Neolymphostin A is a Covalent Phosphoinositide-3-kinase (PI3-K)/Mammalian Target of Rapamycin (mTOR) Dual Inhibitor that Employs an Unusual Electrophilic Vinylogous Ester

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	page
Figure S1. UV chromatogram (254 nm) before and after labeling reaction	S4
Figure S2. UV chromatogram (450 nm) before and after labeling reaction	S4
Figure S3. MS spectra of neolymphostin A/B-cysteine thiol probe adducts (2a/2b/3a/3b)	S5
Table S1. NMR spectral data for neolymphostin A-cysteine thiol probe adduct (2a) ¹ H NMR (500 MHz, DMSO- <i>d</i> ₆)	S6 S7
Table S2. NMR spectral data for neolymphostin A-cysteine thiol probe adduct (2b) ¹ H NMR (500 MHz, DMSO- <i>d</i> ₆) COSY (500 MHz, DMSO- <i>d</i> ₆) HSQC (500 MHz, DMSO- <i>d</i> ₆) HMBC (500 MHz, DMSO- <i>d</i> ₆)	S8 S9 S10 S11 S12
Table S3. NMR spectral data for neolymphostin B-cysteine thiol probe adduct (3a) ¹ H NMR (500 MHz, DMSO- <i>d</i> ₆)	S13 S14
Table S4. NMR spectral data for neolymphostin B-cysteine thiol probe adduct (3b) ¹ H NMR (500 MHz, DMSO- <i>d</i> ₆) COSY (500 MHz, DMSO- <i>d</i> ₆) HSQC (500 MHz, DMSO- <i>d</i> ₆) HMBC (500 MHz, DMSO- <i>d</i> ₆)	S15 S16 S17 S18 S19
¹ H NMR (500 MHz, DMSO- <i>d</i> ₆) of neolymphostin A (4) HPLC chromatogram of neolymphostin A (4)	S20 S21

Description of KINOMEscan	S22
Table S5. Dissociation constants (K _d) for neolymphostin A and selected kinases	S24
Curve images for dissociation constants (K _d) for neolymphostin A and selected kinases	S25
Table S6. Dissociation constants (K _d) for wortmannin and selected kinases	S26
¹ H NMR (500 MHz, phosphate-buffered D ₂ O) of 6	S27
Figure S4a. ¹ H NMR kinetics data for 6 reacting with glutathione, pH 7.4, 25°C Stacked ¹ H NMR spectra of 6 reacting with GSH, pH 7.4, 25°C	S28 S29
Figure S4b. ¹ H NMR kinetics data for 6 reacting with glutathione, pH 7.4, 37°C Stacked ¹ H NMR spectra of 6 reacting with GSH, pH 7.4, 37°C	S30 S31
Table S7. Dissociation constants (K_d) for neolymphostin A and selected kinases under four conditioning experiments (Arms A–D)	S32
Curve images for dissociation constants (K_d) for neolymphostin A and selected kinases under four conditioning experiments (Arms A–D)	S33
Figure S5. Timecourse for covalent modification of PI3K	S35
Figure S6. Neolymphostin docked into PI3Kα	S35
Figure S7. Neolymphostin docked into mTOR	S36
Figure S8. Neolymphostin docked into Pl3Kα (induced fit)	S37
Figure S9. Neolymphostin docked into mTOR (induced fit)	S38
Figure S10. Neolymphostin docked into Pl3Kα (covalent model)	S39
Table S8. HDX-MS data	S40
Figure S11. ¹ H NMR kinetics data for 6 reacting with N $^{\alpha}$ -acetyl lysine, pH 8.1, 25 $^{\circ}$ C Stacked ¹ H NMR spectra of 6 reacting with N $^{\alpha}$ -acetyl lysine, pH 8.1, 25 $^{\circ}$ C	S41 S42
Table S9. NMR spectral data for 8 ¹ H NMR (500 MHz, phosphate-buffered D ₂ O) COSY (500 MHz, phosphate-buffered D ₂ O) HSQC (500 MHz, phosphate-buffered D ₂ O) HMBC (500 MHz, phosphate-buffered D ₂ O)	S43 S44 S45 S46 S47
¹ H NMR (500 MHz, phosphate-buffered D ₂ O) of 6 and hydrolysis product 9	S48
Table S10. Results of competition binding assay for neolymphotin against a panel of 97 kinases	S49

Figure S12. TREEspot™ interaction map for neolymphostin A @ 1000nM	S51
Table S11. Selectivity S-Score for neolymphostin A against a 97 kinase panel	S52
Figure S13. NCI 60-cell line dose response curves for neolymphostin A	S53
Figure S14. NCI 60-cell line testing results for neolymphostin A	S54
Figure S15. NCI 60-cell line mean graphs for neolymphostin A	S55
Figure S16. NCI 60-cell line dose response curves for neolymphostin A (all)	S56
Figure S17. COMPARE results	S57

Figure S1. HPLC chromatogram (450 nm) before and after labeling reaction

C18(2) Phenomenex Luna, 100 x 4.6 mm, 5 μ, 100 Å 10-100% CH₃CN in water (0.1% FA) over 20 min, 0.7 mL min⁻¹

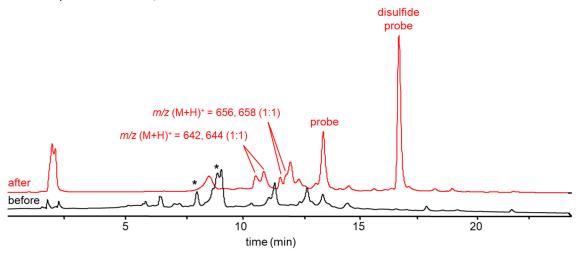
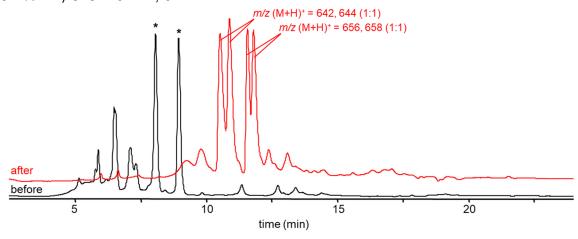


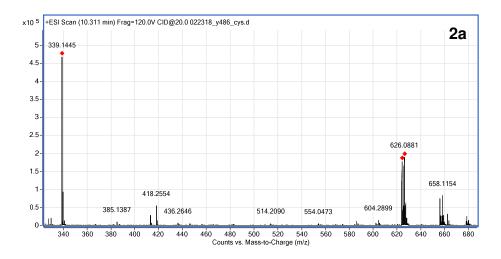
Figure S2. HPLC chromatogram (450 nm) before and after labeling reaction

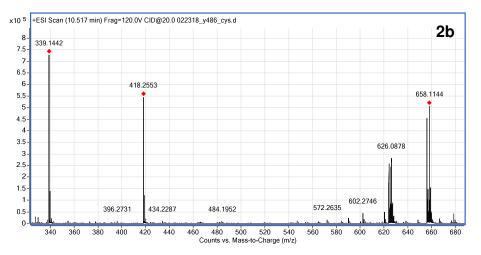
C18(2) Phenomenex Luna, 100 x 4.6 mm, 5 μ , 100 Å 10-100% CH₃CN in water (0.1% FA) over 20 min, 0.7 mL min⁻¹

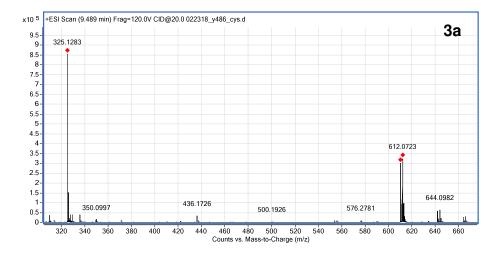


Asterisks (*) signal neolymphostin A (8.96 min) and neolymphostin B (8.08 min)

Figure S3. MS spectra of neolymphostin A/B-cysteine thiol probe adducts (2a/2b/3a/3b)







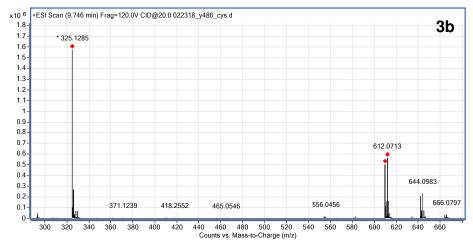


Table S1. NMR spectral data for neolymphostin A–cysteine thiol probe adduct ($\mathbf{2a}$) in DMSO- d_6 at 500 MHz

diastereomer #1

Position #a	δ (mult / I/∐z\\	
	δ _H (mult., (J(Hz))	
1		
2	8.33 (s)	
2a		
2a 3 4	8.47 (s)	
4		
5a		
6		
6-NH	9.82 (s)	
7	8.04 (s)	
8		
8a		
8b		
9		
10	a 3.78 (dd, 16.2, 5.6)	
	b 4.12 (dd, 16.2, 8.0)	
11	5.17 (dd, 7.8, 5.7)	
12	3.27 (s)	
13		
12 13 14 15	2.92 (sept., 6.9)	
15	1.14 (d, 6.8)	
16	1.13 (d, 6.8)	
1'	a 3.02 (dd, 13.7, 9.4)	
	b 3.20 (dd, 13.7, 5.1)	
2'	4.65 (m)	
2'-NH	8.93 (d, 7.8)	
3'		
2'-NH 3' 4' 5' 6' 7' 8'		
5'	7.70 (d, 8.5)	
6'	7.59 (d, 8.5)	
7'		
8'		
9'	3.61 (s)	
	0.0.	

^a Position numbering based on Aotani, et al. *J. Antibiot.* **1997**, *50*, 543–545.

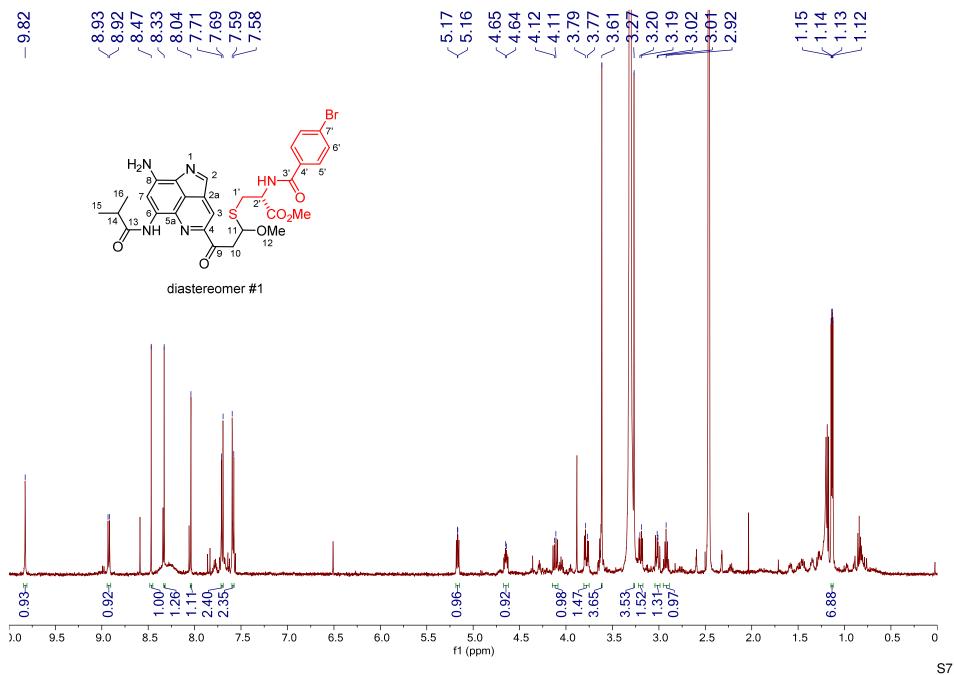


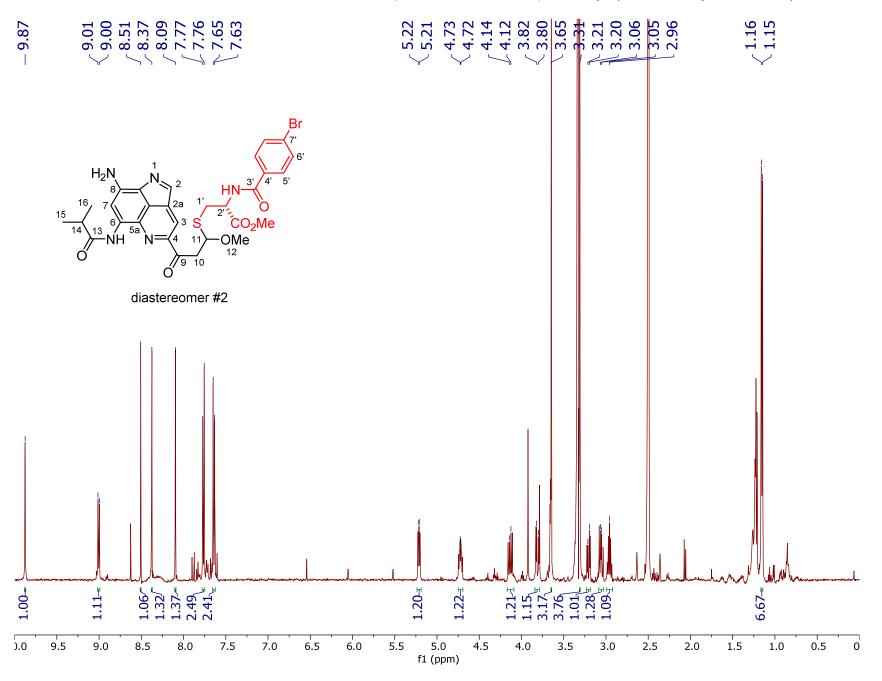
Table S2. NMR spectral data for neolymphostin A–cysteine thiol probe adduct (2b) in DMSO-d₆ at 500 MHz

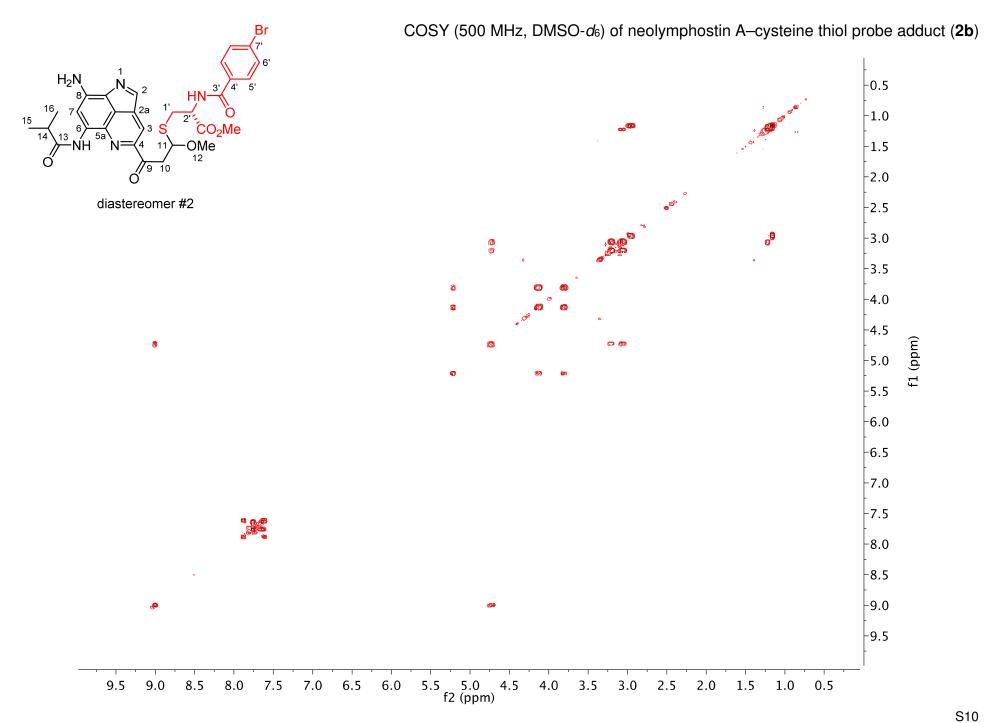
diastereomer#2

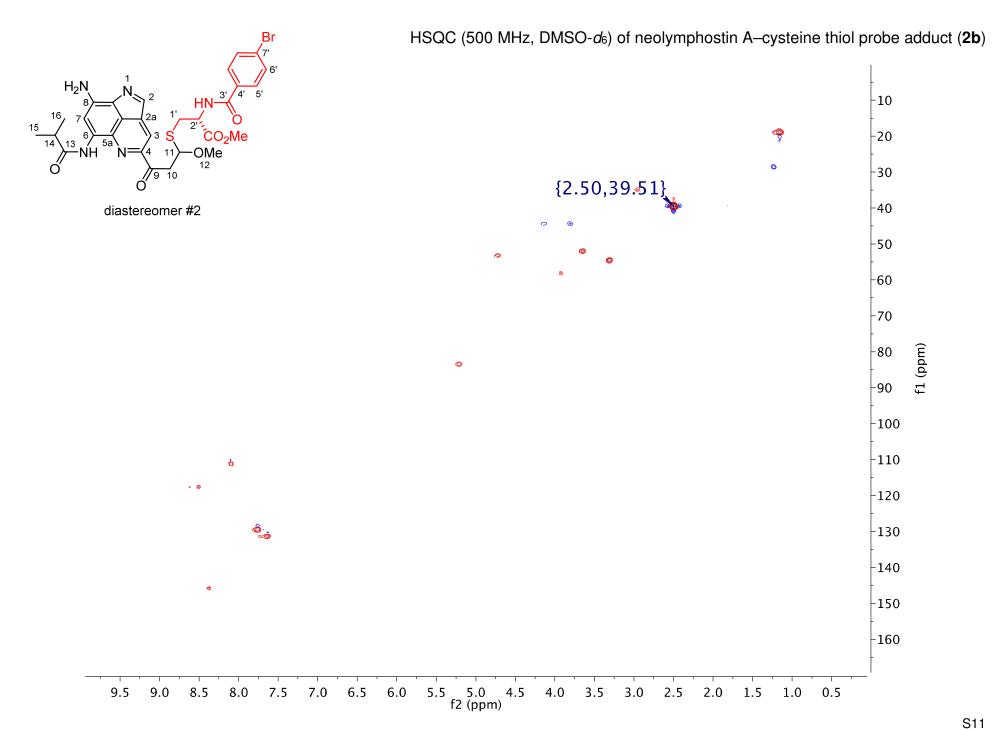
Position #a	δ_{H} (mult., J (Hz))	δ_{c^b}	COSY	HMBC
1				
2	8.37 (s)	145.8		2a, 8b
2a		136.6		
3	8.51 (s)	117.6		4, 8b
4		146.2		
5a		133.7		
6		139.5		
6-NH	9.87 (s)			5a, 7, 13
7	8.09 (s)	111.0		5a, 6, 8a
8		n.d.c		
8a		124.6		
8b		124.8		
9		197.0		
10	a 3.81 (dd, 16.5, 5.1)	44.4	10b, 11	9, 11
	b 4.13 (dd, 16.5, 8.1)		10a, 11	9, 11
11	5.22 (dd, 8.0, 5.1)	83.4	10a, 10b	12
12	3.31 (<i>s</i>)	54.8		11
13		176.1		
14	2.96 (sept., 6.9)	35.1	15, 16	13, 15, 16
15	1.16 (d, 6.6)	19.3	14	13, 14, 16
16	1.16 (d, 6.6)	19.3	14	13, 14, 15
1'	a 3.06 (dd, 13.7, 9.2)	29.5	1'b, 2'	11, 8'
	b 3.21 (dd, 13.7, 5.3)		1'a, 2'	11, 8'
2'	4.73 (m)	53.2	1'a, 1'b, 2'-NH	8'
2'-NH	9.01 (d, 7.8)		2'	1', 3'
3'		165.7		
4'		132.6		
5'	7.77 (d, 8.6)	129.4	6'	3', 7'
6'	7.64 (d, 8.6)	131.3	5'	4', 7'
7'		125.2		
8'		171.1		
9'	3.65 (s)	52.0		8'

^a Position numbering based on Aotani, et al. *J. Antibiot.* **1997**, *50*, 543–545. ^b Carbon chemical shifts were based on HSQC and HMBC data

c n.d. = not determined







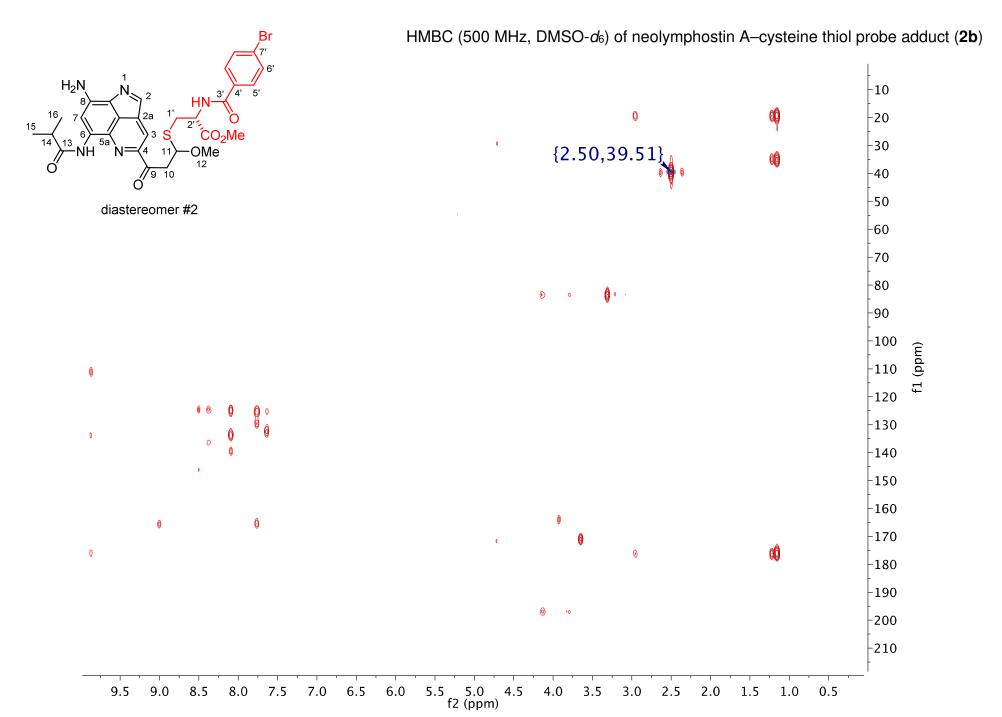


Table S3. NMR spectral data for neolymphostin B–cysteine thiol probe adduct (3a) in DMSO-d6 at 500 MHz

diastereomer #1

Position #a	δ _H (mult., (J(Hz))	
1		
2	8.36 (s)	
2a		
2a 3 4	8.51 (s)	
4		
5a		
6		
6-NH	9.90 (s)	
7	8.12 (s)	
8		
8a		
8b		
9		
10	a 3.83 (dd, 16.2, 5.6)	
	b 4.19 (dd, 16.2, 7.8)	
11	5.18 (dd, 7.6, 5.7)	
12	3.31 (s)	
13 14		
14	2.60 (q, 7.5)	
15 1'	2.60 (q, 7.5) 1.12 (t, 7.4) a 3.07 (dd, 13.7, 9.3)	
1'	a 3.07 (dd, 13.7, 9.3)	
	b 3.27 (dd, 13.7, 5.3)	
2'	4.71 (m)	
2'-NH	8.96 (d, 7.8)	
3'		
4'		
5'	7.74 (d, 8.5) 7.61 (d, 8.5)	
6'	7.61 (d, 8.5)	
2'-NH 3' 4' 5' 6'		
8.		
9'	3.65 (s)	

^a Position numbering based on Aotani, et al. *J. Antibiot.* **1997**, *50*, 543–545.

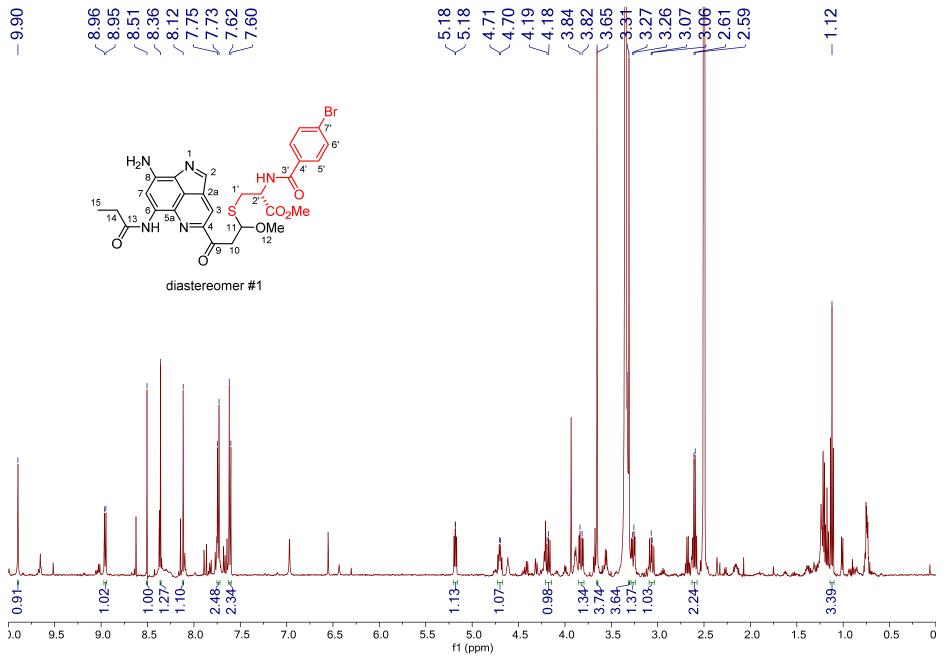


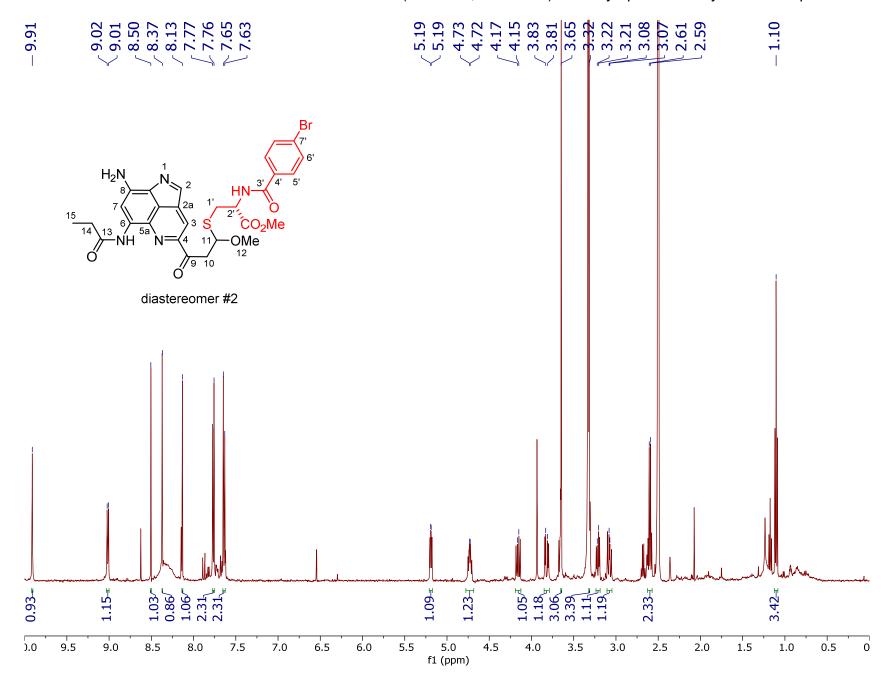
Table S4. NMR spectral data for neolymphostin B-cysteine thiol probe adduct (3b) in DMSO-d₆ at 500 MHz

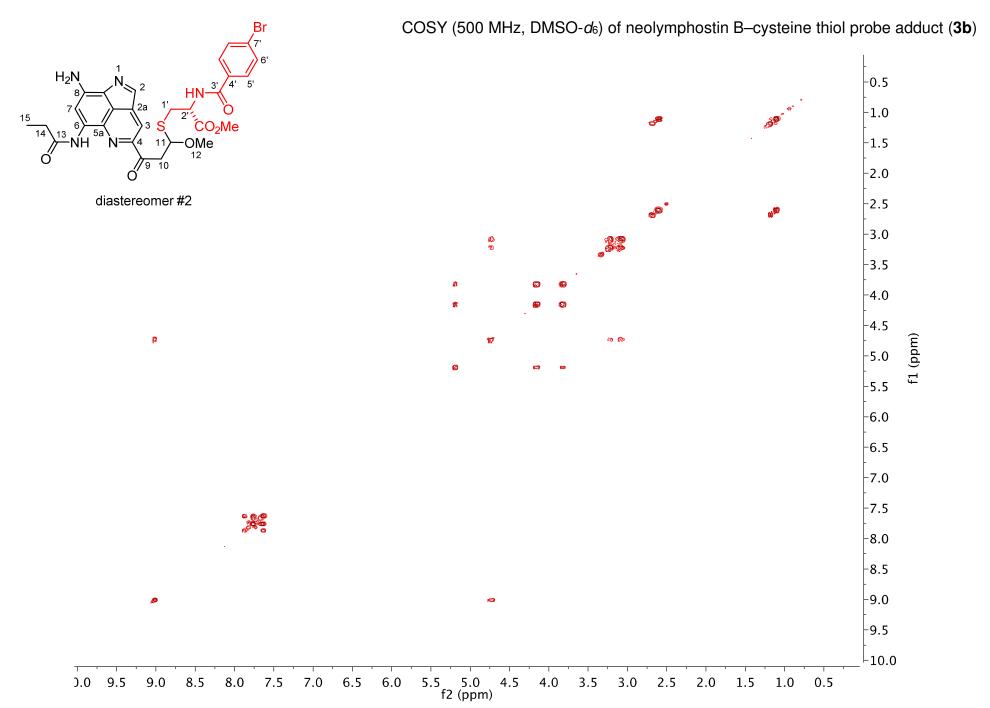
diastereomer #2

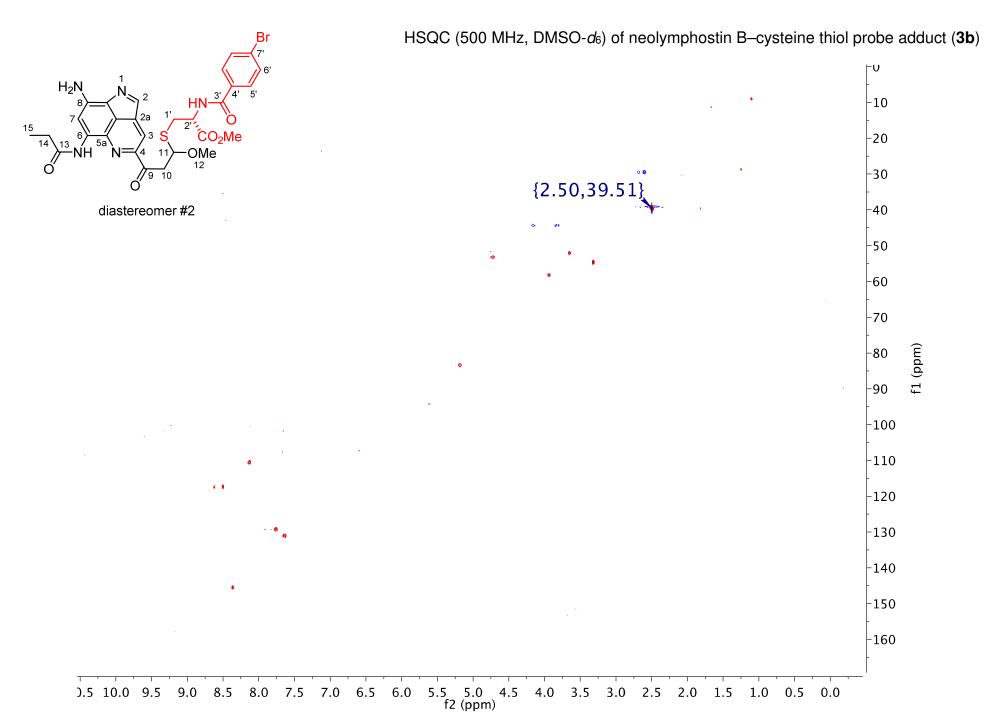
Position #a	δ _H (mult., (J(Hz))	δc ^b	COSY	HMBC
1				
2	8.37 (s)	145.5		2a, 8b
2a		136.3		
3	8.50 (s)	117.4		4, 8b
4		145.4		
5a		133.6		
6		139.3		
6-NH	9.91 (s)			5a, 7, 13
7	8.13 (s)	110.4		5a, 6, 8a
8		n.d.c		
8a		124.8		
8b		124.4		
9		198.0		
10	a 3.82 (dd, 16.8, 5.1)	44.4	10b, 11	9, 11
	b 4.16 (dd, 16.8, 8.2)		10a, 11	9, 11
11	5.19 (dd, 8.0, 5.1)	83.4	10a, 10b	12, 1'
12	3.32 (s)	54.6		11
13		173.2		
14	2.60 (q, 7.5)	29.5	15	13, 15
15	1.10 (t, 7.5)	9.0	14	13, 14
1'	a 3.08 (dd, 13.9, 9.5)	28.9	1'b, 2'	11, 2'
	b 3.22 (dd, 13.9, 5.6)		1'a, 2'	11
2'	4.73 (m)	53.2	1'a, 1'b, 2'-NH	8'
2'-NH	9.02 (d, 7.8)		2'	3'
3'		165.4		
4'		132.6		
5'	7.77 (d, 8.4)	129.2	6'	3', 7'
6'	7.64 (d, 8.4)	131.1	5'	4', 7'
7'		125.4		
8'		171.0		
9'	3.65 (s)	52.0		8'

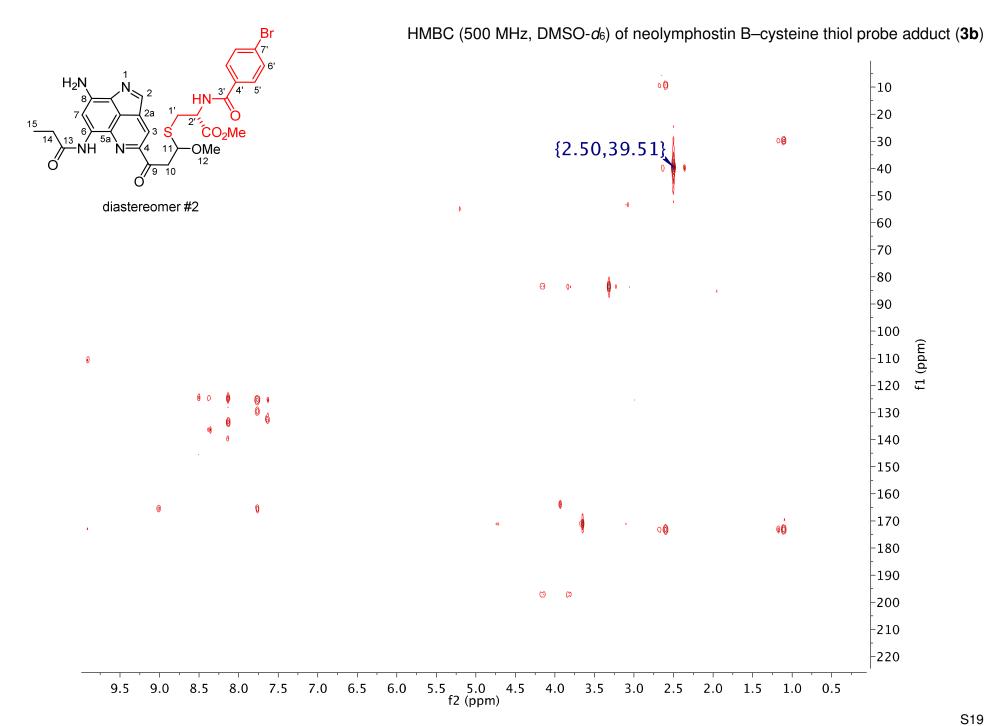
^a Position numbering based on Aotani, et al. *J. Antibiot.* **1997**, *50*, 543–545. ^b Carbon chemical shifts were based on HSQC and HMBC data

c n.d. = not determined

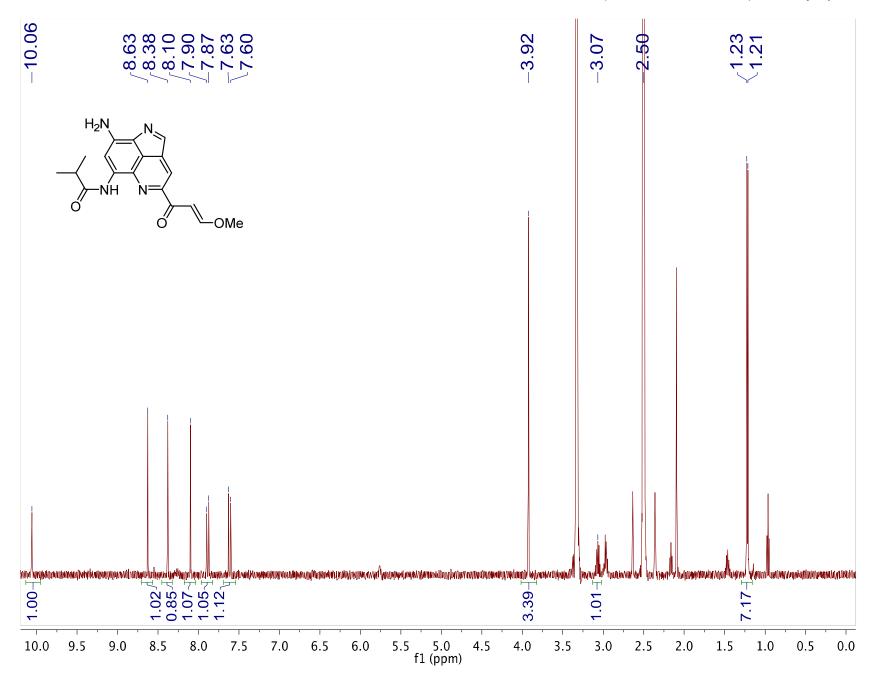




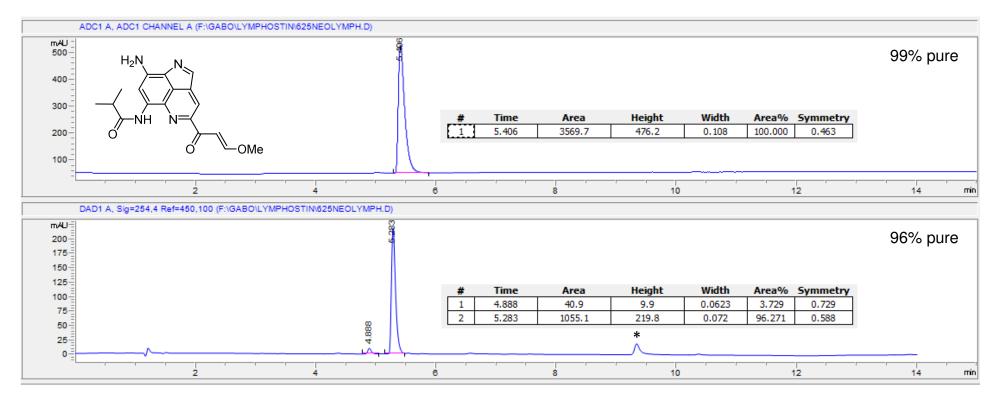


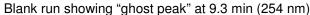


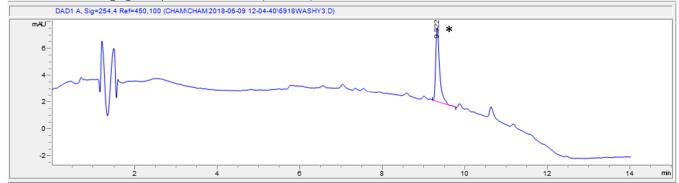
¹H NMR (500 MHz, DMSO-*d*₆) of neolymphostin A (**4**)



C18(2) Luna Phenomenex, 100 x 4.6 mm, 5 μ , 100 Å 10-100% CH₃CN in water (0.1% FA) over 10 min, 1.0 mL min⁻¹ Top: ELSD; Bottom: 254 nm





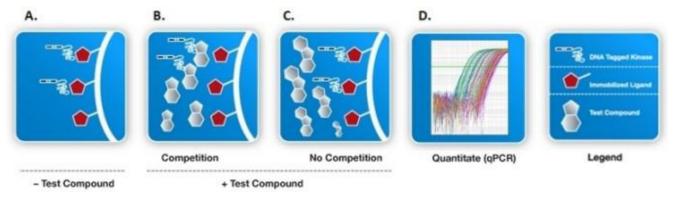


Technology Overview

The KINOMEscan™ screening platform employs a novel and proprietary active site-directed competition binding assay to quantitatively measure interactions between test compounds and more than 450 human kinases and disease relevant mutant variants. This robust and reliable assay technology affords investigators the ability to extensively annotate compounds with accurate, precise and reproducible data. KINOMEscan™ assays do not require ATP and thereby report true thermodynamic interaction affinities, as opposed to IC₅₀ values, which can depend on the ATP concentration.

How KINOMEscan™ Works

Compounds that bind the kinase active site and directly (sterically) or indirectly (allosterically) prevent kinase binding to the immobilized ligand, will reduce the amount of kinase captured on the solid support (A & B). Conversely, test molecules that do not bind the kinase have no effect on the amount of kinase captured on the solid support (C). Screening "hits" are identified by measuring the amount of kinase captured in test versus control samples by using a quantitative, precise and ultra-sensitive qPCR method that detects the associated DNA label (D). In a similar manner, dissociation constants (Kds) for test compound-kinase interactions are calculated by measuring the amount of kinase captured on the solid support as a function of the test compound concentration.



Kinase assays

For most assays, kinase-tagged T7 phage strains were grown in parallel in 24-well blocks in an E. coli host derived from the BL21 strain. E. coli were grown to log-phase and infected with T7 phage from a frozen stock (multiplicity of infection = 0.4) and incubated with shaking at 32°C until lysis (90-150 minutes). The lysates were centrifuged (6,000 x g) and filtered (0.2µm) to remove cell debris. The remaining kinases were produced in HEK-293 cells and subsequently tagged with DNA for qPCR detection. Streptavidin-coated magnetic beads were treated with biotinylated small molecule ligands for 30 minutes at room temperature to generate affinity resins for kinase assays. The liganded beads were blocked with excess biotin and washed with blocking buffer (SeaBlock (Pierce), 1% BSA, 0.05 % Tween 20, 1 mM DTT) to remove unbound ligand and to reduce non-specific phage binding. Binding reactions were assembled by combining kinases, liganded affinity beads, and test compounds in 1x binding buffer (20 % SeaBlock, 0.17x PBS, 0.05 % Tween 20, 6 mM DTT). Test compounds were prepared as 40x stocks in 100% DMSO and directly diluted into the assay. All reactions were performed in polypropylene 384-well plates in a final volume of 0.02 ml. The assay plates were incubated at room temperature with shaking for 1 hour and the affinity beads were washed with wash buffer (1x PBS, 0.05 % Tween 20). The beads were then re-suspended in elution buffer (1x PBS, 0.05 % Tween 20, 0.5 µM non-biotinylated affinity ligand) and incubated at room temperature with shaking for 30 minutes. The kinase concentration in the eluates was measured by aPCR.

Binding Constants (Kds)

Binding constants (K_ds) were calculated with a standard dose-response curve using the Hill equation:

$$Response = Background + \frac{Signal - Background}{1 + \left(K_d^{Hill \, Slope} / Dose^{Hill \, Slope}\right)}$$

The Hill Slope was set to -1.

Curves were fitted using a non-linear least square fit with the Levenberg-Marquardt algorithm.

Compound Handling

An 11-point 3-fold serial dilution of each test compound was prepared in 100% DMSO at 100x final test concentration and subsequently diluted to 1x in the assay (final DMSO concentration = 1%). Most K_ds were determined using a compound top concentration = 30,000 nM. If the initial K_d determined was < 0.5 nM (the lowest concentration tested), the measurement was repeated with a serial dilution starting at a lower top concentration. A K_d value reported as 40,000 nM indicates that the K_d was determined to be >30,000 nM.

scanKINETIC platform

This protocol offers dissociation kinetics to classify inhibitors as irreversible, reversible-slow dissociation, or reversible-rapid dissociation. Kinetic dissociation constants were measured like described above but equilibration time and dilutions were varied in a set of four different experiments, study arms A-D. In study arm A and C, the compound and kinase were combined and equilibrated for 6 h and 1 h, respectively, before measuring the K_d. In study arm B the compound and kinase were combined and equilibrated for 1 h, diluted 30-fold, and re-equilibrated for 5 h. In study arm D the compound and kinase were pre-diluted, combined, and equilibrated for 5 h

Percent Control (%Ctrl)

Neolymphostin A was screened at 1000nm against a panel of kinases and results for primary screen binding interactions are reported as '% Ctrl', where lower numbers indicate stronger hits.

$$\%Ctrl = \left(\frac{\text{test compound signal } - \text{positive control signal}}{\text{negative control signal } - \text{prositive control signal}}\right) X \ 100$$

test compound = neolymphostin A, negative control = DMSO (100%Ctrl), positive control = control compound (0%Ctrl)

Selectivity Score (S-scores)

Selectivity Score or S-score is a quantitative measure of compound selectivity. It is calculated by dividing the number of kinases that compounds bind to by the total number of distinct kinases tested, excluding mutant variants. This value can be calculated using %Ctrl as a potency threshold (below) and provides a quantitative method of describing compound selectivity to facilitate comparison of different compounds.

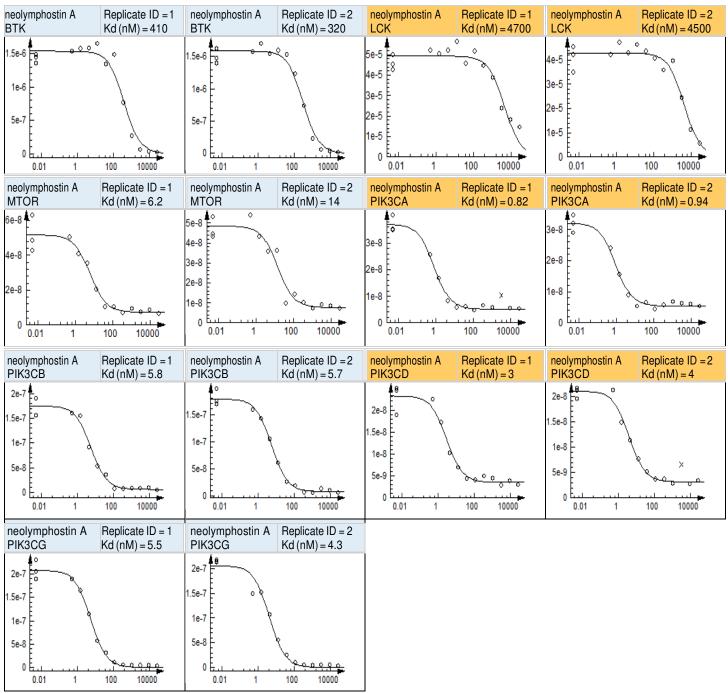
S(35) = (number of non-mutant kinases with %Ctrl <35)/(number of non-mutant kinases tested)

S(10) = (number of non-mutant kinases with %Ctrl < 10)/(number of non-mutant kinases tested)

S(1) = (number of non-mutant kinases with %Ctrl < 1)/(number of non-mutant kinases tested)

 $\textbf{Table S5.} \ \, \text{Dissociation constants } (K_d) \ \, \text{for neolymphostin A and selected kinases}$

Target	neolymphostin A
Gene	K _d (nM)
Symbol	Ka (TIIVI)
BTK	3370
LCK	4600
MTOR	10
PIK3CA	0.88
PIK3CB	5.7
PIK3CD	3.5
PIK3CG	4.9



The amount of kinase measured by qPCR (Signal; y-axis) is plotted against the corresponding compound concentration in nM in log10 scale (x-axis). Data points marked with an "x" were not used for K_d determination.

Table S6. Dissociation constants (K_d) for wortmannin and selected kinases

Target	neolymphostin A
Gene	K _d (nM)
Symbol	Ka (HIVI)
MTOR	9200
PIK3CA	5.4
PIK3CB	7.6
PIK3CD	15
PIK3CG	5.5

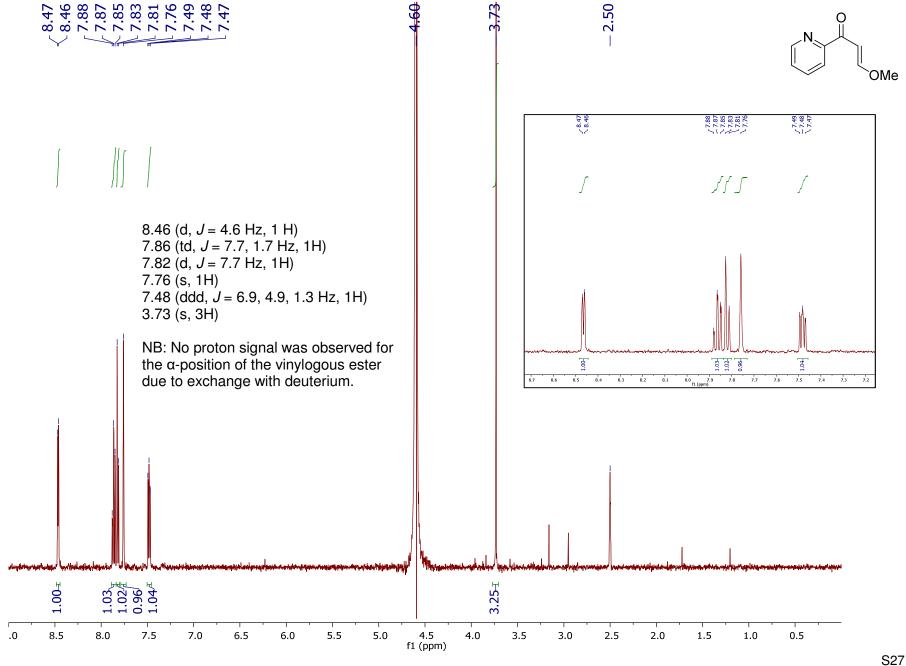
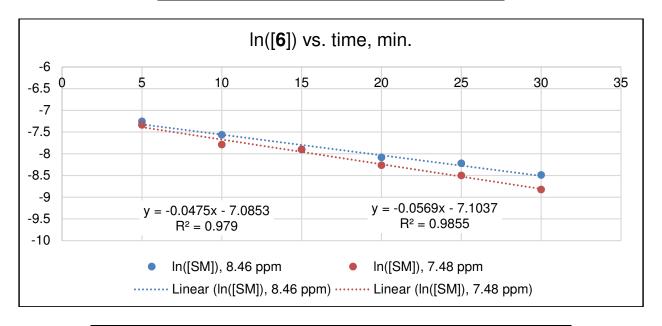


Figure S4a. ¹H NMR kinetics data for 6 reacting with GSH in phosphate-buffered D₂O, pH 7.4, 25°C

time, min.	In([6]), 8.46 ppm	In([6]), 7.48 ppm
5	-7.253256922	-7.331238893
10	-7.56062856	-7.785473736
15	-7.901007052	-7.898153983
20	-8.08047554	-8.259364082
25	-8.218337818	-8.496990484
30	-8.485104877	-8.822574841



	signal at 8.46 ppm	signal at 7.48 ppm	average
slope of In([6]) vs. time	-0.0475	-0.0569	-0.0522ª
calculated half-life, min.	14.6	12.2	13.3 ^b

^aAverage of slopes of each linear regression line

^bCalculated by $t_{1/2} = \ln(2)/(-1*average slope)$

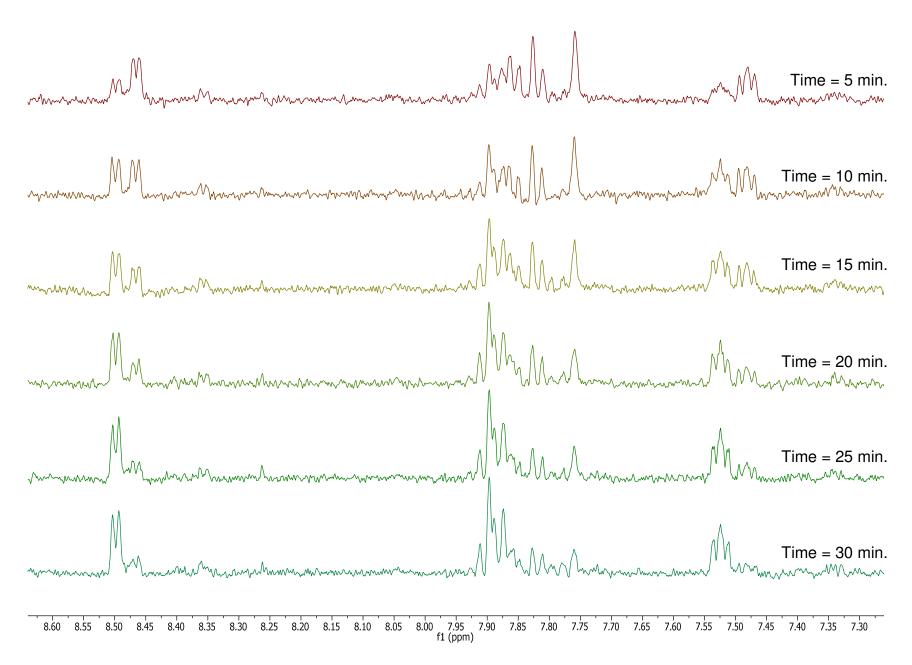
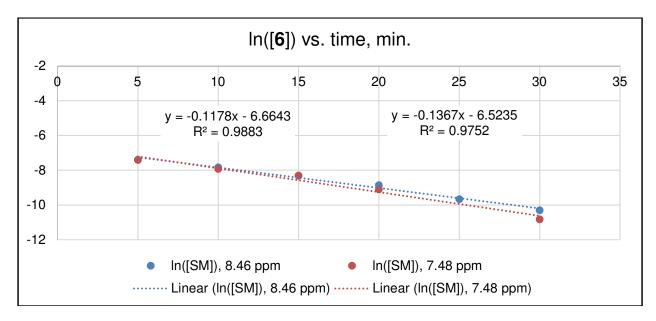


Figure S4b. ¹H NMR kinetics data for 6 reacting with GSH in phosphate-buffered D₂O, pH 7.4, 37°C

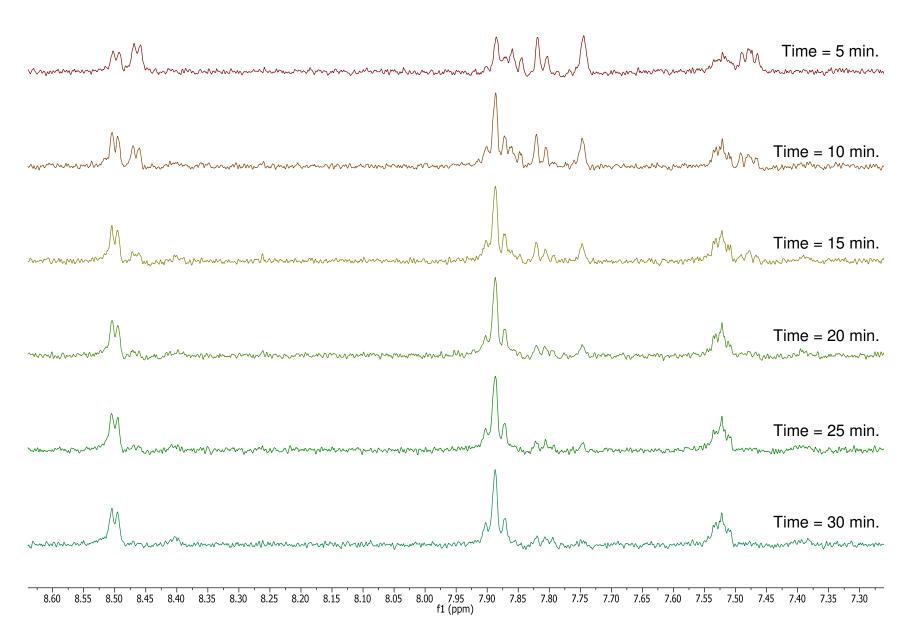
time, min.	In([6]), 8.46 ppm	In([6]), 7.48 ppm
5	-7.385382833	-7.407998793
10	-7.830088325	-7.919356191
15	-8.317580138	-8.303619091
20	-8.853665428	-9.104979856



	signal at 8.46 ppm	signal at 7.48 ppm	average
slope of In([6]) vs. time	-0.1178	-0.1367	-0.12725ª
calculated half-life, min.	5.9	5.1	5.4 ^b

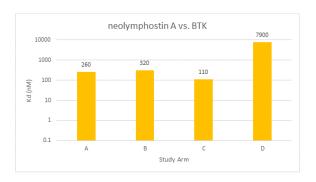
^aAverage of slopes of each linear regression line

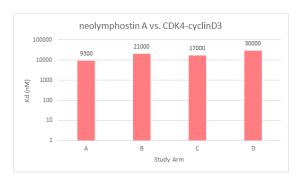
^bCalculated by $t_{1/2} = \ln(2)/(-1*average slope)$

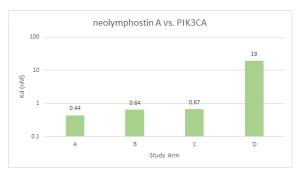


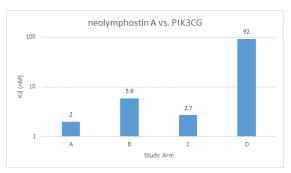
 $\begin{table}{c} \textbf{Table S7.} Dissociation constants (K_d) for neolymphostin A and selected kinases under four conditioning experiments ($Arms A-D$) \\ \end{table}$

Compound	ВТК	CDK4- CYCLIND3	PIK3CA	PIK3CG
Compound name (conditioning)	K _d (nm)	K _d (nm)	K _d (nm)	K _d (nm)
Neolymphostin A (Arm A)	260	9300	0.44	2
Neolymphostin A (Arm B)	320	21000	0.64	5.9
Neolymphostin A (Arm C)	110	17000	0.67	2.7
Neolymphostin A (Arm D)	7900	>30000	19	92

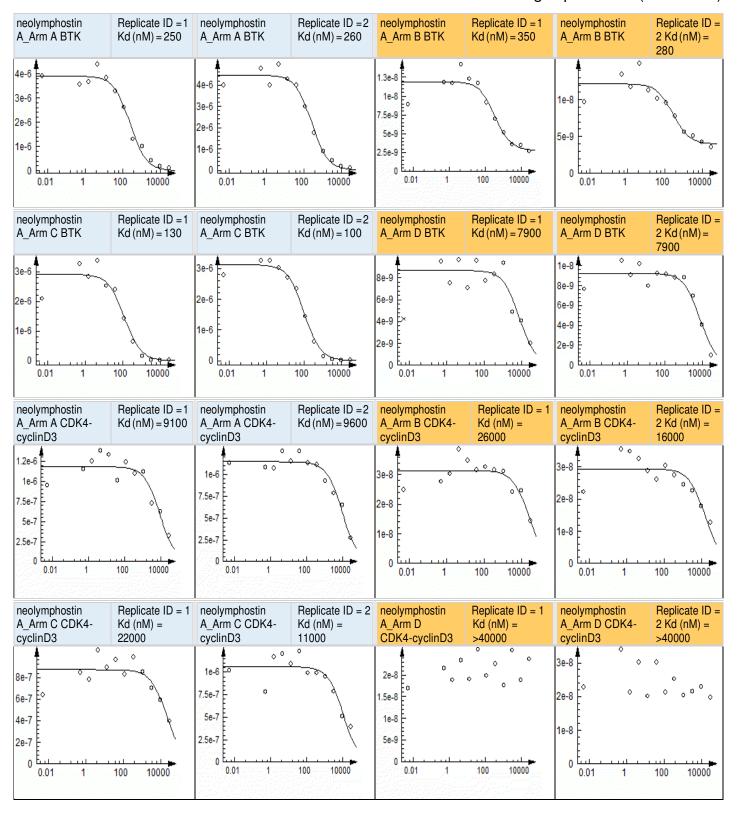








Curve images for dissociation constants (K_d) for neolymphostin A and selected kinases under four conditioning experiments (Arms A–D)



Curve images for dissociation constants (K_d) for neolymphostin A and selected kinases under four conditioning experiments (Arms A–D)...(continued)

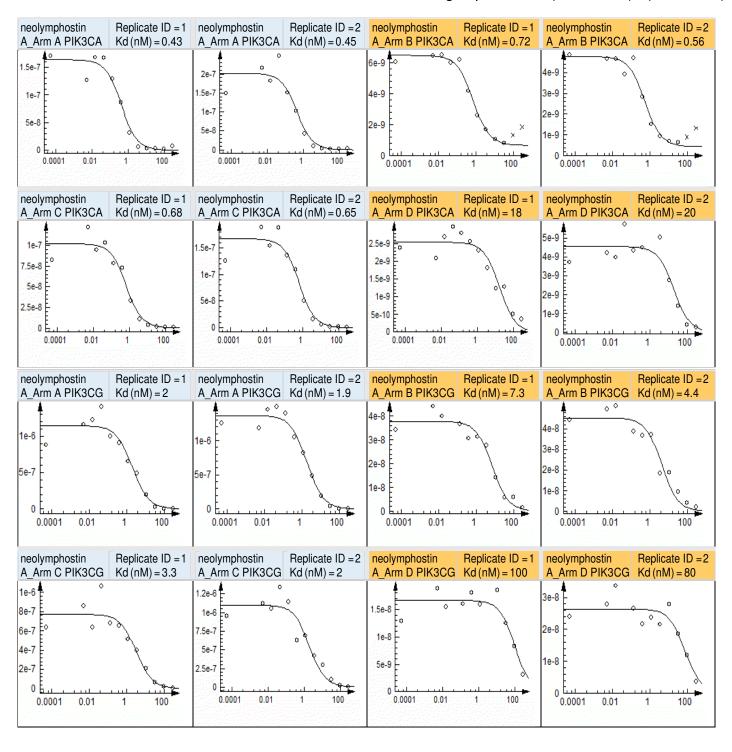
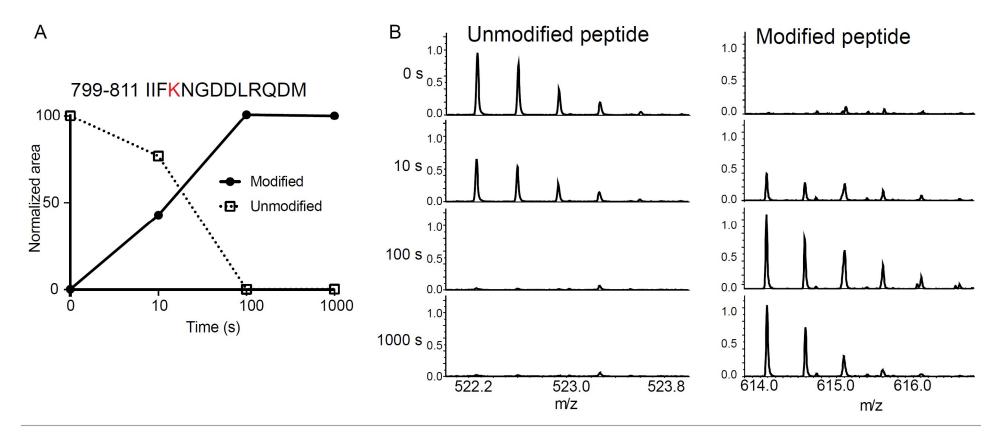


Figure S5. Timecourse for covalent modification of PI3K



A. The intensity for the modified and unmodified variant of the peptide indicated is shown over a time course of incubation with neolymphostin. B. Raw peptide traces of the unmodified and modified peptide over time.

Figure S6. Neolymphostin docked into $PI3K\alpha$

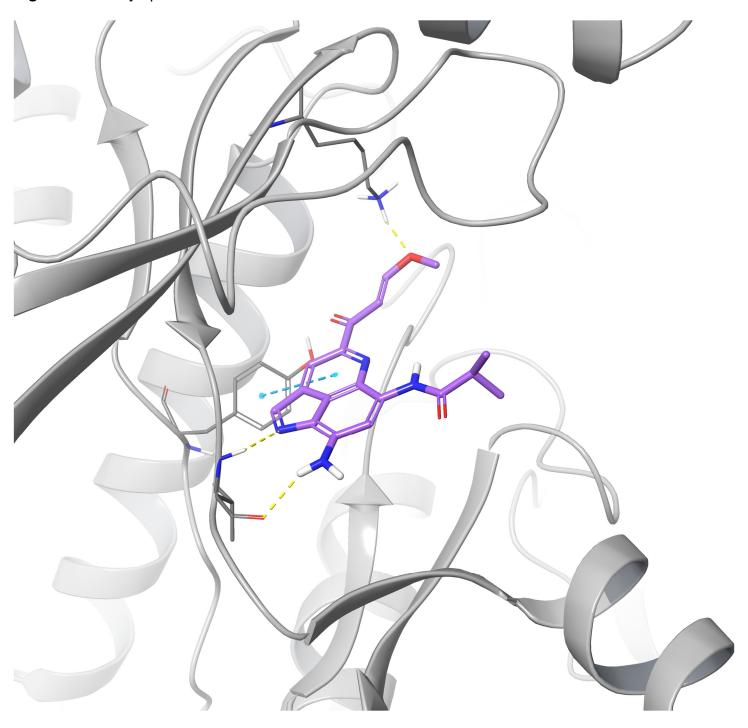


Figure S7. Neolymphostin docked into mTOR

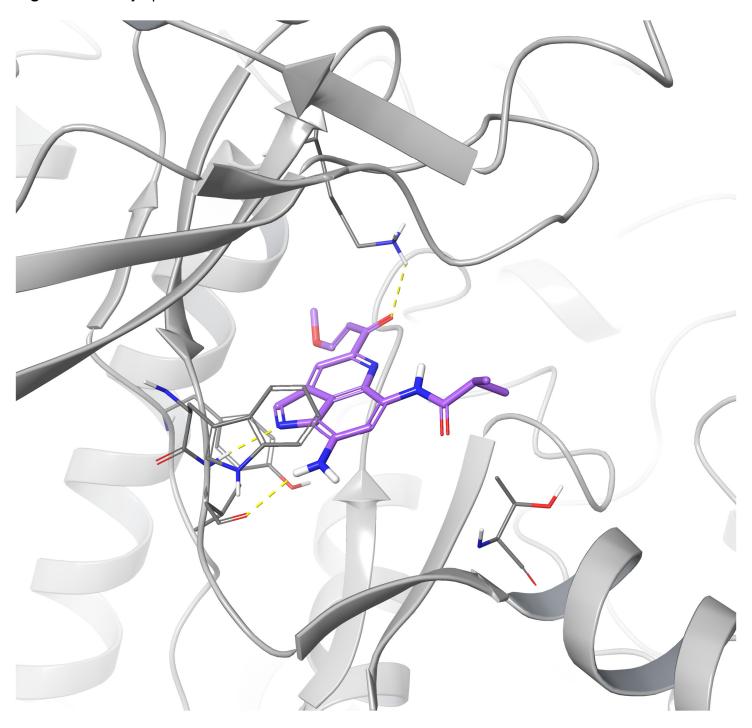


Figure S8. Neolymphostin docked into Pl3K α (induced fit)

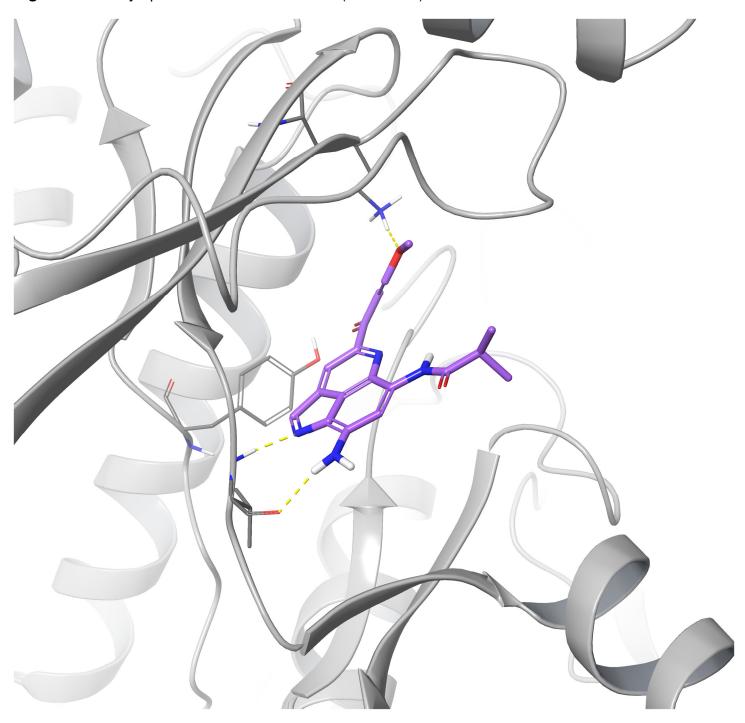


Figure S9. Neolymphostin docked into mTOR (induced fit)

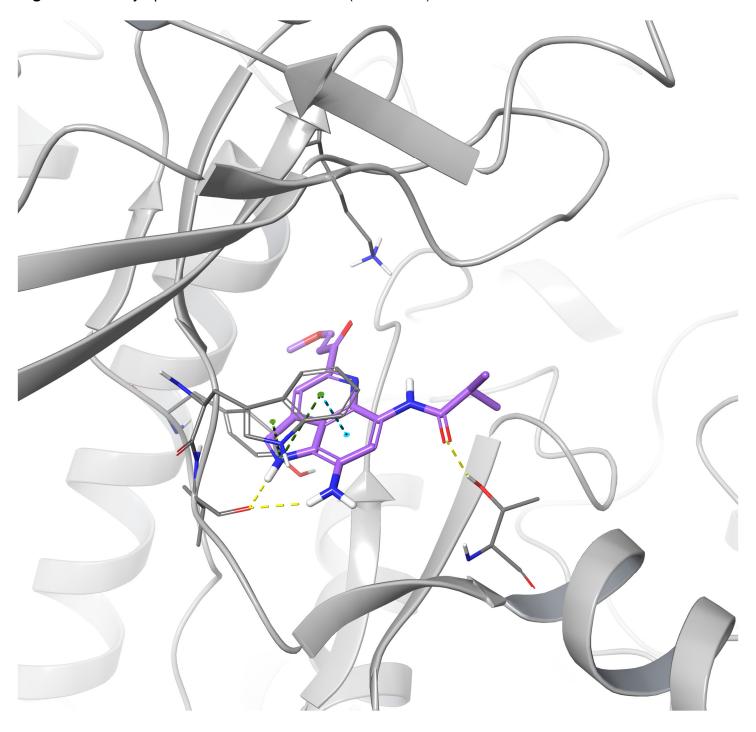


Figure S10. Neolymphostin docked into Pl3Kα (covalent model)

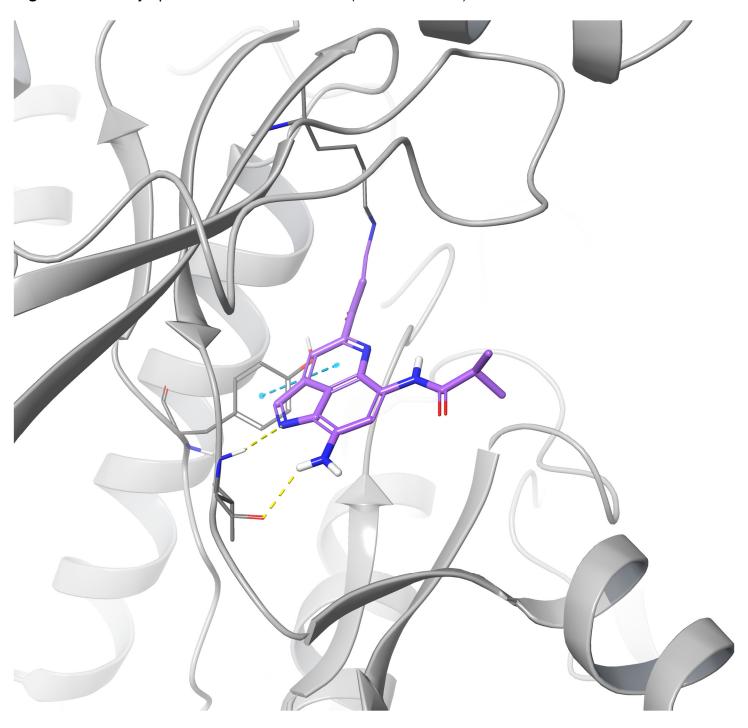


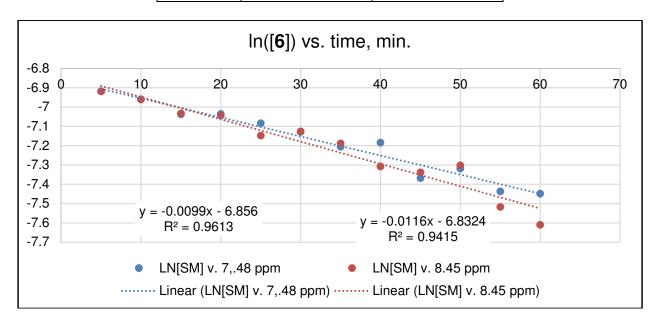
 Table S8. HDX-MS experiment

Start I	End Z	F	T	Sequence	3s on ice		a alone	SD	PI3K 3s on ice		lymphos 300s	tin SD
11	23	2		WGIHLMPPRILVE	8.8	0.1	40.8		8.9	0.5	41.3	1.2
31	35	1		IVTLE	40.5	1.4	55.4	0.4	41.2		56.4	1.1
37 43	42 62	4		LREATL ITIKHELFKEARKYPLHQLL	1.3 2.9	0.2	4.9 21.2	0.3	2.9	0.5	5.4 21.4	0.4
50	67	2	8.67	FKEARKYPLHQLLQDESS	6.9	0.0	34.0	0.5	6.5	0.0	35.5	2.0
69	76	1		IFVSVTQE	2.0	0.4	14.2 21.8	0.1	2.1	0.6	14.6	0.3
71 77	82 82	1		VSVTQEAEREEF AEREEF	1.6 3.1	0.2	21.8	0.2	1.7 3.5	0.3 1.0	22.1 30.6	1.3
83	91	3	0.00	FDETRRLCD	2.2	0.4	6.8	0.3	2.5	0.7	7.9	0.2
83	92	3		FDETRRLCDL	1.0	0.1	3.8	0.1	1.0	0.0	4.4	0.3
92 93	99	2		LRLFQPFL RLFQPFL	0.9	0.3	1.6 1.7	0.0	1.0	0.2	2.0 1.9	0.2
99	103	1	6.67	LKVIE	0.9	0.4	2.7	0.1	1.3	0.3	2.9	0.3
100 120	119 127	4		KVIEPVGNREEKILNREIGF AIGMPVCE	6.7 5.8	0.0	33.8 35.1	0.4	6.7	0.3	34.6 36.6	1.3 0.8
128	139	2		FDMVKDPEVQDF	11.4	0.5	31.2	0.3	11.4		32.2	0.6
145	152	1		NVCKEAVD	7.2	1.3	15.0	2.4	7.6	2.7	15.8	1.0
147 153	152 163	3		CKEAVD LRDLNSPHSRA	5.2 19.6	0.8	13.8 36.8	1.5 0.7	5.8 19.5	0.5	14.6 38.0	0.9
154	164	3		RDLNSPHSRAM	13.3	0.5	34.6		13.5		35.7	1.7
165 193	193 209	5		YVYPPNVESSPELPKHIYNKLDKGQJIVV	12.6	0.1	34.2	0.2	12.6	0.2	34.9	1.5
196	223	5		VIWVIVSPNNDKQKYTL VIVSPNNDKQKYTLKINHDCVPEQVIAE	16.9 9.7	0.4	39.9 22.7		16.4 9.7	0.1	40.4 23.4	1.0
223	233	3	5.12	EAIRKKTRSML	30.1	0.1	43.2	0.5	29.6		44.2	1.7
224	233 252	1		AIRKKTRSML LEYOGKYIL	29.4	1.1 0.3	41.5 52.6		29.6		42.7 54.3	1.6
253	259	1		KVCGCDE	35.1 3.0	0.5	12.9	0.3	34.8	1.0	14.2	0.6
260	269	2		YFLEKYPLSQ	11.8	0.3	18.7	0.1	11.8		19.2	1.0
261 279	275 287	4		FLEKYPLSQYKYIRS LGRMPNLML	10.1 13.0	0.3	16.1 25.0	0.1	9.7	0.3	16.6 25.4	0.3 1.0
294	301	1		YSQLPMDC	36.0	0.4	65.3	0.6	36.0	- 7	66.8	1.5
302	327	4		FTMPSYSRRISTATPYMNGETSTKSL	38.1	0.6	45.3	0.6	37.7		46.6	2.1
328 343	334 369	1		WVINSAL YVNVNIRDIDKIYVRTGIYHGGEPLCD	29.0 12.6	0.2	41.7 23.7	0.2	29.0	0.5	42.7 24.2	0.8
370	389	3	11.81	NVNTQRVPCSNPRWNEWLNY	5.0	0.3	32.5	0.2	4.3	0.5	32.8	2.4
390 405	402	3		DIYIPDLPRAARL SICSVKGRKGAKEEHCPLAWGNINL	1.9	0.4	10.6 6.4	0.2	2.1	0.8	11.1 6.9	0.1
430	436	1		FDYTDTL	3.7 1.7	0.2	9.2	0.1	2.0	0.1	10.5	0.6
437	443	1		VSGKMAL	14.7	0.3	41.5	0.4	15.0		42.4	0.8
444 456	474	2		NLWPVPHGLEDLLNPIGVTGSNPNKETPCLE LNPIGVTGSNPNKETPCLE	5.5 8.6	0.2	30.1 38.7	0.9	5.3 8.7	0.2	31.2 39.5	1.6
474	478	1		ELEFD	3.4	0.5	3.7	0.6	4.8	2.6	5.4	0.5
483	491	2		VVKFPDMSV	21.9	0.0	41.4	0.8	21.8		42.0	1.8
484 492	491 498	1		VKFPDMSV IEEHANW	27.0 9.5	0.1	51.4 39.9	0.6	27.4	0.4 1.4	52.8 39.0	1.6
495	506	3		HANWSVSREAGF	35.2	0.4	45.3	0.3	35.2	0.8	45.9	1.0
507	522	3		SYSHAGLSNRLARDNE	41.5	0.3	43.9		41.1		44.3	1.5
523 532	531 542	2		LRENDKEQL KAISTRDPLSE	14.9 7.1	0.2	44.3 29.9	0.3	15.3 7.2	0.3	45.4 31.4	1.5
546	551	1	9.19	QEKDFL	2.3	0.2	2.8	0.6	2.4	0.7	3.2	0.2
550 552	565 570	4		FLW3HRHYCVTIPEIL WSHRHYCVTIPEILPKLLL	1.4	0.2	16.2 11.6	0.1	0.3	0.4	16.8 12.0	0.8
571	583	2		SVKWNSRDEVAQM	1.4	0.2	7.6	0.1	1.5	0.3	8.1	0.3
584	600	3		YCLVKDWPPIKPEQAME	1.2	0.1	17.5	0.8	1.3	0.3	18.7	0.8
602 631	632	1	13.87	LDCNYPDPMVRGFAVRCLEKYLTDDKLSQYL YLIOL	3.0 0.4	0.1	18.2	0.7	1.0	0.1 1.8	18.6	0.4
636	649	2		VQVLKYEQYLDNLL	0.9	0.2	15.2	0.2	0.8	0.2	15.6	0.3
649	666	4		LVRFLLKKALTNQRIGHF	0.3	0.0	0.6	0.1	0.5	0.2	0.7 2.0	0.1
667 672	671 687	2		FFWHL KSEMHNKTVSQRFGLL	1.7 2.5	0.5	16.9	0.9	0.8 2.5	0.3	17.1	0.6
688	692	1	7.65	LESYC	1.8	0.6	2.1	0.6	2.6	0.9	2.7	0.6
691 698	697 709	2		YCRACGM YLKHLNRQVEAM	13.9	0.2	30.3	0.5	14.1	1.1 0.3	30.7	0.7
716	734	3		TDILKQEKKDETQKVQMKF	27.3	0.1	54.0	0.4	27.1		55.5	1.0
735	744	2		LVEQMRRPDF	5.2	0.4	61.6		4.8	0.6	63.3	3.3
745 769	766 781	4		MDALQGFLSPLNPAHQLGNLRL CRIMSSAKRPLWL	2.5 11.3	0.3	8.7 26.2	0.1	10.8	0.5	8.9 24.0	1.0
782	791	1		NWENPDIMSE	23.0	0.2	51.4	0.2	22.3		52.3	1.5
793	797	1		LFQNN	48.0	0.8	64.9	0.2	45.7	2.5	65.0	1.8
794 798	798 806	2		FQNNE EIIFKNGDD	22.8 5.0	0.9	60.0 18.3	2.1 0.4	20.6	0.4	57.5	2.4
799	807	1		IIFKNGDDL	6.3	0.4	22.7	0.8				
799 799	811 813	3		IIFKNGDDLRQDM IIFKNGDDLRODMLT	3.2	0.2	24.6 16.5	0.3				
802	811	2		KNGDDLRQDM	4.3	0.1	30.4	1000				
815	821	2	9.55	QIIRIME	0.3	0.1	0.4	0.1	0.4	0.1		0.2
822 830	830 839	1 2		NIWQNQGLD DLRMLPYGCL	9.2	0.2	26.8 10.8	0.3 1.6	9.9	1.1	27.9 9.4	0.6
840	847	1	10.93	SIGDCVGL	2.9	0.4	8.1	0.3	3.0	777	8.6	0.3
847	858	2		LIEVVRNSHTIM	9.6	0.4	21.7	0.2	4.5	0.5	20.5	1.2
847 848	859 858	3		LIEVVRNSHTIMQ IEVVRNSHTIM	8.6 10.4	0.2	23.6 26.0		4.1 5.4	0.5	20.7	1.5 0.7
848	859	2	7.12	IEVVRNSHTIMQ	9.7	0.2	27.4	0.4	5.2	0.7	24.1	0.6
860	872 893	3		IQCKGGLKGALQF NSHTLHQWLKDKNKGEIYDAA	63.6	0.3	66.5	0.2	63.3	0.9	66.4	1.3
873 880	893 893	2		NSHTLHQWLKDKNKGEIYDAA WLKDKNKGEIYDAA	6.5 8.6	0.1	23.5 32.4		6.9 9.0	-	24.4 33.2	0.6
892	896	1	9.46	AAIDL	2.3	1.1	2.8	0.8	2.4	1.6	3.1	0.4
909	922			FILGIGDRHNSNIM	1.8	0.2	7.7	0.1	1.8	0.3	7.8	0.4
921 930	930 956	5		IMVKDDGQLF FHIDFGHFLDHKKKKFGYKRERVPFVL	3.2 4.1	0.5	4.5 17.0	0.8	3.4 3.9	1.0 0.3	5.4 16.9	0.5
957	961	1	10.45	TQDFL	1.5	0.1	10.4	0.1	1.8	0.5	10.7	0.5
961 977	980 983	4		LIVISKGAQECTKTREFERF FERFOEM	11.7 2.0	1.3	23.4	0.5	10.9		24.3	2.0
977	983 989	1		FERFQEM CYKAYL	2.0	1.6 0.7	3.4 1.6	0.9	2.8	0.6	4.3 2.6	0.6
990	997	2	5.07	AIRQHANL	2.8	0.2	30.7	0.5	2.8	0.4	32.0	1.2
1002 1006	1006	1		FSMML LGSGMPEL	0.9 9.2	0.4	10.5 40.0	0.2	1.1 8.8	0.8	14.2 40.2	0.4
1014	1013	1		QSFDDIA	16.2	0.8	18.9	0.7	16.4			1.2
1017	1021	1	7.84	DDIAY	3.5	0.5	24.0	0.2	3.3	1.1	24.5	0.8
1029	1037	2		DKTEQEALE	10.3 15.7	0.1	32.8 23.4	0.8	10.3	0.4	33.3	1.0
1039 1039	1055	4		FMKQMNDAHHGGWTTKM FMKQMNDAHHGGWTTKMDW	15.7	0.1	23.4	0.2	15.8	0.1	24.0 22.7	0.7
1060	1068	2		HTIKQHALN	46.2	0.2	52.2	0.8	45.8	0.1	53.4	1.9

Figure S11. 1 H NMR kinetics data for **6** reacting with N^{α} -acetyl lysine in phosphate-buffered $D_{2}O$, pH 8.1, 25°C

$$\begin{array}{c} O \\ \hline N \\ \hline O \\ \hline$$

time, min.	In([6]), 8.46 ppm	In([6]), 7.48 ppm
5	-6.918118066	-6.918566195
10	-6.96067768	-6.960123264
15	-7.032918422	-7.037289331
20	-7.044091723	-7.03558865
25	-7.148035113	-7.083645945
30	-7.125757431	-7.128426641
35	-7.188657664	-7.205387682
40	-7.308078988	-7.184008656
45	-7.338538195	-7.3678788
50	-7.301659565	-7.318497444
55	-7.517132576	-7.437014604
60	-7.6102922	-7.448322872



	signal at 8.46 ppm	signal at 7.48 ppm	average
slope of In([6]) vs. time	-0.0116	-0.0099	-0.01075ª
calculated half-life, min.	59.8	70.0	64.5 ^b

^aAverage of slopes of each linear regression line

^bCalculated by $t_{1/2} = \ln(2)/(-1*average slope)$

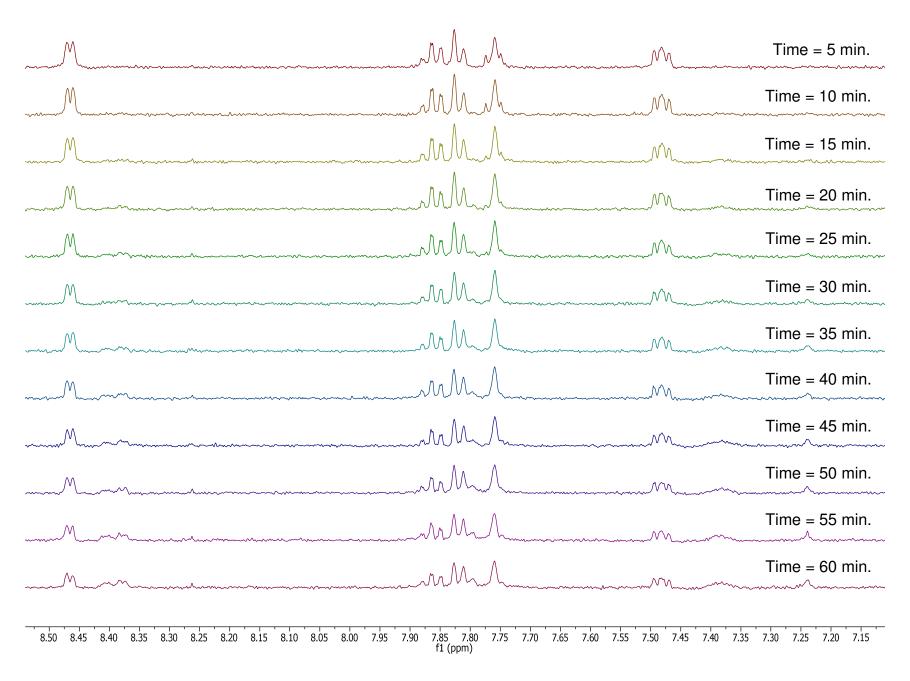
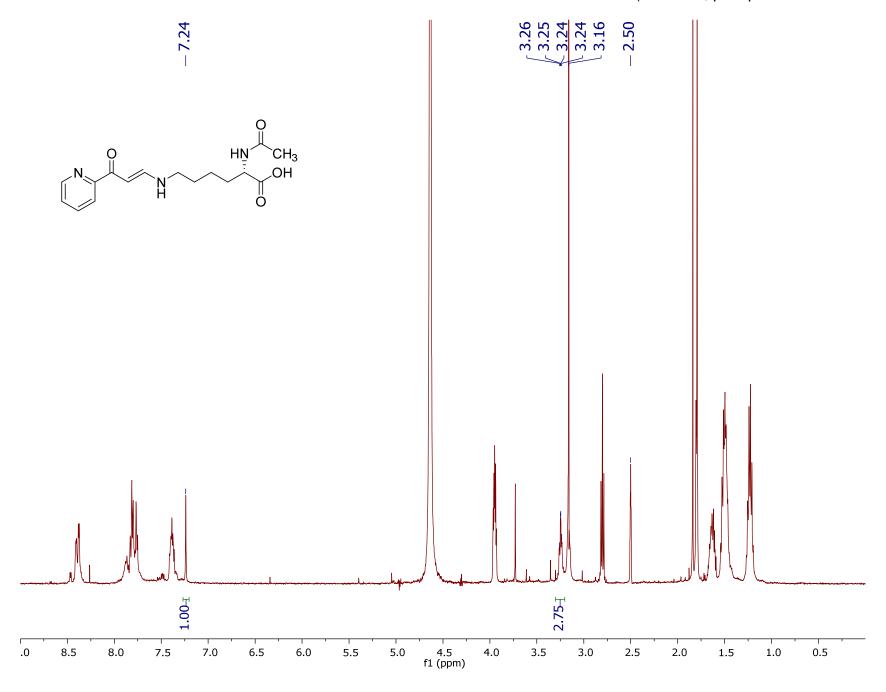
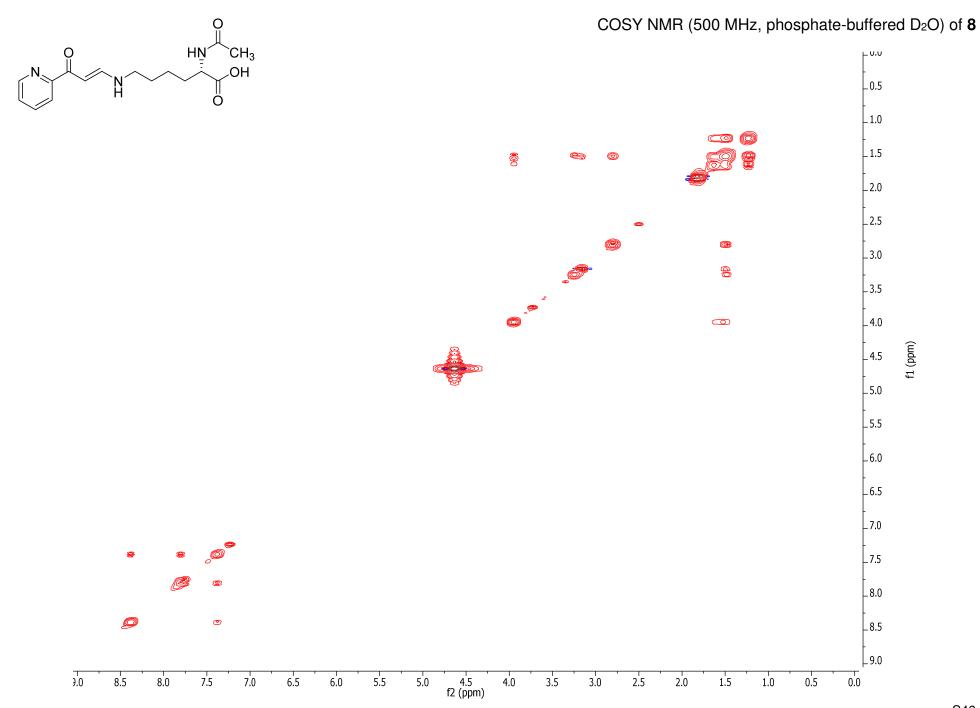


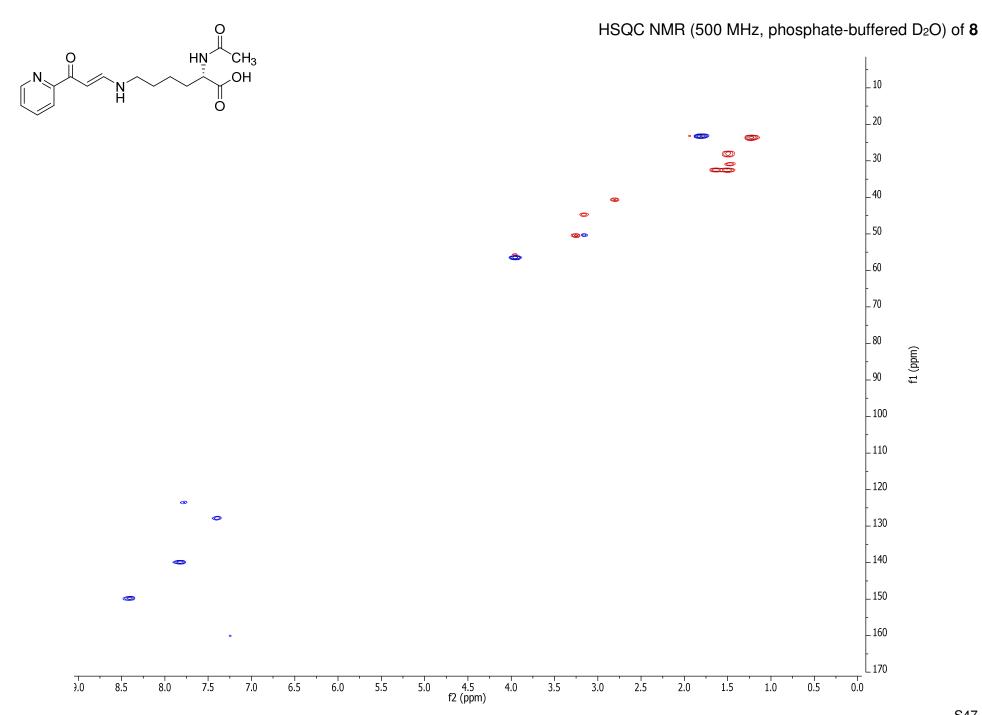
Table S9. NMR spectral data for 8 in phosphate-buffered D2O, pH 8.1, at 500 MHz

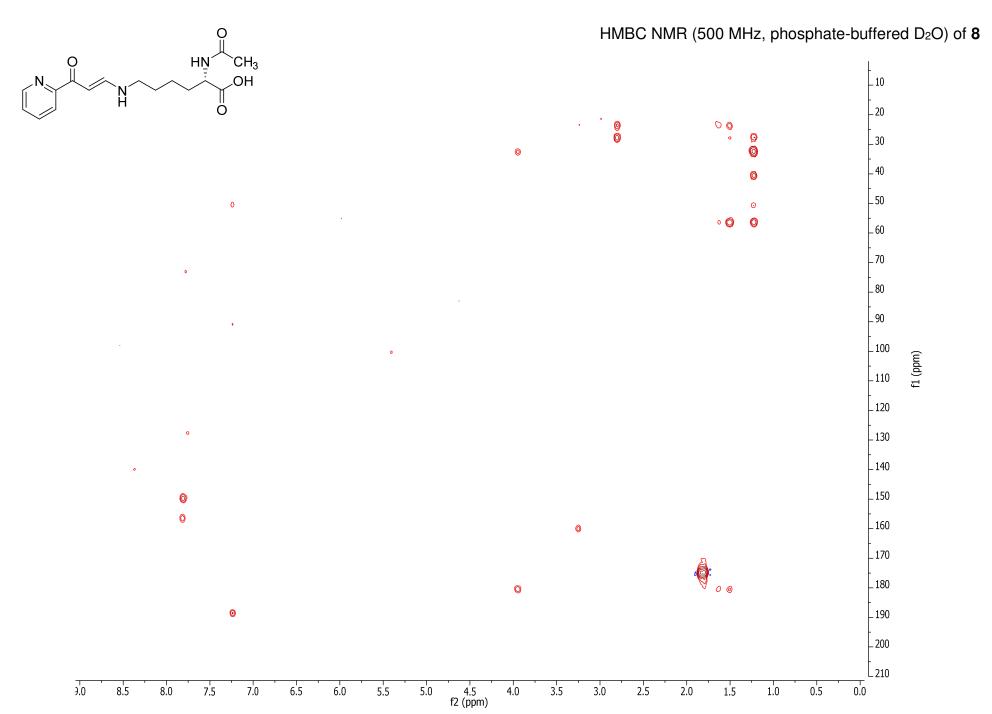
Position #	δ _H (mult., J(Hz))	δc ^b	COSY	HMBC
1	8.39 (dd, 13.5, 4.4)	149.8	2	3
2	7.39 (m)	127.8	1, 3	4
3	7.80 (m, 7.5)	139.9	2, 4	1, 5
4	7.76 (m, 7.5)	123.5	3	2
5		156.0		
6		188.0		
7	a	90.6		
8	7.24 (s)	160.0		6, 7, 10
9-NH	a			
10	3.25 (td, 6.5, 2.3)	50.4	11	8, 11, 12
11	1.52 (m)	27.8	10, 12	10, 12
12	1.23 (m)	23.6	11, 13	10, 11, 13, 14
13	a 1.52 (m) b 1.63 (m)	32.5	14 12, 13a, 14	11, 12, 14, 15 12, 14, 15
14	3.95 (m)	56.4	13a, 13b	12, 13, 15
15		180.4		
16-OH	a			
17-NH	a			
18		174.8		
19	1.81 (s)	23.2		18

^a Proton signals were not observed due to exchange with deuterium. ^b Carbon chemical shifts were based on HSQC and HMBC data.









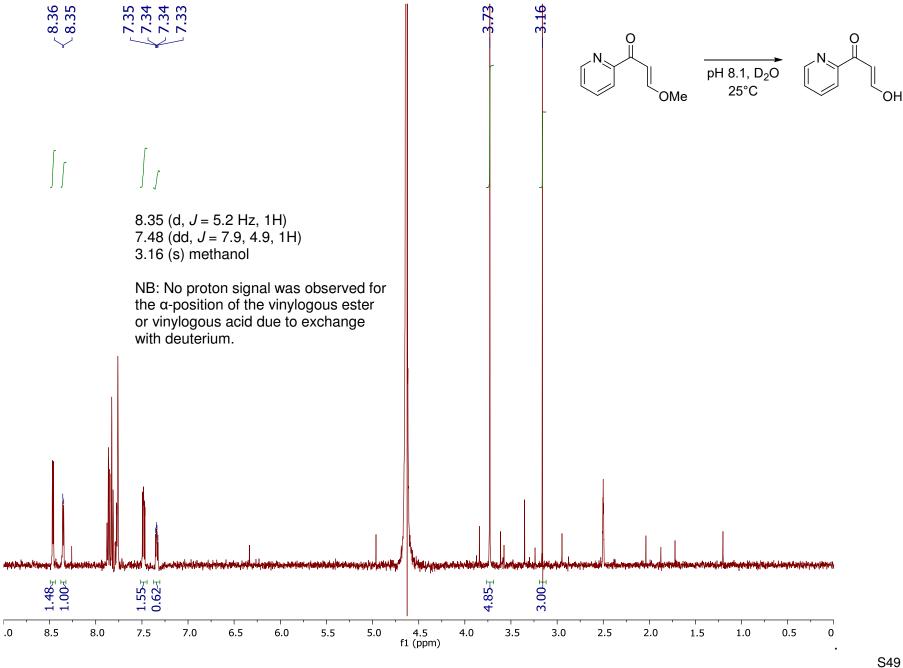


Table S10. Results of competition binding assay for neolymphotin against a panel of 97 kinases

Target	Neolymphostin A
Gene Symbol	%Ctrl @ 1000nM
ABL1(E255K)-phosphorylated	70
ABL1(T315I)-phosphorylated	75
ABL1-nonphosphorylated	39
ABL1-phosphorylated	76
ACVR1B	96
ADCK3	87
AKT1	100
AKT2	97
ALK	100
AURKA	1.5
AURKB	1.9
AXL	39
BMPR2	57
BRAF	77
BRAF(V600E)	77
BTK	15
CDK11	59
CDK2	87
CDK3	96
CDK7	50
CDK9	89
CHEK1	88
CSF1R	82
CSNK1D	75
CSNK1G2	33
DCAMKL1	83
DYRK1B	80
EGFR	58
EGFR(L858R)	57
EPHA2	96
ERBB2	28
ERBB4	90
ERK1	90
FAK	98
FGFR2	94
FGFR3	91
FLT3	74
GSK3B	89
IGF1R	96
IKK-alpha	78
IKK-beta	83
INSR	93
JAK2(JH1domain-catalytic)	62
JAK3(JH1domain-catalytic)	18
JNK1	0.25
JNK2	0.25
JNK3	0.13
KIT	84
KIT(D816V)	91
141(55157)	J 1

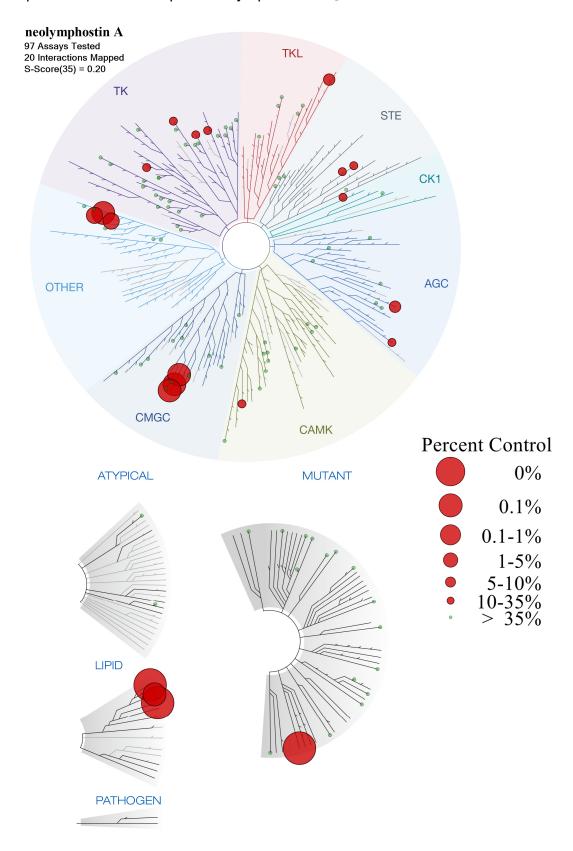
Table S10. (continued)

Target	Neolymphostin A
Gene Symbol	%Ctrl @ 1000nM
KIT(V559D,T670I)	73
LKB1	93
MAP3K4	88
MAPKAPK2	98
MARK3	100
MEK1	10
MEK2	12
MET	60
MKNK1	87
MKNK2	97
MLK1	8
p38-alpha	100
p38-beta	100
PAK1	100
PAK2	98
PAK4	100
PCTK1	70
PDGFRA	61
PDGFRB	95
PDPK1	100
PIK3C2B	0
PIK3CA	0
PIK3CG	0.55
PIM1	100
PIM2	100
PIM3	91
PKAC-alpha	89
PLK1	100
PLK3	76
PLK4	0.95
PRKCE	18
RAF1	90
RET	86
RIOK2	41
ROCK2	93
RSK2(Kin.Dom.1-N-terminal)	6.1
SNARK	27
SRC	89
SRPK3	69
TGFBR1	97
TIE2	61
TRKA	12
TSSK1B	93
TYK2(JH1domain-catalytic)	60
ULK2	75
VEGFR2	68
YANK3	99
ZAP70	72

%Ctrl Legend:

0≤x<.1 .1≤x<1 1s	<10 10≤x<35 x≥35
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Figure S12. TREEspot™ interaction map for neolymphostin A @ 1000nM

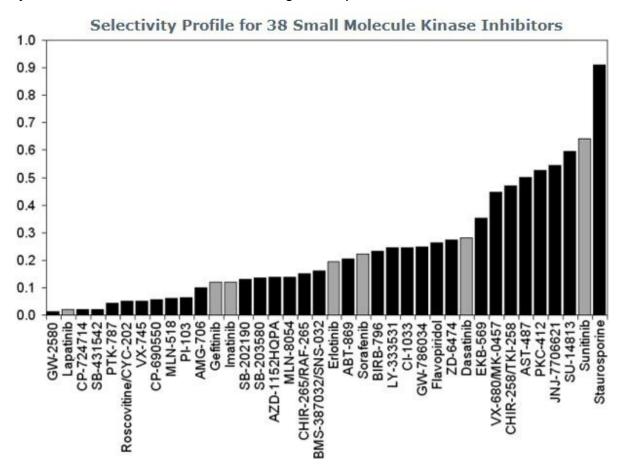


TREEspot™ is a proprietary data visualization software tool developed by KINOMEscan. Kinases found to bind are marked with red circles, where larger circles indicate higher-affinity binding

Table S11. Selectivity S-Score for neolymphostin A against a 97 kinase panel

Compound Name	Selectivity Score Type	Number of Hits	Number of Non-Mutant Kinases	Screening Concentration (nM)	Selectivity Score
Neolymphostin A	S(35)	20	90	1000	0.222
Neolymphostin A	S(10)	11	90	1000	0.122
Neolymphostin A	S(1)	7	90	1000	0.078

Selectivity S35-Score for 38 kinase inhibitors against a panel of 287 kinases



KINOMEscan's in vitro competition binding assay was used to evaluate 38 kinase inhibitors against a panel of 287 distinct human protein kinases (~55% of the predicted human protein kinome), and three lipid kinases. The compounds tested included 21 tyrosine kinase inhibitors, 15 serine-threonine kinase inhibitors, 1 lipid kinase inhibitor and staurosporine. S(35) = (number of non-mutant kinases with %Ctrl <35)/(290 kinases tested; 27 mutant variants were excluded from this analysis). Compounds approved for use in humans (as of August, 2007) are highlighted (gray bars).

Figure S13. NCI 60-cell line dose response curves for neolymphostin A

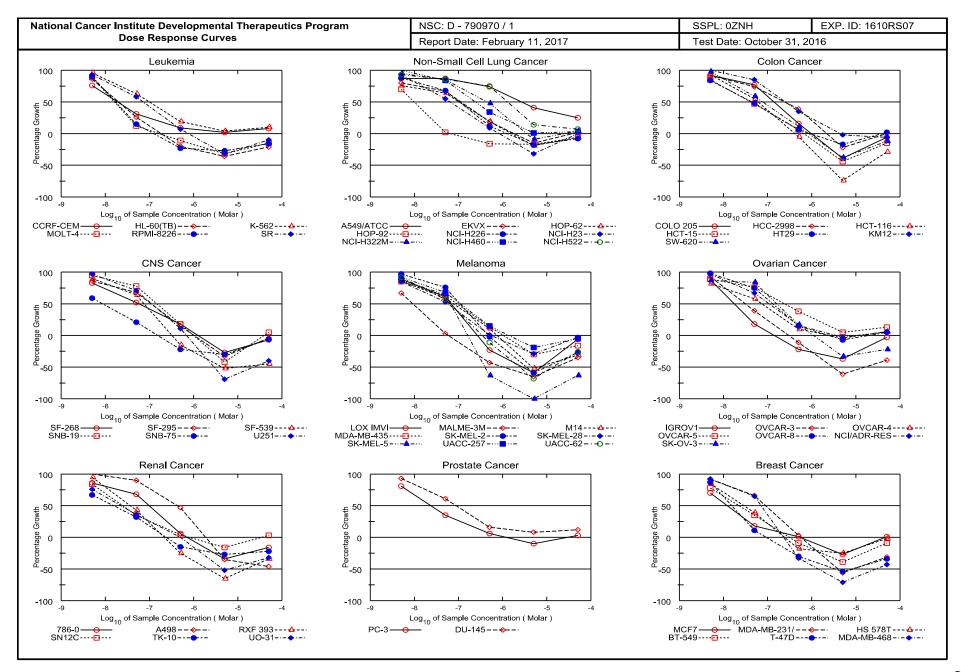


Figure S14. NCI 60-cell line testing results for neolymphostin A

		Nati	onal (Cand	er Ir			evelop Testir				peutio	s Progra	m	
NSC : D - 79	NSC : D - 790970 / 1					Experiment ID: 1610RS07						Test	Test Type : 08		1olar
Report Date	: Februaı	∕y 11, 20)17		Tes	t Date	: Octob	er 31, 20	16			QNS	:	MC :	
COMI : Y486	.338				Sta	in Rea	gent : S	RB Dual-	Pass I	Related	i	SSPL	: 0ZNH		
Panel/Cell Line	Time Zero	Ctrl	-8.3	Mear -7.3	Optica -6.3	Lo I Densiti -5.3	-	centration	P -7.3	ercent G -6.3	Frowth -5.3	-4.3	GI50	TGI	LC50
Leukemia CCRF-CEM HL-60(TB) K-562 MOLT-4 RPMI-8226 SR	0.382 0.809 0.228 0.563 0.926 0.568	1.593 3.021 2.145 2.041 2.303 2.337	1.305 2.720 2.069 1.872 2.162 2.220	0.763 1.384 1.434 0.737 1.139 1.601	0.492 0.649 0.595 0.503 0.717 0.692	0.404 0.521 0.301 0.383 0.675 0.387	0.476 0.640 0.416 0.477 0.777 0.511	76 86 96 89 90	31 26 63 12 15 58	9 -20 19 -11 -23 7	2 -36 4 -32 -27 -32	8 -21 10 -15 -16 -10	1.93E-8 2.00E-8 9.86E-8 1.59E-8 1.71E-8 7.28E-8	> 5.00E-5 1.84E-7 > 5.00E-5 1.66E-7 1.28E-7 7.57E-7	> 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5
Non-Small Cell Lur A549/ATCC EKVX HOP-62 HOP-92 NCI-H226 NCI-H23 NCI-H322M NCI-H460 NCI-H522	ng Cancer 0.470 0.853 0.945 1.002 0.768 0.648 0.967 0.183 0.922	1.819 2.298 2.068 1.574 1.285 2.032 2.254 1.894 2.081	1.645 2.016 1.793 1.405 1.222 1.938 2.185 1.889 2.006	1.639 1.824 1.668 1.014 1.122 1.403 2.061 1.617 1.924	1.470 1.121 1.156 0.837 0.832 0.771 1.582 0.771 1.791	0.770 0.834 0.627 0.441 0.884 0.195	0.807 0.882 0.892 0.939 0.705 0.649 1.017 0.241 1.006	87 80 76 70 88 93 95 100	87 67 64 2 68 55 85 84 86	74 19 19 -16 12 9 48 34 75	41 -15 -19 -17 -18 -32 -9 1	25 2 -6 -6 -8 4 3 7	2.73E-6 1.13E-7 1.03E-7 9.94E-9 1.06E-7 6.29E-8 4.36E-7 2.41E-7 1.29E-6	> 5.00E-5 1.59E-6 6.42E-8 1.26E-6 > 5.00E-5 > 5.00E-5	> 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5
Colon Cancer COLO 205 HCC-2998 HCT-116 HCT-15 HT29 KM12 SW-620	0.580 0.864 0.252 0.330 0.228 0.566 0.254	1.888 2.666 2.145 2.224 1.362 2.852 1.734	1.785 2.528 2.016 1.941 1.181 2.866 1.699	1.587 2.206 1.296 1.213 0.778 2.518 1.131	0.787 1.568 0.240 0.442 0.293 1.358 0.415	0.357 0.677 0.065 0.185 0.189 0.555 0.157	0.546 0.858 0.179 0.281 0.256 0.535 0.223	92 92 93 85 84 101 98	77 74 55 47 48 85 59	16 39 -5 6 6 35	-39 -22 -74 -44 -17 -2	-6 -29 -15 2 -6 -12	1.38E-7 2.45E-7 6.09E-8 4.08E-8 4.53E-8 2.49E-7 7.76E-8	9.76E-7 2.20E-6 4.16E-7 6.56E-7 4.40E-6 8.33E-7	> 5.00E-5 > 5.00E-5 - > 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5
CNS Cancer SF-268 SF-295 SF-399 SNB-19 SNB-75 U251	0.752 0.611 1.194 0.265 0.941 0.495	2.376 2.825 3.050 1.047 1.875 1.932	2.096 2.510 2.861 1.007 1.496 1.896	1.594 2.123 2.375 0.878 1.134 1.519	1.048 0.937 1.019 0.404 0.734 0.656	0.546 0.408 0.578 0.154 0.663 0.154	0.699 0.585 0.662 0.302 0.889 0.297	83 86 90 95 59	52 68 64 78 21 71	18 15 -15 18 -22	-27 -33 -52 -42 -30 -69	-7 -4 -45 5 -6 -40	5.67E-8 1.10E-7 7.46E-8 1.47E-7 8.74E-9 1.13E-7	1.25E-6 1.01E-6 3.25E-7 1.52E-7 6.90E-7	> 5.00E-5 > 5.00E-5 -> 5.00E-5 > 5.00E-5
Melanoma LOX IMVI MALME-3M M14 MDA-MB-435 SK-MEL-2 SK-MEL-28 SK-MEL-5 UACC-257 UACC-62	0.462 0.588 0.406 0.495 1.000 0.560 0.727 1.025 0.899	2.614 1.044 1.751 2.356 2.074 1.718 2.593 1.941 2.891	2.386 0.892 1.558 2.084 2.037 1.543 2.345 1.866 2.729	1.769 0.602 1.200 1.672 1.816 1.177 1.932 1.648 2.025	0.333 0.549 0.759 0.982 0.571 0.270 1.161	-0.002 0.834	0.435 0.385 0.281 0.415 0.729 0.543 0.266 0.969 0.663	89 67 86 85 97 85 87 92	61 3 59 63 76 53 65 68 57	-23 -43 11 14 -2 1 -63 15 -11	-59 -65 -52 -30 -59 -29 -100 -19 -68	-6 -35 -31 -16 -27 -3 -63 -5	6.72E-8 9.11E-9 7.69E-8 9.31E-8 1.08E-7 5.77E-8 6.50E-8 1.09E-7 6.25E-8	2.67E-7 5.79E-8 7.39E-7 1.05E-6 4.74E-7 5.38E-7 1.61E-7 1.38E-6 3.42E-7	> 5.00E-5 > 5.00E-5 3.96E-7 > 5.00E-5
Ovarian Cancer IGROV1 OVCAR-3 OVCAR-4 OVCAR-5 OVCAR-8 NCI/ADR-RES SK-OV-3	0.532 0.441 0.705 0.733 0.522 0.647 1.104	1.905 1.409 1.594 2.242 2.037 2.289 2.092	1.341 1.438 2.077 2.009	1.219 1.876 1.658	0.393 0.798 1.304 0.747 0.907	0.170	0.271 0.759 0.932 0.592	88 93 82 89 98 97 87	18 39 58 76 75 67 84	-22 -11 10 38 15 16 17	-37 -61 -4 5 -7 -3	-3 -39 6 13 5 6	1.73E-8 3.15E-8 7.30E-8 2.39E-7 1.30E-7 1.07E-7 1.61E-7	1.39E-7 3.03E-7 > 5.00E-5 1.11E-6	> 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5 > 5.00E-5
Renal Cancer 786-0 A498 RXF 393 SN12C TK-10 UO-31	0.636 1.153 0.641 0.522 1.429 0.508	2.392 1.897 1.212 2.076 2.506 1.898	1.931 1.186 1.835 2.149	1.823	0.480 0.588 1.218	0.744 0.224 0.438 1.040	0.533 0.626 0.426 0.564 1.113 0.345	86 105 95 84 67 76	68 90 43 35 32 36	6 47 -25 4 -15	-34 -35 -65 -16 -27 -52	-16 -46 -34 3 -22 -32	9.81E-8 4.27E-7 3.65E-8 2.50E-8 1.51E-8 2.26E-8	6.95E-7 1.86E-6 2.13E-7 2.41E-7 5.04E-7	> 5.00E-5 > 5.00E-5 -> 5.00E-5 > 5.00E-5
Prostate Cancer PC-3 DU-145	0.588 0.346	1.649 1.584		0.957 1.099		0.530 0.447	0.623 0.497	81 93	35 61	6 16	-10 8	3 12	2.32E-8 8.75E-8	> 5.00E-5	> 5.00E-5 > 5.00E-5
Breast Cancer MCF7 MDA-MB-231/ATC HS 578T BT-549 T-47D MDA-MB-468	0.360	1.954 1.251 2.145 2.212 1.610 1.855	1.472 1.199 1.989 1.967 1.513	0.642 1.023 1.506 1.496	0.373 0.595 0.904 1.013 0.592	0.262 0.255 0.832 0.681 0.392	0.384 0.394 1.087 1.017 0.563	70 92 85 78 87 92	18 66 39 35 11 65	1 3 -18 -9 -30 -33	-27 -56 -25 -39 -54 -71	1 -31 -2 -9 -34 -43	1.20E-8 9.08E-8 2.83E-8 2.21E-8 1.53E-8 7.12E-8	5.68E-7 2.39E-7 3.11E-7 9.13E-8 2.32E-7	> 5.00E-5 > 5.00E-5 > 5.00E-5

Figure S15. NCI 60-cell line mean graphs for neolymphostin A

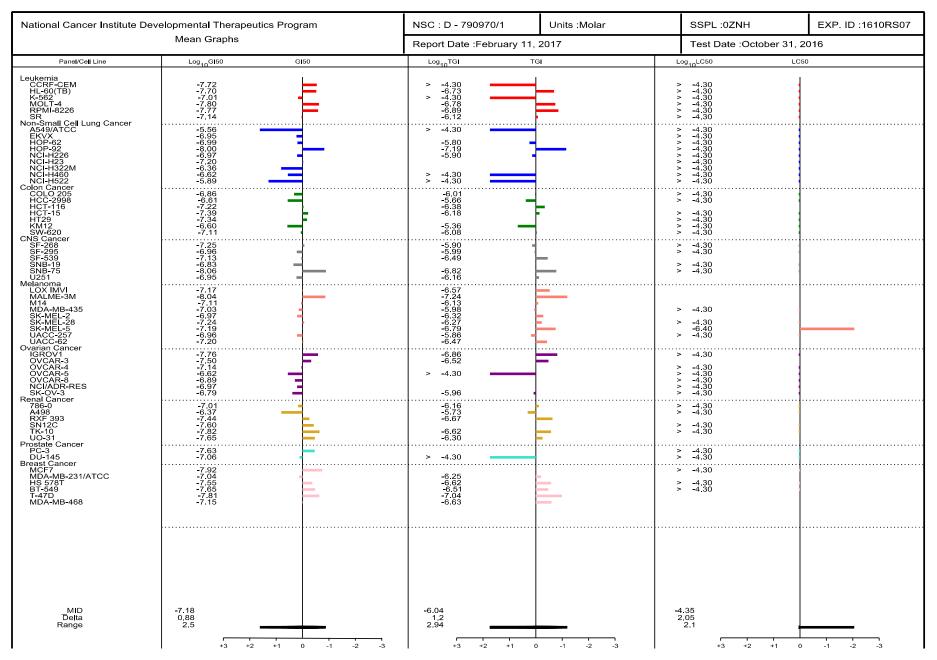


Figure S16. NCI 60-cell line dose response curves for neolymphostin A (all)

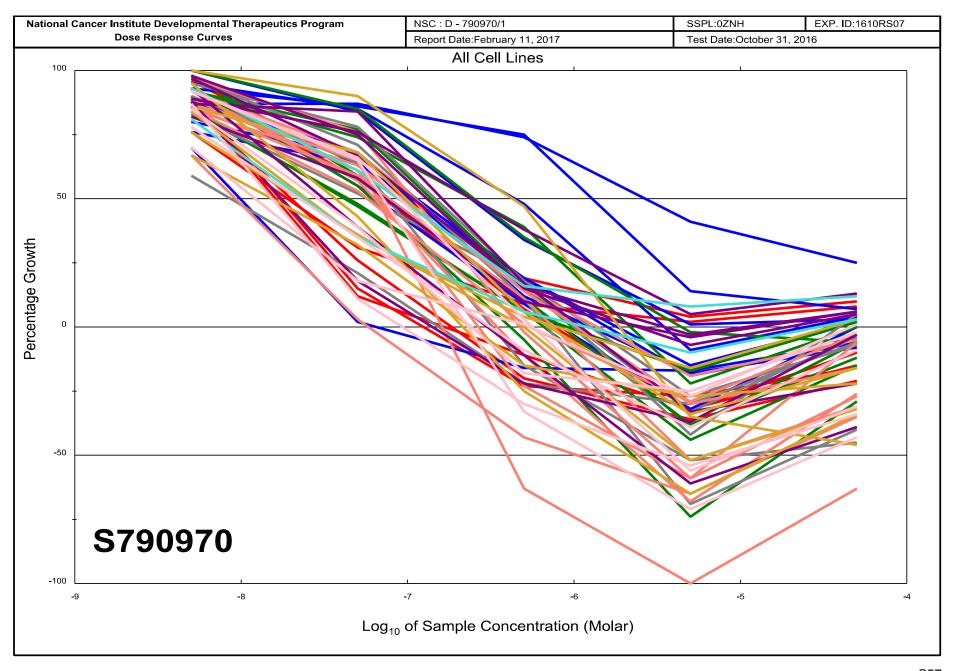


Figure S17. COMPARE results

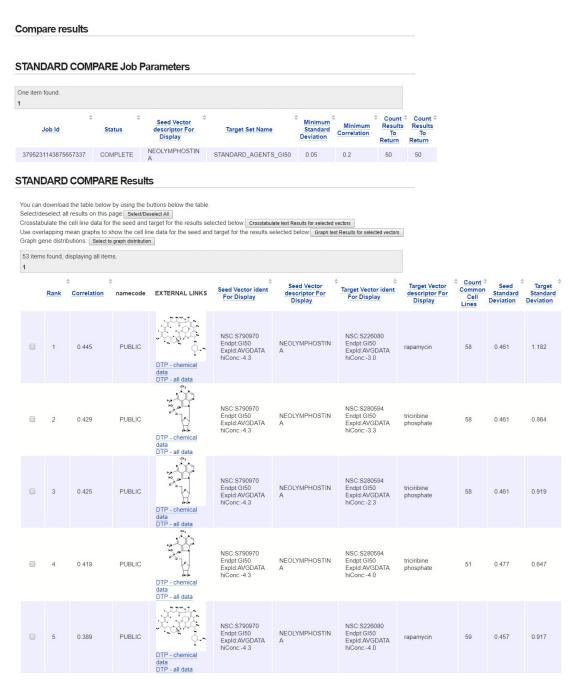


Figure S17. COMPARE results (continued)

6	0.378	PUBLIC	DTP - chemical data	NSC:S790970 Endpt:GI50 Expld:AVGDATA hiConc:-4.3	NEOLYMPHOSTIN A	NSC:S73754 Endpt:GI50 Expld:AVGDATA hiConc:-2.9	fluorodopan	43	0.492	0.195
7	0.367	PUBLIC	DTP - chemical data DTP - all data	NSC:S790970 Endpt:GI50 Expld:AVGDATA hiConc:-4.3	NEOLYMPHOSTIN A	NSC:S226080 Endpt:GI50 Expld:AVGDATA hiConc:-7.0	rapamycin	59	0.457	0.778
8	0.358	PUBLIC	H ₃ C O TN - CH ₃ H ₃ C O TN - CH ₃ DTP - chemical data DTP - all data	NSC:S790970 Endpt:GI50 Expld:AVGDATA hiConc:-4.3	NEOLYMPHOSTIN A	NSC:S253272 Endpt:GI50 Expld:AVGDATA hiConc:-2.0	caracemide	58	0.461	0.336
9	0.356	PUBLIC	structure not found or not known DTP - chemical data DTP - all data	NSC:S790970 Endpt:GI50 Expld:AVGDATA hiConc:-4.3	NEOLYMPHOSTIN A	NSC:S167780 Endpt:GI50 Expld:AVGDATA hiConc:-3.9	asaley	58	0.461	0.431
10	0.337	PUBLIC	DTP - chemical data DTP - all data	NSC:S790970 Endpt:GI50 Expld:AVGDATA hiConc:-4.3	NEOLYMPHOSTIN A	NSC:S291643 Endpt:GI50 Expld:AVGDATA hiConc:-2.3	pyrimidine-5- glycodialdehyde	58	0.461	0.167