H ₄ -HCN) |

^aRelative intensities are provided in parentheses.^bOnly ions with relative intensities higher than 5% are reported.

Table S4. ^{13}C NMR data of mitraphylline (**1**) isolated from *U. guianensis* shoots after incorporation of 1- ^{13}C -D-glucose into cultures (DMSO-d₆).

| C | δ_{c} | δ_{c}^* | Relative intensity of signal | | |
|---------------------------|---------------------|-----------------------|------------------------------|----------|------------------|
| | | | L | U | ΔC |
| 2 | 180.5 | 180.9 | 1.5 | 1.4 | 1.18 |
| 3 | 74.1 | 74.6 | 1.8 | 2.3 | 0 |
| 5 | 53.3 | 54.3 | 2.1 | 1.9 | 1.21 |
| 6 | 34.7 | 35.1 | 2.8 | 1.9 | 1.62 |
| 7 | 55.4 | 55.5 | 2.0 | 2.6 | 0 |
| 8 | 134.0 | 133.3 | 0.9 | 0.9 | 0 |
| 9 | 123.6 | 123.0 | 2.5 | 1.3 | 2.11 |
| 10 | 122.3 | 122.6 | 1.1 | 1.3 | 0 |
| 11 | 128.2 | 128.0 | 1.7 | 1.5 | 1.24 |
| 12 | 109.5 | 109.5 | 1.5 | 1.8 | 0 |
| 13 | 142.1 | 140.6 | 1.3 | 0.6 | 2.38 |
| 14 | 28.4 | 28.3 | 2.1 | 1.6 | 1.44 |
| 15 | 30.1 | 30.4 | 1.6 | 2.4 | 0 |
| 16 | 106.9 | 106.9 | 1.0 | 1.6 | 0 |
| 17 | 154.2 | 154.0 | 2.1 | 1.6 | 1.44 |
| 18 | 15.1 | 14.8 | 5.1 | 3.4 | 1.65 |
| 19 | 73.8 | 73.8 | 1.9 | 2.4 | 0 |
| 20 | 40.6 | 40.5 | 1.7 | 2.1 | 0 |
| 21 | 53.8 | 54.3 | 2.0 | 1.7 | 1.30 |
| 22 | 166.7 | 167.1 | 1.5 | 1.9 | 0 |
| 23-OCH₃ | 51.1 | 50.7 | 5.3 | 3.3 | 1.77 |

* literature¹

U: control experiments with unlabeled precursor; L: labeling experiment with ^{13}C precursor;
 $\Delta\text{C} = 1.1\% \times \text{L}/\text{U}$: increase in the relative intensity (significant increases in bold for enriched carbons).

Table S5. ^{13}C NMR data of isomitraphylline (**3**) isolated from *U. guianensis* shoots after incorporation of 1- ^{13}C -D-glucose into cultures (DMSO-d₆).

| C | δ_{c} | δ_{c}^* | Relative intensity of signal | | |
|---------------------------|---------------------|-----------------------|------------------------------|----------|------------------|
| | | | L | U | ΔC |
| 2 | 179.9 | 181.6 | 1.6 | 1.5 | 1.17 |
| 3 | 71.1 | 71.6 | 2.1 | 2.2 | 0 |
| 5 | 52.7 | 53.3 | 2.2 | 1.9 | 1.27 |
| 6 | 34.8 | 35.3 | 3.9 | 1.9 | 2.26 |
| 7 | 55.7 | 56.4 | 2.4 | 2.2 | 1.20 |
| 8 | 133.6 | 133.7 | 0.8 | 1.2 | 0 |
| 9 | 124.3 | 124.7 | 2.9 | 1.5 | 2.13 |
| 10 | 121.5 | 122.2 | 1.4 | 1.6 | 0 |
| 11 | 127.6 | 127.4 | 1.9 | 1.5 | 1.39 |
| 12 | 109.3 | 109.6 | 1.9 | 1.7 | 1.22 |
| 13 | 141.5 | 140.3 | 1.7 | 1.1 | 1.70 |
| 14 | 28.7 | 29.1 | 2.6 | 1.7 | 1.68 |
| 15 | 29.4 | 29.9 | 2.0 | 2.1 | 1.05 |
| 16 | 106.7 | 107.2 | 1.2 | 1.5 | 0 |
| 17 | 153.7 | 153.7 | 2.2 | 1.5 | 1.61 |
| 18 | 14.7 | 14.8 | 5.7 | 3.2 | 1.96 |
| 19 | 73.3 | 73.8 | 2.4 | 2.4 | 0 |
| 20 | 40.6 | 40.8 | 2.0 | 2.2 | 0 |
| 21 | 53.4 | 54.2 | 2.7 | 1.6 | 1.86 |
| 22 | 166.1 | 167.0 | 1.5 | 1.5 | 0 |
| 23-OCH₃ | 50.7 | 50.7 | 5.4 | 3.2 | 1.86 |

* literature¹

U: control experiments with unlabeled precursor; L: labeling experiment with ^{13}C precursor;
 $\Delta\text{C} = 1.1\% \times \text{L}/\text{U}$: increase in the relative intensity (significant increases in bold for enriched carbons).

Table S6. ^{13}C NMR data of isorhynchophylline (**4**) isolated from *U. guianensis* shoots after incorporation of 1- ^{13}C -D-glucose into cultures (DMSO-d₆).

| C | δ_c | δ_c^* | Relative intensity of signal | | |
|----------------------------|------------|--------------|------------------------------|-----|------------|
| | | | L | U | ΔC |
| 2 | 180.2 | 182,4 | 2.5 | 2.3 | 1.19 |
| 3 | 72.0 | 72.2 | 1.0 | 1.0 | 0 |
| 5 | 53.5 | 54.2 | 4.5 | 4.5 | 0 |
| 6 | 34.7 | 36.5 | 2.0 | 0.9 | 2.4 |
| 7 | 56.0 | 57.0 | 4.5 | 6.4 | 0 |
| 8 | 133.8 | 134.2 | 0.9 | 1.1 | 0 |
| 9 | 124.4 | 125.2 | 5.8 | 3.9 | 1.6 |
| 10 | 121.4 | 122.1 | 0.7 | 0.7 | 0 |
| 11 | 127.4 | 127.1 | 4.0 | 4.0 | 0 |
| 12 | 109.0 | 109.6 | 3.5 | 4.2 | 0 |
| 13 | 141.3 | 140.7 | 2.0 | 2.0 | 0 |
| 14 | 30.0 | 30.1 | 1.1 | 0.5 | 2.4 |
| 15 | 37.3 | 38.3 | 1.7 | 1.7 | 0 |
| 16 | 110.8 | 113.0 | 1.0 | 1.1 | 0 |
| 17 | 160.2 | 159.5 | 1.0 | 0.7 | 1.6 |
| 18 | 11.0 | 11.2 | 2.2 | 1.5 | 1.6 |
| 19 | 23.8 | 24.3 | 3.4 | 4.1 | 0 |
| 20 | 37.3 | 38.3 | 1.7 | 1.7 | 0 |
| 21 | 57.7 | 58.2 | 1.7 | 1.1 | 1.7 |
| 22 | 166.6 | 168.4 | 0,3 | 0.3 | 0 |
| 23-OCH₃ | 50.4 | 50.9 | 1.6 | 0.7 | 2.5 |
| 24- OCH₃ | 61.3 | 61.2 | 1.7 | 0.7 | 2.7 |

* literature¹

U: control experiments with unlabeled precursor; L: labeling experiment with ^{13}C precursor;
 $\Delta C = 1.1\% \times L/U$: increase in the relative intensity (significant increases in bold for enriched carbons).

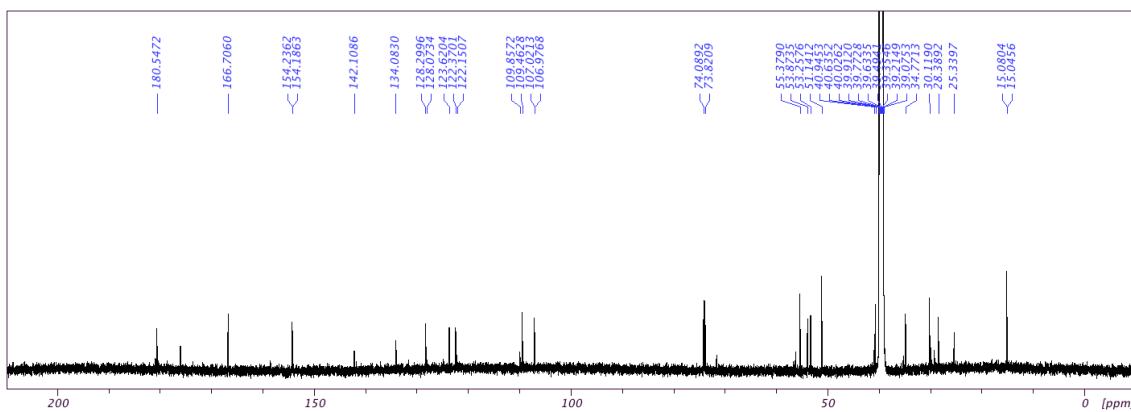


Figure S38. ^{13}C NMR (150 MHz, DMSO-d₆) spectrum of **1** with natural ^{13}C -isotopic abundance.

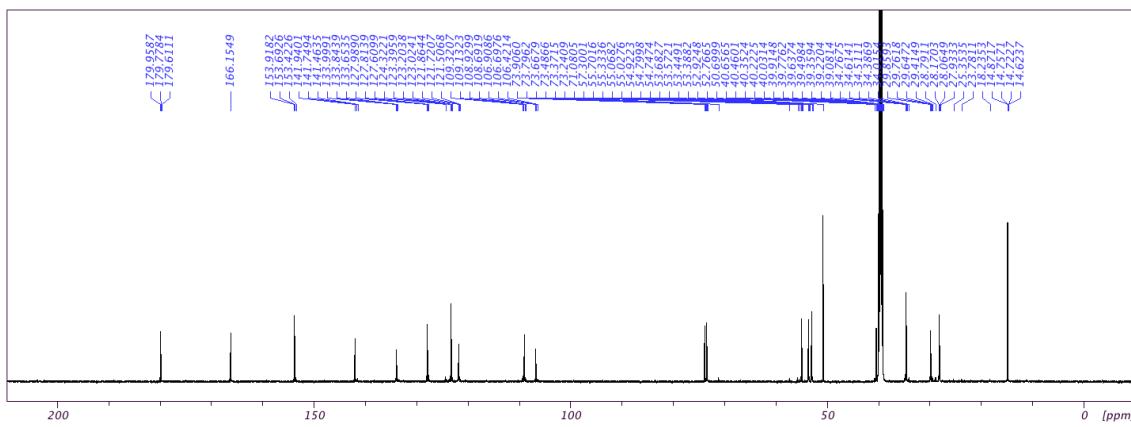


Figure S39. ^{13}C NMR (150 MHz, DMSO-d₆) spectrum of **1** after *U. guianensis* plantlets grew on culture with 1- ^{13}C -D-glucose.

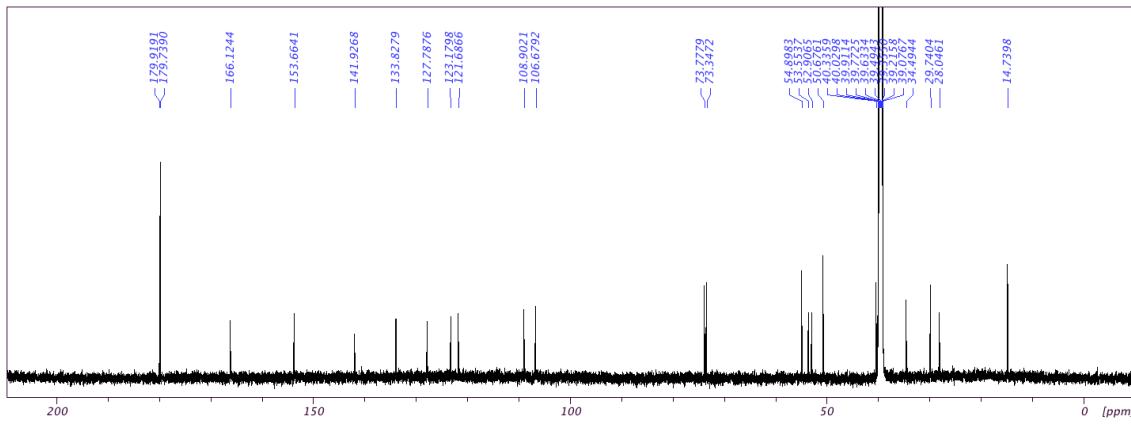


Figure S40. ^{13}C NMR (150 MHz, DMSO-d₆) spectrum of **1** after *U. guianensis* plantlets grew on culture with 2- ^{13}C -tryptophan.

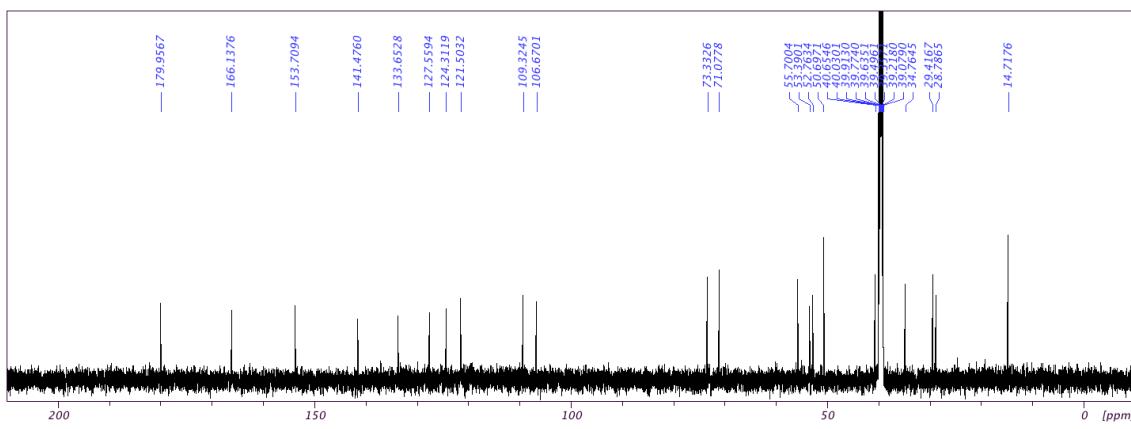


Figure S41. ^{13}C NMR (150 MHz, DMSO-d₆) spectrum of **3** with natural ^{13}C -isotopic abundance.

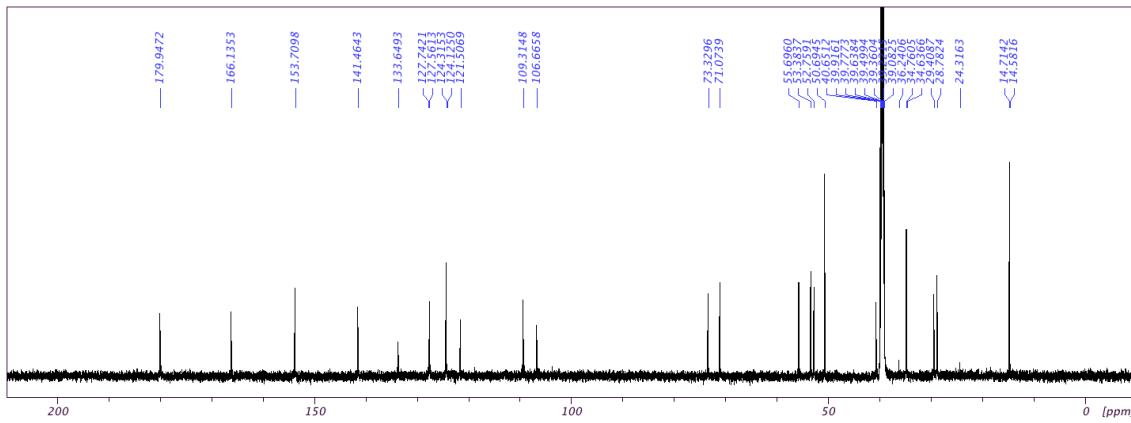


Figure S42. ^{13}C NMR (150 MHz, DMSO-d₆) spectrum of **3** after *U. guianensis* plantlets grew on culture with 1- ^{13}C -D-glucose.

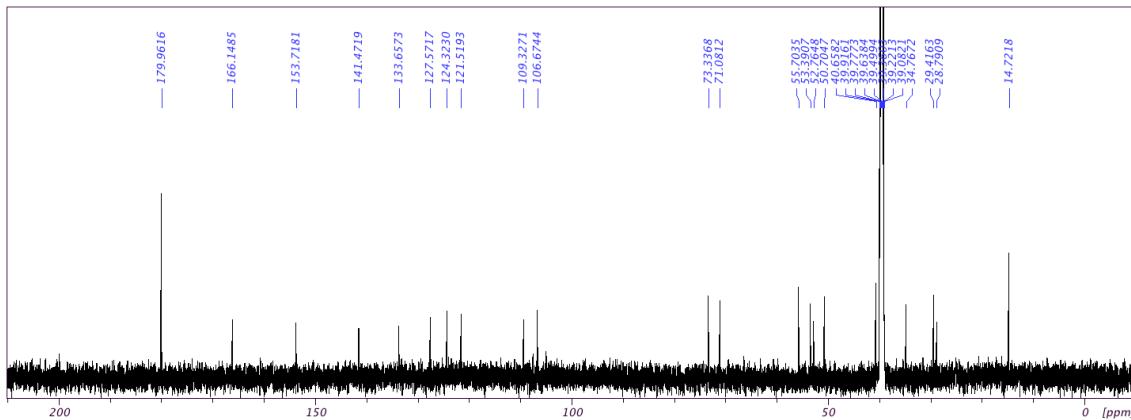


Figure S43. ^{13}C NMR (150 MHz, DMSO-d₆) spectrum of **3** after *U. guianensis* plantlets grew on culture with 2- ^{13}C -tryptophan.

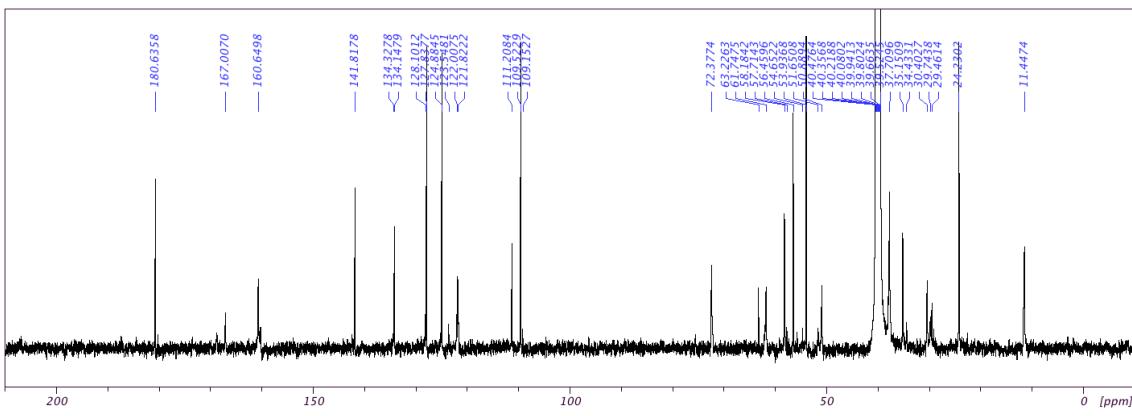


Figure S44. ^{13}C NMR (150 MHz, DMSO-d₆) spectrum of **4** with natural ^{13}C -isotopic abundance.

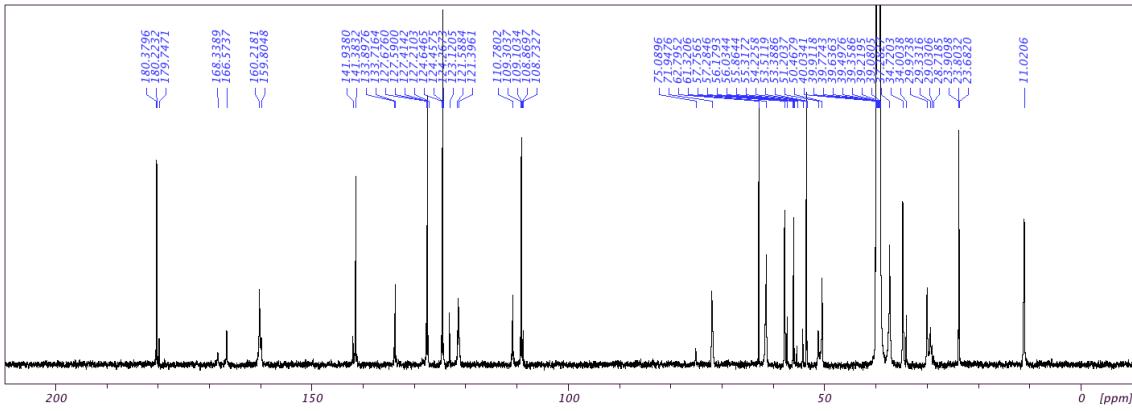


Figure S45. ^{13}C NMR (150 MHz, DMSO- d_6) spectrum of **4** after *U. guianensis* plantlets grew on culture with 1- ^{13}C -D-glucose.

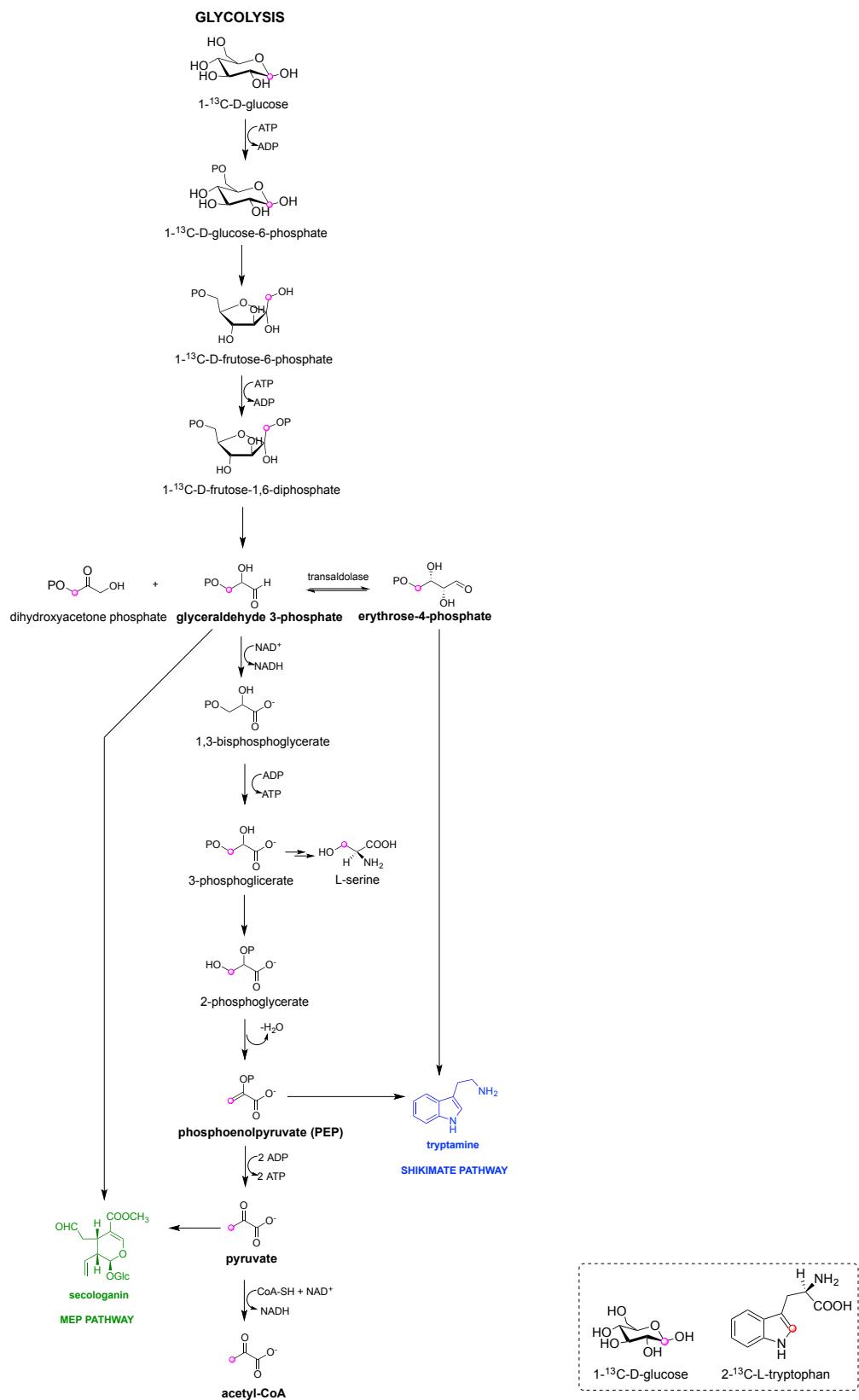


Figure S46. Biosynthetic pathway to oxindole alkaloids scaffolds (**1-4**) derived from $1\text{-}^{13}\text{C}$ -D-glucose and $2\text{-}^{13}\text{C}$ -tryptophan.

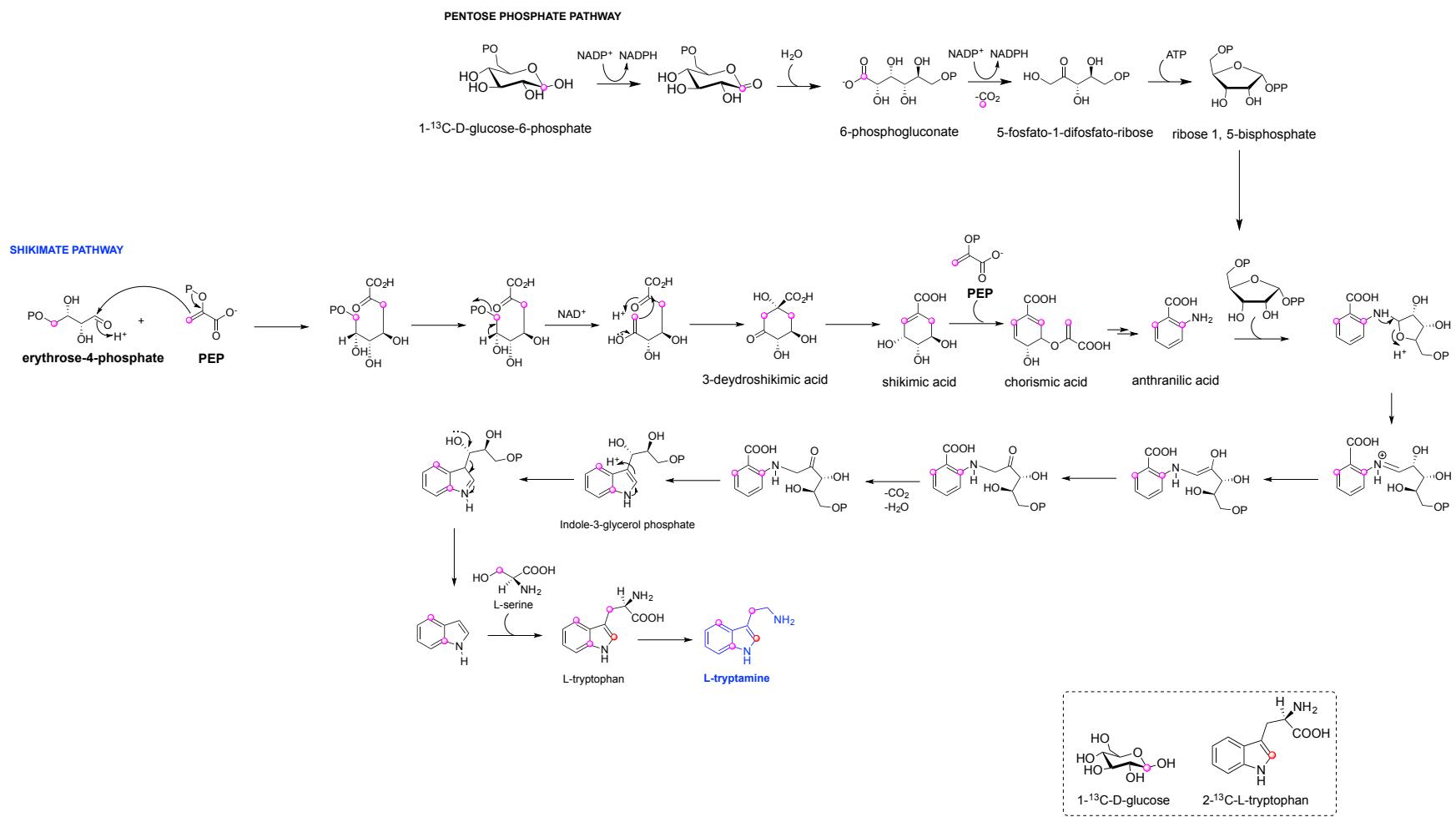


Figure S46 (continued)

VIA DOS TERPENOS (MEP)

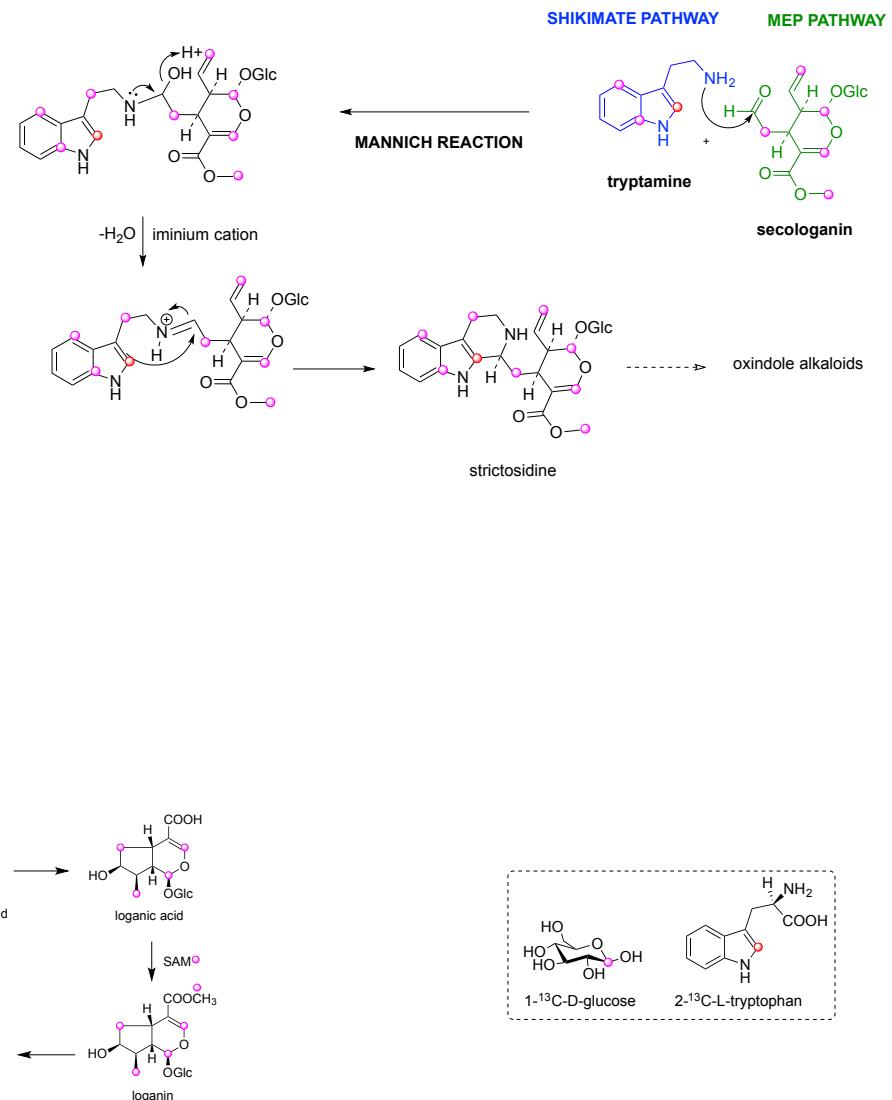
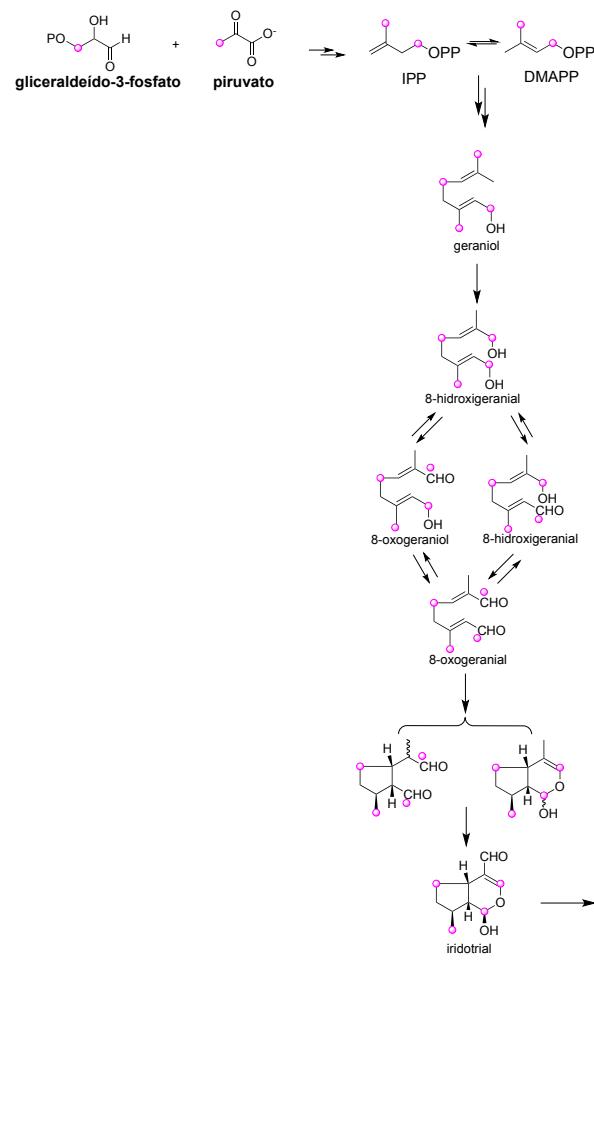


Figure S46 (continued)

Reference

- 1) a) Toure, H., Babadjamian, A., Balansard, G., Faure, R., Houghtons, P. J. Complete ^1H and ^{13}C NMR chemical shift assignments for some pentacyclic oxindole alkaloids. *Spectrosc. Lett.*, **25**, 293-300 (1992); b) Seki, H., Takayama, H., Aimi, N., Sakal, S., Ponglux, D. A nuclear magnetic resonance study on the eleven stereoisomers of heteroyohimbine-type oxindole alkaloid. *Chem. Pharm. Bull.* **41**, 12, 2077-2086 (1993); c) Carbonezi, C. A., Hamrski, L. H., Junior, O. A. F., Furlan, M., Bolzani, V. Da S., Young, M. C. M. Determinação por RMN das configurações relativas e conformações de alcalóides oxindólicos isolados de *Uncaria guianensis*. *Quim. Nova* **27**, 6, 878-881 (2004).