







































































Supplementary Information

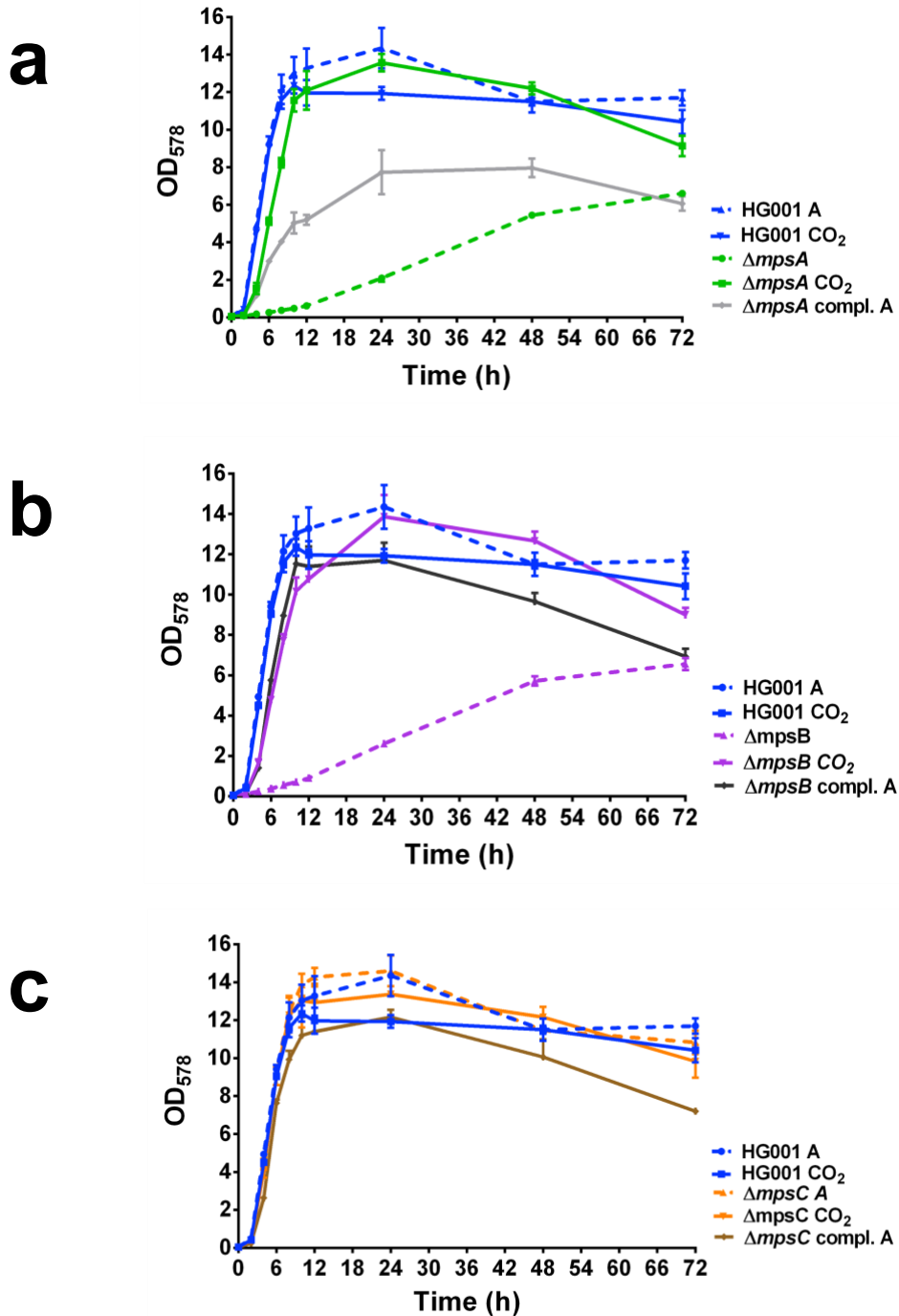
MpsAB is important for *Staphylococcus aureus* virulence and growth under atmospheric CO₂ levels

Fan et al.

Strains		Day 1	Day 2	Day 3	Day 4	Day 5
HG001	A					
	CO ₂					
$\Delta mpsA$	A					
	CO ₂					
pRB473-mpsA	A					
	CO ₂					
$\Delta mpsB$	A					
	CO ₂					
pctuf-mpsB	A					
	CO ₂					
$\Delta mpsC$	A					
	CO ₂					
pRB473-mpsC	A					
	CO ₂					

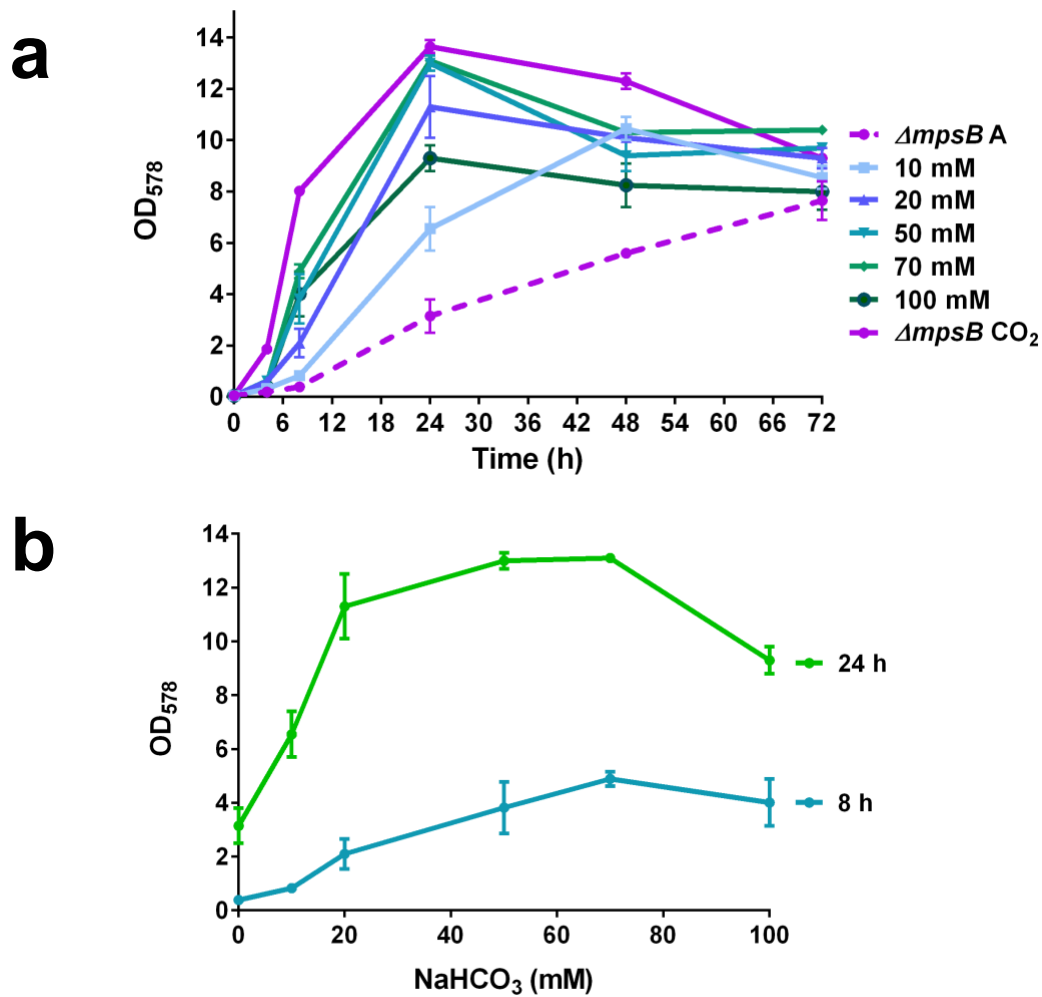
Supplementary Figure 1: Growth of HG001 $\Delta mpsA$, $\Delta mpsB$ and $\Delta mpsC$

Colony appearance of wild type HG001, $\Delta mpsA$, $\Delta mpsB$ and $\Delta mpsC$ and its corresponding complemented mutant during five days of incubation in atmospheric conditions and 5% CO₂. White bar represents a scale of 1 mm.



Supplementary Figure 2: Growth of HG001 *mps* deletion mutants

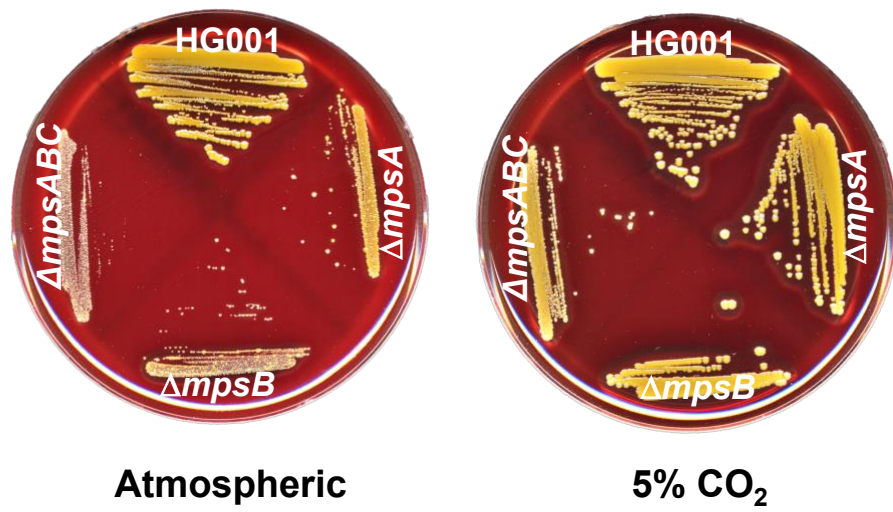
Growth of HG001, (a) $\Delta mpsA$, (b) $\Delta mpsB$ and (c) $\Delta mpsC$ and their corresponding complemented mutant in atmospheric conditions (A) and 5% CO₂. The growth of the deletion mutant *mpsA* and *mpsC* can also be complemented by plasmid pRB473 carrying *mpsA* and *mpsC* respectively. The growth of deletion mutant *mpsB* was complemented by plasmid pctuf carrying *mpsB*. Each point shown in the graph represents the mean value \pm standard error mean (SEM) from three independent experiments. Source data are provided as a Source Data file.



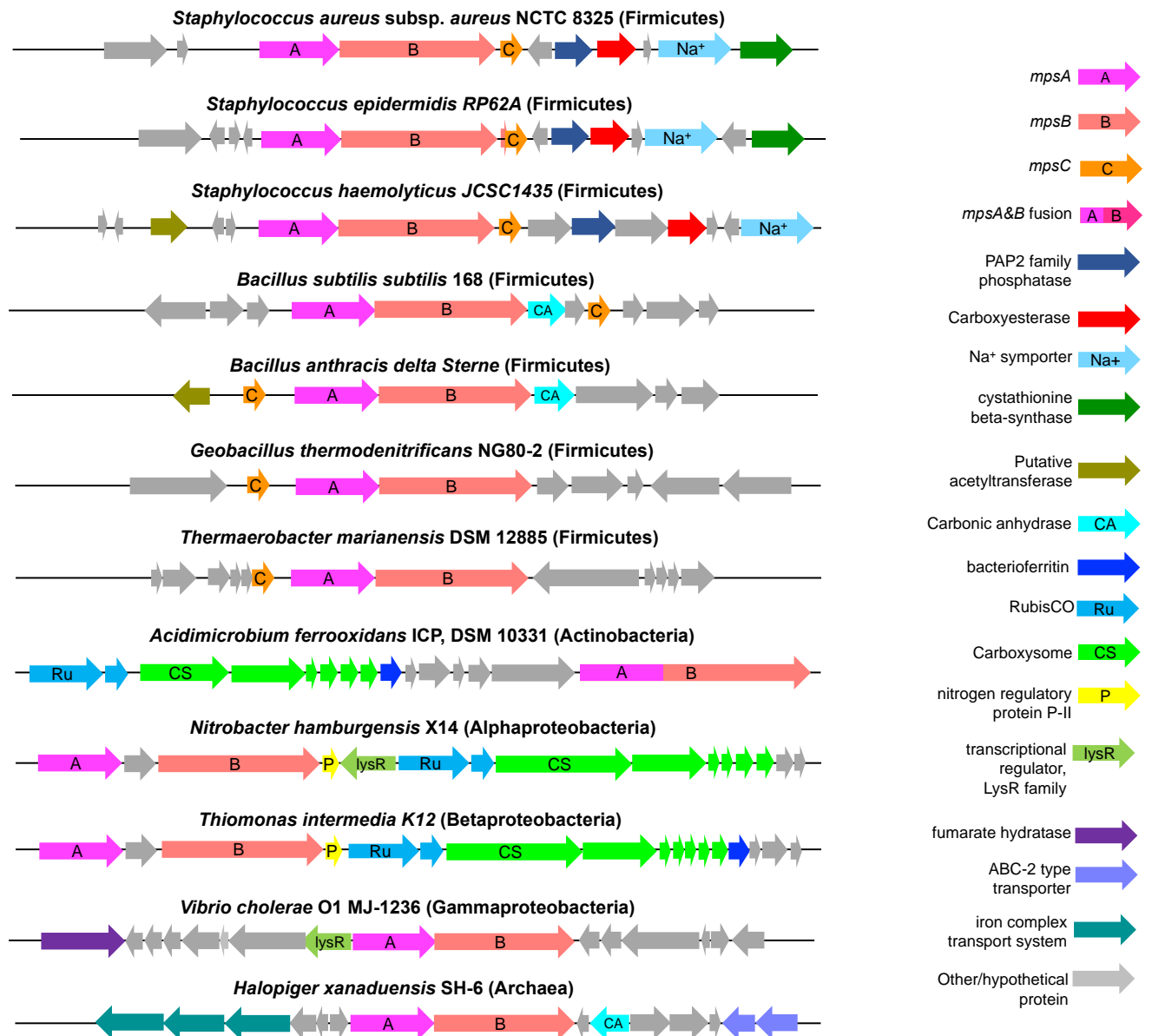
Supplementary Figure 3: Growth of HG001 $\Delta mpsB$ can be complemented with NaHCO_3

(a) Growth of $\Delta mpsB$ in atmospheric (A) and with additions of NaHCO_3 and also 5% CO_2 . Each concentration of NaHCO_3 is indicated in the graph, ranging from 10 mM to 100 mM. Additions of NaHCO_3 improved the growth of $\Delta mpsB$ in a concentration dependent manner up to 70 mM. Each point shown in the graph represents the mean value \pm standard error mean (SEM) from two independent experiments.

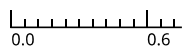
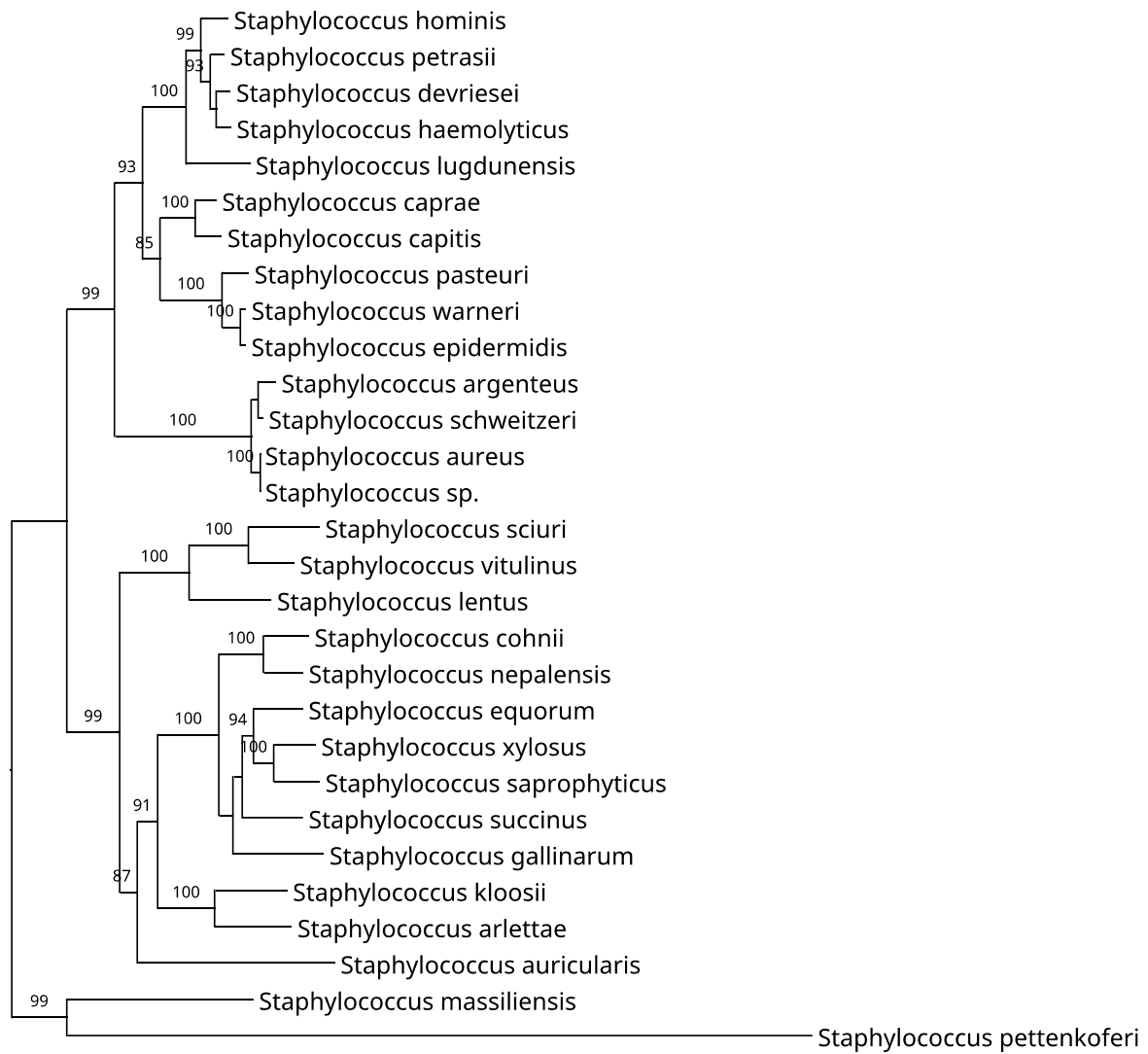
(b) The same graph illustrated with OD as a function of bicarbonate concentration at 8 h and 24 h. Source data are provided as a Source Data file.



Supplementary Figure 4: The production of hemolytic toxin in HG001 and its three *mps* mutant strains ($\Delta mpsA$, $\Delta mpsB$ and $\Delta mpsABC$) grown under atmospheric and 5% CO₂ conditions.

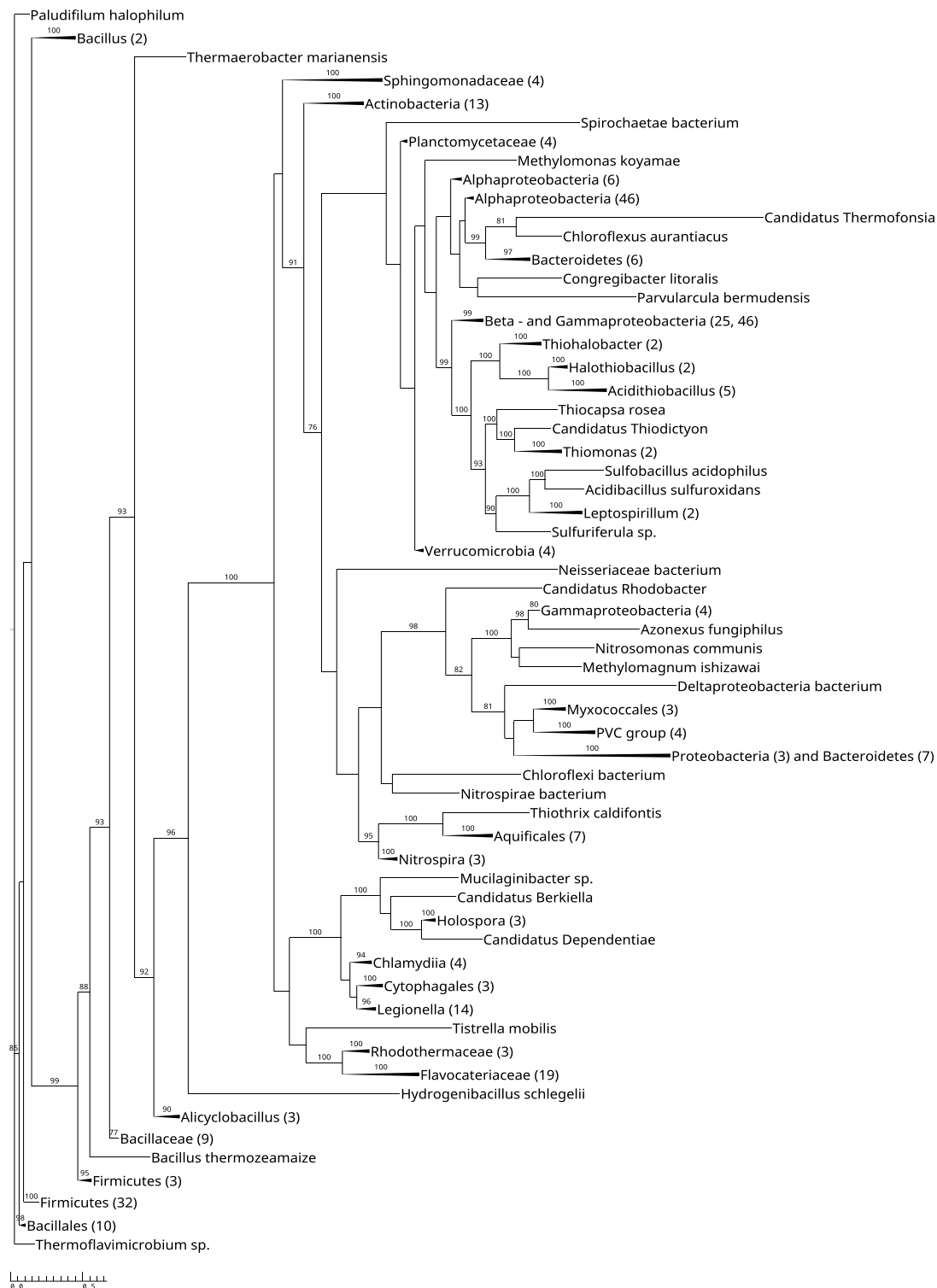


Supplementary Figure 5: Gene synteny presenting the genome organization around *mpsAB* in *S. aureus* and its homologs in selected genomes
mpsAB homologs are almost always collocated. Homologs of *mpsAB* exist as a single fused gene in *Acidimicrobium ferrooxidans*. *mpsAB* always points to the right, even if it is located on the complementary strand.



Supplementary Figure 6: Phylogenetic tree of Staphylococci

Maximum likelihood phylogenetic analysis of concatenated alignments of *mpsA* and *mpsB* homologs amongst the genus *Staphylococcus*. Node support is indicated by bootstrap values from 100 resamplings of the alignment when they exceeded 70%. Source data are provided as a Source Data file.



Supplementary Figure 7: Phylogenetic tree of all bacterial phyla

Maximum likelihood phylogenetic analysis of concatenated alignments of *mpsA* and *mpsB* homologs amongst the bacterial domain. Node support is indicated by bootstrap values from 100 resamplings of the alignment when they exceeded 70%. Collapsed clades are labeled by the shared taxonomic rank of associated taxa. The number of taxa belonging to the respective clade is indicated in brackets. Source data are provided as a Source Data file.

Supplementary Table 1: Bacterial strains used in this study

Strain	Description	Reference or source
<i>E. coli</i> EDCM636	<i>E. coli</i> MG1655 ($\Delta canA1:: FLK2(lacZ,kan)$, λ^- , $\Delta fnr-267,rph-1$) replacement of <i>can</i> by kanamycin resistance marker	¹
<i>E. coli</i> EDCM636 <i>mpsA</i>	<i>E. coli</i> EDCM636 (pRB473- <i>mpsA</i>) carrying <i>mpsA</i>	This study
<i>E. coli</i> EDCM636 <i>mpsB</i>	<i>E. coli</i> EDCM636 (pRB473- <i>mpsB</i>) carrying <i>mpsB</i>	This study
<i>E. coli</i> EDCM636 <i>mpsAB</i>	<i>E. coli</i> EDCM636 (pRB473- <i>mpsAB</i>) carrying <i>mpsA</i> and <i>mpsB</i>	This study
<i>E. coli</i> EDCM636 <i>mpsABC</i>	<i>E. coli</i> EDCM636 (pRB473- <i>mpsABC</i>) carrying <i>mpsA</i> , <i>mpsB</i> and <i>mpsC</i>	This study
<i>S. aureus</i> HG001	NCTC8325 derivative with repaired <i>rsbU</i>	²
<i>S. aureus</i> HG001 $\Delta mpsA$	$\Delta mpsA$ (deletion of SAOUHSC_00412)	This study
<i>S. aureus</i> HG001 $\Delta mpsB$	$\Delta mpsB$ (deletion of SAOUHSC_00413)	This study
<i>S. aureus</i> HG001 $\Delta mpsC$	$\Delta mpsC$ (deletion of SAOUHSC_00414)	This study
<i>S. aureus</i> HG001 $\Delta mpsABC$	$\Delta mpsABC$ (deletion of SAOUHSC_00412, SAOUHSC_00413, SAOUHSC_00414)	This study
<i>S. aureus</i> HG001 $\Delta mpsA$ (pRB473- <i>mpsA</i>)	<i>S. aureus</i> HG001 $\Delta mpsA$ complemented with <i>mpsA</i>	This study
<i>S. aureus</i> HG001 $\Delta mpsB$ (pctuf- <i>mpsB</i>)	<i>S. aureus</i> HG001 $\Delta mpsB$ complemented with <i>mpsB</i>	This study
<i>S. aureus</i> HG001 $\Delta mpsC$ (pRB473- <i>mpsC</i>)	<i>S. aureus</i> HG001 $\Delta mpsC$ complemented with <i>mpsC</i>	This study
<i>S. aureus</i> HG001 $\Delta mpsABC$ (pRB473- <i>mpsABC</i>)	<i>S. aureus</i> HG001 $\Delta mpsABC$ complemented with <i>mpsABC</i>	This study
<i>S. aureus</i> HG001 $\Delta mpsABC$ (pTX30- <i>can</i>)	<i>S. aureus</i> HG001 $\Delta mpsABC$ with insertion of carbonic anhydrase, <i>can</i> from <i>E. coli</i> MG1655)	This study
<i>S. aureus</i> USA300	Community-associated methicillin resistant clinical isolate, pulsed- field type USA300	³
<i>S. aureus</i> USA400(MW2)	Community-associated methicillin resistant clinical isolate, pulsed- field type USA400	⁴

Supplementary Table 2: Oligonucleotides used in this study

Primer name	Sequence (5'→3')
For the construction of <i>S. aureus</i> HG001 Δ <i>mpsB</i>	
mpsB_up_fwd	CCGGGCTAGCGCGCAGATCTGTA ACTATTTATATTGCG ACTG
mpsB_up_rev	CCA ACTTTTTAACTGTGTTGTCATACCTTTC
mpsB_down_fwd	ACAACACAGTTAAAAAGTTGGATTTAACGATGATATG
mpsB_down_rev	GCTTGATATCGTCGACAGATCTTATTTTTGTCTATATAG CGTTAATCATTAC
For the construction of <i>S. aureus</i> HG001 Δ <i>mpsC</i>	
mpsC_up_fwd	GGTACCCGGGCTAGCGCGCAGATCTCAAGCGGTTTTAA TGCTAAG
mpsC_up_rev	TATGCTTTTCTTTCGTTCTTTTCATATCACTAAC
mpsC_down_fwd	AAGAACGAAAGAAAAGCATATTTAAGAAGGTTAGG
mpsC_down_rev	CAAGCTTGATATCGTCGACAGATCTCCAACCGCTAAAT GCCTC
For the construction of <i>S. aureus</i> HG001 Δ <i>mpsB</i> complementation plasmid	
pc_mpsB_fwd	GTTTCGAGGAGGTTTAATTAATGACAACACAGTTAAATAT CAATTC
pc_mpsB_rev	TAAGTACTTCAGCTAATTAAGCTTAAATCCA ACTTTTTAAA ACGCC
For the construction of <i>S. aureus</i> HG001 Δ <i>mpsC</i> complementation plasmid	
pmpsA_com_fwd	ATCCCCGGGTACCGAGCTCGAATTCTTATTTTTAAAAGGA CGGGAAATAC
pmpsA_com_rev	ACTAACTCCATATTTCTAATCTCTCGCATAATTG
mpsC_com_fwd	GATTAGAAATATGGAGTTAGTGATATGAAAAGAAC
mpsC_com_Rev	CAAGCGCTCATCGCAGTGCGAATTCATTCATCCACGTA ATAAACAC
For the construction of plasmids pRB473- <i>mpsAB</i> and pRB473- <i>mpsB</i>	
PmpsABC Fwd	CCTGCAGGTCTGACTCTAGAGGATCCGGACGGGAAATA CTGCCTAAC
PmpsABC Rev	TTCACCTCCTTATTTCTAATCTCTCGCATAATTGCTTATG
SD+mpsB Fwd	AGAGATTAGAAATAAGGAGGTGAAAGGTATGAC
SD+mpsB Rev	TCGCAGTGCGAATTCGAGCTCTTAAATCCA ACTTTTTAAA ACGC
For the construction of plasmid pTX30- <i>can</i>	
BgIII_SD_can_fwd	AATGTAAGATCTAGGAGGTTTAATTAATGAAAGACATAG ATACACTCATCAG
can_rev_EcoRI	ATTATAGAATTCATATTATTTGTGGTTGGCGTGTTTCAG
For qRT-PCR	
mpsA qpcr Fwd	TTTGCAGTCATTGGCTTTTTTCGGT
mpsA qpcr Rev	AAGATGCAAACGACGTAATCGCAGT
gyrB Fwd	AAGGTATTATGGCGGCACGTG
gyrB Rev	ATCGCCTGCGTTCTAGAGTCAC

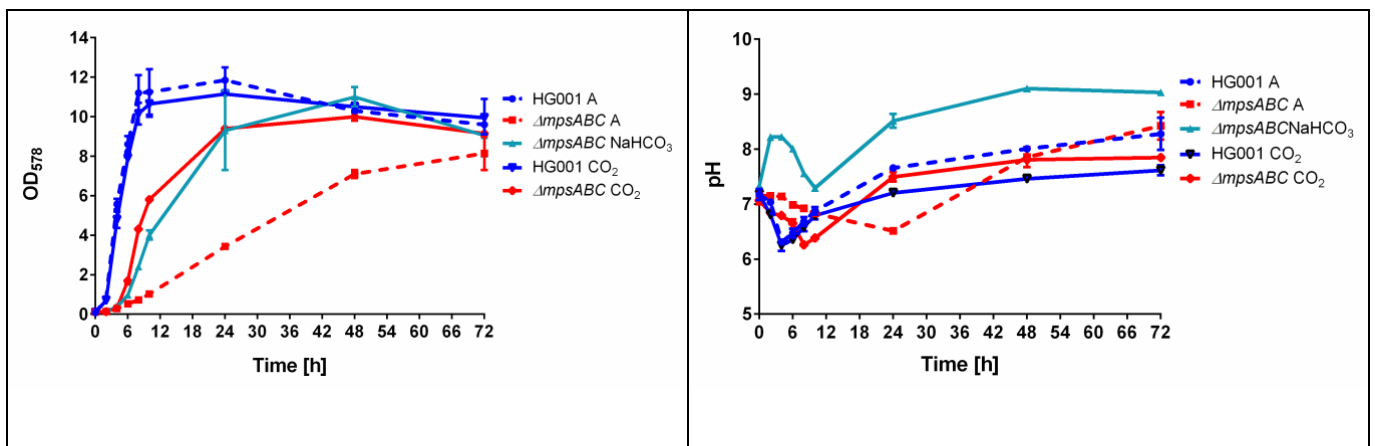
Supplementary Note 1

For Figure 2b in the manuscript, the pH levels were checked at the beginning of the experiment. The pH of the medium (TSB) increased with increased concentration of NaHCO_3 , as shown in the Supplementary Table 3.

Supplementary Table 3: pH of medium with addition of NaHCO_3

Supplementation	pH of medium (TSB) at the beginning
Aerobic	7.173
1 mM NaHCO_3	7.199
5 mM NaHCO_3	7.306
10 mM NaHCO_3	7.380
50 mM NaHCO_3	7.782
CO_2	7.156

In addition, we checked the effect of pH on the growth of ΔmpsABC (see figures below). For this reason, we used NaHCO_3 at 20 mM which gave an initial pH of 7.582. As can be seen from Supplementary Figure 8, the growth of the ΔmpsABC in CO_2 and NaHCO_3 were similar (left), although the pH for the medium with NaHCO_3 was higher (right), indicating that pH of the medium does not play a crucial role in promoting the growth.



Supplementary Figure 8: Growth (left) and extracellular pH (right) of wild type HG001 and ΔmpsABC in atmospheric conditions (A) and 5% CO_2 and 20 mM NaHCO_3 .

Each point shown in the graph represents the mean value \pm standard error mean (SEM) from two independent experiments.

Supplementary References

1. Merlin C, Masters M, McAteer S, Coulson A. Why is carbonic anhydrase essential to *Escherichia coli*? *J Bacteriol* **185**, 6415-6424 (2003).
2. Herbert S, *et al.* Repair of global regulators in *Staphylococcus aureus* 8325 and comparative analysis with other clinical isolates. *Infect Immun* **78**, 2877-2889 (2010).
3. Diep BA, *et al.* Complete genome sequence of USA300, an epidemic clone of community-acquired methicillin-resistant *Staphylococcus aureus*. *Lancet* **367**, 731-739 (2006).
4. Baba T, *et al.* Genome and virulence determinants of high virulence community-acquired MRSA. *Lancet* **359**, 1819-1827 (2002).