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| Last updated by author(s): | 7-8-19           |

## **Reporting Summary**

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| For all statistical ana | lyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.   |
|-------------------------|--|
| n/a Confirmed           |  |
| ☐ ☐ The exact s         | ample size $(n)$ for each experimental group/condition, given as a discrete number and unit of measurement   |
| A statemen              | t on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly  |
| I X I I I               | cal test(s) used AND whether they are one- or two-sided<br>n tests should be described solely by name; describe more complex techniques in the Methods section.  |
| A description           | on of all covariates tested  |
| A description           | on of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons   |
| A full descr            | iption of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) on (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)  |
|                         | pothesis testing, the test statistic (e.g. $F$ , $t$ , $r$ ) with confidence intervals, effect sizes, degrees of freedom and $P$ value noted as exact values whenever suitable.  |
| For Bayesia             | n analysis, information on the choice of priors and Markov chain Monte Carlo settings  |
| For hierarcl            | nical and complex designs, identification of the appropriate level for tests and full reporting of outcomes  |
| Estimates o             | of effect sizes (e.g. Cohen's $d$ , Pearson's $r$ ), indicating how they were calculated   |
| '                       | Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.  |
| Software and            | code   |
| Policy information al   | pout <u>availability of computer code</u>  |
| Data collection         | In-house software was used to collect the electrophysiologycal data (ionic or gating currents) from the experiments for this work.   |
| Data analysis           | In-house software was used to analyze the data. Microsoft Excel was used to organize the data and for the calculation of means and standard error of the means. GraphPad Prism (GraphPad Software, Inc.) was used to plot the data.  |
|                         | ustom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. de deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information. |

## Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The data that support the findings of this study are available on reasonable request from the corresponding author FB.

| Field-spe  | ecific reporting  |  |  |  |
|--|---|--|--|--|
| Please select the o  | ne below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.   |  |  |  |
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|  |   |  |  |  |
| Life scier   | nces study design   |  |  |  |
| All studies must dis   | sclose on these points even when the disclosure is negative.  |  |  |  |
| Sample size  | We determined the sample size to be between 4-6 repetitions of the same experiment, independently. No repeated measurement was performed in the same oocyte.  |  |  |  |
| Data exclusions  | No data was excluded from analysis other than those that clearly indicated unsuccessful expression of the protein by the oocytes. We defined those cases when the amplitude of the current peak to be recorded was smaller than 500 nA.                               |  |  |  |
| Replication  | Different batches of oocytes were used for the heterologous expression of the channels.   |  |  |  |
| Randomization  | The frogs we used as oocytes donors were wild type frogs and they were selected from our colony for the day of oocytes surgical extraction without any criteria that would bias the outcomes. We believe this introduces a high level of randomization to our assays. |  |  |  |
| Blinding   | We did not performed blind experiments.   |  |  |  |
| Reporting for specific materials, systems and methods  We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, |   |  |  |  |
| system or method list  | ted is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.   |  |  |  |
| Materials & ex   | perimental systems Methods  |  |  |  |
| n/a Involved in th   | ne study n/a Involved in the study  |  |  |  |
| Antibodies   | ChIP-seq  |  |  |  |
| Eukaryotic   |   |  |  |  |
| Palaeontol   | ogy MRI-based neuroimaging  |  |  |  |
|  | d other organisms   |  |  |  |
|  | search participants   |  |  |  |
| Clinical dat   |   |  |  |  |
| Animals and  | other organisms   |  |  |  |
| Policy information   | about studies involving animals; ARRIVE guidelines recommended for reporting animal research  |  |  |  |
| Laboratory anima   | We used Xenopus laevis females as oocytes donors.   |  |  |  |

Note that full information on the approval of the study protocol must also be provided in the manuscript.

No wild animal was used in the present study.

No field-collected sample was used in the present study.

The University of Chicago IACUC approved the protocols followed by the present study.

Wild animals

Ethics oversight

Field-collected samples