

Supplementary information

A universal approach to investigate circRNA protein coding function

Running title: Intron-mediated enhancement boosts circRNA translation

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Supplementary Fig. 1

A.

MEDIDQSSLV	SSSADSPPRP	PPAFKYQFVT	EPEDEEDEED	EEEEDEDEDL
60	70	80	90	100
EELEVLERKP	AAGLSAAPVP	PAAAPLLDFS	SDSVPPAPRG	PLPAAPPTAP
110	120	130	140	150
ERQPSWERSP	AASAPSLPPA	AAVLPSKLPE	DDEPPARPPA	PAGASPLAEP
160	170	180	190	200
AAPPSTPAAP	KRRGSGSVDE	TLFALPAASE	PVIPSSAEKI	MDLKEQPGNT
210	220	230	240	250
VSSGQEDFPS	VLFETAASLP	SLSPLSTVSF	KEHGYLEGNS	AVASTEETIE
260	270	280	290	300
ETLNEASREL	PERATNPFVN	RESAEFSVLE	YSEMGSFNG	SPKGESAMLV
310	320	330	340	350
ENTKEEVIVR	SKDKEDLVCS	AALHNPQESP	ATLTKVVKED	GVMSPKMTD
360	370	380	390	400
IFNEMKMSVV	APVREEYADF	KPFEQAWEVK	DTYEGSRDVL	AARANMESKV
410	420	430	440	450
DKKCFEDSLE	QKGGKDSSES	RNENASFPRT	PELVKDGSR	YITCDSFSSA
460	470	480	490	500
TESTAANIFP	VLEDHTSENK	TDEKKIEERK	AQIITEKTSP	KTSNPFLVAI
510	520	530	540	550
HDSEADYVTT	DNLSKVTEAV	VATMPEGLTP	DLVQEACESE	LNEATGTKIA
560	570	580	590	600
YETKVDLVQT	SEAIQESIYP	TAQLCPSFEE	AEATPSPVLP	DIVMEAPLNS
610	620	630	640	650
LLPSTGASVA	QPSASPLEVP	SPVSYDGIKL	EPENPPPYEE	AMSVALKTSD
660	670	680	690	700
SKEEIKEPES	FNAAAQEAEA	PYISIACDLI	KETKLSTEPS	PEFSNYSEIA
710	720	730	740	750
KFEKSVDPHC	ELVDDSSPES	EPVDLFSDDS	IPEVPQTQEE	AVMLMKESLT
760	770	780	790	800
EVSETVTQHK	HKERLSASPO	EVGKPYLESF	QPNLHITKDA	ASNEIPTLTK
810	820	830	840	850
KETISLQMEE	FNTAIYSNDD	LLSSKEDKMK	ESETFSDSSP	IEIIDEFPTF
860	870	880	890	900
VSAKDDSPKE	YTDLEVSNKS	EIANVQSGAN	SLPCSELPCD	LSFKNTYPKD
910	920	930	940	950
EAHVSDEFSK	SRSSSVKVPL	LLPNVSALES	QIEMGNIVKP	KVLTKEAEEK
960	970	980	990	1000
LPSDTEKEDR	SLTAVLSAEL	NKTSVVDLLY	WRDIKKTGVV	FGASLFLLLS
1010	1020	1030	1040	1050
LTVFSIVSVT	AYIALALLSV	TISFRIYKGV	IQAIQKSDEG	HPFRAYLESE
1060	1070	1080	1090	1100
VAISEELVQK	YSNSALGHVN	STIKELRRLF	LVDDLVDLSL	FAVLMWVFTY
1110	1120	1130	1140	1150
VGALFNGLTL	LILALISLFS	IPVIYERHQA	QIDHYLGLAN	KSVKDAMAKI
1160				
QAKIPGLKRR	AE			

A. Full protein sequence of mouse Rtn4.

The contributions of exon 2 and 3 are highlighted in yellow and green, respectively. The glutamic acid codon (represented by E at position 188 of the protein sequence) is contributed by both exons, while the aspartic acid residue D at position 169 (highlighted in blue) resulted from circularization and remains D in the product of circular translation.

B.

MDLKEQPGNT	VSSGOEDFPS	VLFFETAASLP	SLSPLSTVSF	KEHGylGNLS
10	20	30	40	50
AVASTEgTIE	ETLNEASREL	PERATNPfVN	RESAEfSVLE	YSEMgSSFNG
60	270	80	90	100
SPKGESAMLV	ENTKEEVIVR	SKDKEDLVCS	AALHNPQESP	ATLTKVVKED
110	120	130	140	150
GVMSPEKTMD	IFNEMKMSVV	APVREEYADF	KPFEQAWEVK	DTYEGSRDVL
160	170	180	190	200
AARANMESKV	DKKCFEDSLE	QKGHGKDSES	RNENASFPRT	PELVKDGsRA
210	220	230	240	250
YITCDSfSSA	TESTAANIFP	VLEDHTSENK	TDEKKIEERK	AQIITEKTSP
260	270	280	290	300
KTSNPFVLVAI	HDSEADYVTT	DNLSKVTEAV	VATMPEGLTP	DLVQEACESE
310	320	330	340	350
LNEATGTkIA	YETKVDLVQT	SEAIQESIYP	TAQLCPSFEE	AEATPSPVLP
360	370	380	390	400
DIVMEAPLNS	LLPSTGASVA	QPSASPLEVP	SPVSyDGIKL	EPENPPPYYE
410	420	430	440	450
AMSVAlKtSD	SKEEIKEPES	FNAAAQEAEA	PYISIAcdLI	KETKLSTEPS
460	470	480	490	500
PEFSNYSEIA	KFEKSVPDHC	ELVDDSSPES	EPVDLFSDDS	IPEVPQTQEE
510	520	530	540	550
AVMLMkESLT	EVSETVTQHK	HKERLSASPO	EVGKPYLESF	QPNLHITKDA
560	570	580	590	600
ASNEIPTLTK	KETISLQMEE	FNTAIYSNDD	LLSSKEDKMK	ESETFSdSSP
610	620	630	640	650
IEIIDEfPTE	VSAKDDSPKE	YTDLEVSNKS	BIANVQSGAN	SLPCSELPCD
660	670	680	690	700
LSFKNTYPKD	EAHVSDefSK	SRSSVSKVPL	LLPNVSALES	QIEMGNIVKP
710	720	730	740	750
KVLTKEAEeK	LPSDTEKEDR	SLTAVLSAEL	NKTSARAETH	LRRGSGSK
760	770	780	790	798

B. Protein sequence of linear counterpart mRNA expressed by pCMV-Rtn4-Exon2-Exon3.

The contributions of exon 3 are highlighted in green. The amino acid sequence from the vector is in gray.

C.

13 ↓

AUGAGACCCUUU UUGCUCUuccUG CUGCAUCUGAGC CUGUGAUACCCU CCUCUGCAGAAA 60
 E T L F A L P A A S E P V I P S S A E K

→

AAAUUAUGGAUU UGAAGGAGCAGC CAGGUAAACACUG UUUCGUCUGGUC AAGAGGAUUUCC 120
 I M D L K E Q P G N T V S S G Q E D F P..

CAUCUGUCCUGU UGA AACUGCUG CCUCUCUuccUU CUCUAUCUCCUC UCUCAACUGUUU 180

CUUUUAAAGAAC ACGGAUACCUUG GUAA CUUAUCAG CAGUGGCAUCCA CAGAAGGAACUA 240

15

UUGAAGAAACUU UAAAUGAAGCUU CUAGAGAAUUGC CAGAGAGGGCAA CAAAUCCAuuUG 300

→

UAAAUAGAGAGU CAGCAGAGUUUU CAGUAUUAAGAAU ACUCAGAA AUG GAUCAUCUUUCA
 10

→

AUGGCUCCCAA AAGGAGAGUCAG CC AUGUAGUAG AAAACACUAAGG AAGAAGUAAUUG

UGAGGAGUAAAG ACAAAGAGGAUU UAGUUUGUAGUG CAGCCCUUCAUA AUCCACAAGAGU

CACCUGCGACCC UUACUAAAGUGG UJAAAGAAGACG GAGUU[→]AUGUCUC CAGAAAAGACAA[→]
 UG²GACAUUUUUA AUGAAUGAAA UGUCAGUGGJAG CACCUGUGAGGG AAGAGUAUGCAG 600
 AUUUUAAGCCAU UUGACAAGCAU⁶GGGAAGUGAAAG AUACUU⁵¹AUGAGG GAAGUAGGGAUG⁴⁶
 UGCUGGCUGCUA GAGCUAAU[→]AUGG AAAGUAAAGUGG ACAAAAAAUGCU UUGAAGAUGCC²
 UGGAGCAAAAAa GUC²⁰AUGGGAAGG AUJAGUGAAAGCA GAA¹²AUGAGAUG¹⁰ CUUCUUUCCCA
 GUACCCAGAAC UUGUGAAGGACG GCUCCAGAGCGU ACAUCACCUGUG AUUCCUUUA^cCU
 CAGCAACCGAGA GUACUGCAGCAA ACAUUUCCUG UGC⁴UAGAAGAUC ACACUUCAGAAA 900
 AUAAAACAGA⁴AUG AAAAAAAAAA[→]JAG AAGAAAGGAAG CCCAAAUUA¹⁴UAA CAGAGAAGACUA⁹
 GCCCAAACGU CAAUCCUUUCC UUGUAGCAAUAC AUGAUUCCGAGG CAGAUUAUGUCA
 CAACAGA¹⁴UAAUU UAUCAAGGUGA CUGAGGCAGUAG UGGCAACC[→]AUGC CUGAAGGUCUAA
 CGCCAGAUU⁴⁶UAG UUCAGGAAGCAU¹⁴GUGAAAGUGAAC UGAACGAAGCCA CAGGUACAAAGA
 UUGCUUAUGAAA CAAAAGUGGACU UGGUCCAGACAU CAGAAGCUAUAC AAGAGUCAUUU 1200
 ACCCCACAGCAC AGCUUUGCCCAU CAUU[→]UGAGGAAG CUGAAGCAACUC CGUCACCAGUUU
 UGCCUGAUAUUG UUAUGGAAGCUC CAUUAAAUUCUC UCCUCCAAGCA CUGGUGCUUCUG
 UAGCGCAGCCCA GUGCAUCCCCAC UAGAAGUACCGU CUCCAGU³UAGUU AUGACGGUAUAA
 AGCUUGAGGCCUG AAAAUCCCCAC CAUAUGAAGAAG CCAUGAGUGUAG CACUAAAACAU[→]
 CGGACgCAAAGG AAGAAU¹⁷UAAAG AGCCUGAAAGUU UJAAUGCAGCUG UUCAGGAAGCAG 1500
 AAGCUCCUUAUA UAUCCAUUGC¹AUGUGAUUJAAUUA AAGAAACAAAGC UCUCCACUGAGC
 CAAGUCCAGAGU UCUC²⁸UAAUUAUU CAGAAA¹⁴UAGCAA AAUUUGAGAAGU CGGUGCCUGAUC
 ACUGUGAGCUCG UGGAUGAUUCCU CACCgGAAUCUG AACCAGUUGACU UAUU¹⁴UAGUGAUG
 AUUCAAUUCCUG AcGUCCACAAA CACAAGAGGAGG CUG[→]UGAUGCUAA[→]UGAAGGAGAGUC
 UCACUGAAGUGU CUGAGACAGUAA CACAACACAAAC AUAAAGGAGAGAC UUAGUGCUUCAC 1800
 CUCAGGAGGUG GAAAGCCAUAAU UAGAGUCUUUUC AGCCCAUUUAC AUUUUACAAAAG
 AUGCUGCAUCUA AUGAAAUCCAA CAUUGA⁸CCAAA AGGAGACAAUUU CUUUGCAA[→]AUGG
 AAGAGUU¹¹UAAUA CUGCAAUUUAUU CCAUGAUGACU UACUUUCUUCUA AGGAAGACAAA[→]
 19

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UGA AAGAAAGUG AAACAUUUUCCG AUUCAUCUCCCA UUGAGA UAAUAG AUGAGUUUCCCA
          9
CAUUUGUCAGUG C UAAAG AUGAUU CUCC U AAGGAGU ACAC UGACC UAG AAGUAUCCAACA 2100
          58
AAAGUGA AAUUG C U A AUGUCCAGA GCGGGGCCAAUU CGUUGCCUUGCU CAGAAUUGCCCU
          32          29          26
GUGACC UUUCUU UCAAGAAUACAU AUCC U A A GAUG AAGCACA AUGUCU CAG AUGA AUUCU
          8
CCAAAAG UAGGU CCAGUGUAUC U A A GGUGCCCUU AU UGCUUCCAA AUG UUUCUGCUUUGG
          →
AAUCUCAAA UAG AA AUGGGCAACA UAG U A A ACCCA AAGUACUUACGA AAGAAGCAGAGG

AAAAACUUCUU CUGA UACAGAGA AAGAGGACAGAU CCC UGA CAGCUG UAUUGUCAGCAG 2400
  K L P S D T E K E D R S L T A V L S A E
AGC UGA A U A A A CUUCAG 2418
  L N K T S [D]

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C. Nucleotide sequence of circRtn4.

The nucleotides contributed by exon 2 are shown in blue and those from exon 3 in black lettering; the junction is depicted by a vertical arrow. Due to the circular nature of the RNA, the nucleotide at position 2418 is fused to nucleotide 1. If this circular RNA is translated, it generates a GAU codon (highlighted in yellow) coding for aspartic acid, denoted by a [D]. All possible AUG start codons are shown in green and stop codons in red. Where start and stop codons overlap, only the non-overlapping parts of stop codons are red. The numbers above AUG codons indicates the amino acid lengths of hypothetical reading frames, while the 16 AUG codons marked by horizontal arrows are hypothetically proceeding uninterrupted by any stop codon, thus circling the RNA template. Circumstantial evidence points to a start at the second AUG marked by a green arrow. Part of the amino acid sequence of the open reading frame is shown in the one-letter code beneath the nucleic acid sequence. Nucleotides are arranged in blocks of 12.

D.

Frame 3

```

ETL FALPAASEPVIPSSAEKI MDLKEQPGNTVSSGQEDFPSVLFETAASLPSLSPLSTV SFKEHG YLGN
LSAVASTE GTIEETLNEASRELPERATNPFVNRESAEFSVLEYSEM GSSFN GSPKGESAM LVENTKEEV
IVRSKDKEDLVCSAALHNPOES PATLTKVVKEDGVM SPEKTM DIFNEMKMSV VAPVREEYADFKPFEQA
WEVKD TYEGSRDVL AARANMESKV DKKCFEDSLEQKSHGKDSESRNENASFPSTPELVKDGSRAYITCD
SFTSATESTAA NIFPVLEDHTSENKTDEKKIEERKAQII TEKTS PKTSNPFLVAIHDSEADYVTTDNLS
KVTEAVVATM PEGLTPDLVQEACESELNEATGTKIAYETKVDLVQ TSEAIQESIYPTAQLCPSFEEAEA
TPSPVLPDIVMEAPLNSLLPSTGASVAQPSASPLEVPSVSYDGIKLEPENPPP YEEAMSVALKTS DAK
EEIKEPESFNAAVQEA EAPYIS IACDLIKETKLSTEPSPEFSNYSEIAKFEKSVPDHCELVDDSSPESE
PVDLFSDD SIPDVPQTQEEAVMLM KESL TEVSETVTQHKKERLSASPQEVGKPYLESFQPNLHITKDA
ASNEIPTLTKKETISLQMEEFNTAIYSNDDLSSKEDKM KESETFSDSSPIEIID EFPFTFVSAKDDSPK
EYTDLEVSNKSEIANVQSGANSLPCSELPCDLSFKNTYPKDEAHVSDEF SKSRSSVSKVPLLLPNVSAL
ESQIEMGNIVKPKVLTKEAEKLP SDTEKEDRSLTAVLSAELNKTS

```

Frame 2

```

-DPFCSSCCI-ACDTLLCRKNYGFEGAAR-HCFVWSRGFPICPV-NCCLSSFSISSLNCFF-RTRIPW-
LISSGIHRRNY-RNFK-SF-RIAREGNKSICK-RVSRVFSIRILRNGIIFQWLPKRRVSHVSRKH-

```

GRSNCEE-RQRGFSL-CSPS-STRVTCDPY-SG-RRRSYVSRKDNHGF--NENVSGSTCEGRVCRF-
AI-TSMGSELYL-GK-GCAGC-S-YGK-SGQKML-R-PGAKKSWEG--KQK-
ECFFPQYPRTEGRLQSVHHL-FLYLSNREYCSKHFPKARRSHFRK-NR-KKNRRKEGPNYNRED-
PQNVKSFPCSNL-FRGRLLCHNR-FIKGD-GSSGNHA-RSNARFSSGSM-K-TERSRYKDCL-
NKSGLGPDIRSITRVNLPSTALPII-GS-SNSVTSFA-
YCYGSSIKFSPSKHWCFCSAQAQCIPTTRSTVSS-L-RYKA-A-KSPTI-RSHECSTKNIGRKGGRN-RA-
KF-CSCSGSRSSLYIHC-M-FN-RNKALH-AKSRVL-LFRNSKI-EVGA-SL-ARG-FLTGI-TS-LI--
-FNS-RPTNTRGGCDANEGESH-SV-DSNTTQT-GET-CFTSGGRKAI FRVFSQAQFTYYKRCCI--
NSNIDQKGDNFFANGRV-YCNLFQ--LTFE-GRQNERK-NIFRFISH-DNR-VSHICQC-R-FS-GVH-
PRSIQOK-NC-CPERGQFVALLRIAL-PFFQEQEYIS-R-STCLR-ILQK-VQCI-
GALIASKCFCFGISNRNGQHS-TQSTYERSRGKTSF-YRERGOIPDSCIVSRAE-NF

Frame 1

MRPFLLELLHLSL-YPPLQKKLWI-
RSSQVTLFRLVKRISHLSCLKLLPLFLLYLLSOLFLLKNTDTLVTYQQWHPQKELLKKL-
MKLLENCQRGQQIHL-IESQQSFQY-NTQKWDHLSMAPQKESQPC--KTLRKK-L-GVKTKRI-
FVVQPFIIHKSHLRPLLKWLKKTTELCLQKRQWTFMK-KCQW-HL-GKSMQILSHLNKHGK-
KILMREVGMCWLELEIWKVKWTKNALKIAWSKKVMGRIVKAEMRMLLSPVPQNL-
RTAPERTSPVIPLPQQPRVLQQTFSLC-KITLQKIKQMKKK-KKGRPKL-QRRLAPKRQILSL-
QYMI PRQIM SQQIIYQR-LRQ-WQPCLKV-RQI-FRKHVKVN-
TKPQVQRLLMKQKWTWSRHQKLYKSQFTPOHSFAHHLRKLKQLRHQFCLILLWKLH-ILSFQALVLL-
RSPVHPH-KYRLQLVMTV-SLSLKIPHHMKKP-V-H-
KHRTQRKKLKS LKVLMLQFRKQKLLIYPLHVI-LKKQSSPLSQVQSSLI IQK-
QNLRSRCLITVSSWMI PHRNLNQLTYLVMIQFLTSHKHKRRL-C--RRVSLKCLRQ-
HNTNIRDLVHLRR-ESHI-SLFSPIYILQKMLHLMKFQH-
PKRRQFLCKWKS LILQFIPMMTYFLLRKT-KKVKHFPIHLPLR--MSFPHLSVLKMI LLRSTLT-
KYPTKVKLLMSRAGPIRCLAQNCPVTFLSRIHILKMKHMSQMNSPKVGVPYLRCPCYCFQMFLLWNLK-
KWAT-LNPKYL RKKQRKNFLLIQRKRTDP-QLYCOQS- IKLQ

D. circRtn4 computationally translated in all three reading frames.

Only frame 3 is open. Methionine residues are highlighted in green and longer potential ORFs in other reading frames of hypothetical polypeptides are highlighted in blue.

A. circRtn4-FLAG-Stop

```
AUGAGACCCUUU UUGCUCUCCUG CUGCAUCUGAGC CUGUGAUACCCg actacaaggacg 60
..E T L F A L P A A S E P V I P D Y K D D

acgatgacaagc caccgtgaUCCU CUGCAGAAAAAA UUAUGGAUUUGA AGGAGCAGCCAG 120
D D K P P - M D L K E Q P G...
```

B. circRtn4-FLAG-ac

```
AUGAGACCCUUU UUGCUCUCCUG CUGCAUCUGAGC CUGUGAUACCCg actacaaggacg 60
E T L F A L P A A S E P V I P D Y K D D

acgatgacaagc caccgacUCCUC UGCAGAAAAAAU UAUGGAUUUGcA GGAGCAGCCAGG 120
D D K P P T P L Q K K L W I C R S S Q V
M D L Q E Q P G

UAACACUGUUUC GUCUGGUCAAGA GGAUUUCCCAUC UGUCCUGUUUGA AACUGCUGCCUC 180
T L F R L V K R I S H L S C L K L L P L
N T V S S G Q E D F P S V L F E T A A S
UCUCCUUCUCU AUCUCCUCUCUC AACUGUUUCUUU UAAAGAACACGG AUACCUUGGUAA 240
F L L Y L L S Q L F L L K N T D T L V T
L P S L S P L S T V S F K E H G Y L G N
CUUAUCAGCAGU GGCAUCCACAGA AGGAACUAUUGA AGAAACUUUAAA UGAAGCUUCUAG 300
Y Q Q W H P Q K E L L K K L -
L S A V A S T E G T I E E T L N E A S ... etc.
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Supplementary Fig. 2

Additional constructs circRtn4-Stop (A) and circRtn4_FLAG-ac (B). Blue lettering indicates nucleotide sequence contributions of *Rtn4* exon 2 and black upper-case lettering contributions of *Rtn4* exon 3. Nucleotides are arranged in blocks of 12. The nucleotides for the FLAG sequence are in black lower-case letters. The corresponding amino acids are depicted in bold and purple (IUPAC one-letter amino acid code). The putative start codon in green. A) A translation stop codon was introduced after the sequence encoding the FLAG peptide, presumably after almost a full circle of translation (indicated in red). The predicted Rtn4 polypeptide variant (IUPAC one-letter amino acid code) is ~800 amino acids in length. B) The insertion of two nucleotides (ac) at positions 78/79 and a single nucleotide exchange at position 107 (c) are shown in lower case letters highlighted in blue. The correspondingly changed amino acid Q at position 4 (highlighted in blue) of the assumed Rtn4-derived amino acid sequence is also highlighted in blue. The latter was changed in order to remove a stop codon early in the second round of translation. The predicted polypeptide sequence is given in the IUPAC one-letter amino acid code. Translation product for the first round on the circle is shown in the second aa sequence row. Translation could theoretically proceed for about another ~69 amino acid into a next round of translation, provided that AUG start codon 98-100 is being used for translation initiation. The amino acid sequence of the second round of translation would leave the reading frame after the FLAG peptide at the glutamine

residue and proceed (top amino acid sequence immediately under the nucleotide sequence) until the stop codon at position 285-287 (red) is encountered. The predicted protein would be ~869 amino acids in length.

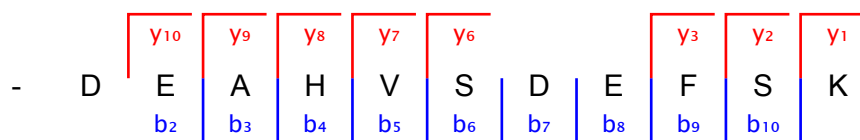
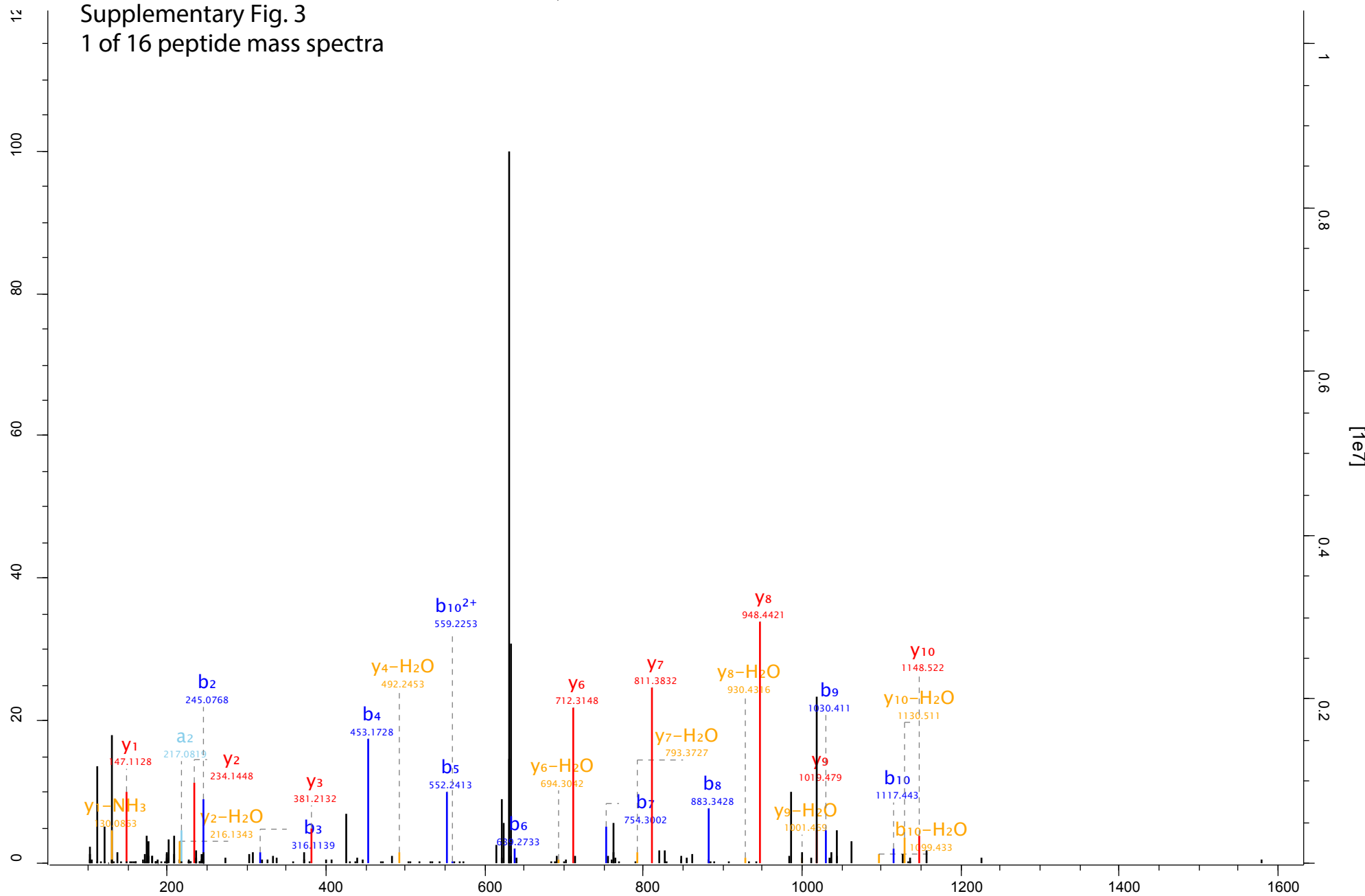
Supplementary Fig. 3

16 mass spectra of the tryptic peptides from circRtn4 derived protein

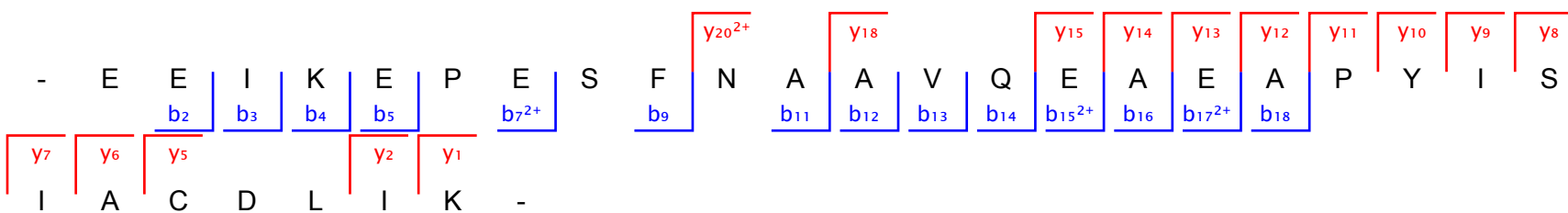
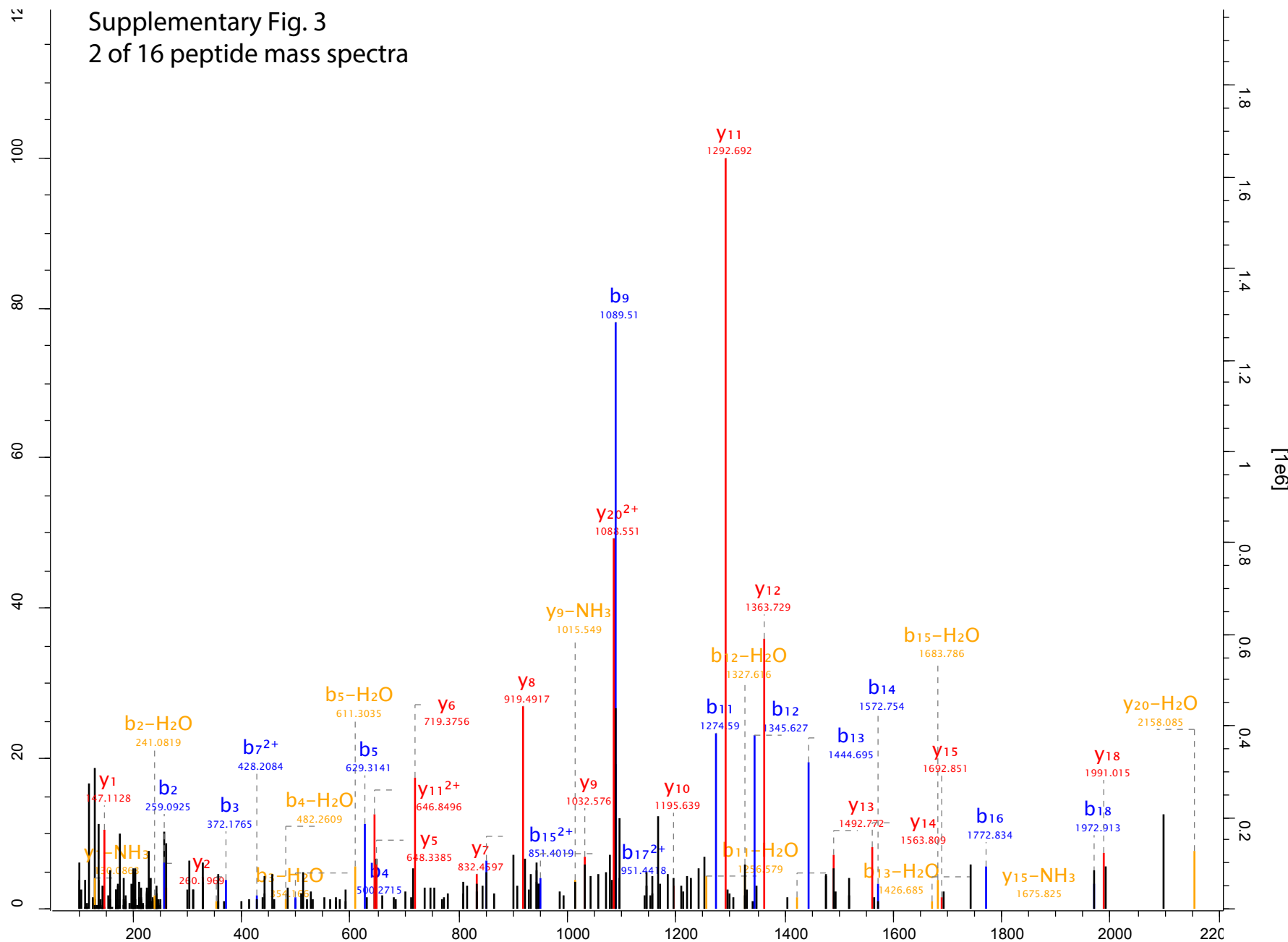
The left coordinate is the relative intensity, the right coordinate is the absolute intensity, the horizontal coordinate is m/z.

Scan 16346 Method FTMS; HCD Score 144.1 m/z 632.28

Supplementary Fig. 3
1 of 16 peptide mass spectra

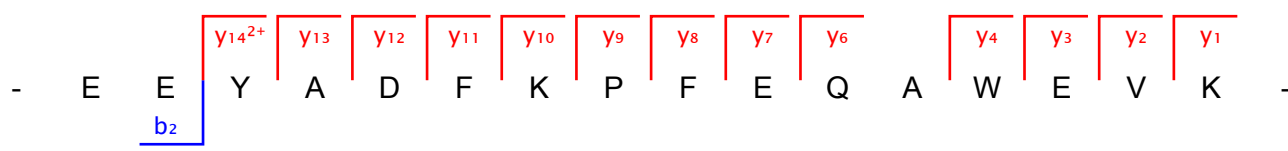
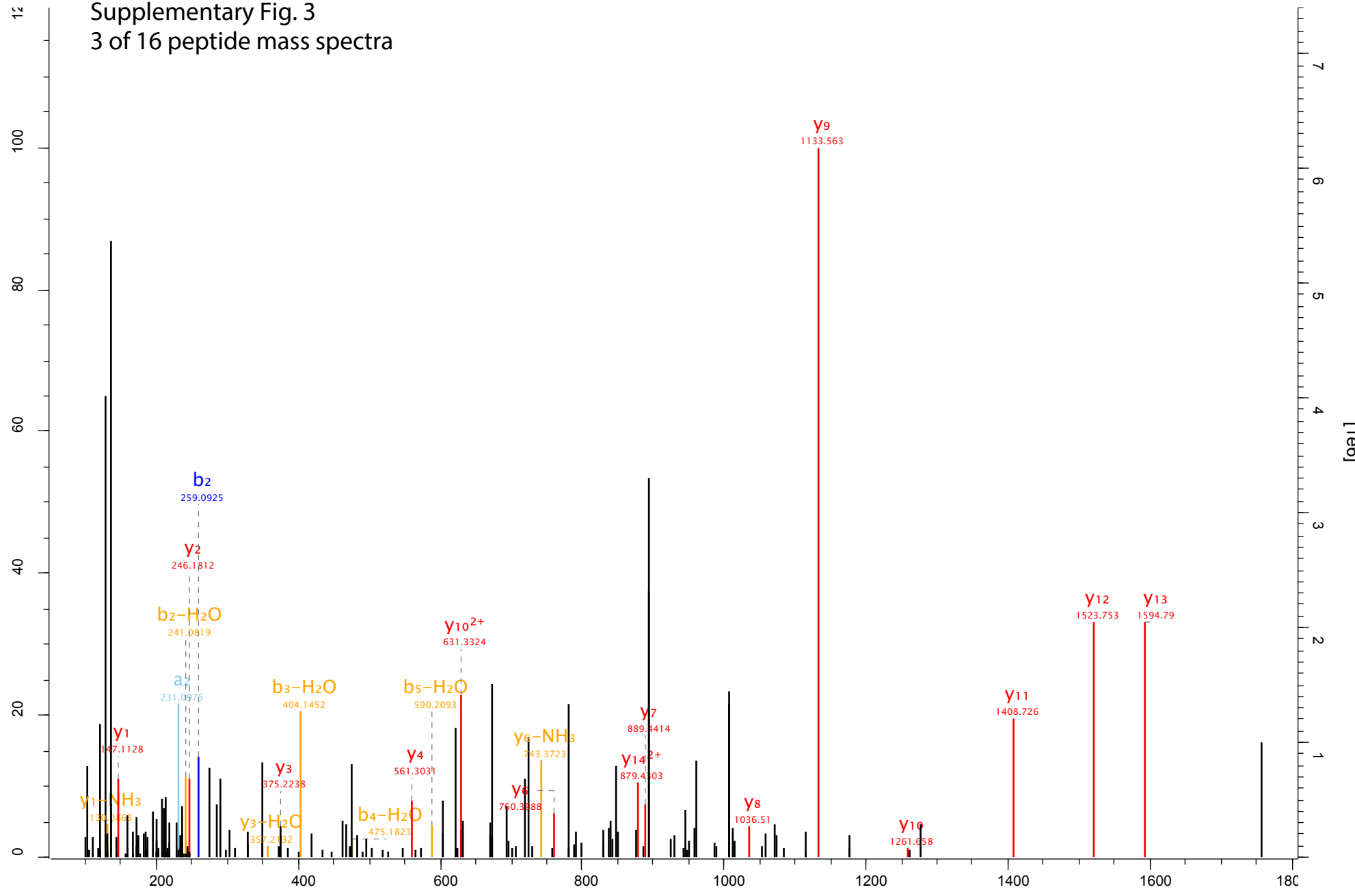


Supplementary Fig. 3
2 of 16 peptide mass spectra



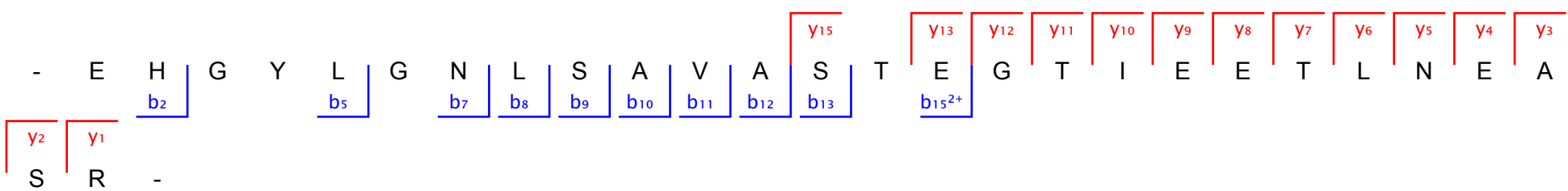
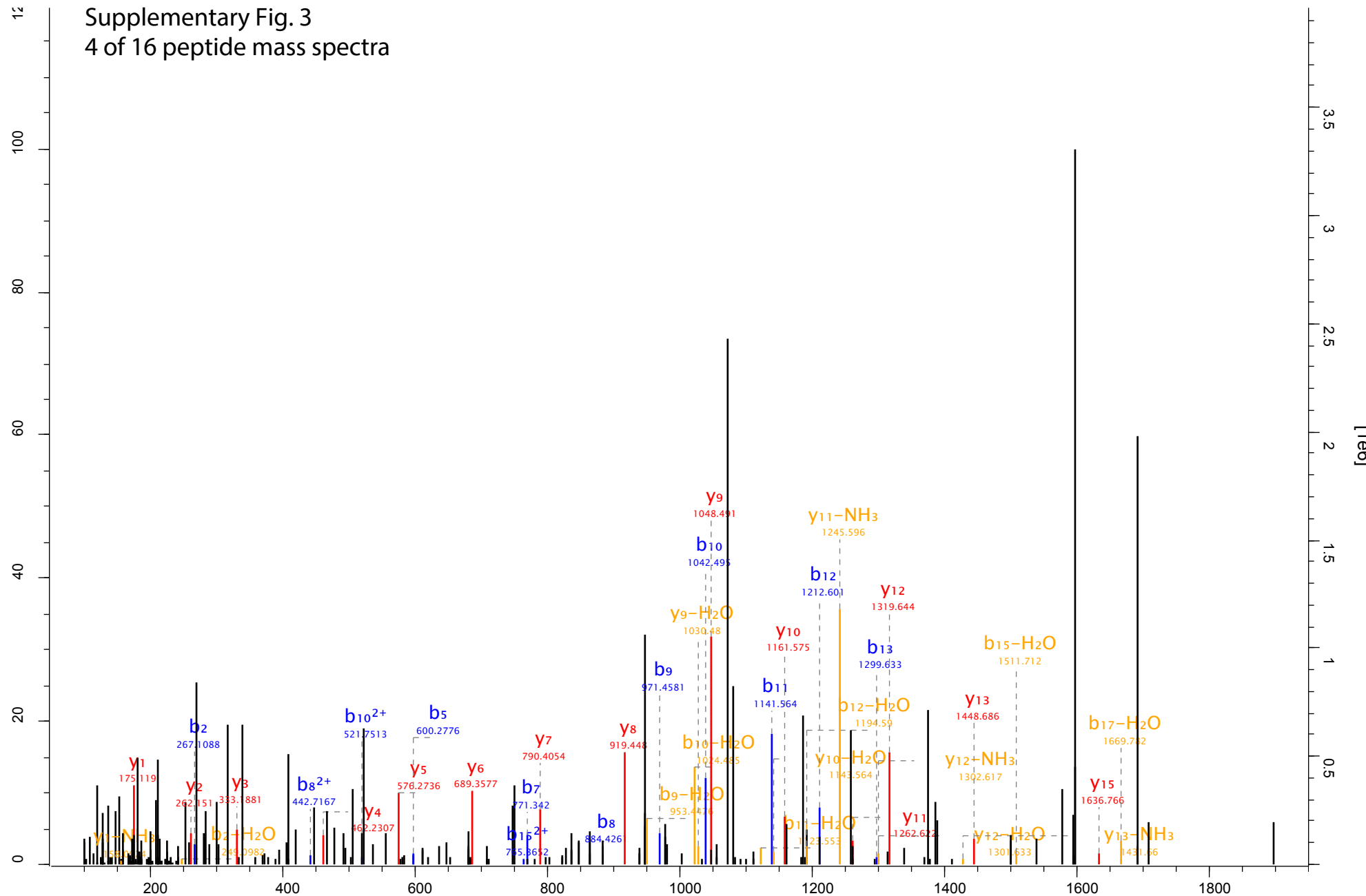
Scan 88482 Method FTMS; HCD Score 62.81 m/z 672.99

Supplementary Fig. 3
3 of 16 peptide mass spectra



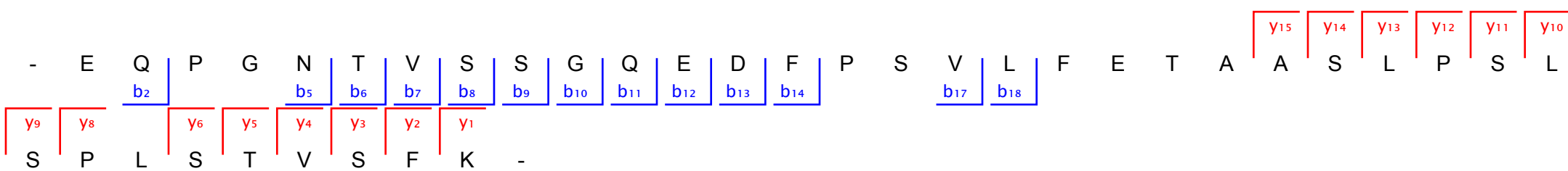
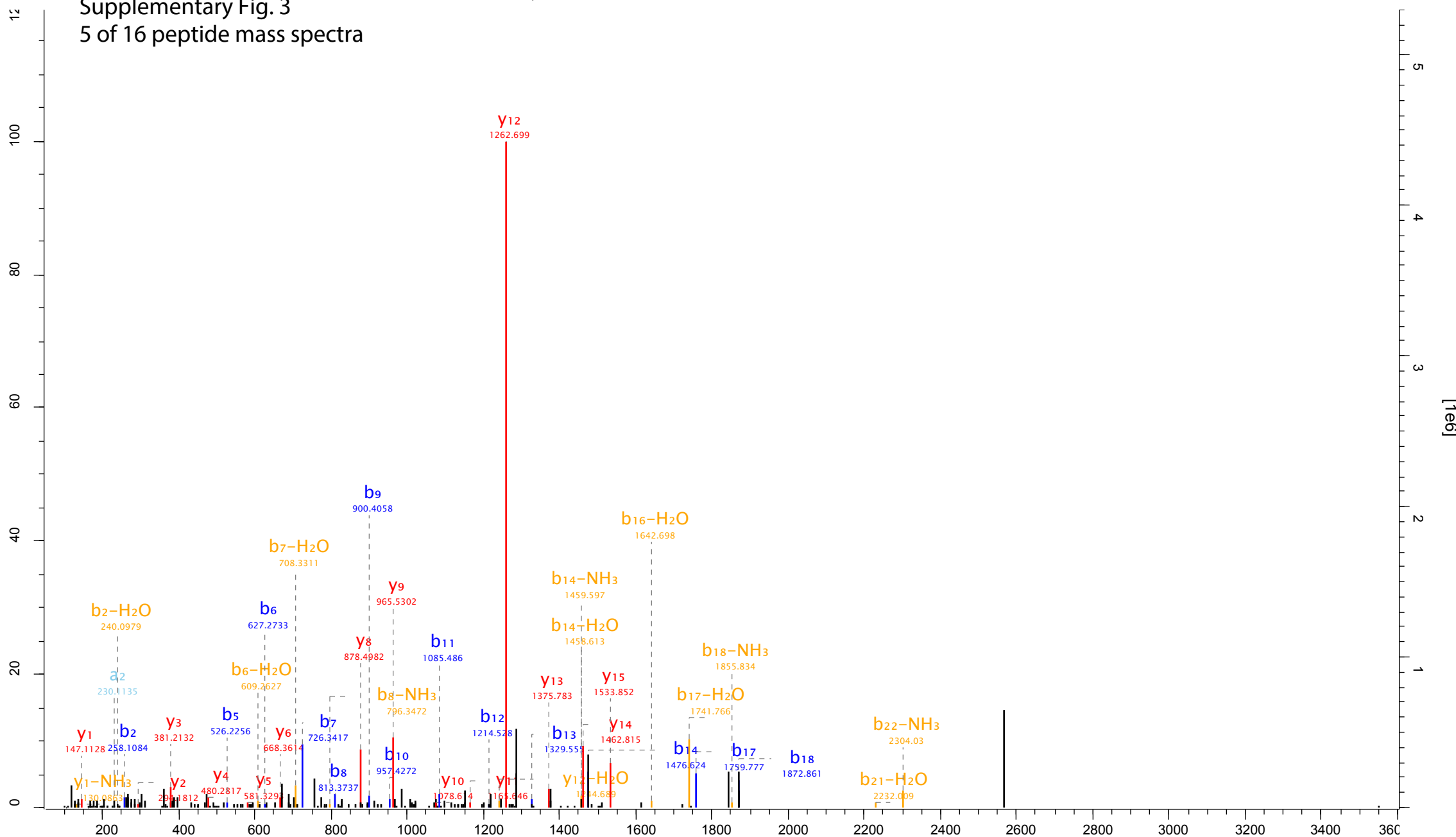
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Supplementary Fig. 3
4 of 16 peptide mass spectra



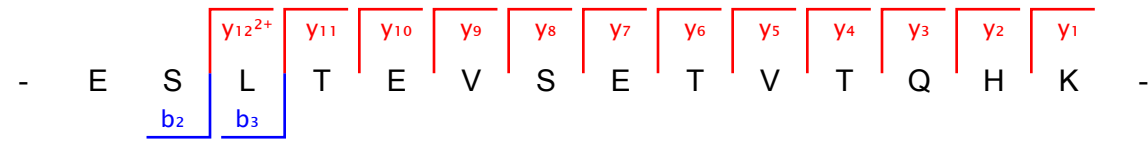
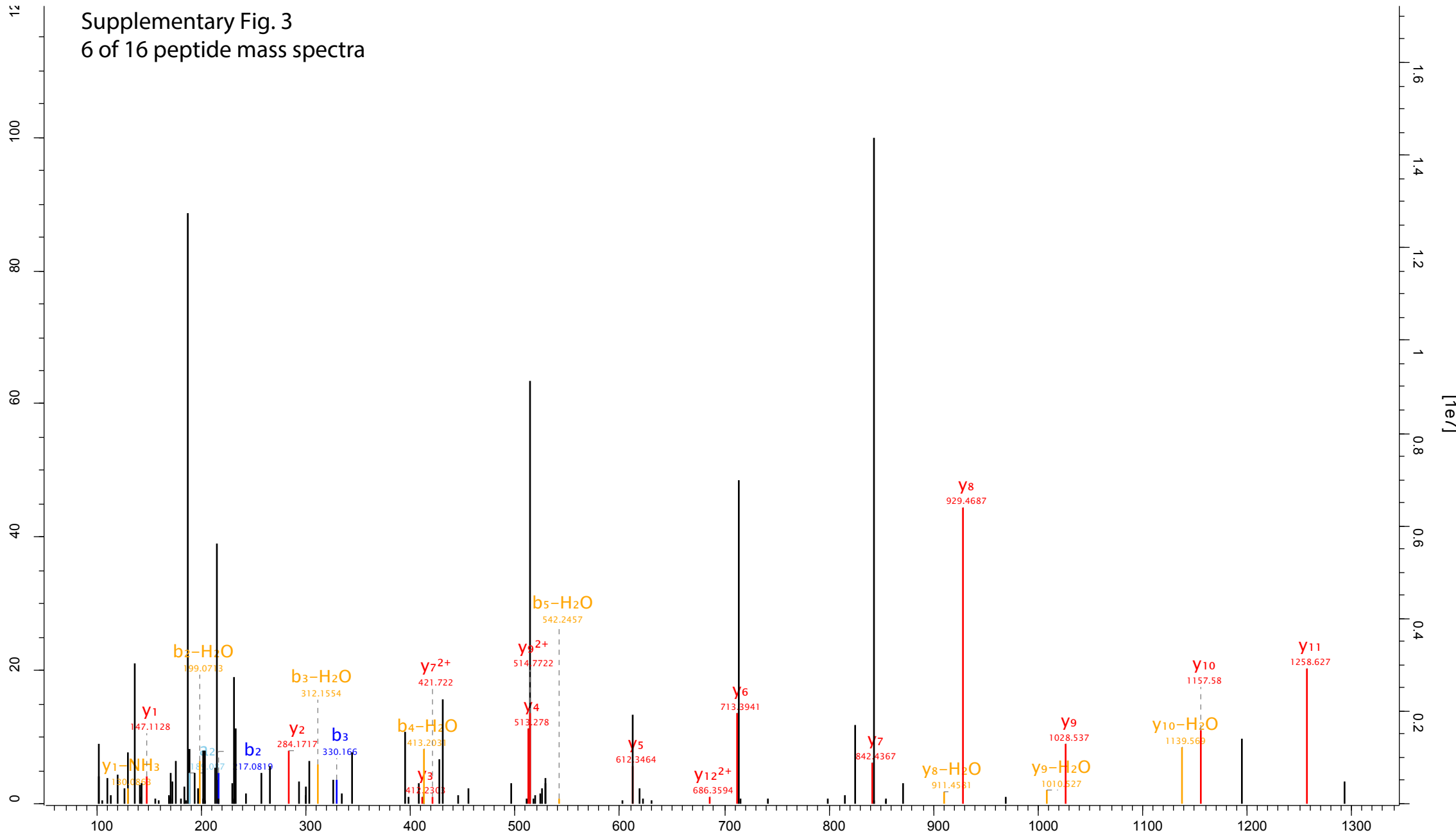
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Supplementary Fig. 3
5 of 16 peptide mass spectra



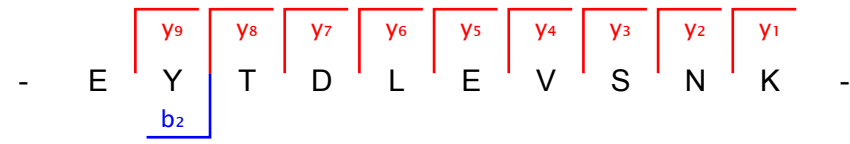
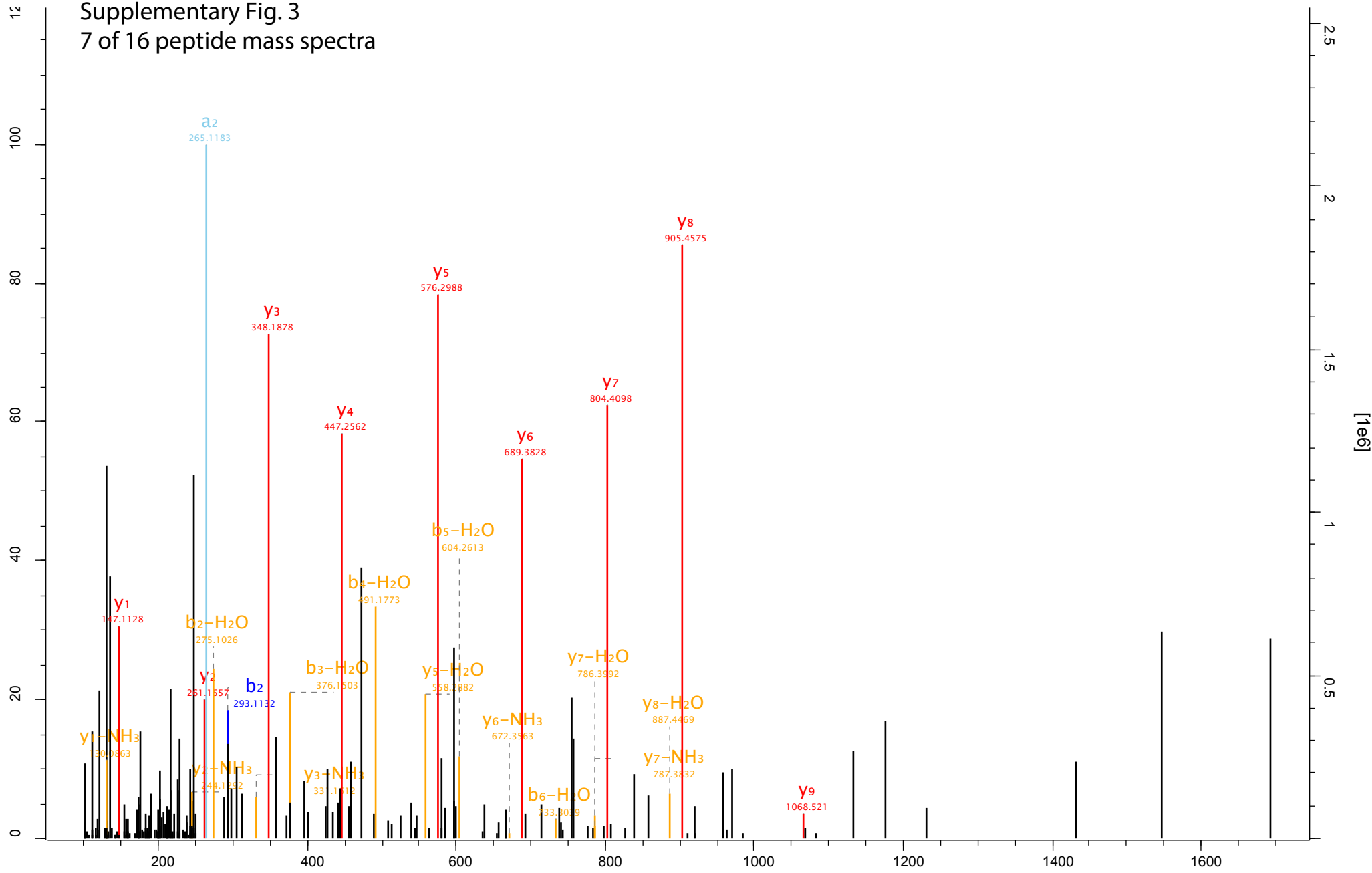
Scan 40472 Method FTMS; HCD Score 69.26 m/z 529.93

Supplementary Fig. 3
6 of 16 peptide mass spectra



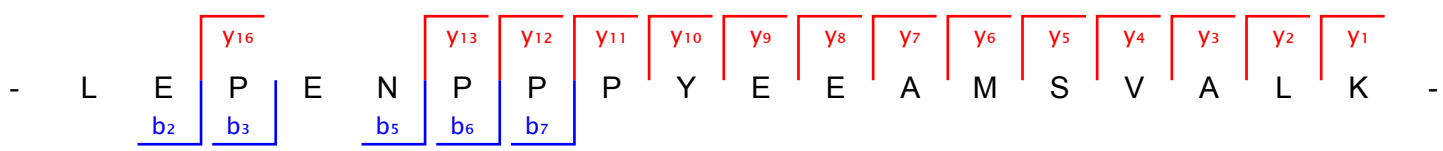
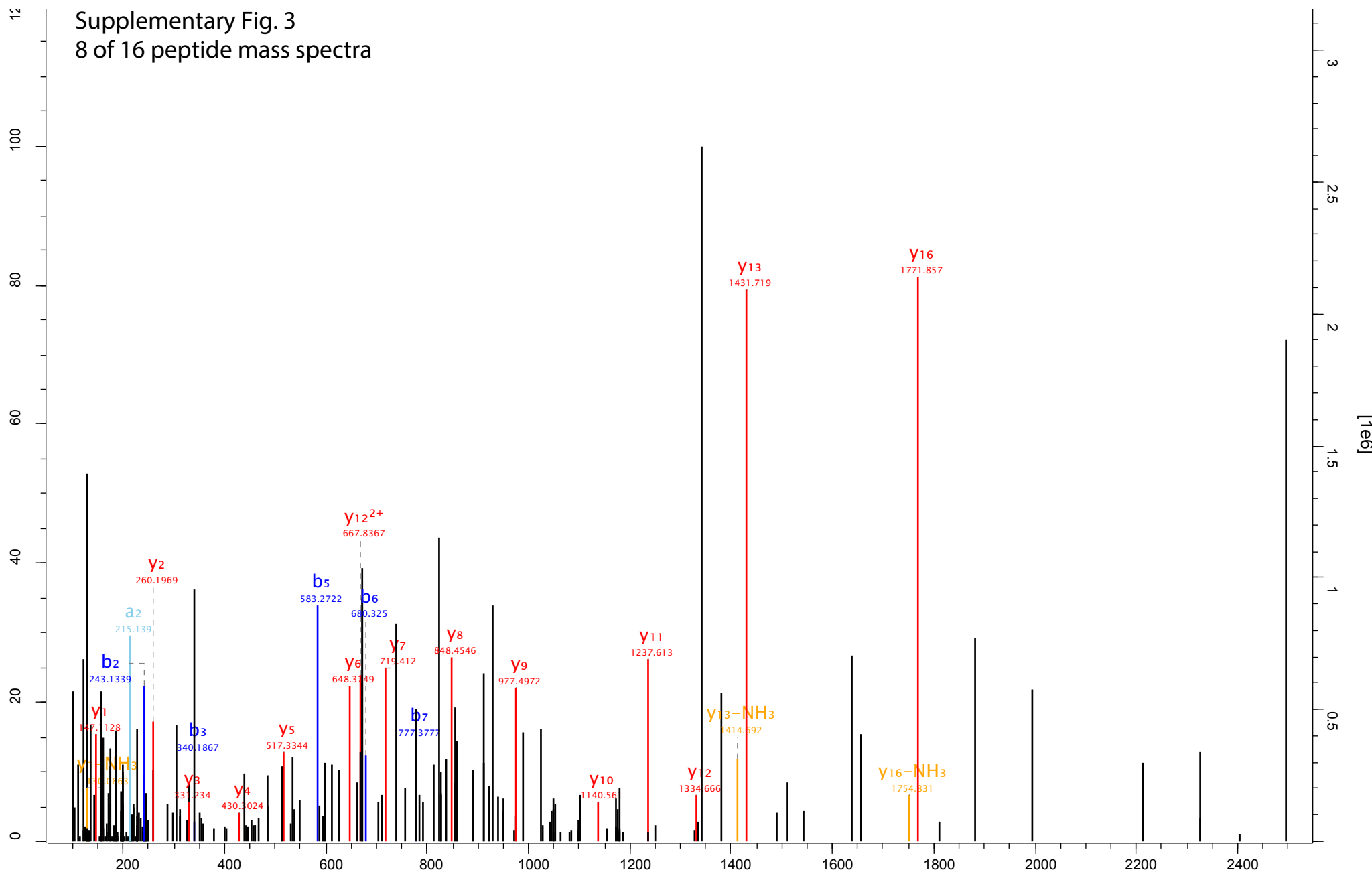
Scan 26668 Method FTMS; HCD Score 88.06 m/z 599.29

Supplementary Fig. 3
7 of 16 peptide mass spectra



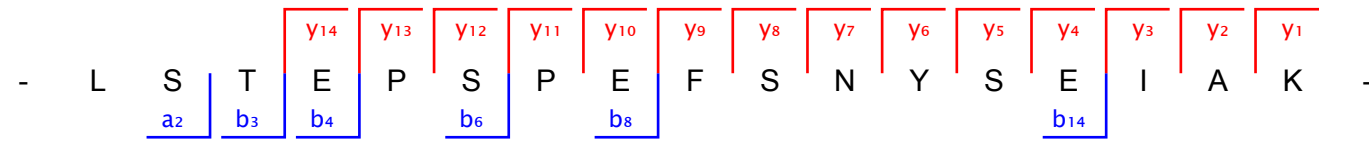
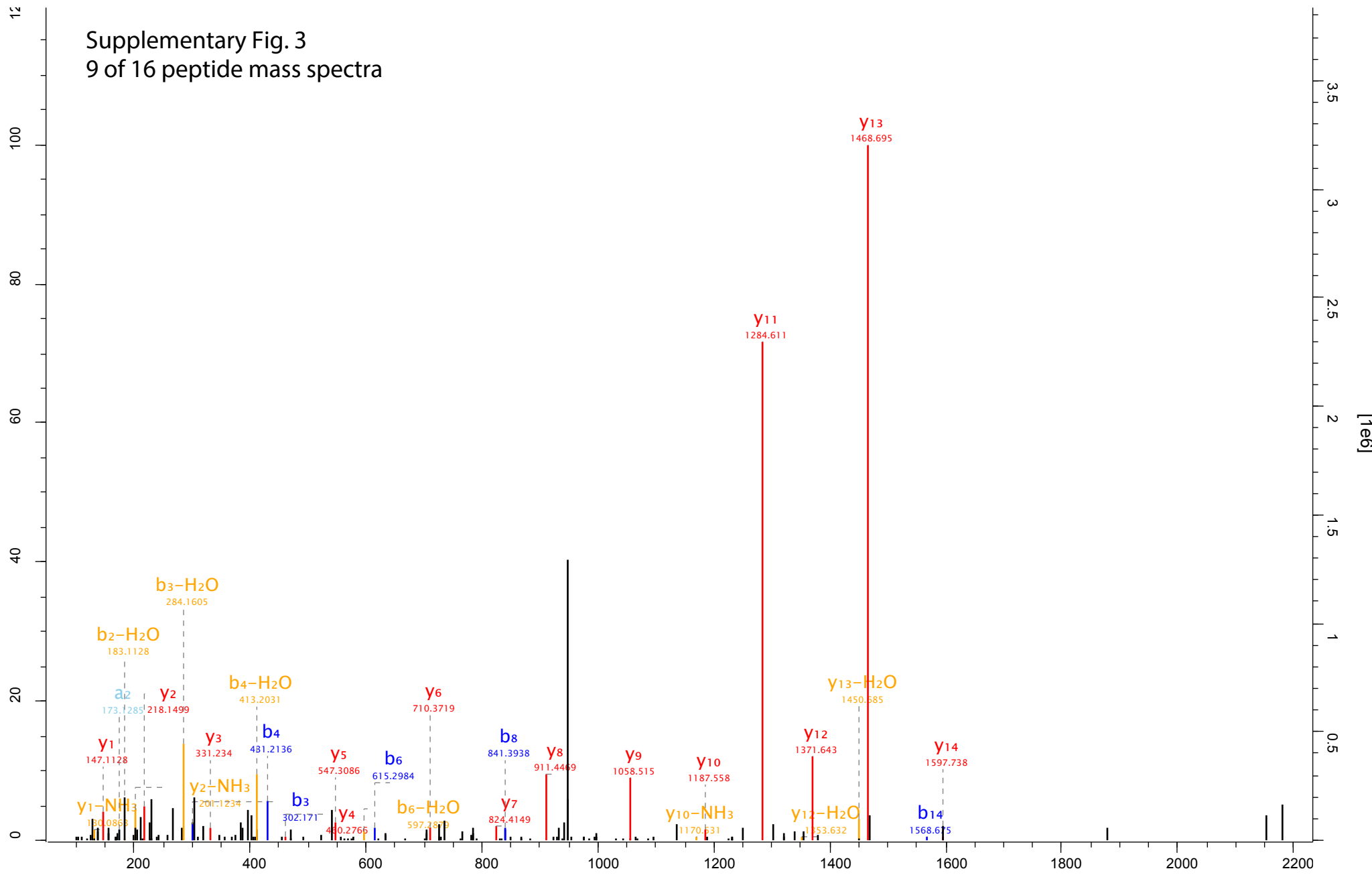
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Supplementary Fig. 3
 8 of 16 peptide mass spectra

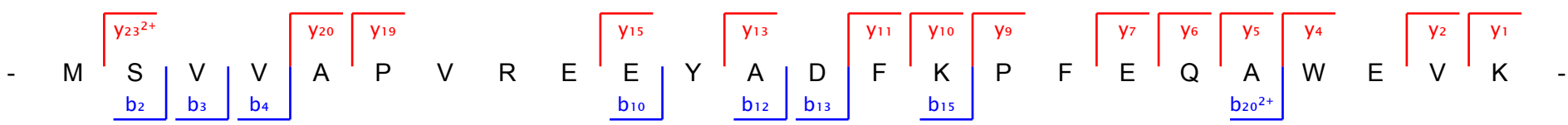
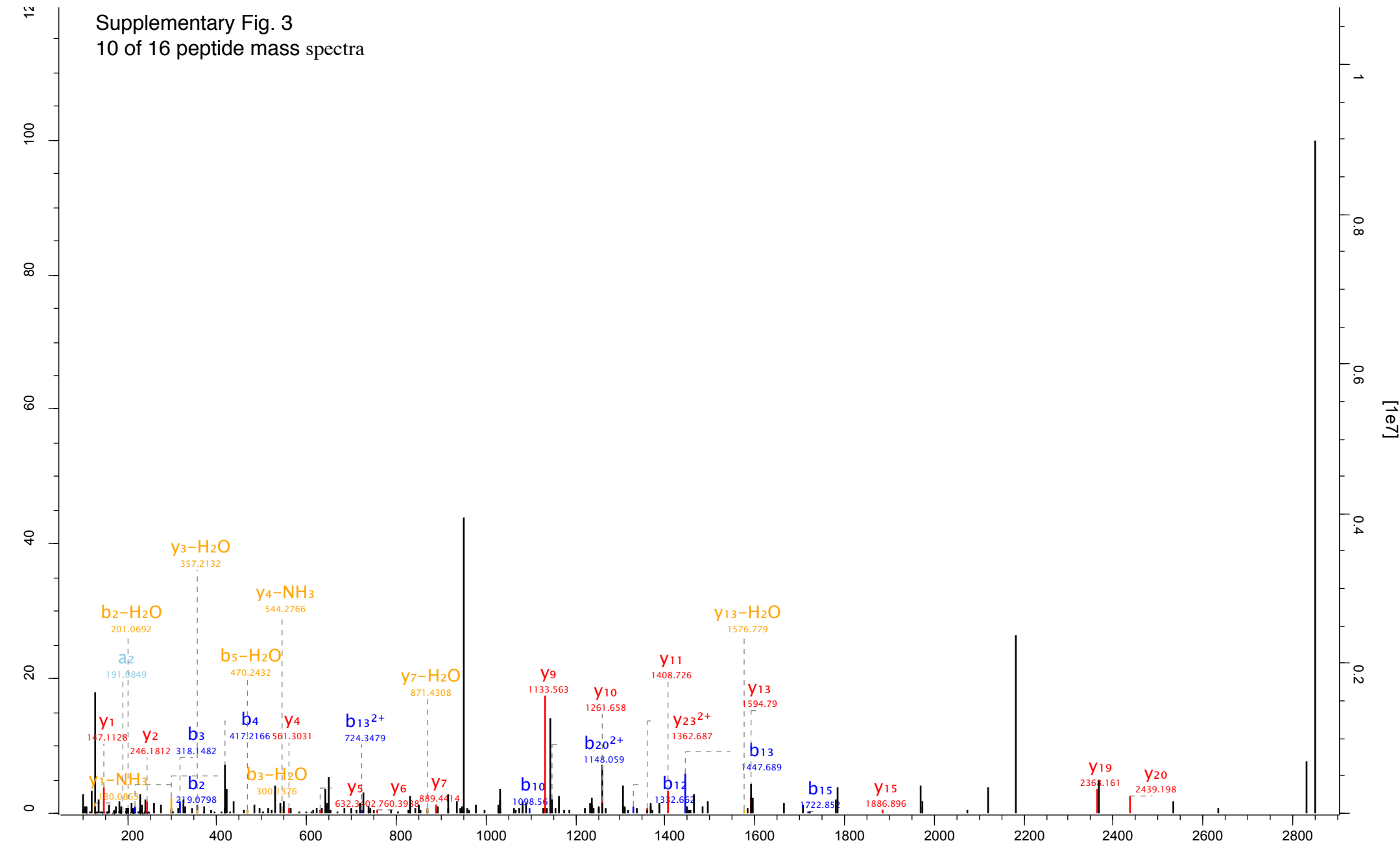


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Supplementary Fig. 3
 9 of 16 peptide mass spectra

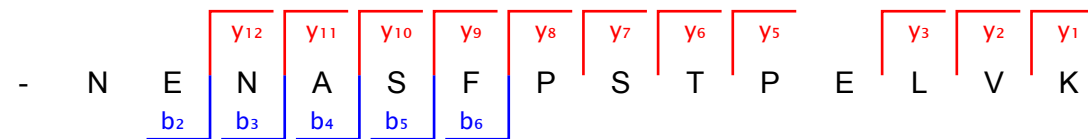
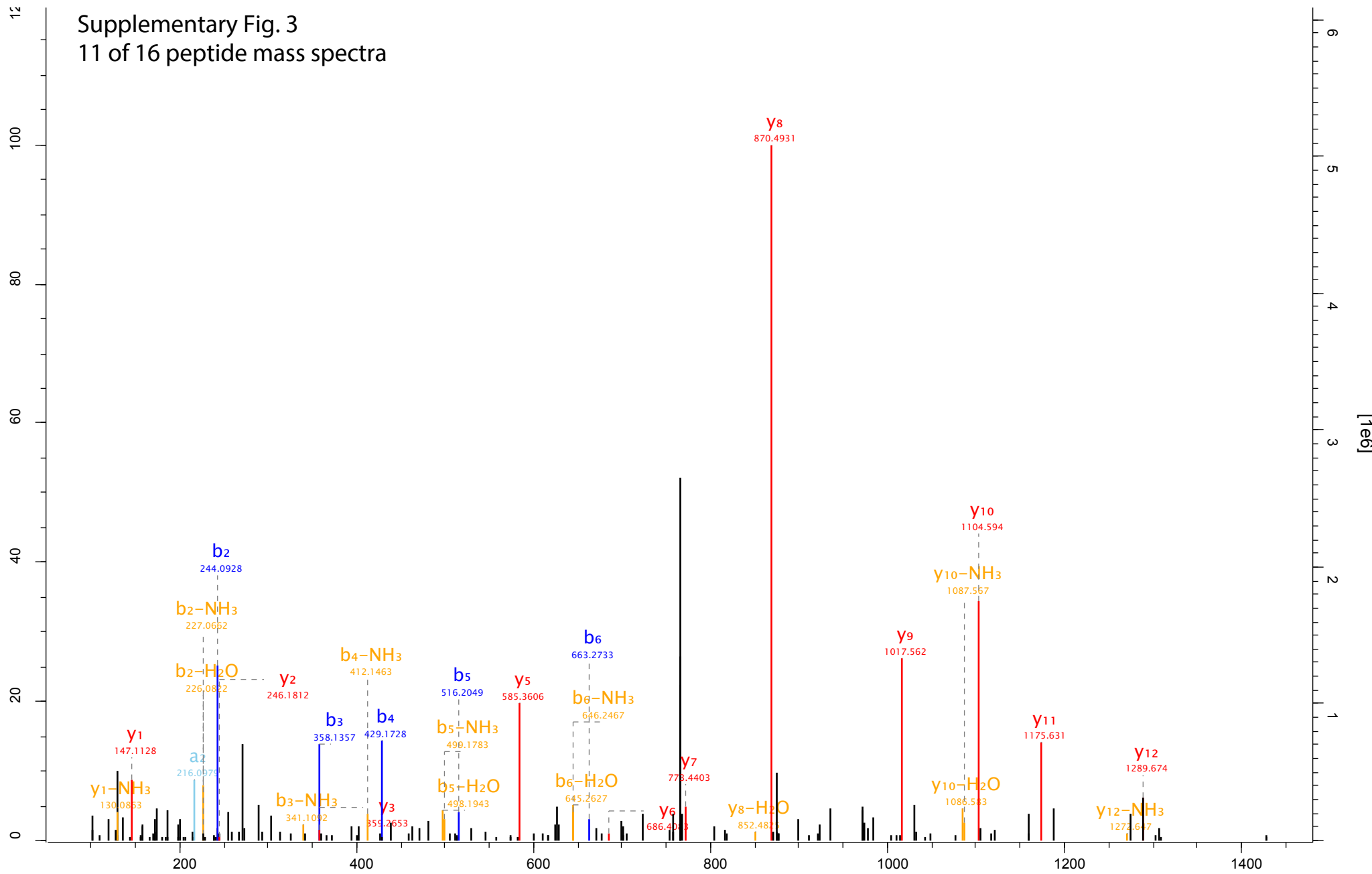


Supplementary Fig. 3
10 of 16 peptide mass spectra



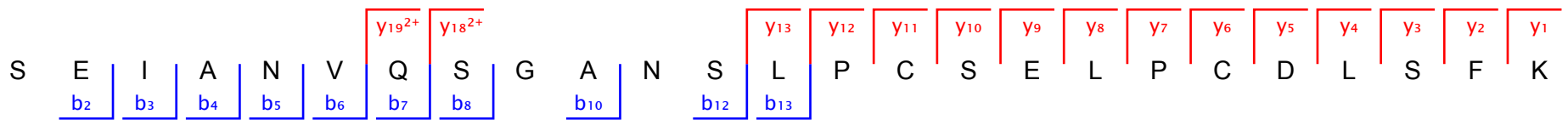
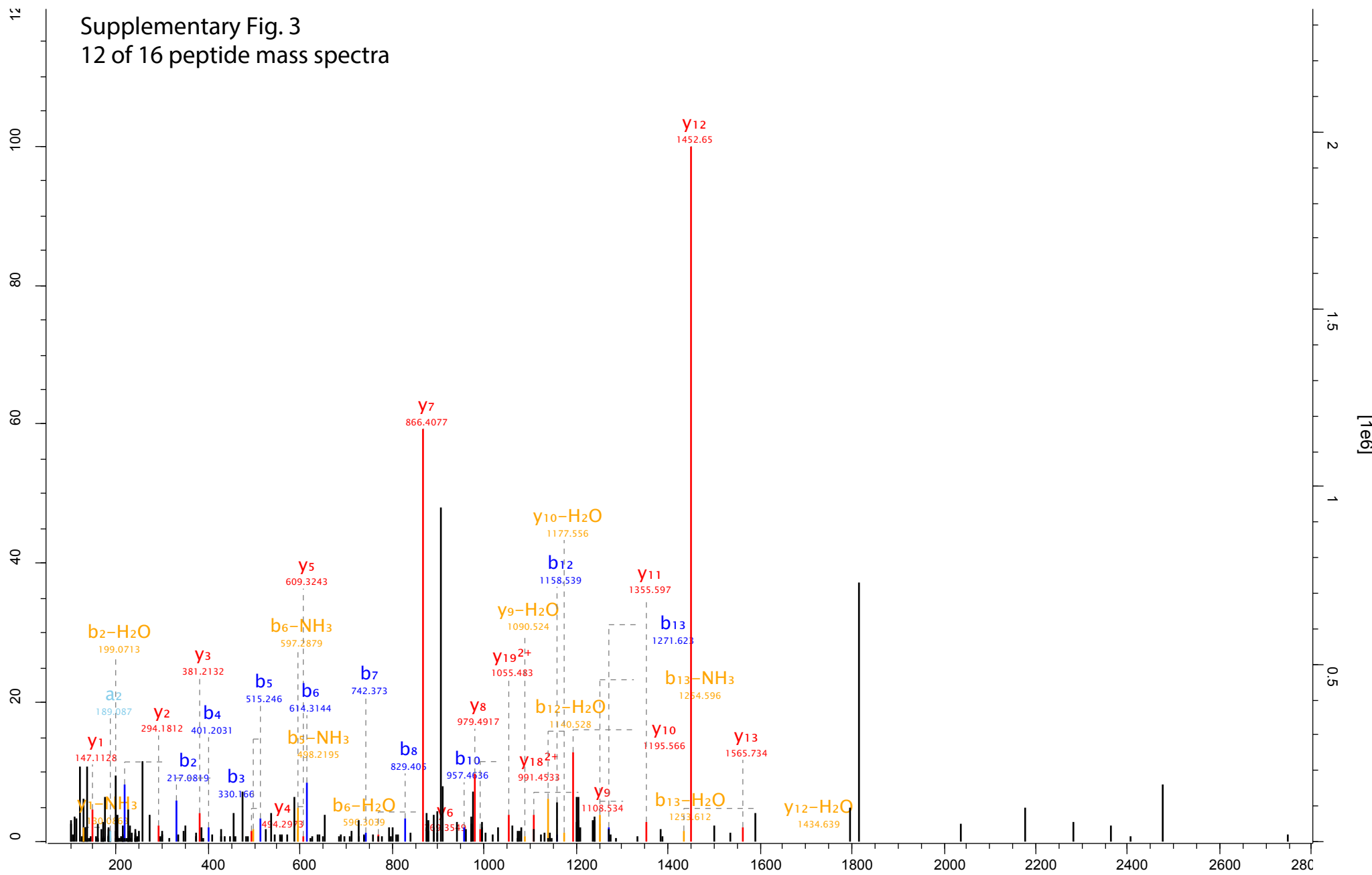
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Supplementary Fig. 3
 11 of 16 peptide mass spectra



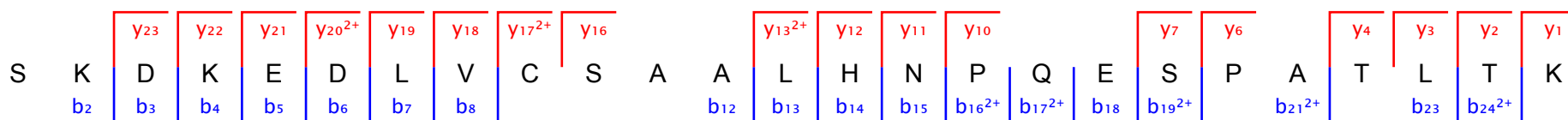
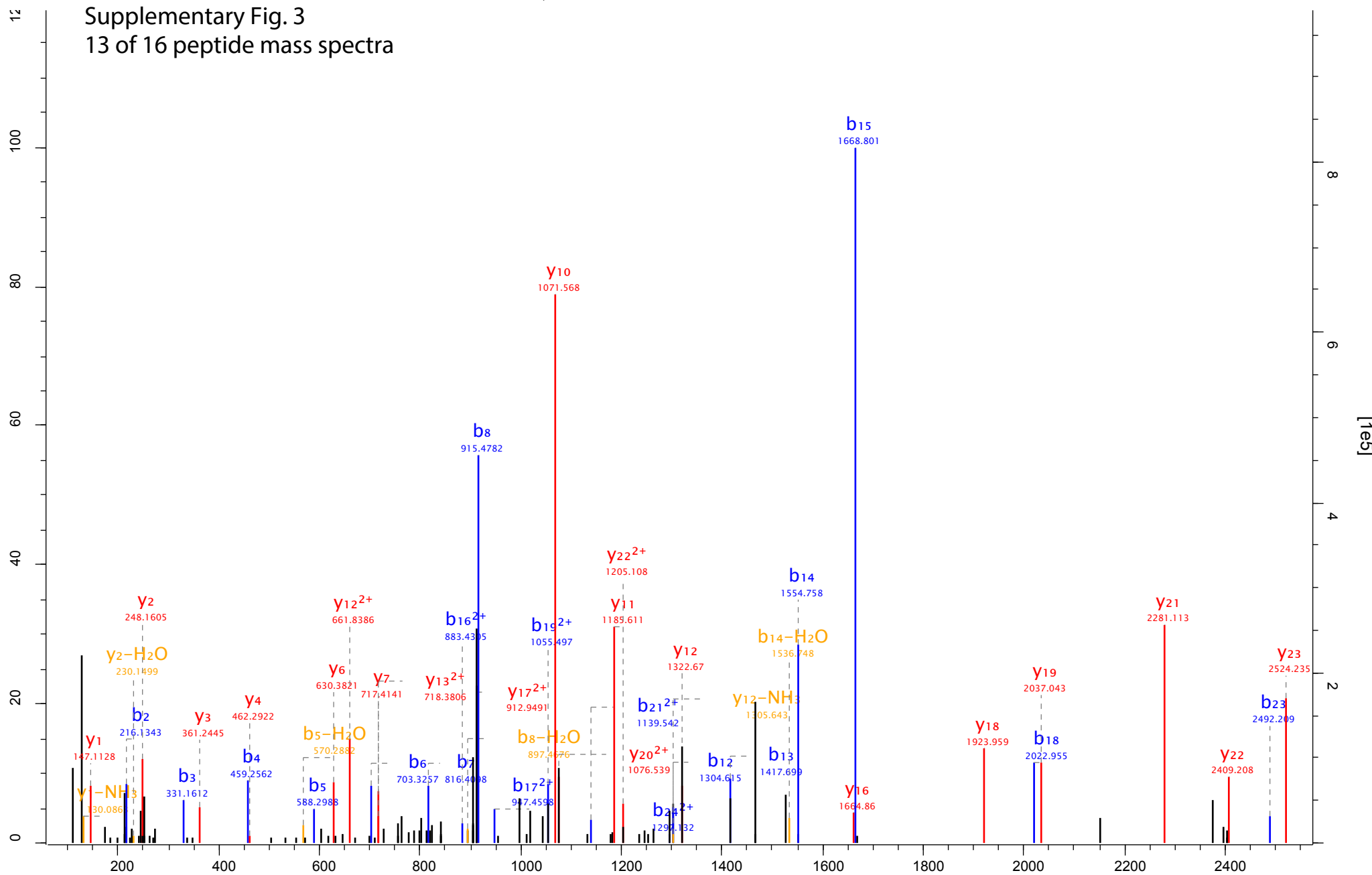
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Supplementary Fig. 3
12 of 16 peptide mass spectra



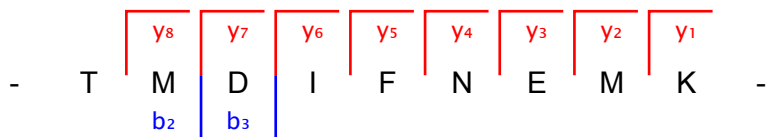
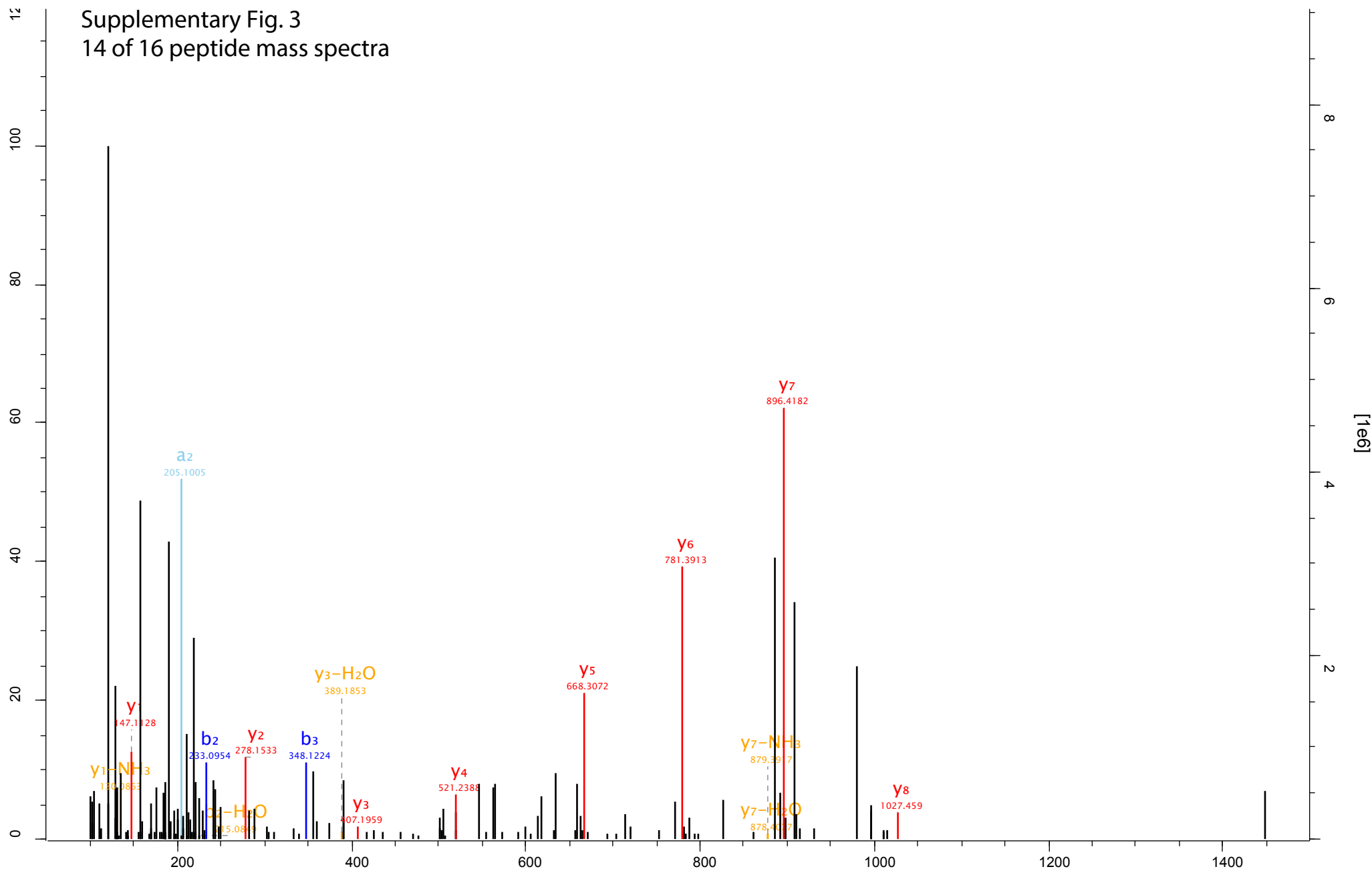
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Supplementary Fig. 3
 13 of 16 peptide mass spectra

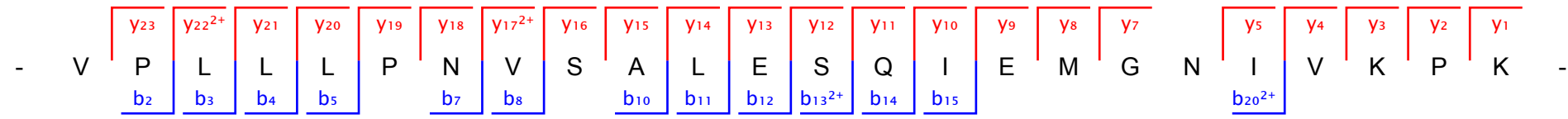
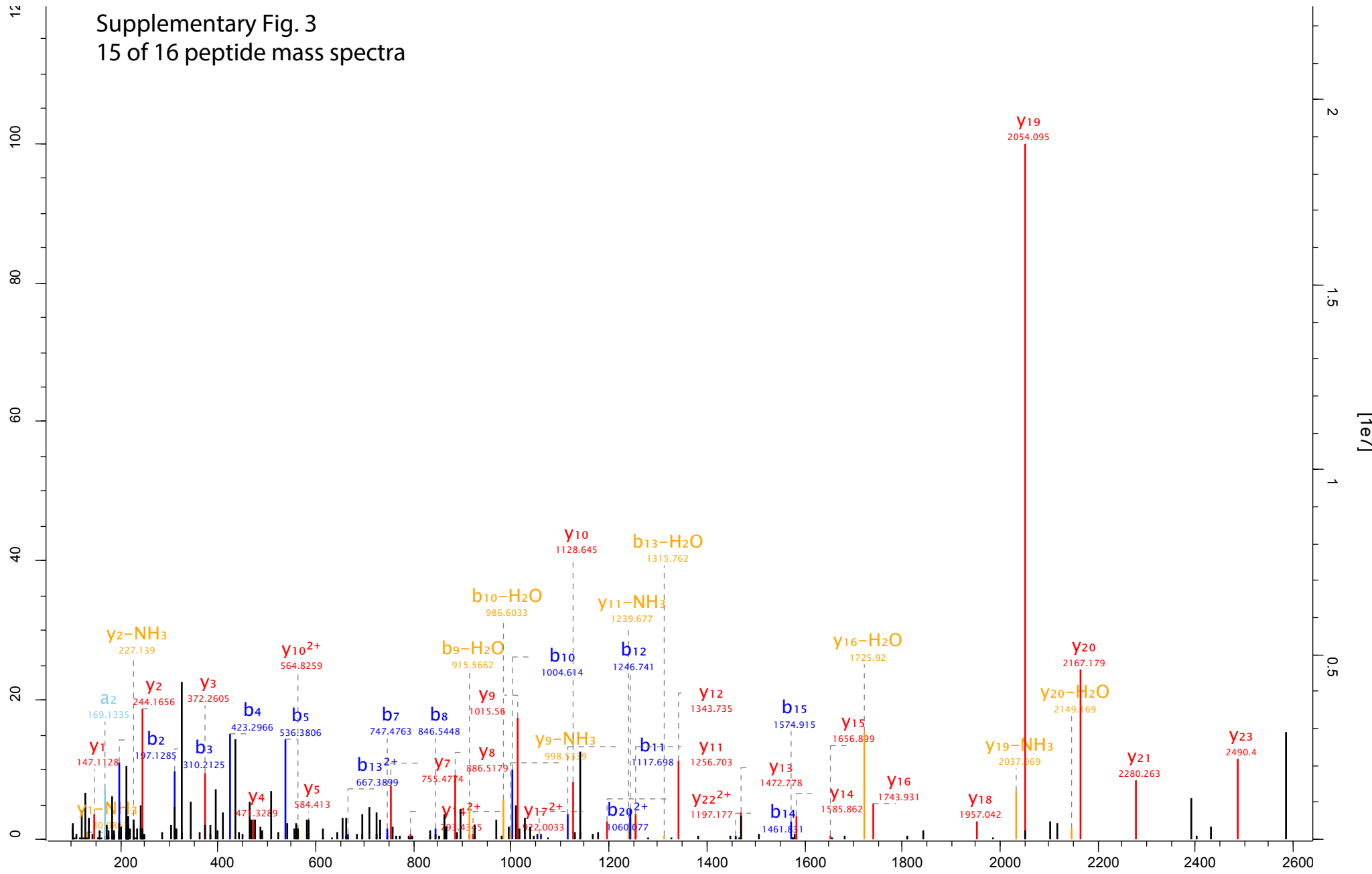


Scan 68376 Method FTMS; HCD Score 50.3 m/z 564.75

Supplementary Fig. 3
14 of 16 peptide mass spectra



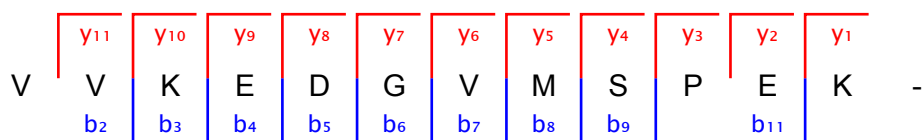
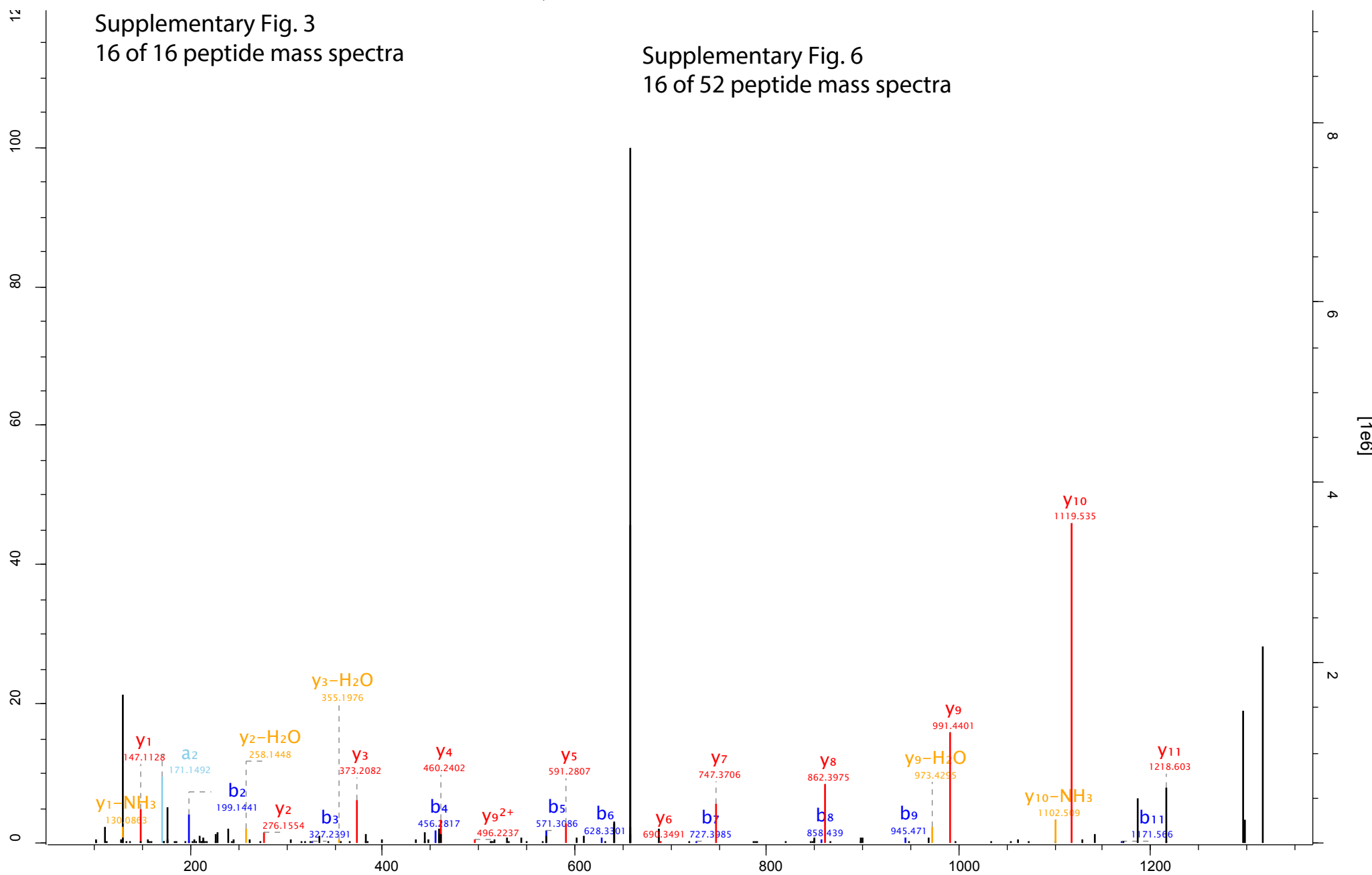
Supplementary Fig. 3
15 of 16 peptide mass spectra



Scan Method Score m/z
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Supplementary Fig. 3
 16 of 16 peptide mass spectra

Supplementary Fig. 6
 16 of 52 peptide mass spectra



A.

MDLK_EQPGNTVSSGQEDFPSVLFETAASLPSLSPLSTVSVFK_EHGylGNLSAVASTEgTIEE
TLNEASR_ELPERATNPFVNRESAEFSVLEYSEMGSsfNGSPKGESAMLVENTKeeVIVR_S
KDKEDLVCSAALHNPQESpATLTK_VVKEDGVMsPEK_TMDIFNEMK_MsVVAPVREEYAD
FKPFEQAWEvK_DTYEGSRDVLAArANMESKVDKkCFEDSLEQKSHGkdSESR_NENASf
PSTPELVK_DGSRAYITCDSfTSATEStAANIFPVLEDHTSENKTDEKKIEERKAQIIITEKTSP
KTSNPFLVAIHdSEADYVTTDNLSKVTEAVVATMPEGLTPDLVQeACESELNEATGTKIA
YETKVDLVQTSerAIQESIYPTAQLCPSFEeAEATPSPVLPDIVMEAPLNsLLPSTGASVAQP
SASPLeVSPVSYDGIK_LEPENPPPYYEEAMsVALK_TSDAK_EEIKEPESfNAAVQeAEAPYIS
IACDLIK_ETK_LSTEPSPEFSNYSEIAK_FEKSVPDHCELVDdSSPESEPVDFsDDSIpDVPQT
QEEAVMLMK_ESLTEVSETVTQHK_HKERLSASPQEVGKPYLESfQPNLHITKDAASNEIPT
LTKKETISLQMEEFNTAIYSNDDLLSSKEDKMKESeTfSDSSPIEIIDeFPTfVSAKDDSPK_
EYTDLEVSnk_SEIANVQSGANSLPCSELPCDLsFK_NTYPK_DEAHVSDEFSK_SRSSVSK_VPLLL
PNVSALESQIEMGNIVKPK_VLTKEAEeKLPsDTEKEDRSLTAVLSAElnKTSDETLfALP
AASEPVIPSSAEKI

B.

MDLK.EQPGNTVSSGQEDFPSVLFETAASLPSLSPLSTVSVFK_EHGylGNLSAVASTEgTIEE
TLNEASR_ELPER_ATNPFVNR_ESAEFSVLEYSEMGSsfNGSPK_GESAMLVENTK.EEVIVR_S
K.DKEDLVCSAALHNPQESpATLTK_VVK.EDGVMsPEK_TMDIFNEMK_MsVVAPVR.EEYAD
FKPFEQAWEvK.DTYEGSR.DVLAAR_ANMESKVDKk_CFEDSLEQK_SHGK.DSESR.NENASf
PSTPELVK.DGSR_AYITCDSfTSATEStAANIFPVLEDHTSENKTDEK.K_IIEER_KAQIIITEKTSP
K_TSNPFLVAIHdSEADYVTTDNLSK_VTEAVVATMPEGLTPDLVQeACESELNEATGTK_IA
YETKVDLVQTSerAIQESIYPTAQLCPSFEeAEATPSPVLPDIVMEAPLNsLLPSTGASVAQP
SASPLeVSPVSYDGIK_LEPENPPPYYEEAMsVALK_TSDAK_EEIKEPESfNAAVQeAEAPYIS
IACDLIK.ETK_LSTEPSPEFSNYSEIAK.FEK_SVPDHCELVDdSSPESEPVDFsDDSIpDVPQT
QEEAVMLMK_ESLTEVSETVTQHK_HKERLSASPQEVGKPYLESfQPNLHITK_DAASNEIPT
LTK.K_ETISLQMEEFNTAIYSNDDLLSSK.EDK_MK.ESETfSDSSPIEIIDeFPTfVSAK.DDSPK_
EYTDLEVSnk_SEIANVQSGANSLPCSELPCDLsFK_NTYPK.DEAHVSDEFSK_SRSSVSK_VPLLL
PNVSALESQIEMGNIVK.PK_VLTK_EAEeK.LPSDTEKEDR_SLTAVLSAElnK_TSDETLfALP
AASEPVIPdykdddkppSSAEK_I

Supplementary Fig. 4 Tryptic peptides identified by mass spectrometry.

A, B. Peptides derived from construct circRtn4 and from circRtn4-FLAG are shown in red and blue, respectively. For mass spectrograms, see Supplementary Figs. 3 and 6, respectively. The amino acids contributed by the FLAG sequence are in lower case. The glutamic acid residue (E) contributed by the splice site between exons 2 and 3 (last row) and the aspartic acid residue (D) at the site of circularization (penultimate row) are shown in bold.

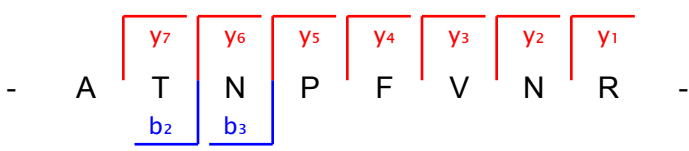
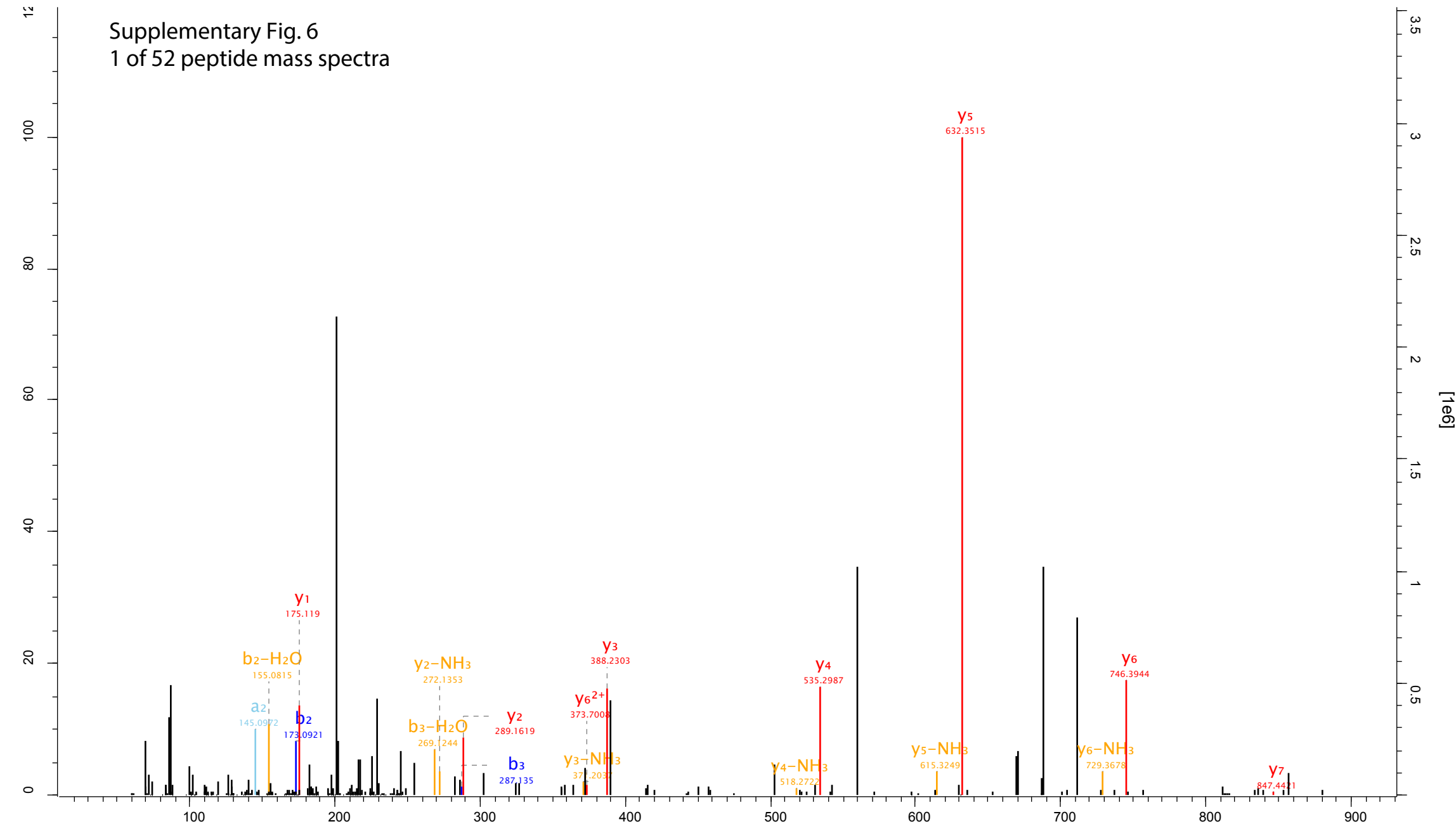
Complete digestion is marked by an underline symbol and observed incomplete digestion yielding two or more overlapping products is marked by a dot, i.e., there were additional sequences (N-terminally and/or C-terminally) in identified peptides beyond the dots.

Supplementary Fig. 6

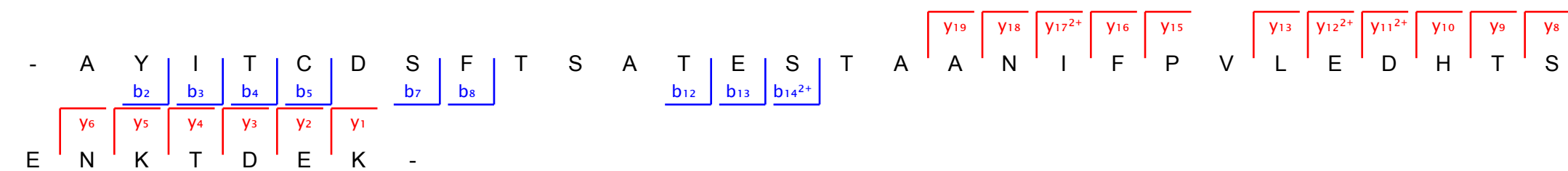
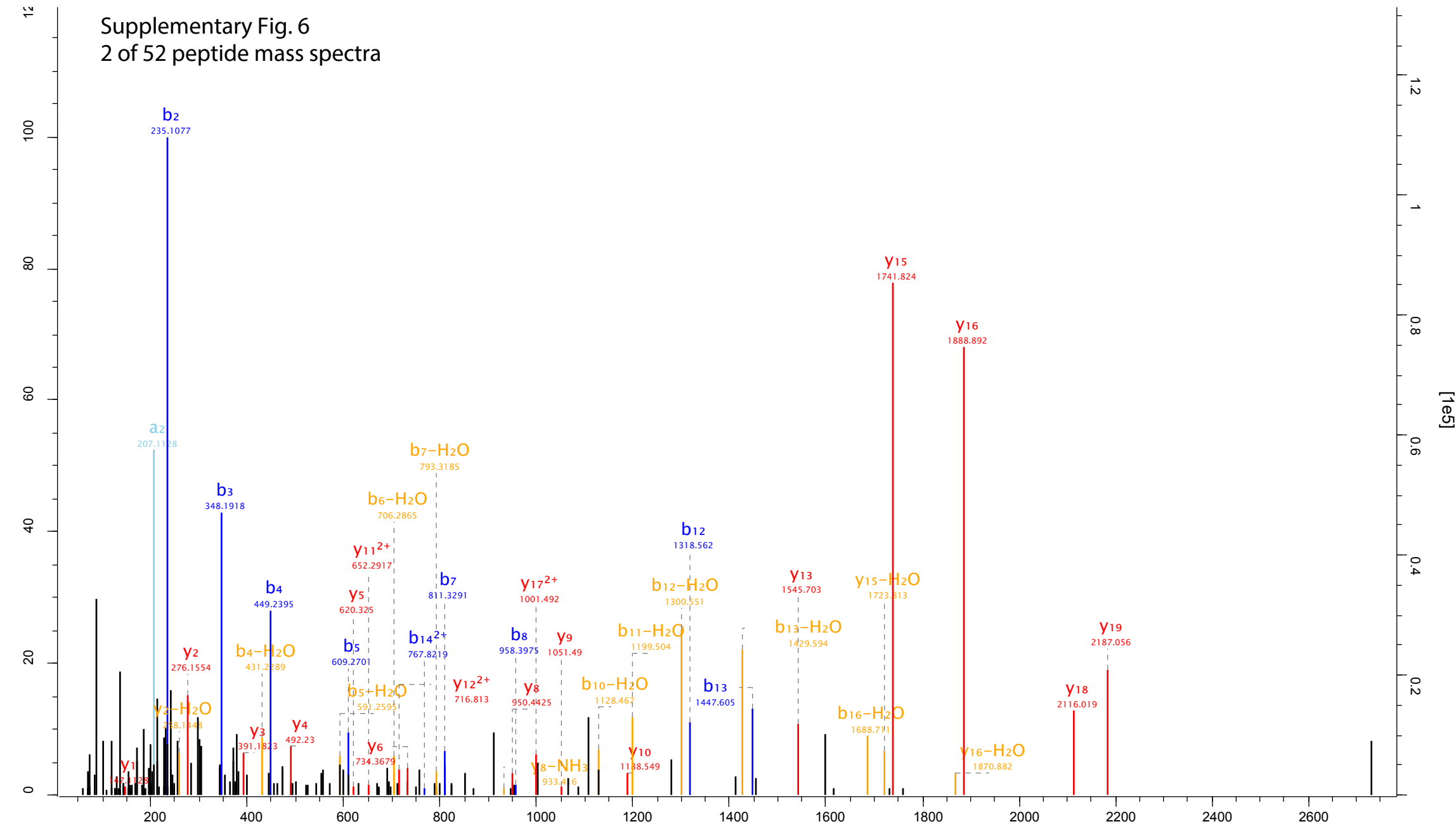
52 mass spectra of the tryptic peptides from circRtn4-FLAG derived protein

The left coordinate is the relative intensity, the right coordinate is the absolute intensity, the horizontal coordinate is m/z .

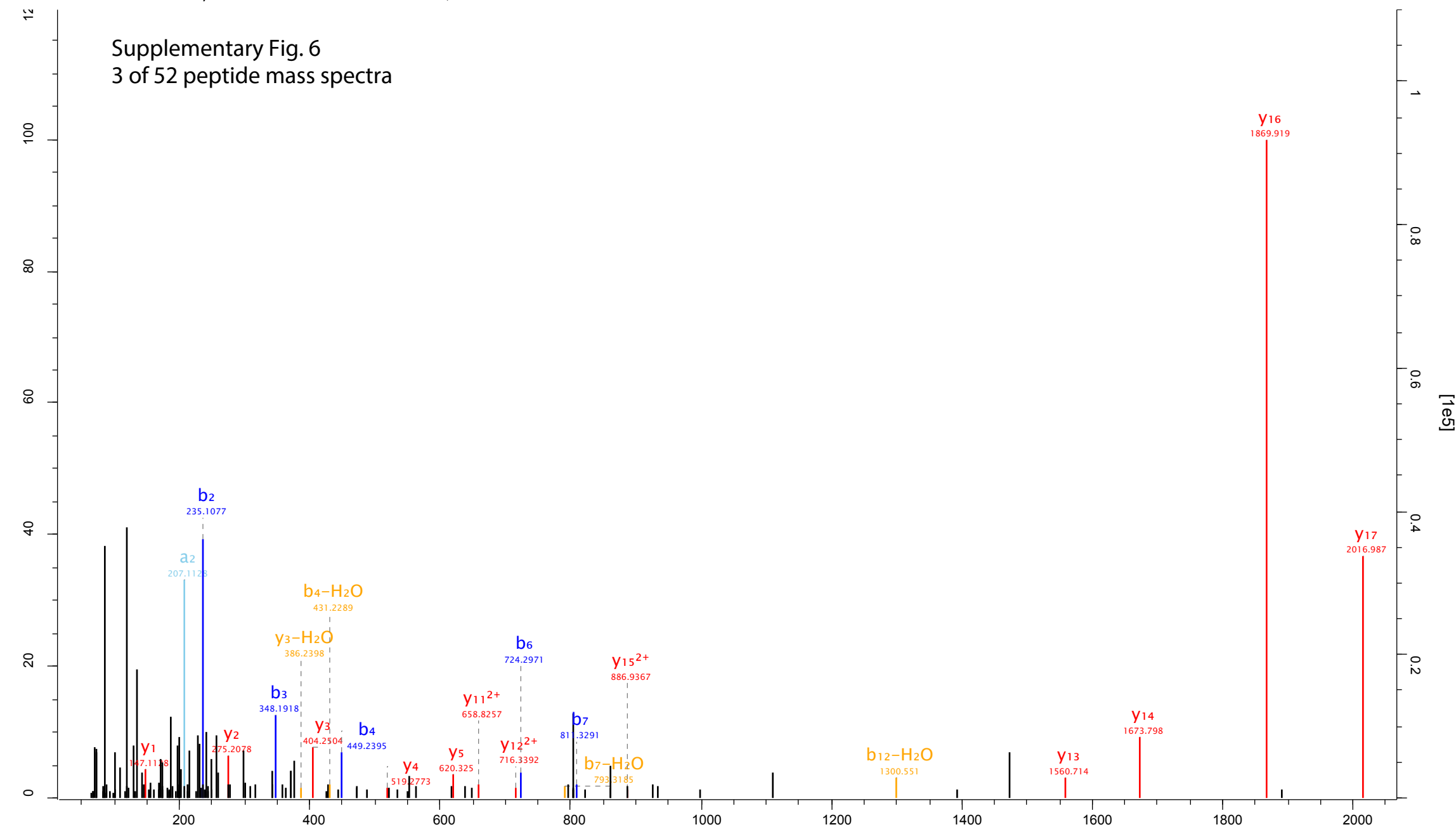
Supplementary Fig. 6
1 of 52 peptide mass spectra



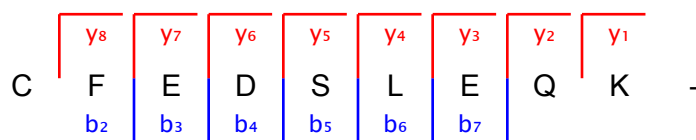
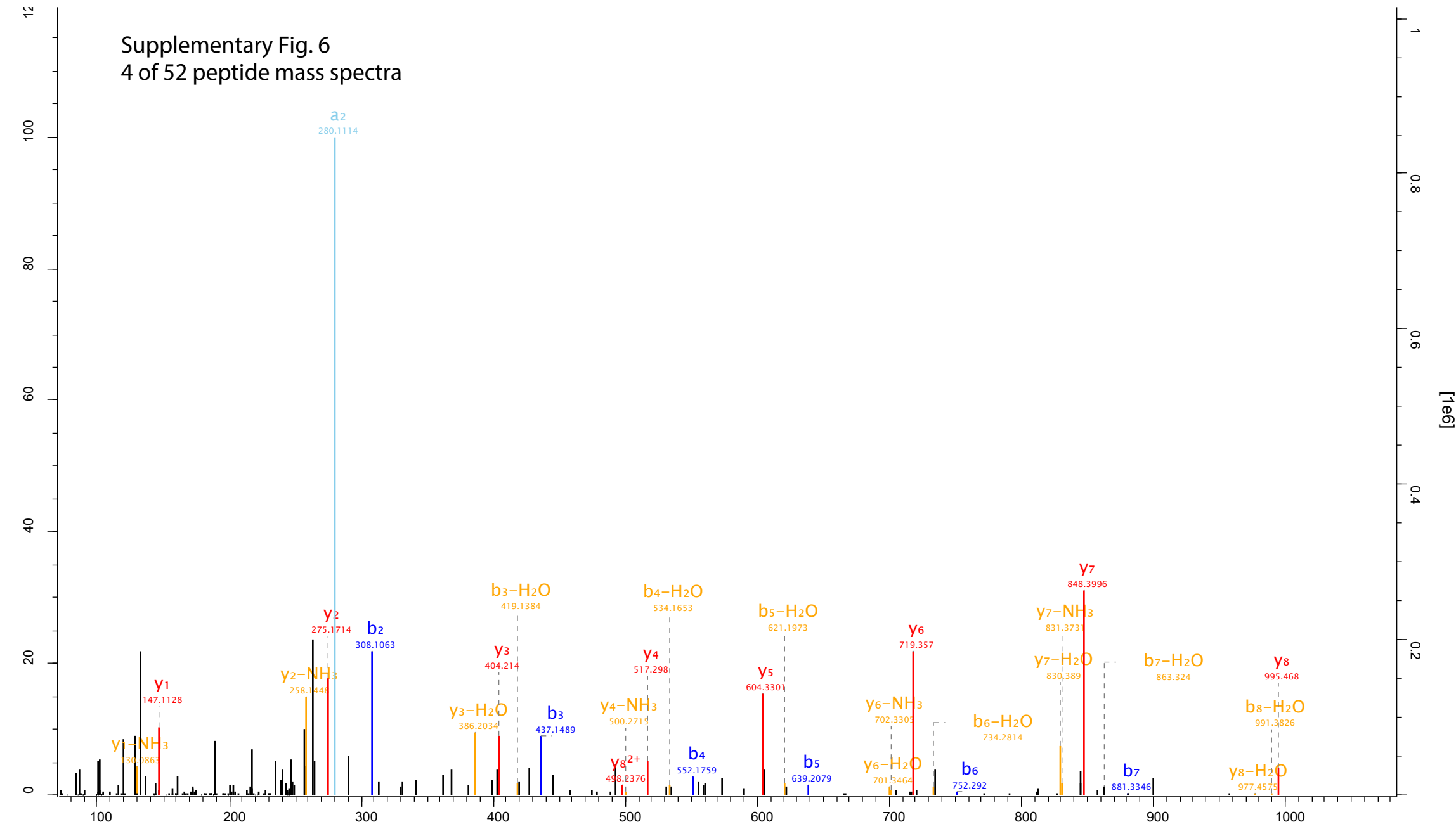
Supplementary Fig. 6
 2 of 52 peptide mass spectra



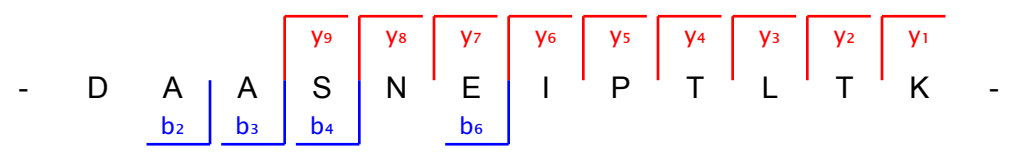
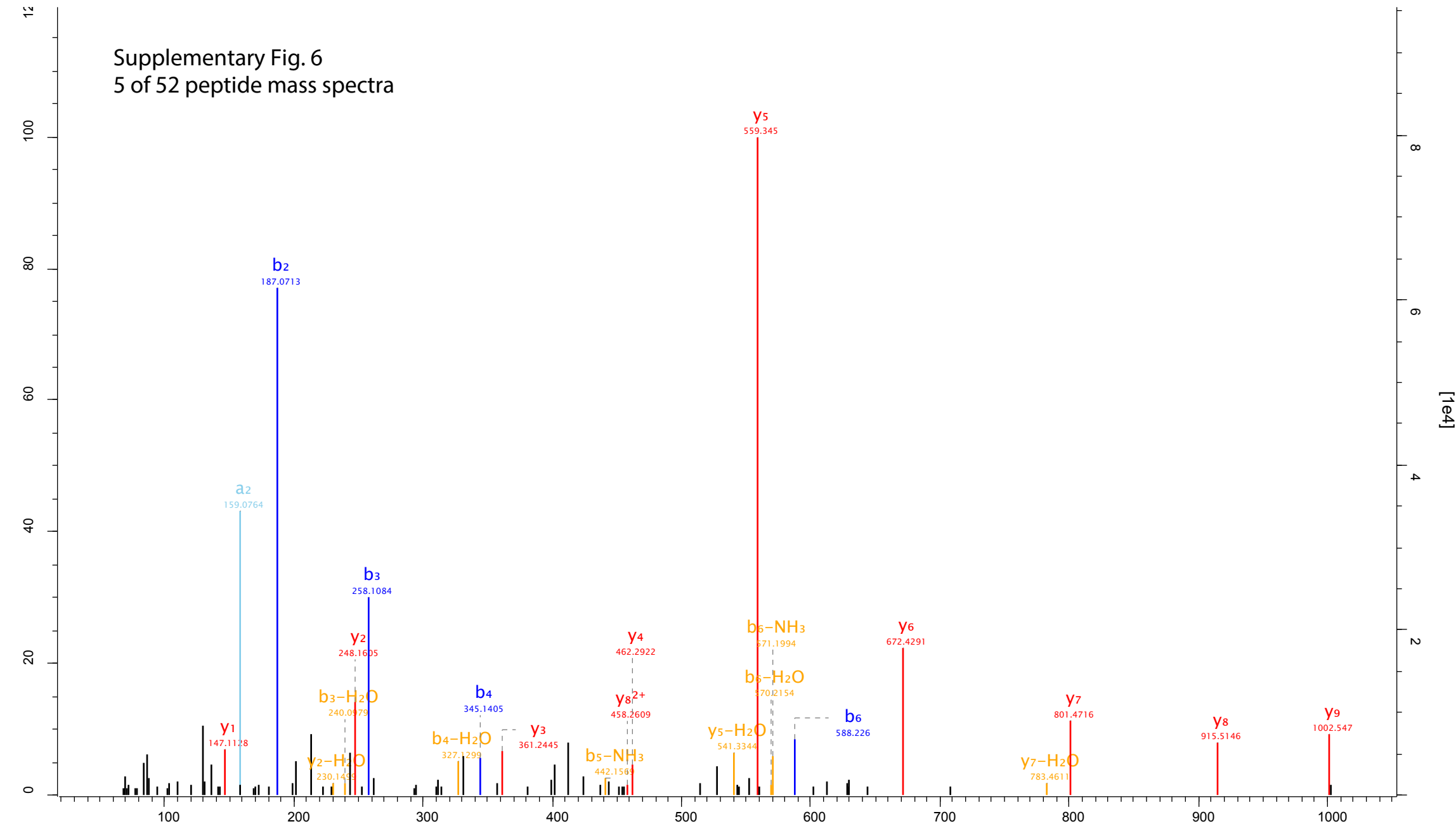
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3 of 52 peptide mass spectra



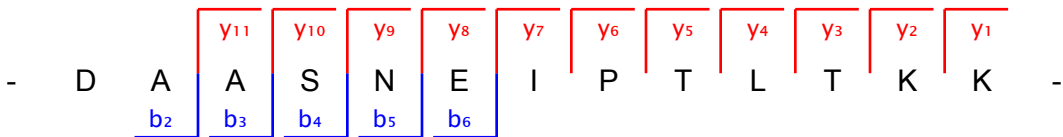
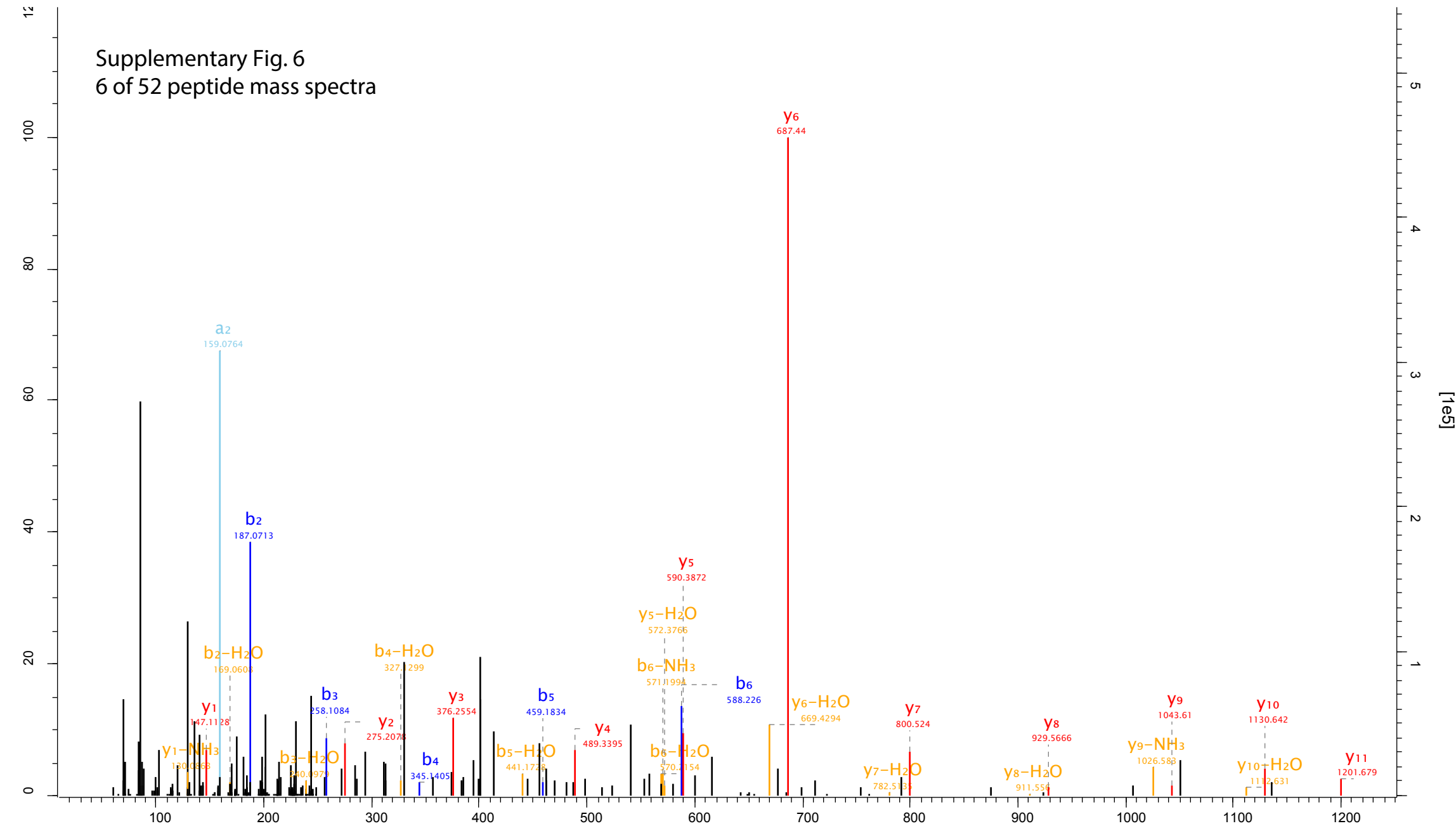
Supplementary Fig. 6
4 of 52 peptide mass spectra



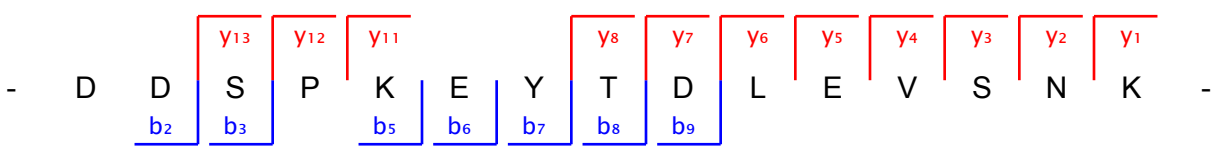
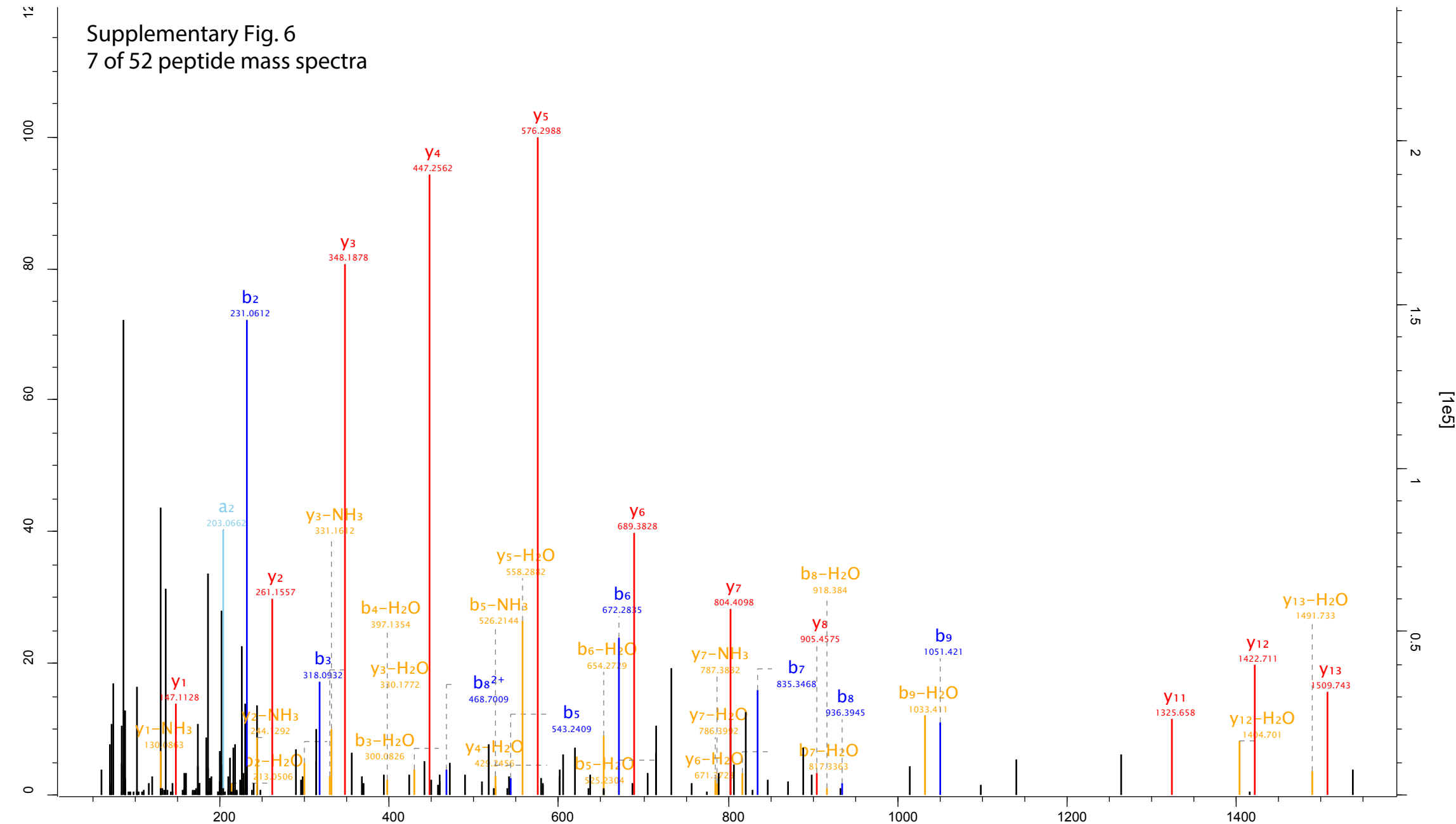
Supplementary Fig. 6
5 of 52 peptide mass spectra



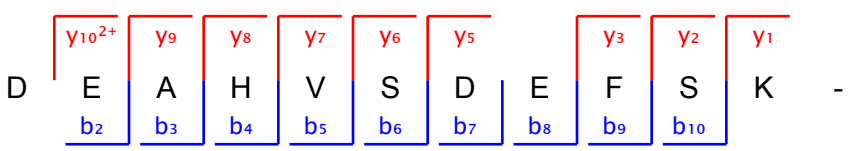
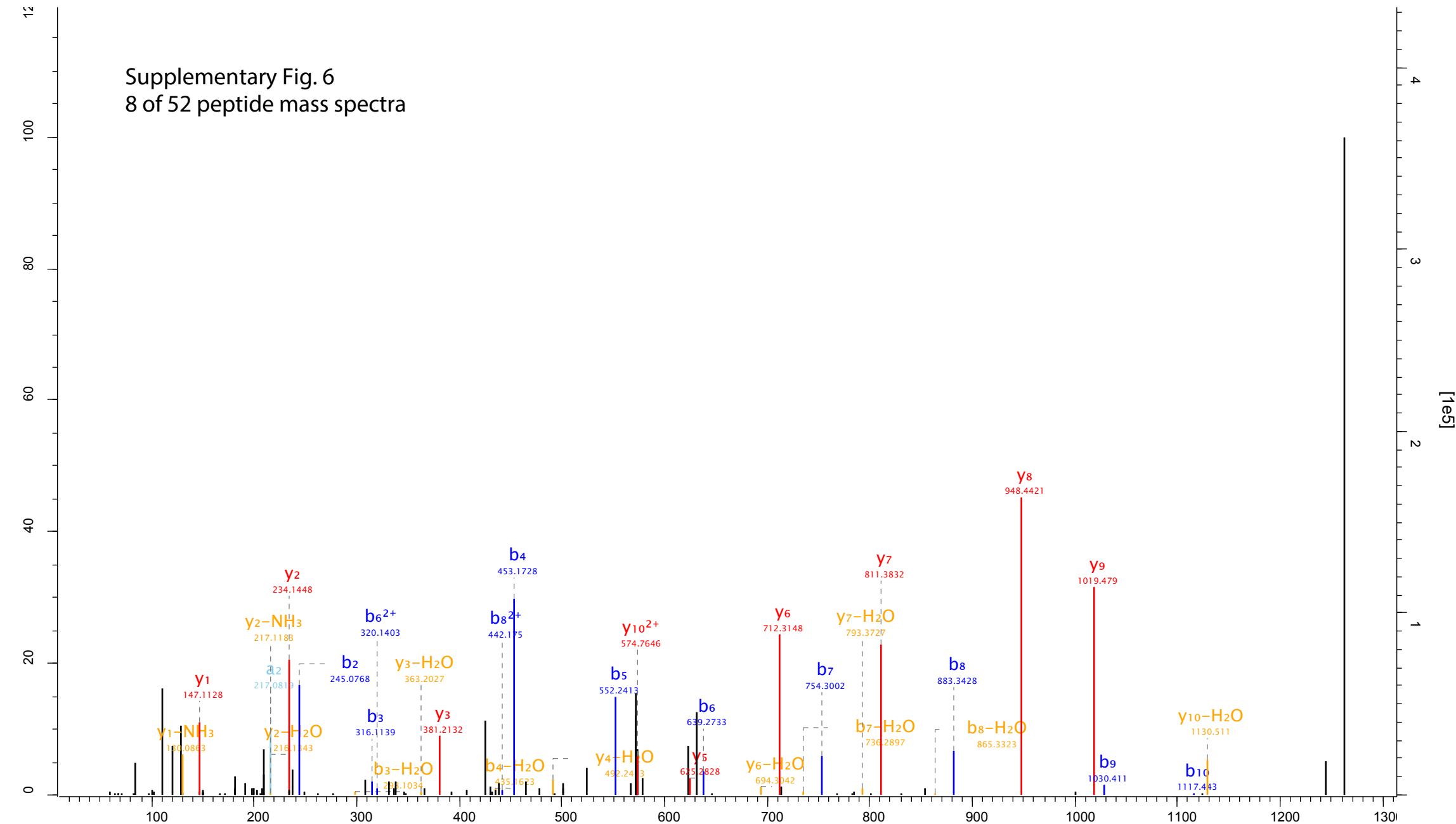
Supplementary Fig. 6
6 of 52 peptide mass spectra



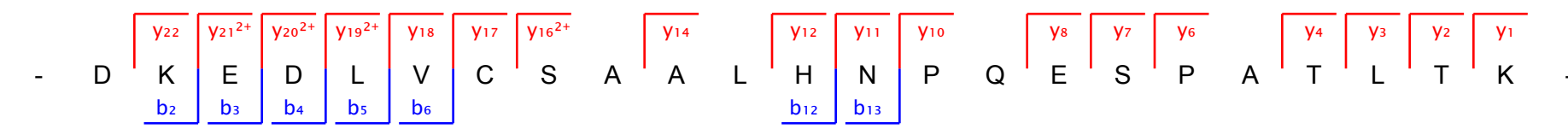
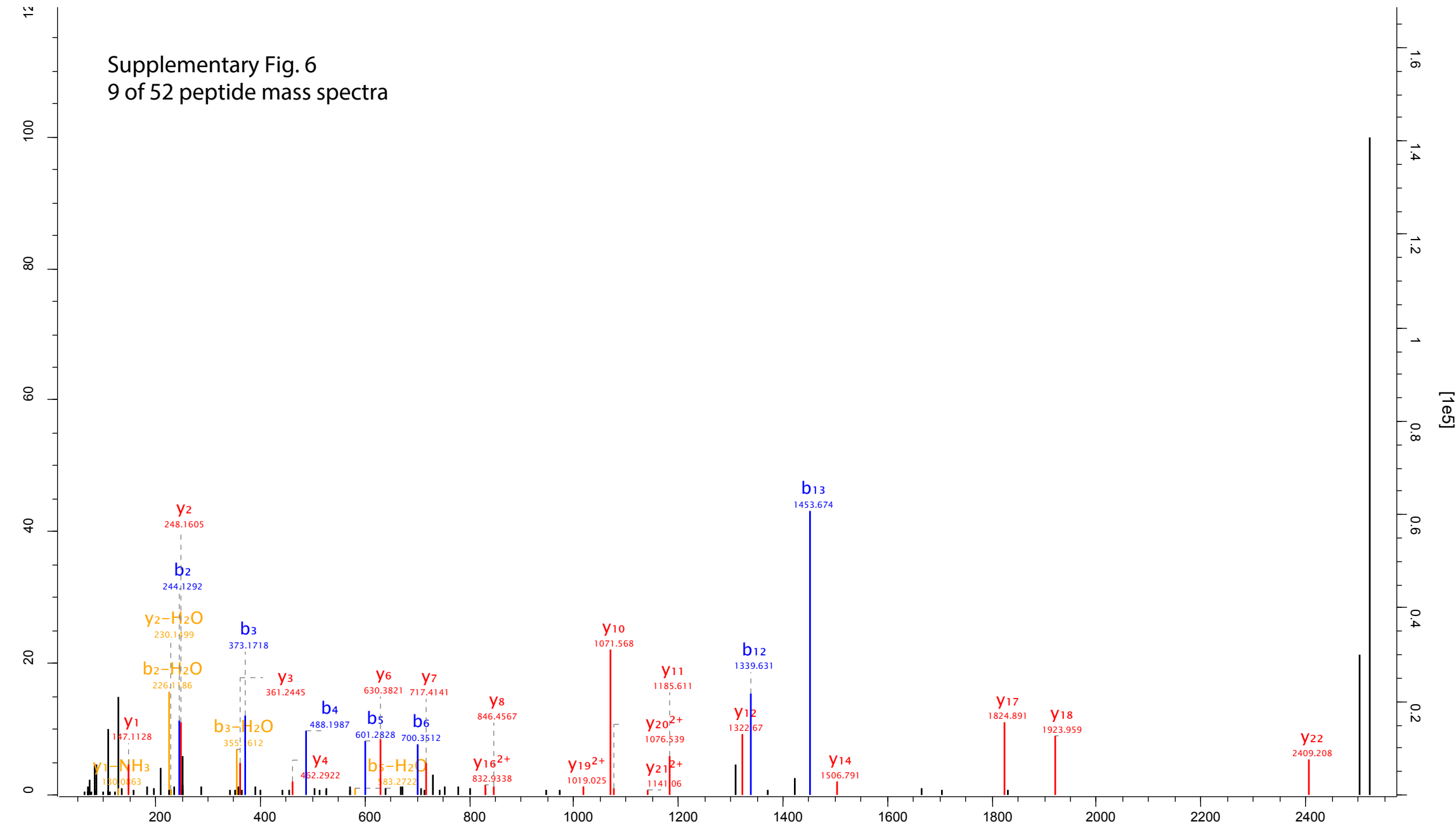
Supplementary Fig. 6
7 of 52 peptide mass spectra



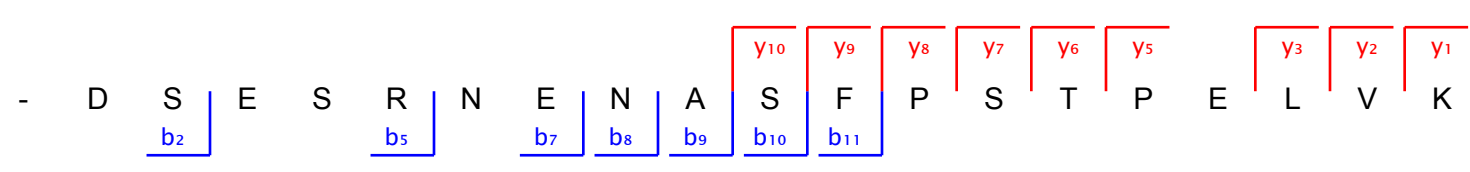
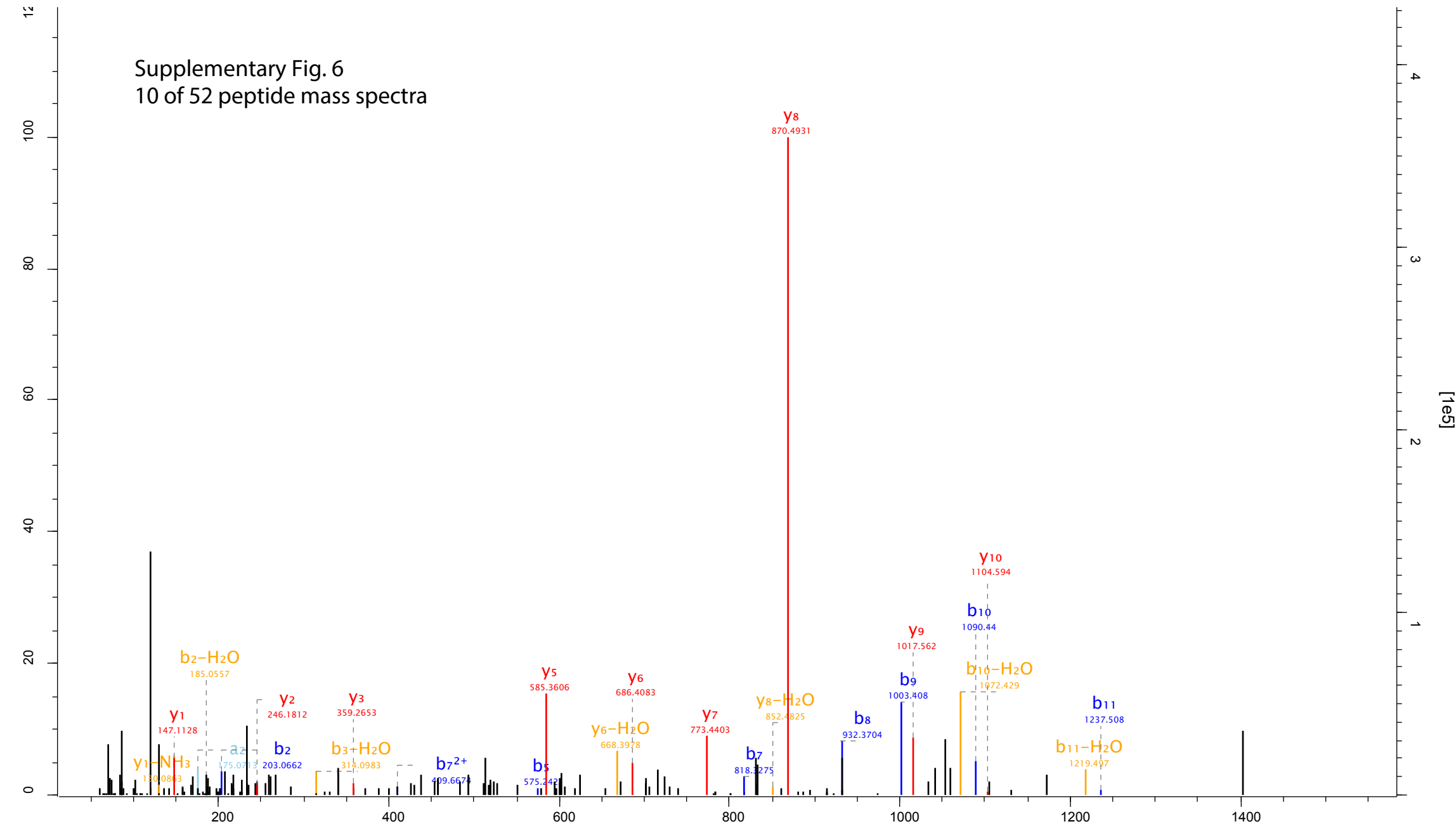
Supplementary Fig. 6
8 of 52 peptide mass spectra



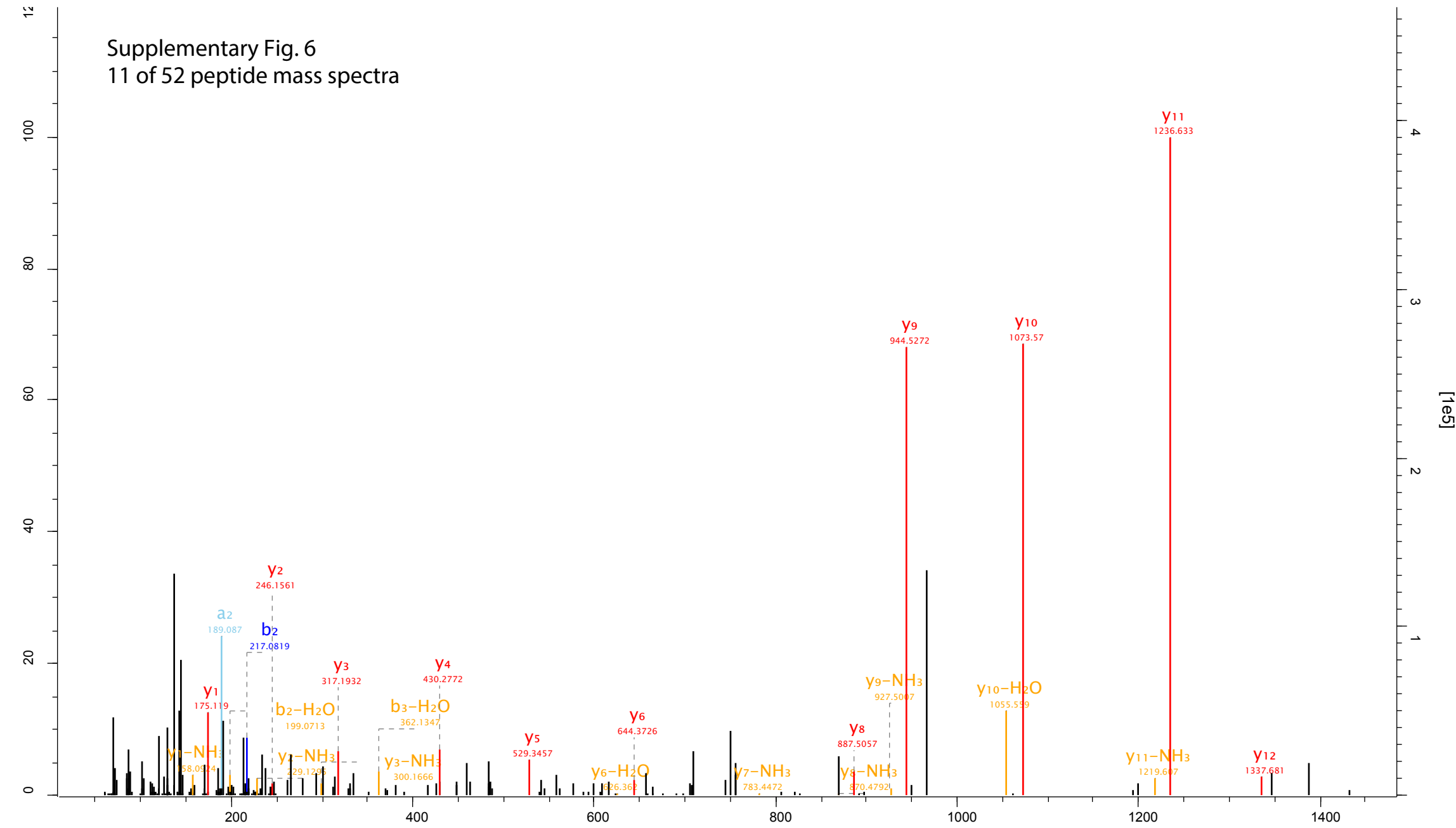
Supplementary Fig. 6
9 of 52 peptide mass spectra



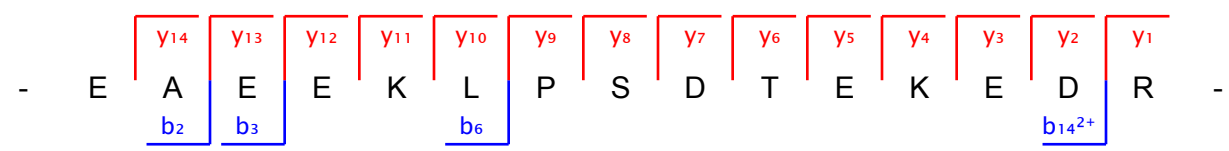
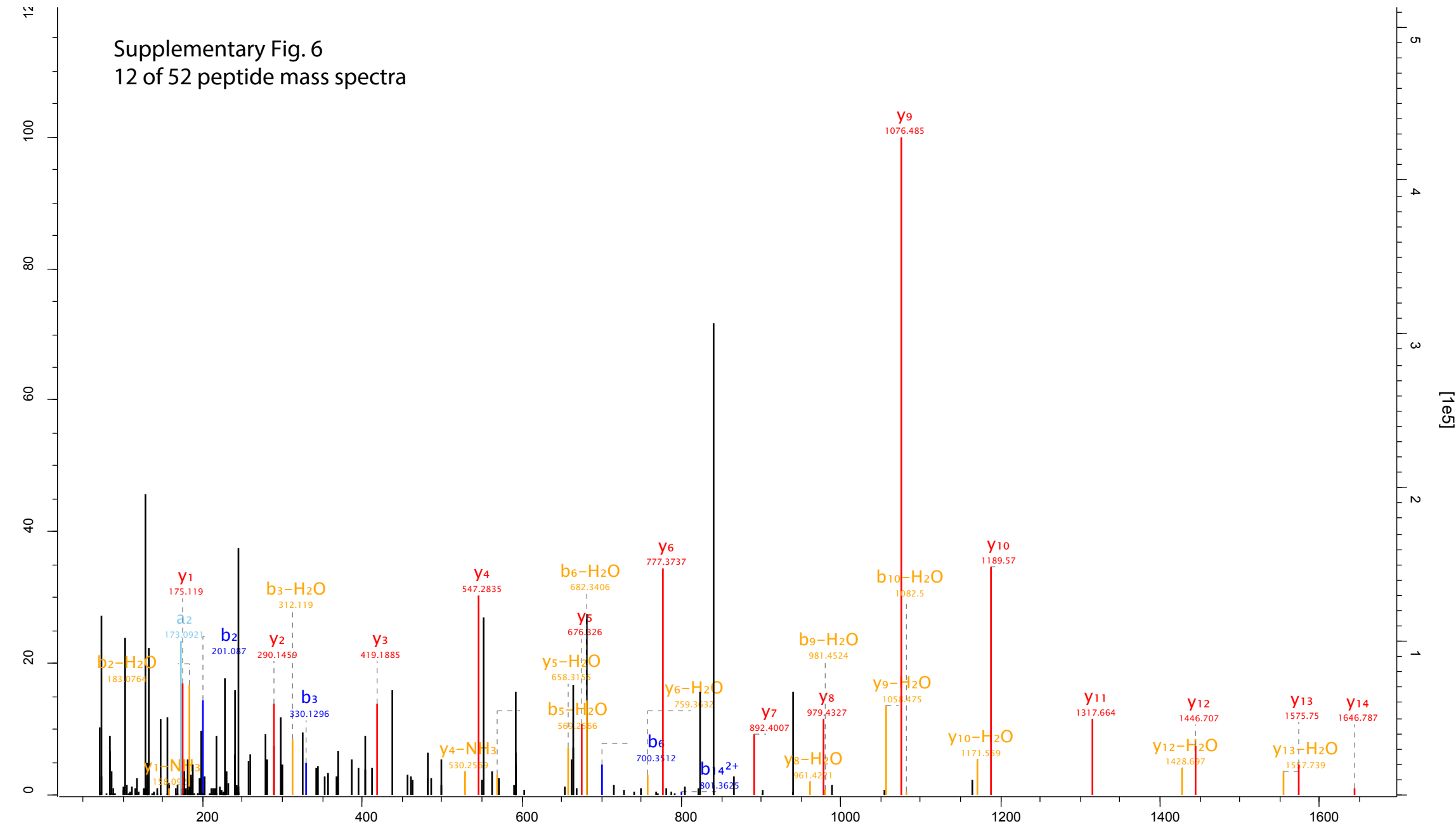
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10 of 52 peptide mass spectra



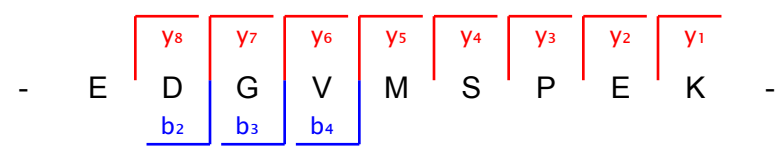
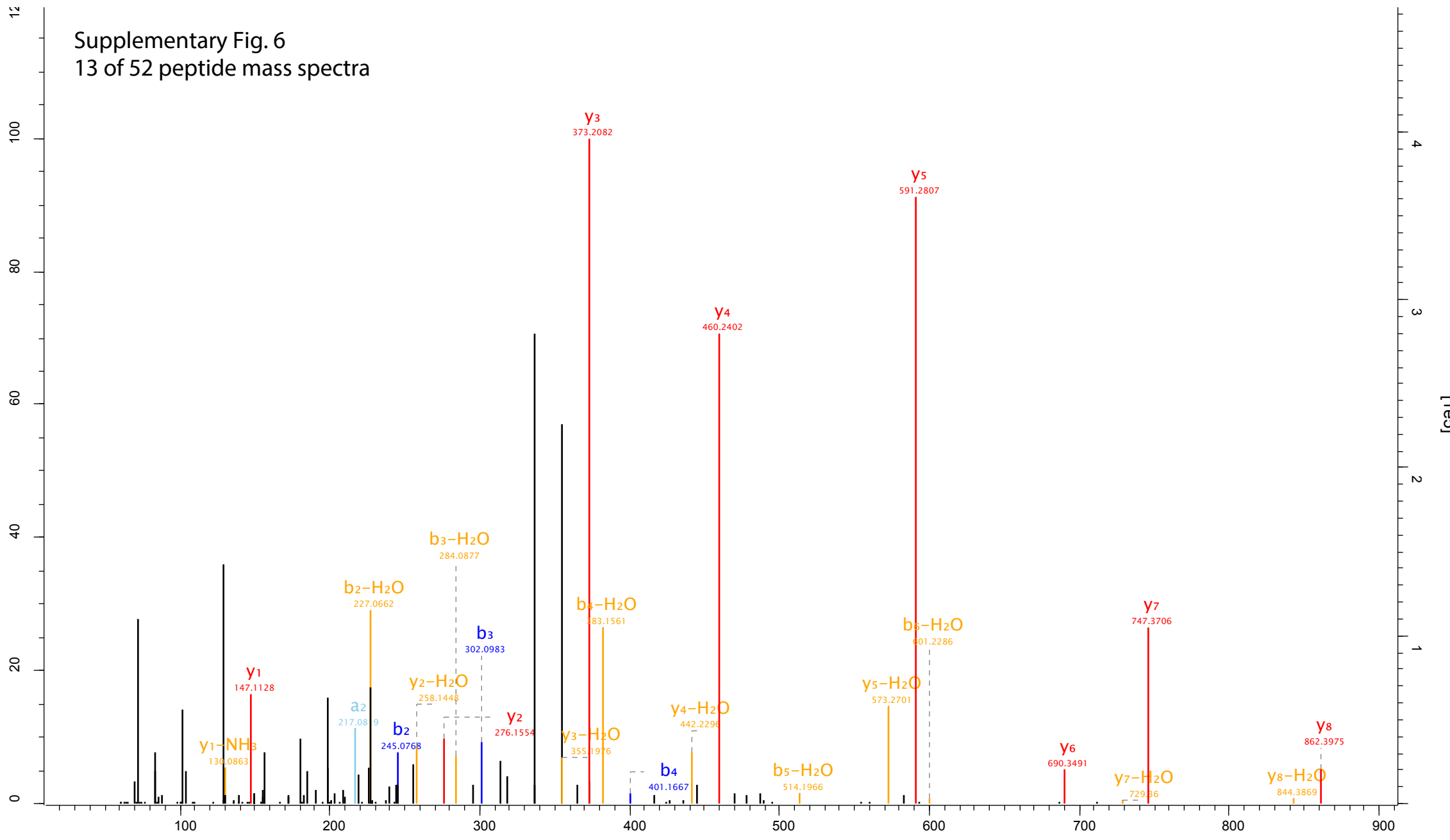
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11 of 52 peptide mass spectra



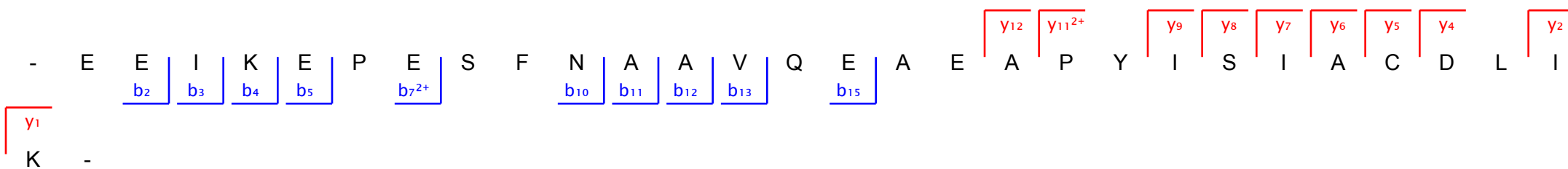
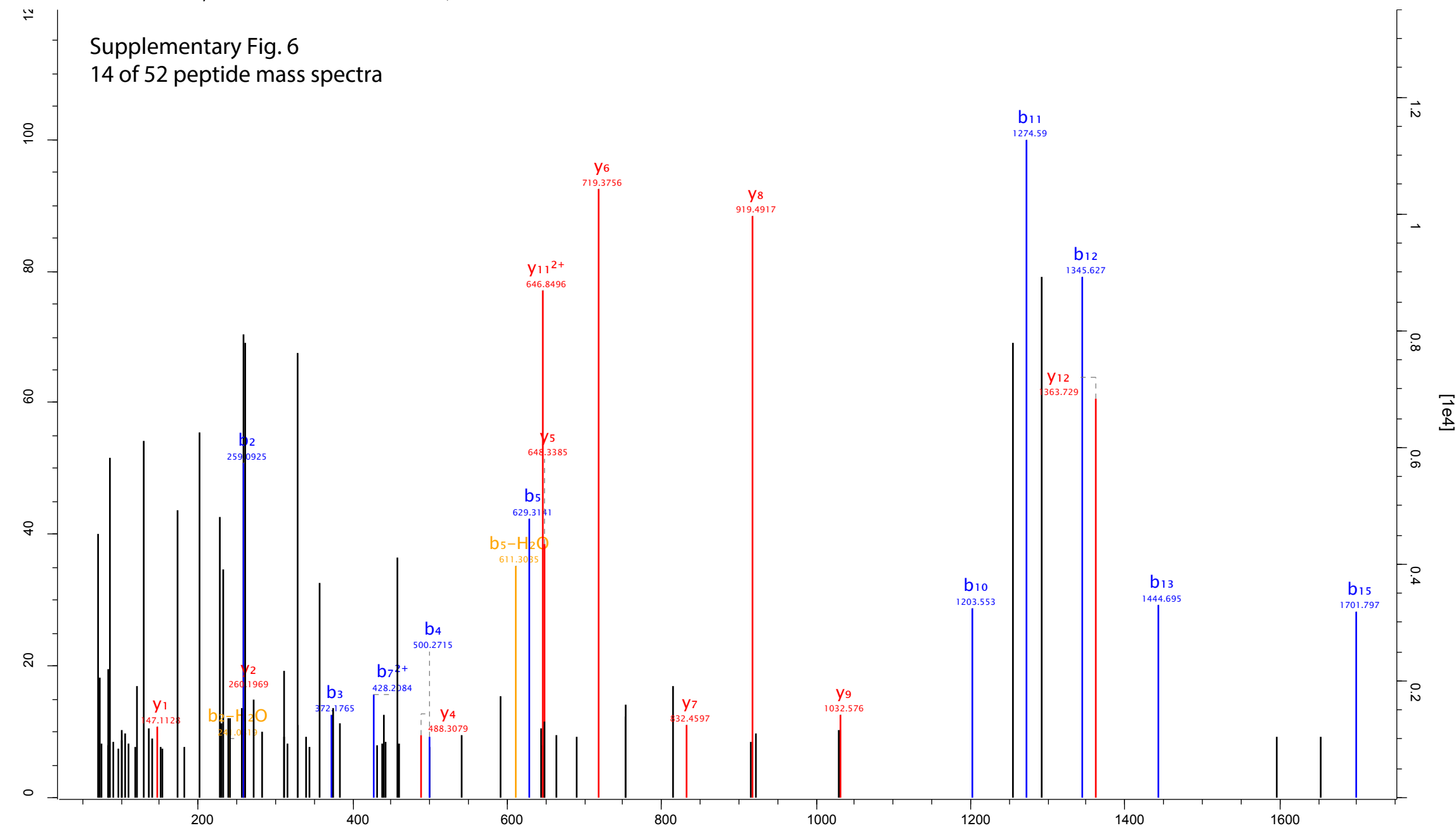
Supplementary Fig. 6
12 of 52 peptide mass spectra



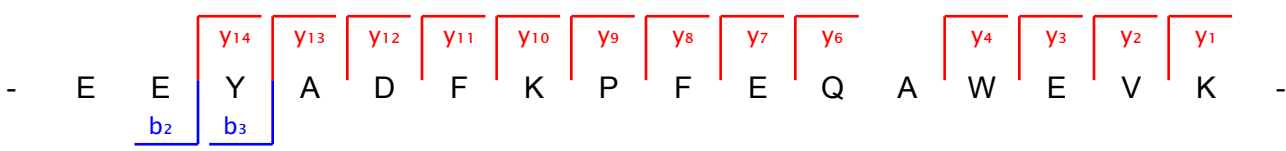
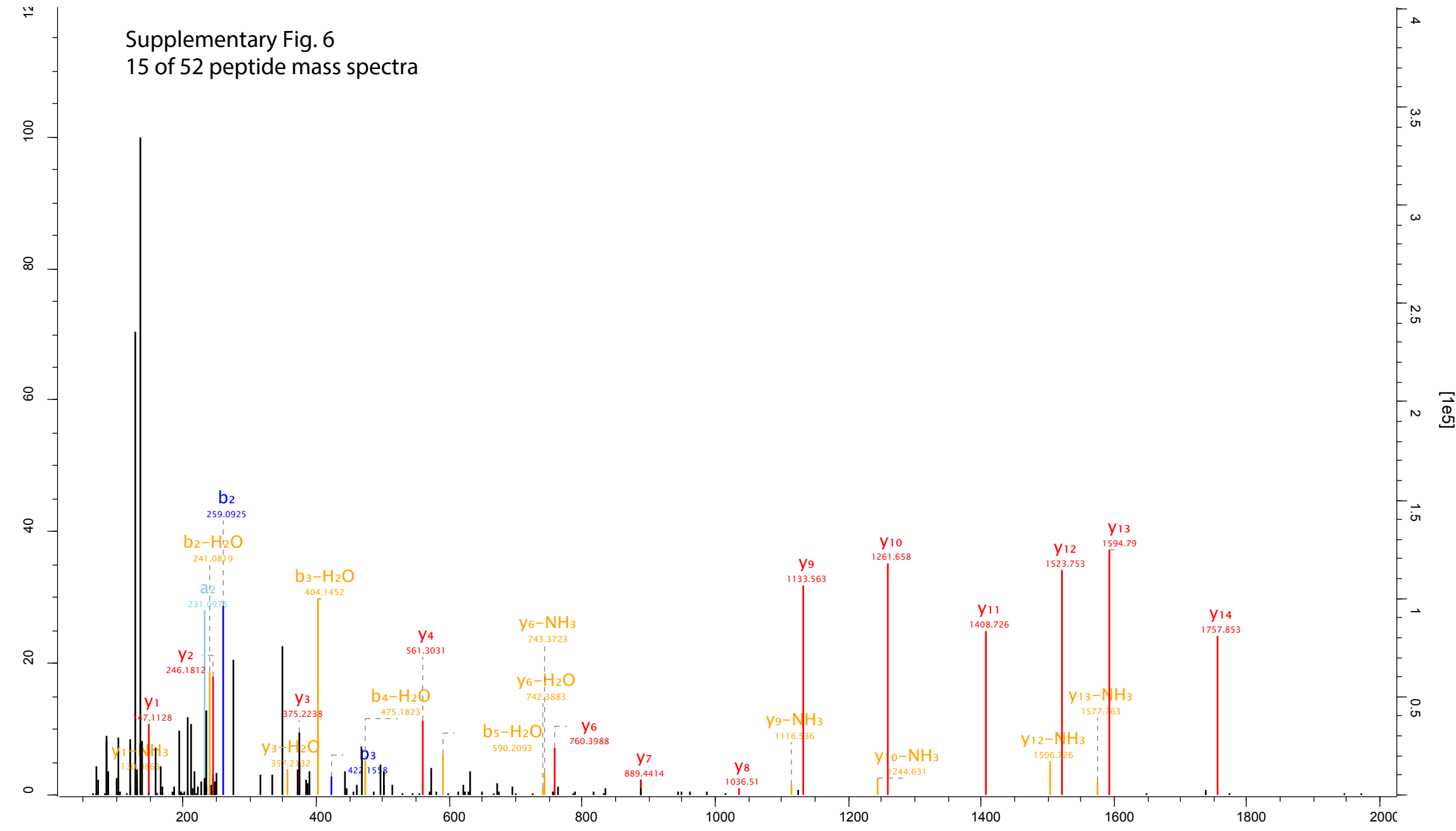
Supplementary Fig. 6
13 of 52 peptide mass spectra



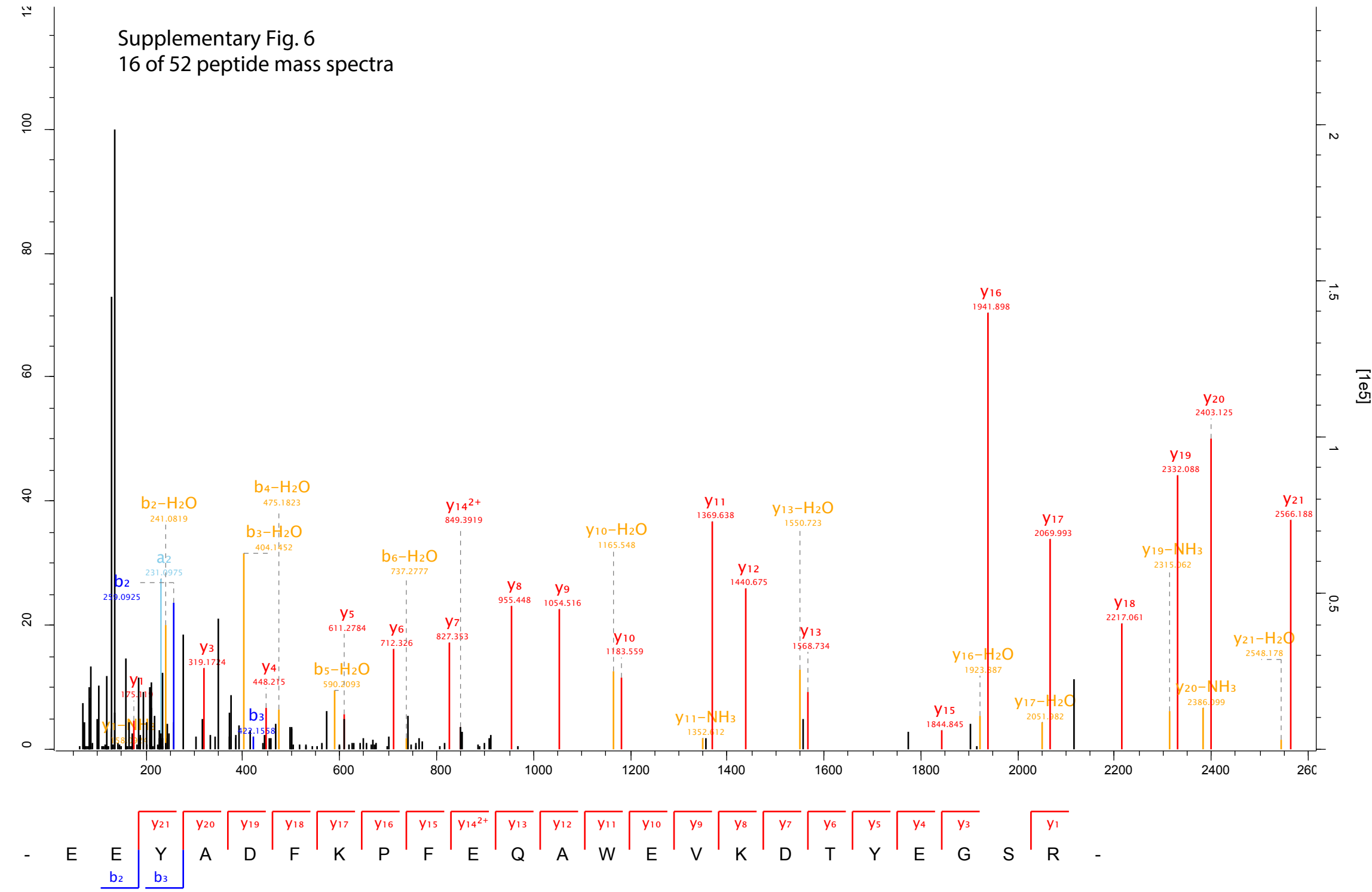
Supplementary Fig. 6
14 of 52 peptide mass spectra



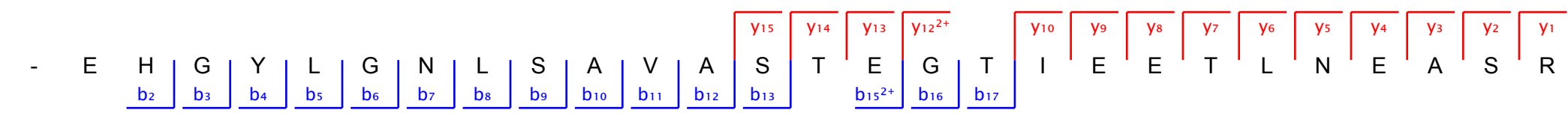
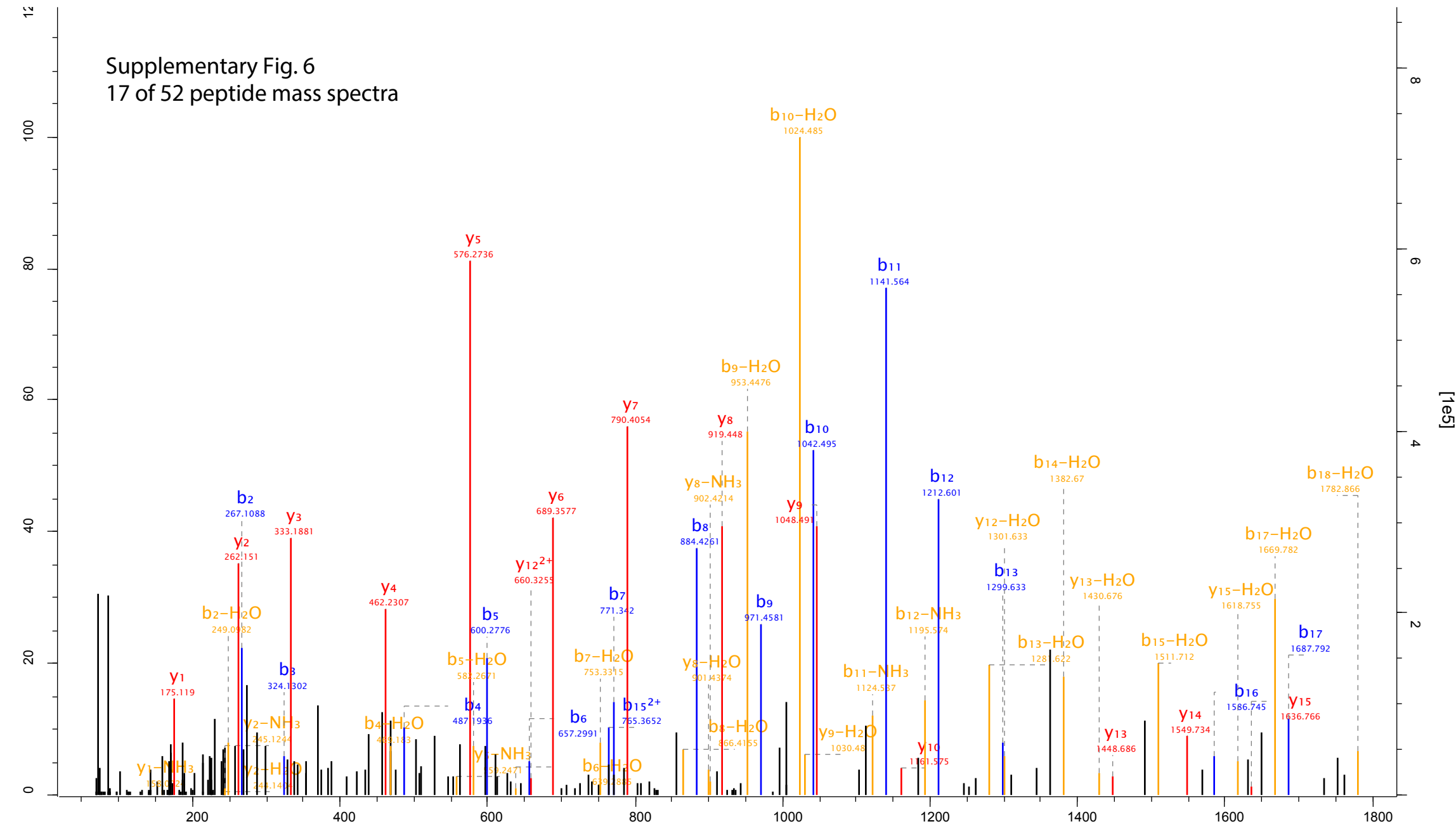
Supplementary Fig. 6 15 of 52 peptide mass spectra



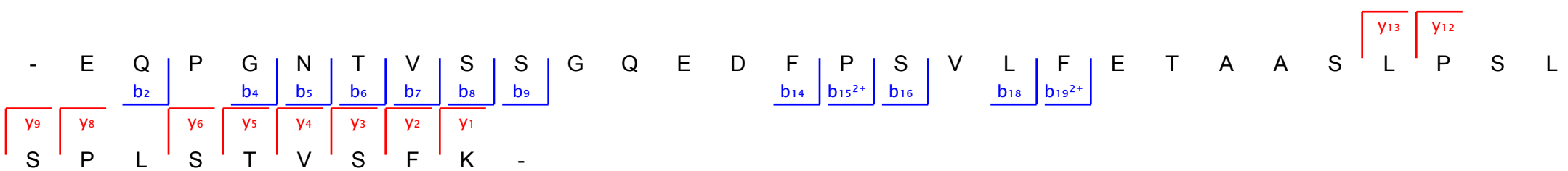
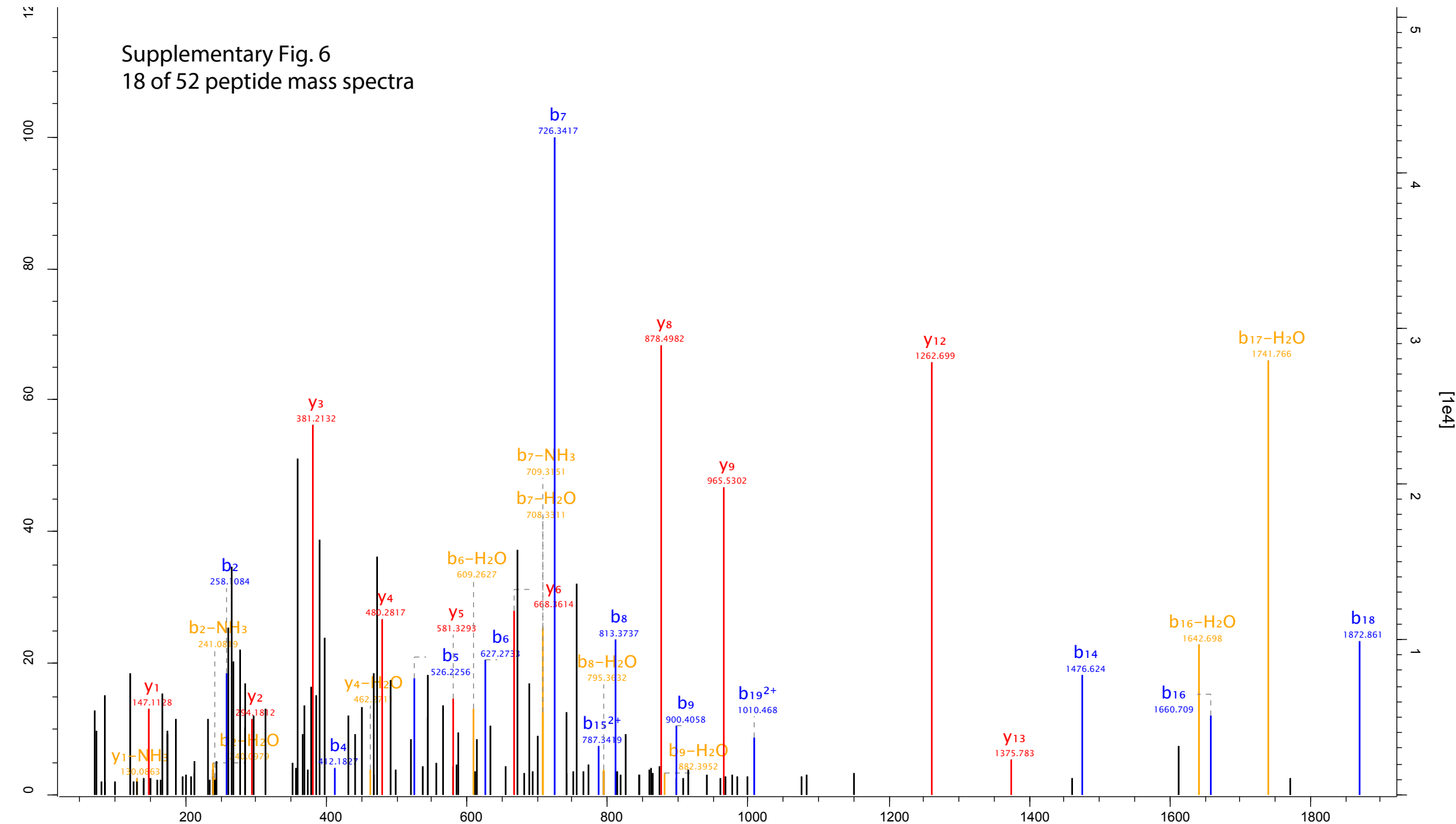
Supplementary Fig. 6
16 of 52 peptide mass spectra



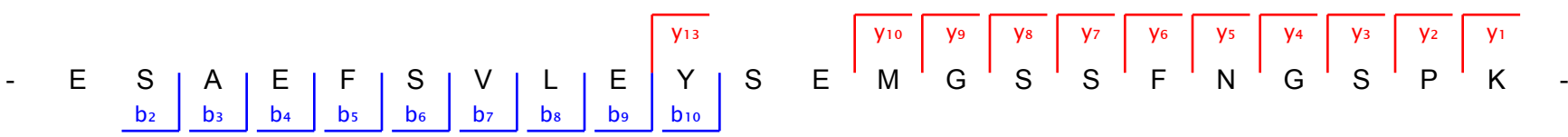
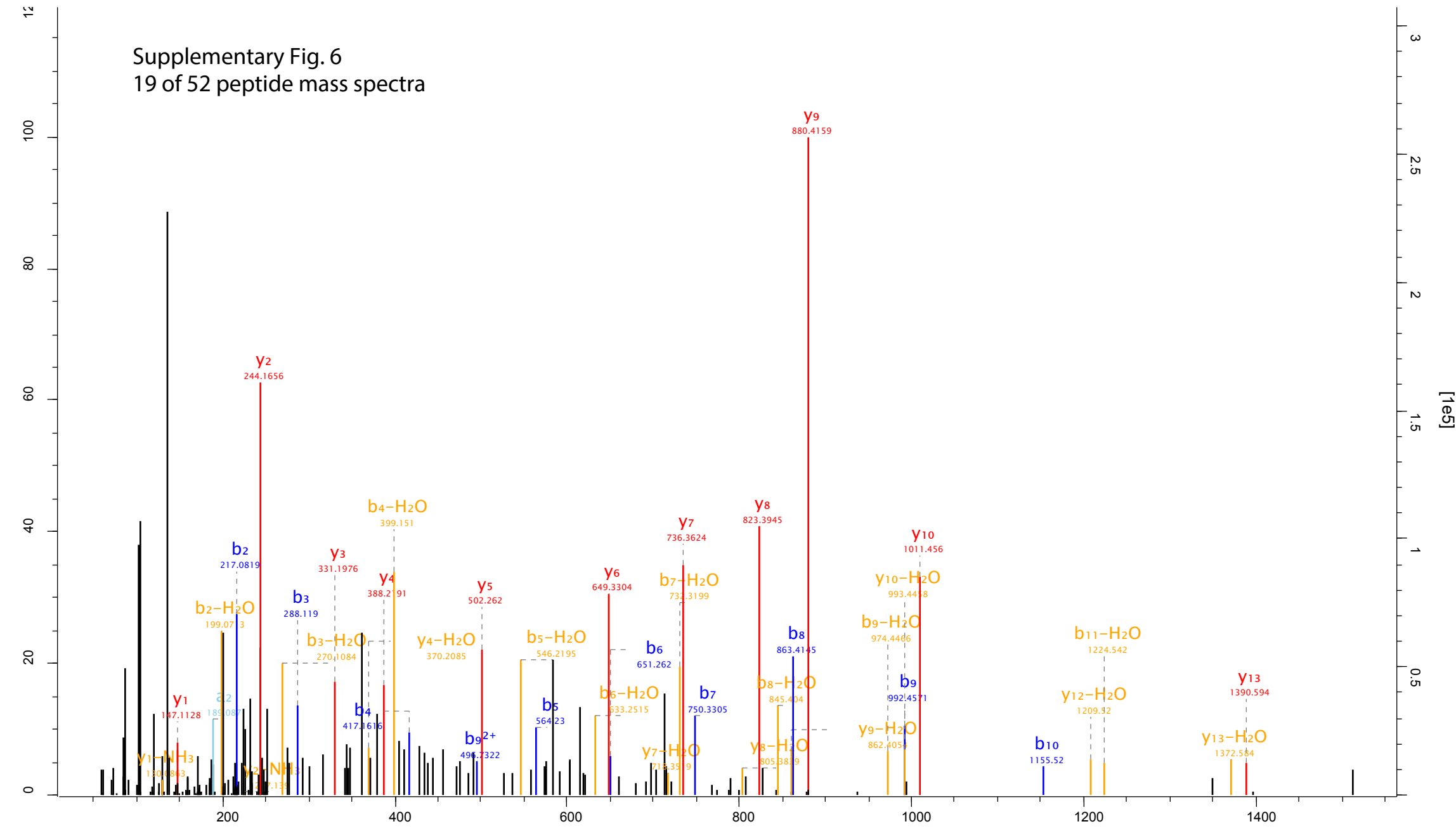
Supplementary Fig. 6
17 of 52 peptide mass spectra



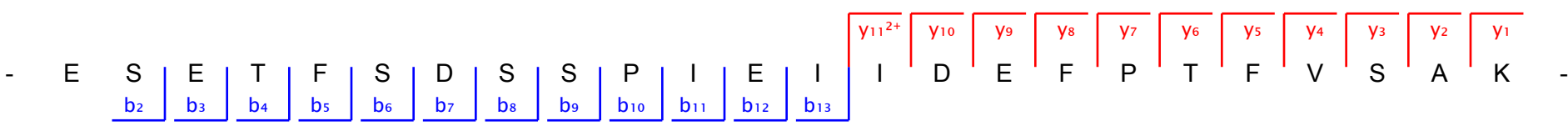
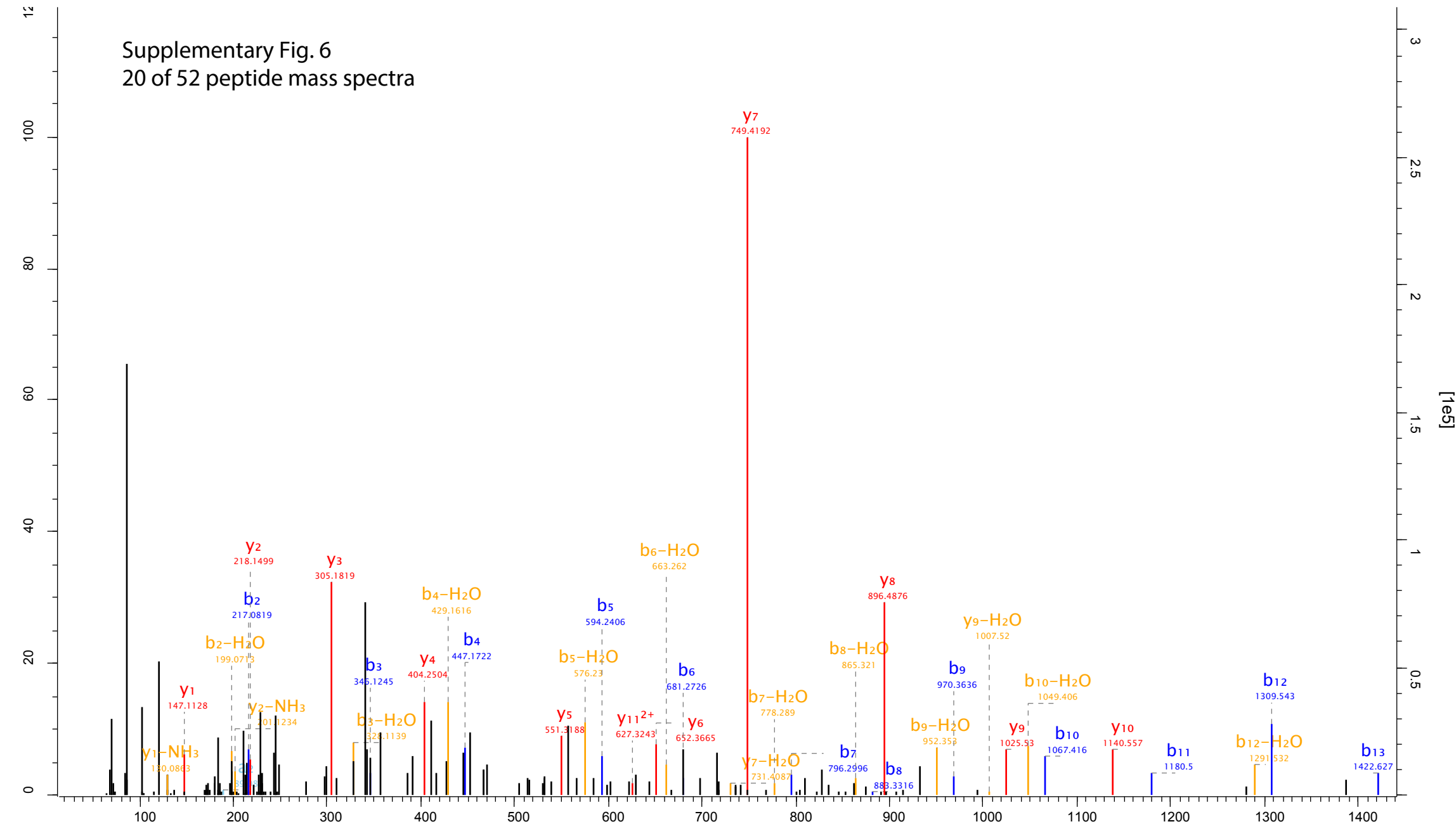
Supplementary Fig. 6
18 of 52 peptide mass spectra



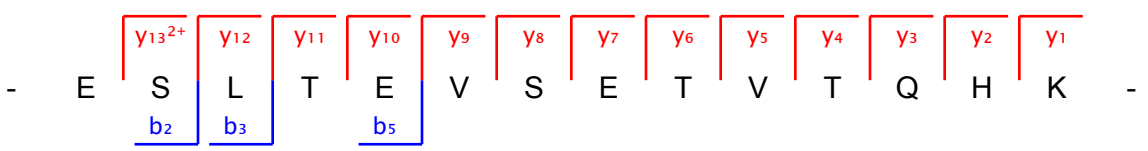
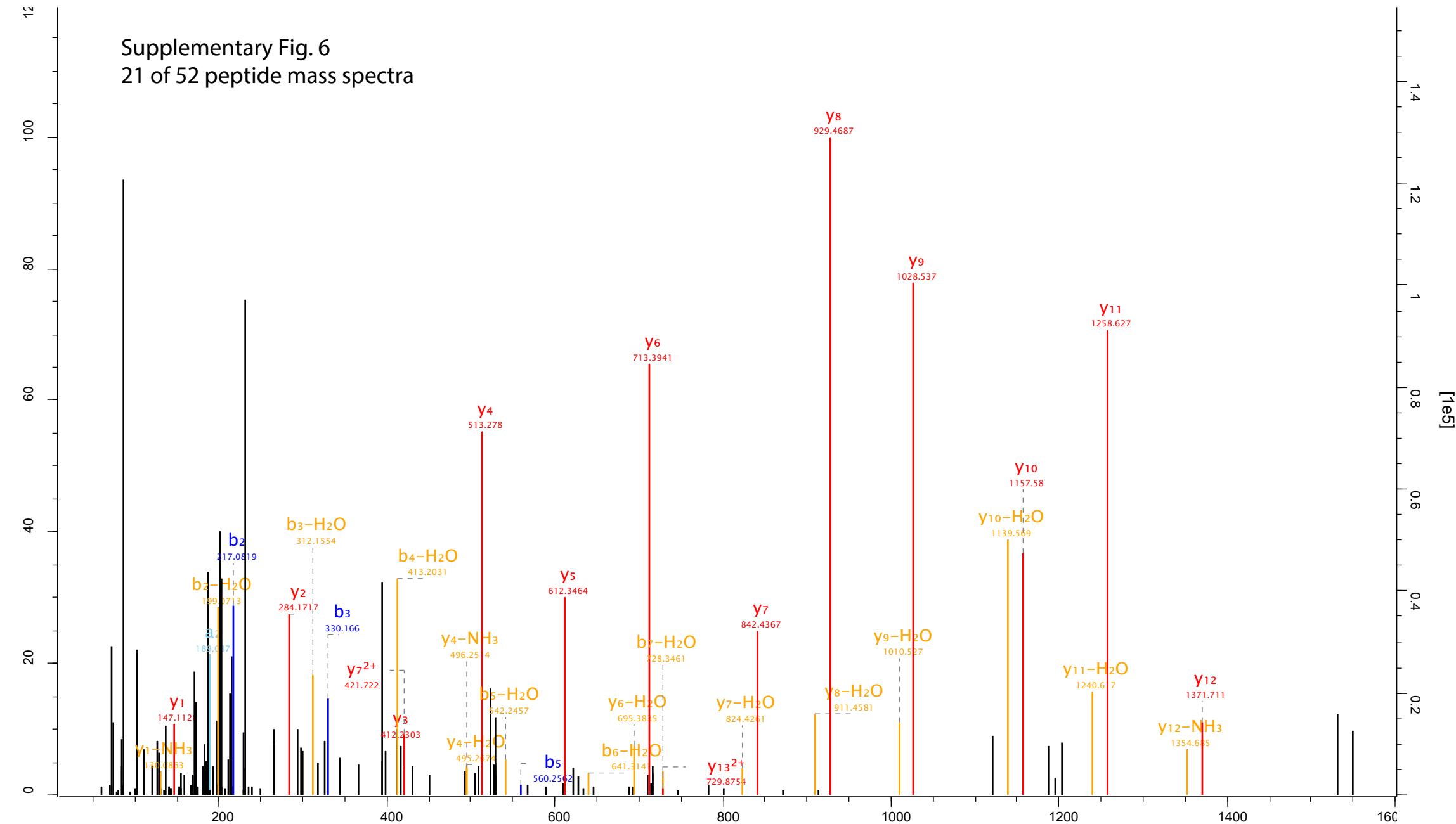
Supplementary Fig. 6
19 of 52 peptide mass spectra



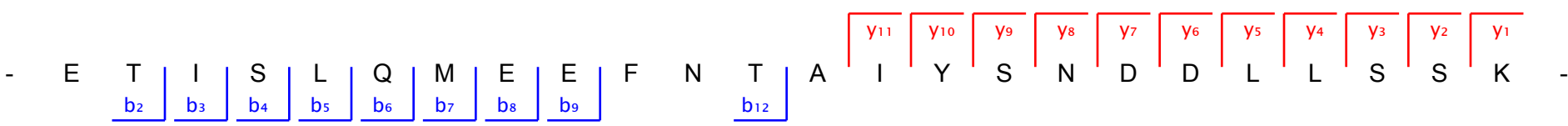
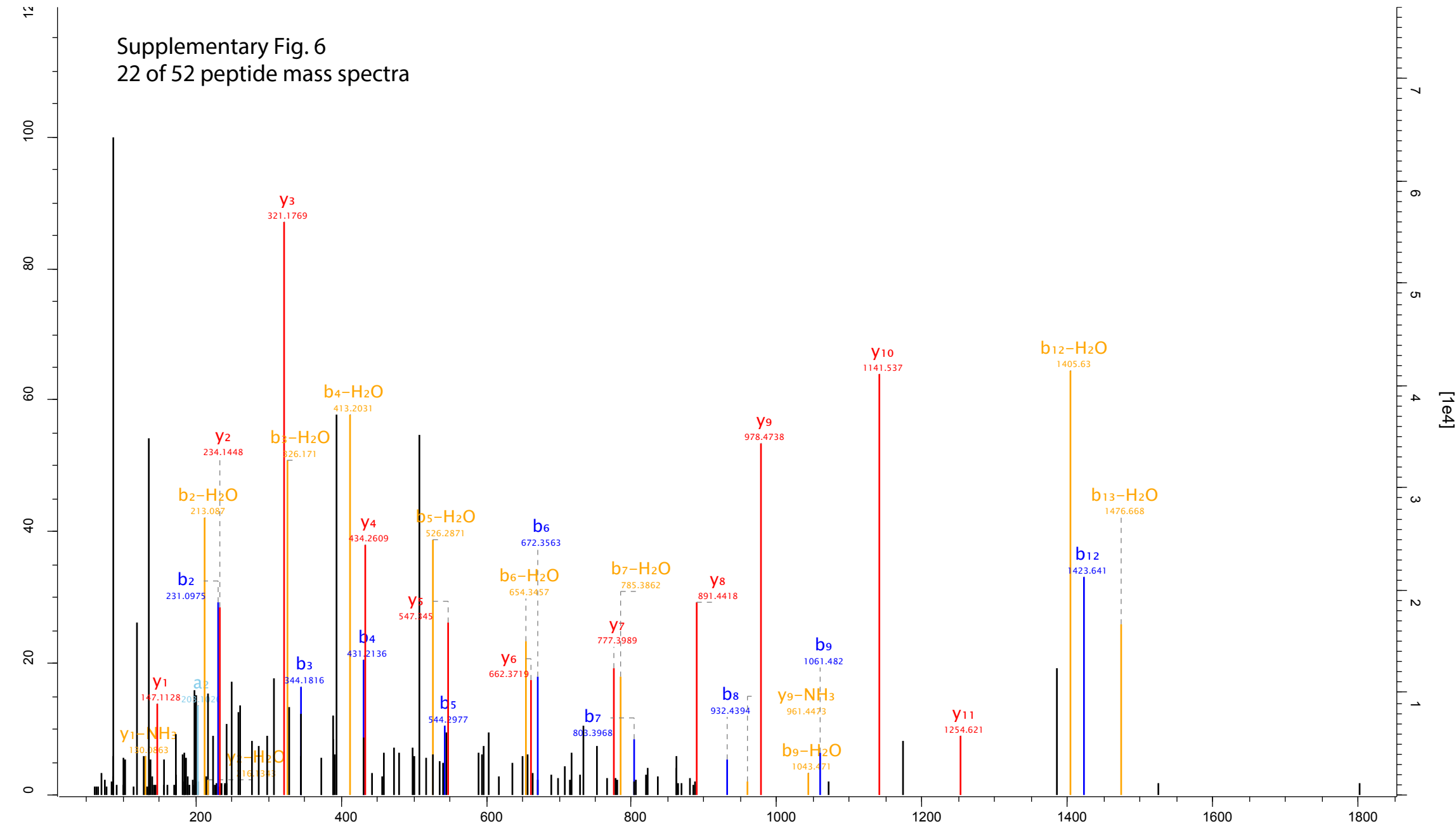
Supplementary Fig. 6
20 of 52 peptide mass spectra



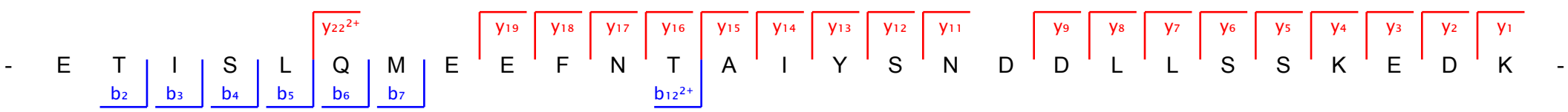
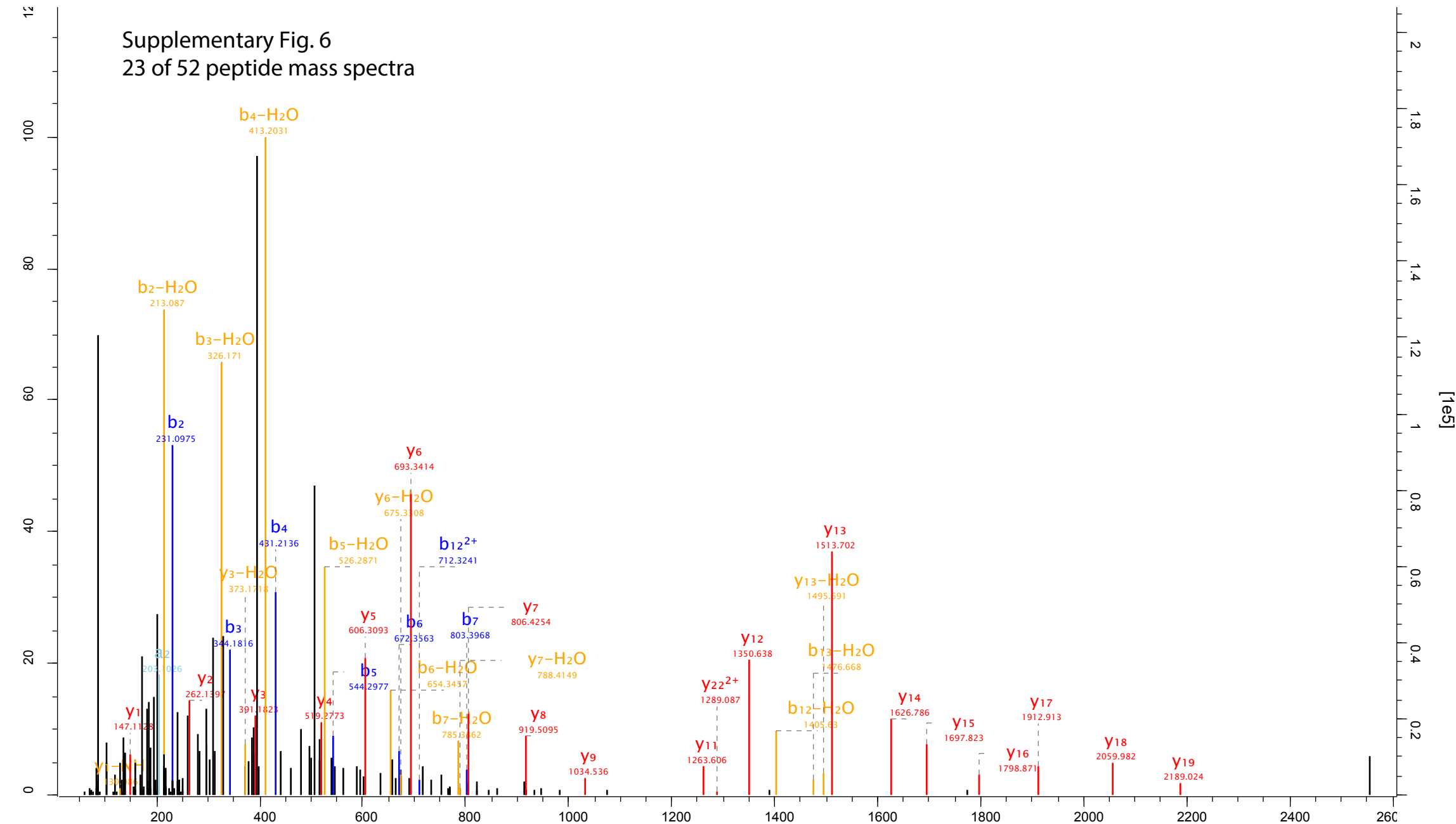
Supplementary Fig. 6
21 of 52 peptide mass spectra



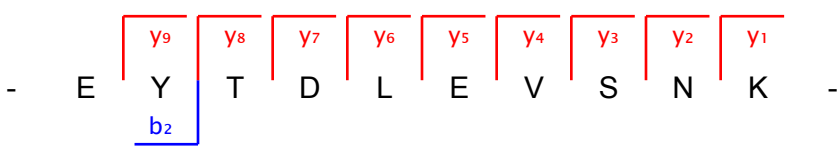
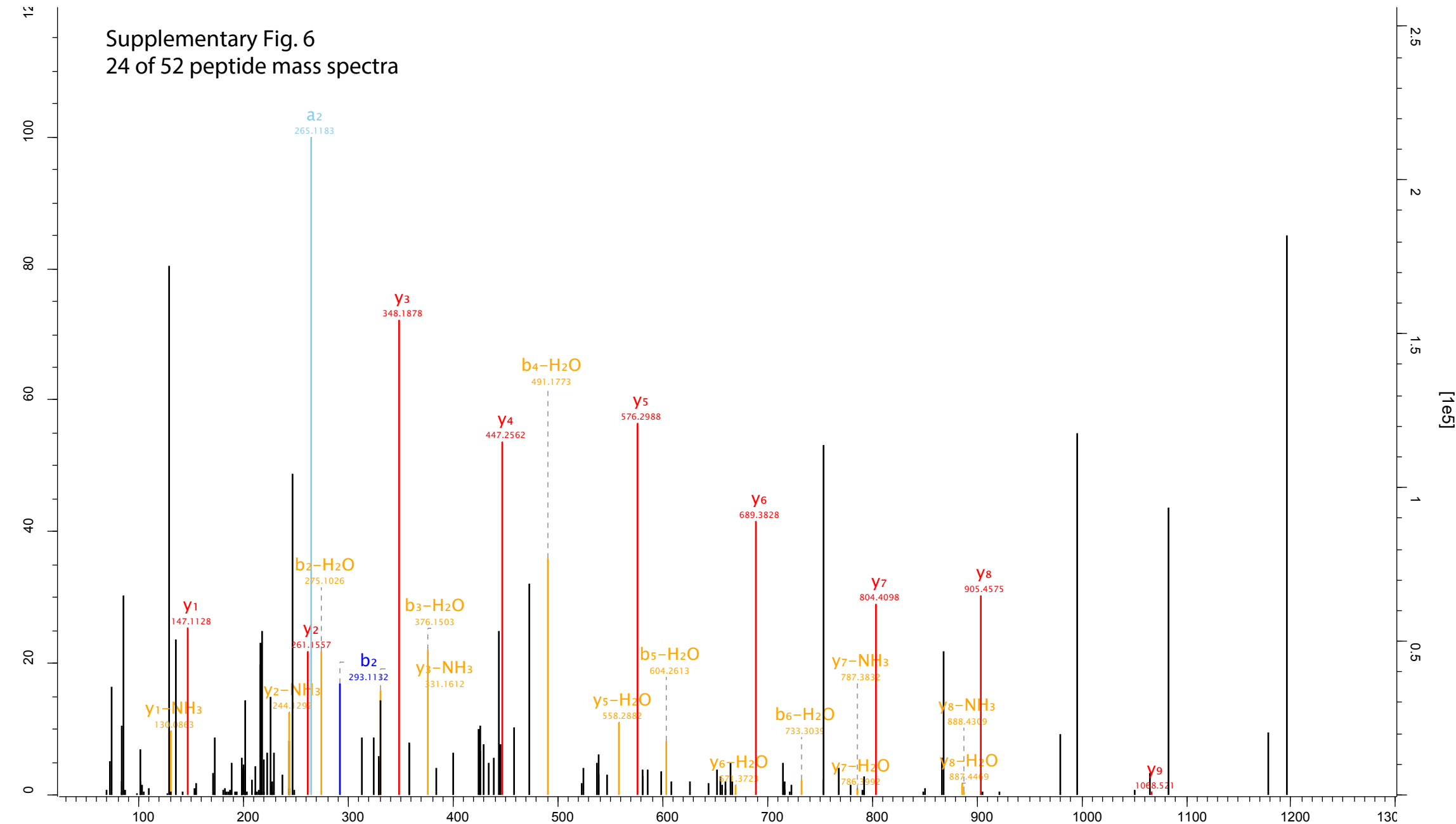
Supplementary Fig. 6
22 of 52 peptide mass spectra



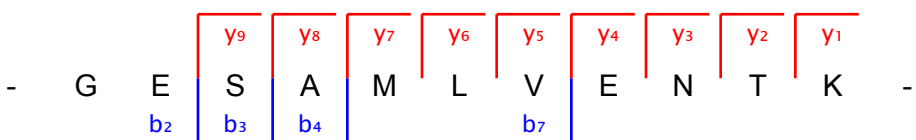
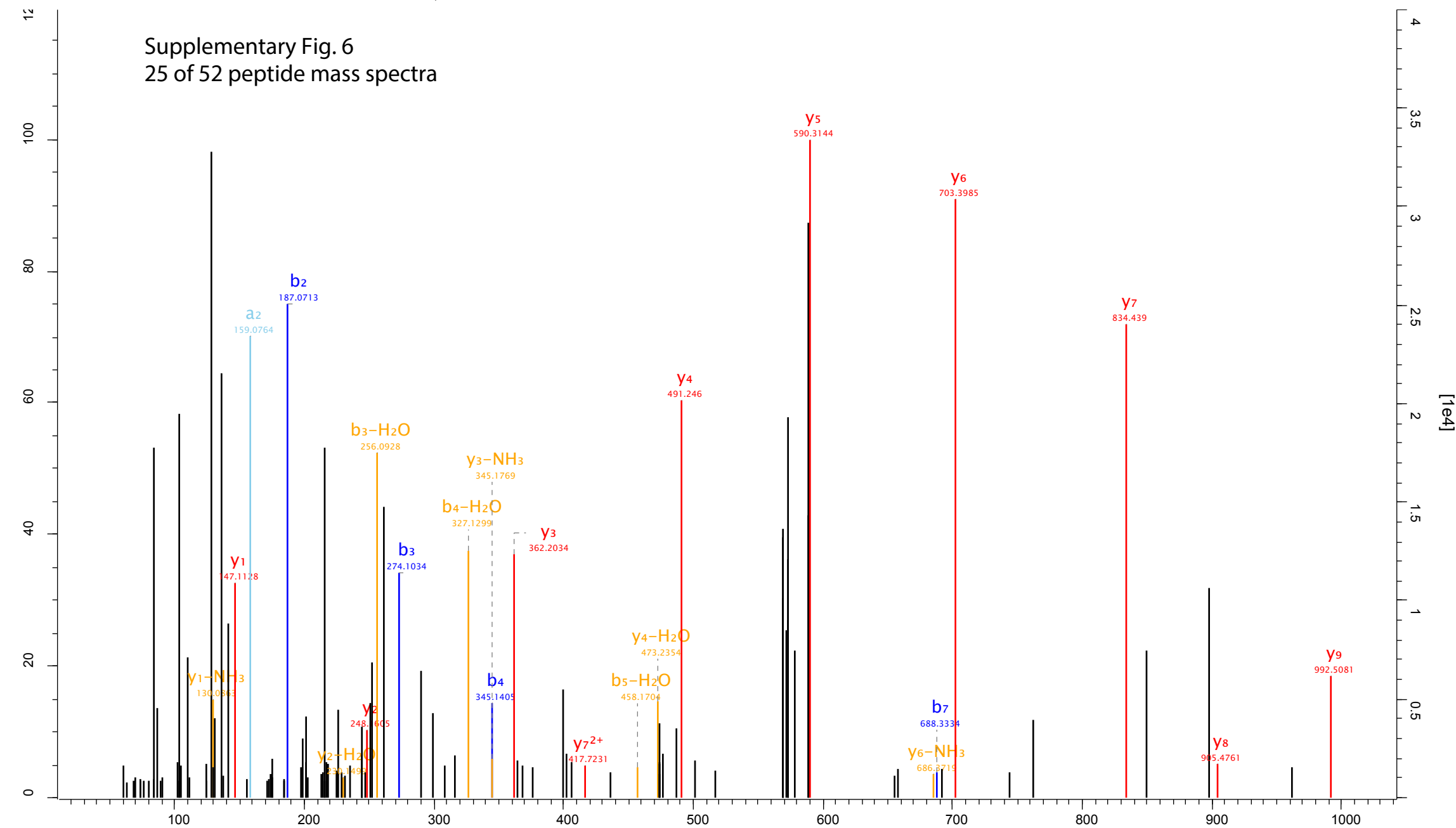
Supplementary Fig. 6
23 of 52 peptide mass spectra



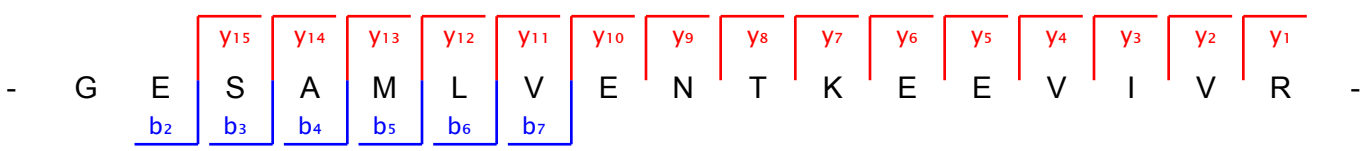
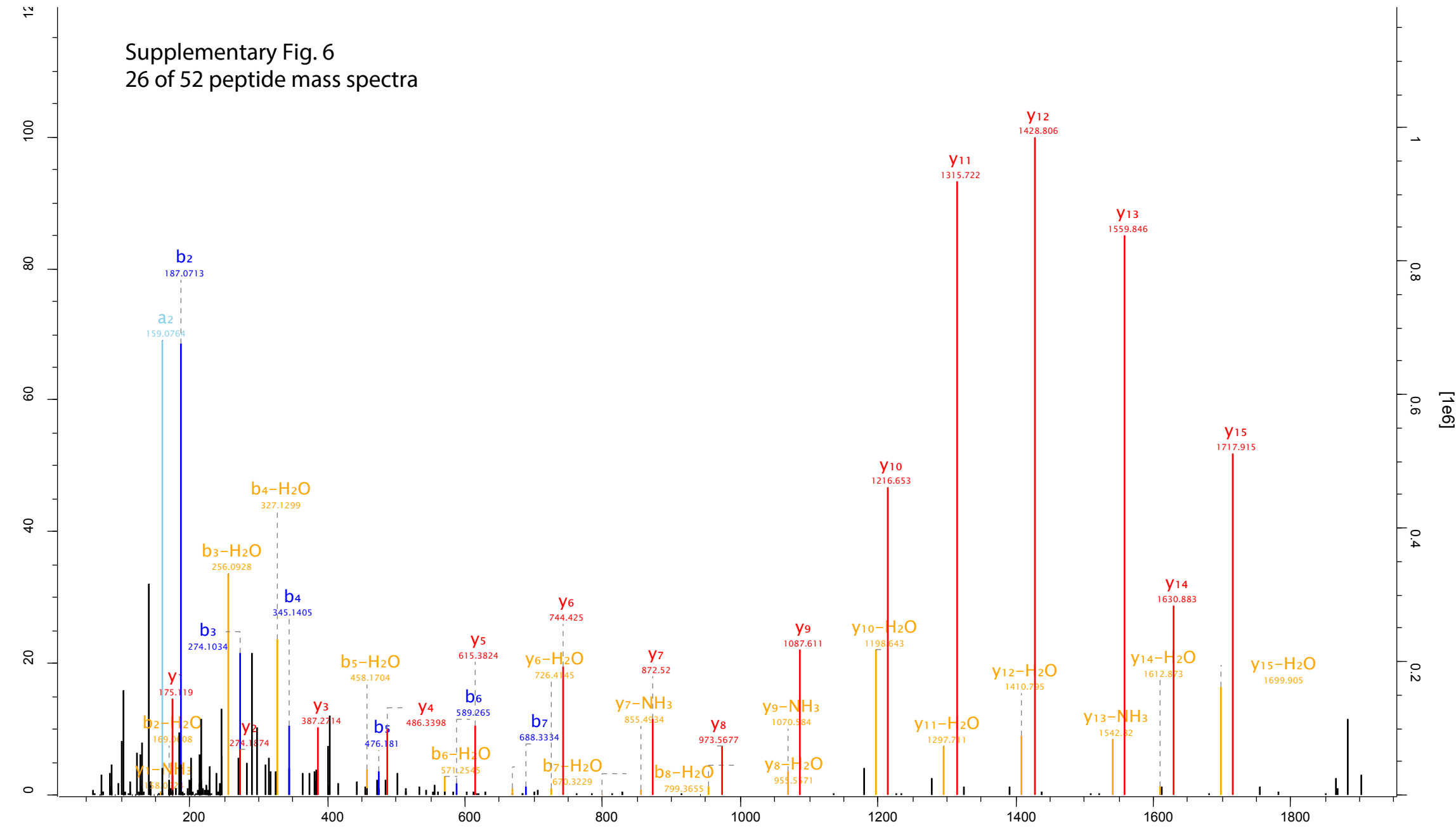
Supplementary Fig. 6 24 of 52 peptide mass spectra



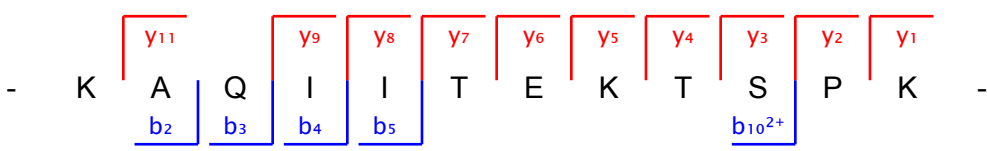
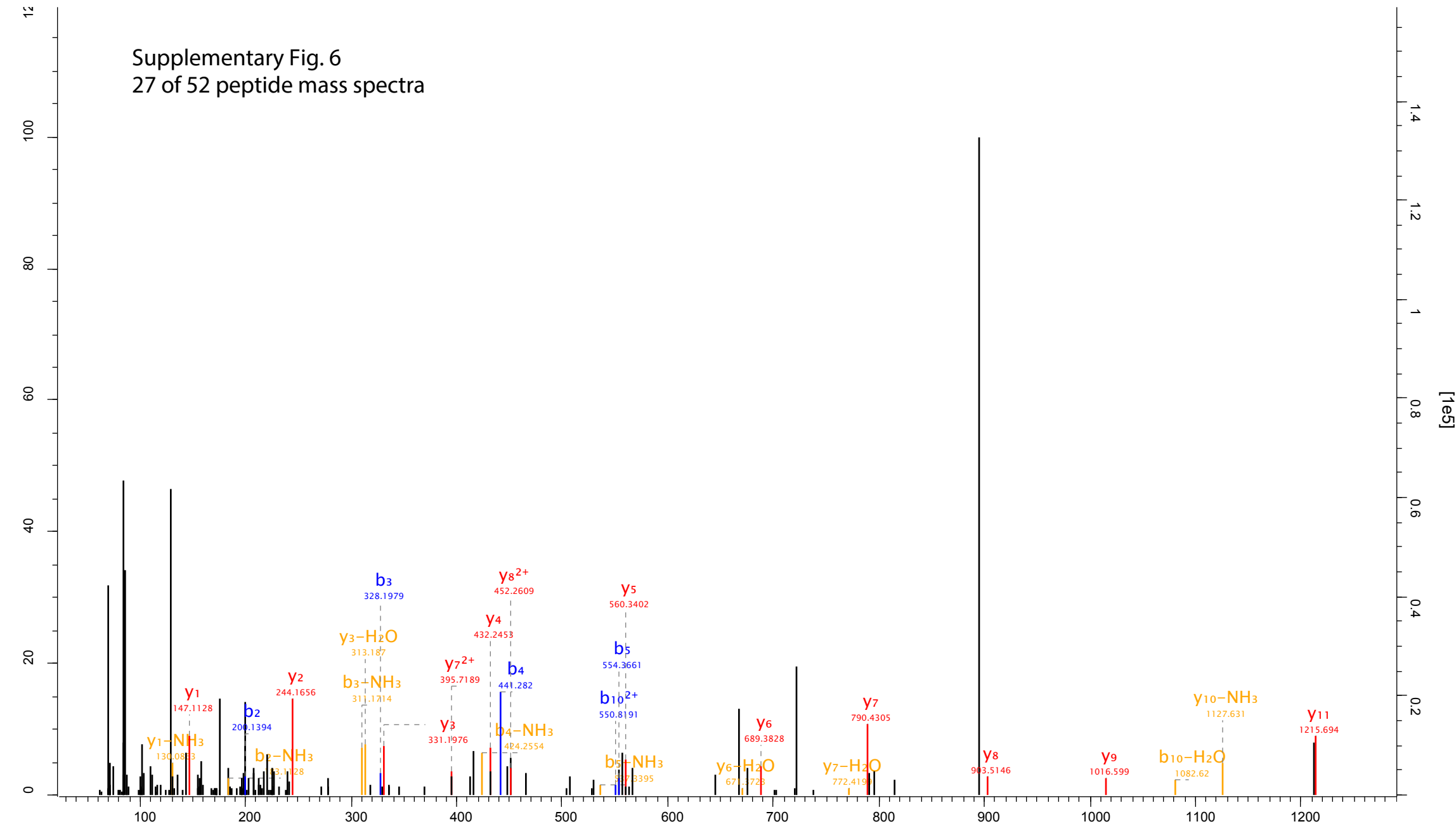
Supplementary Fig. 6
25 of 52 peptide mass spectra



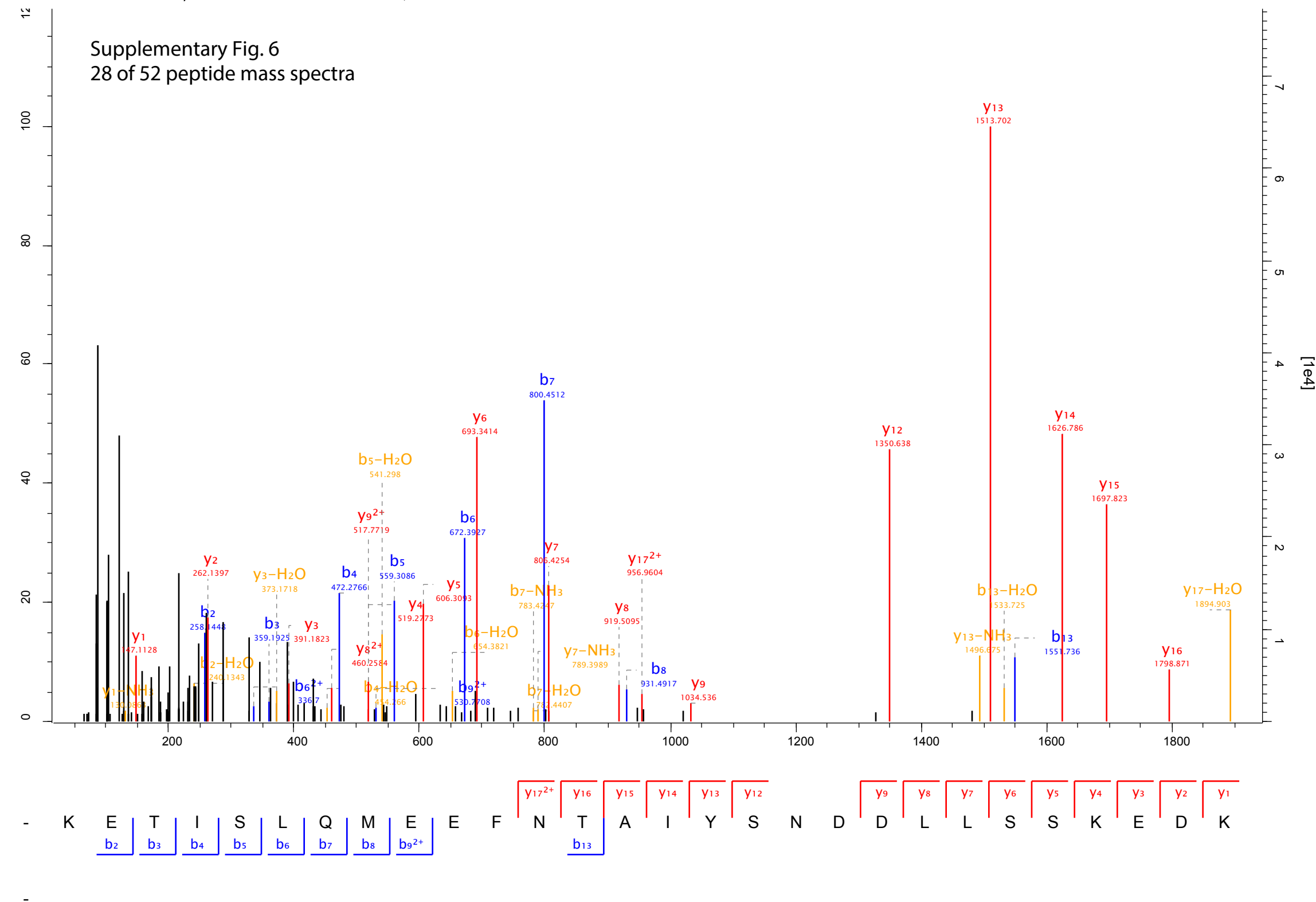
Supplementary Fig. 6
26 of 52 peptide mass spectra



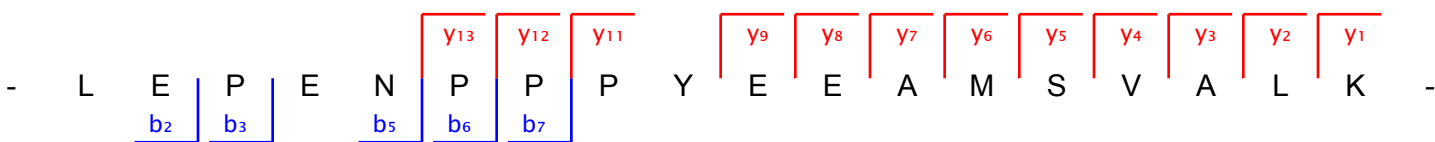
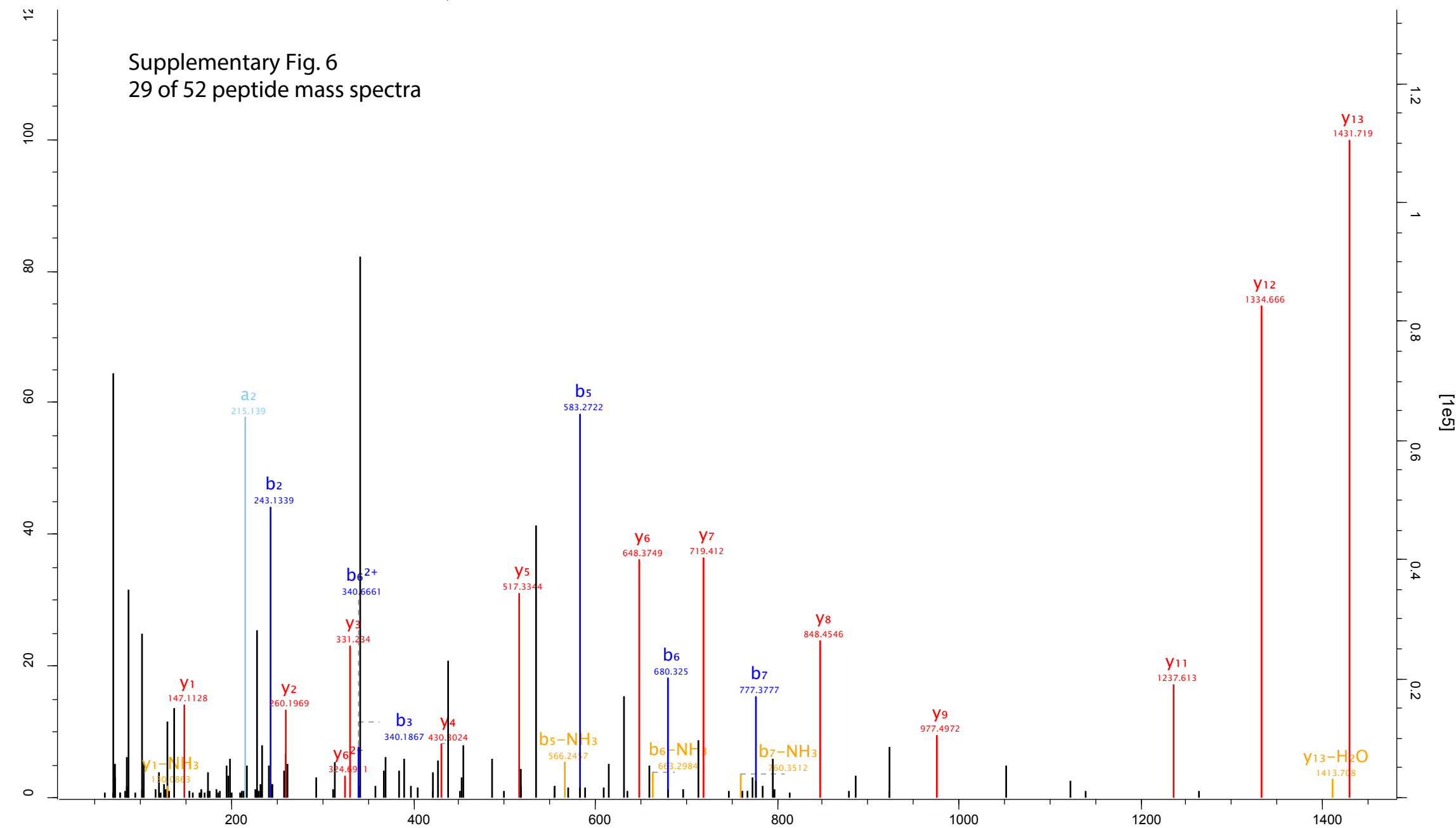
Supplementary Fig. 6
27 of 52 peptide mass spectra



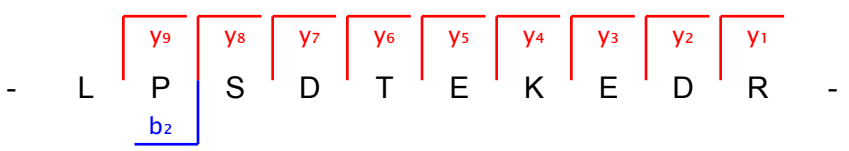
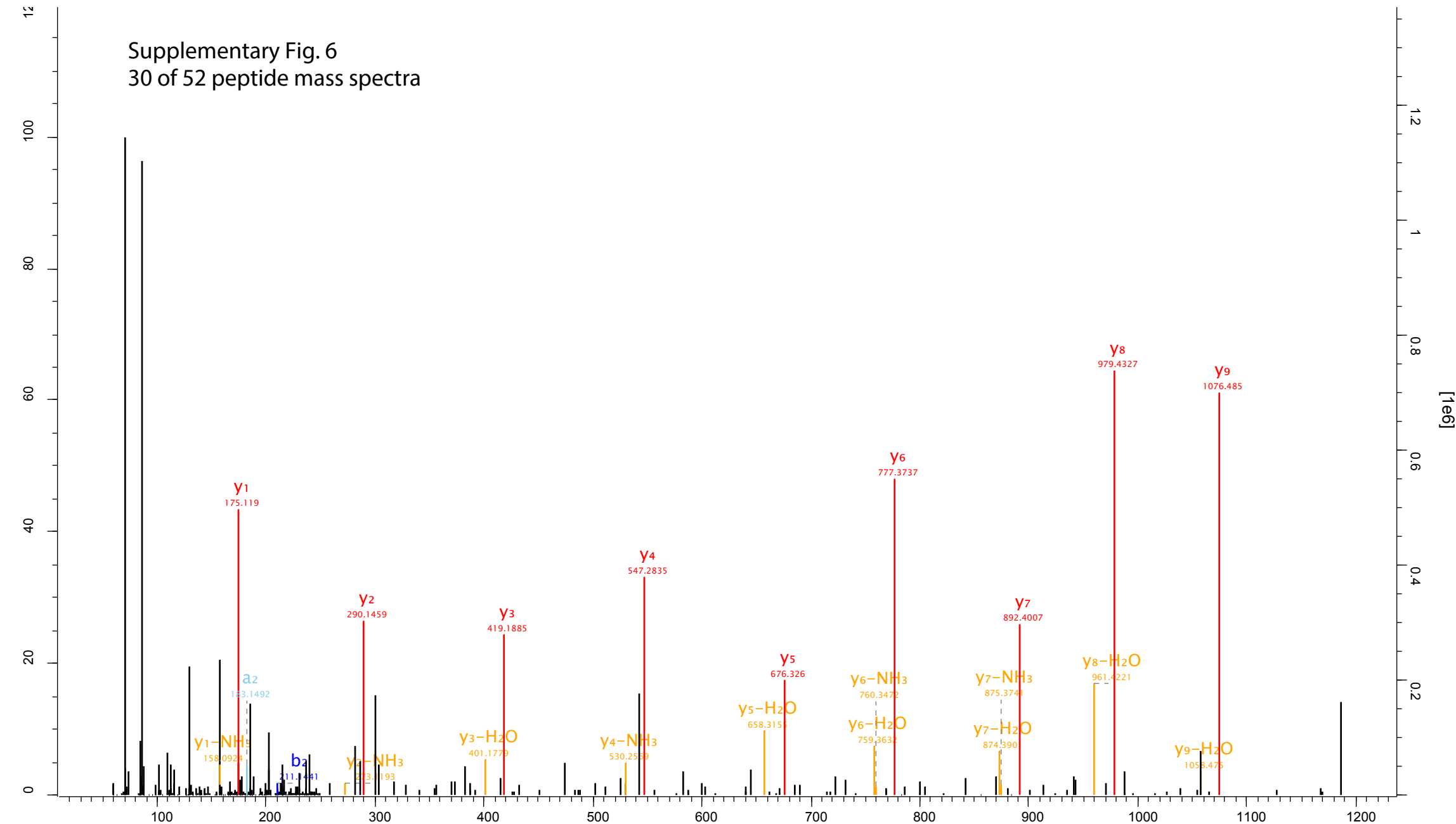
Supplementary Fig. 6
28 of 52 peptide mass spectra



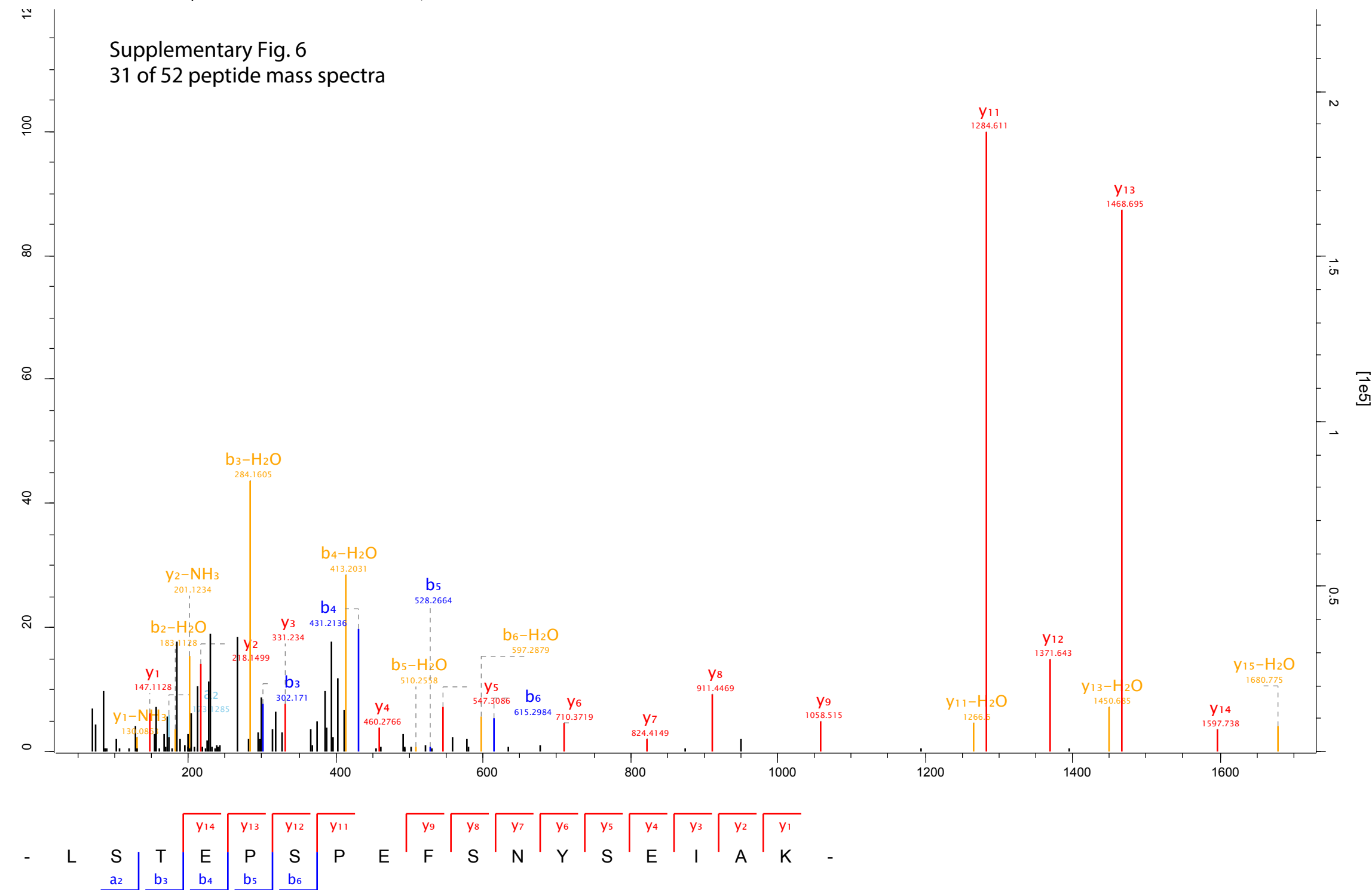
Supplementary Fig. 6
29 of 52 peptide mass spectra



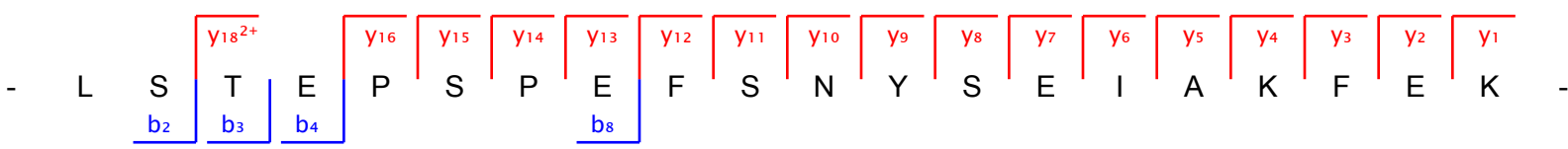
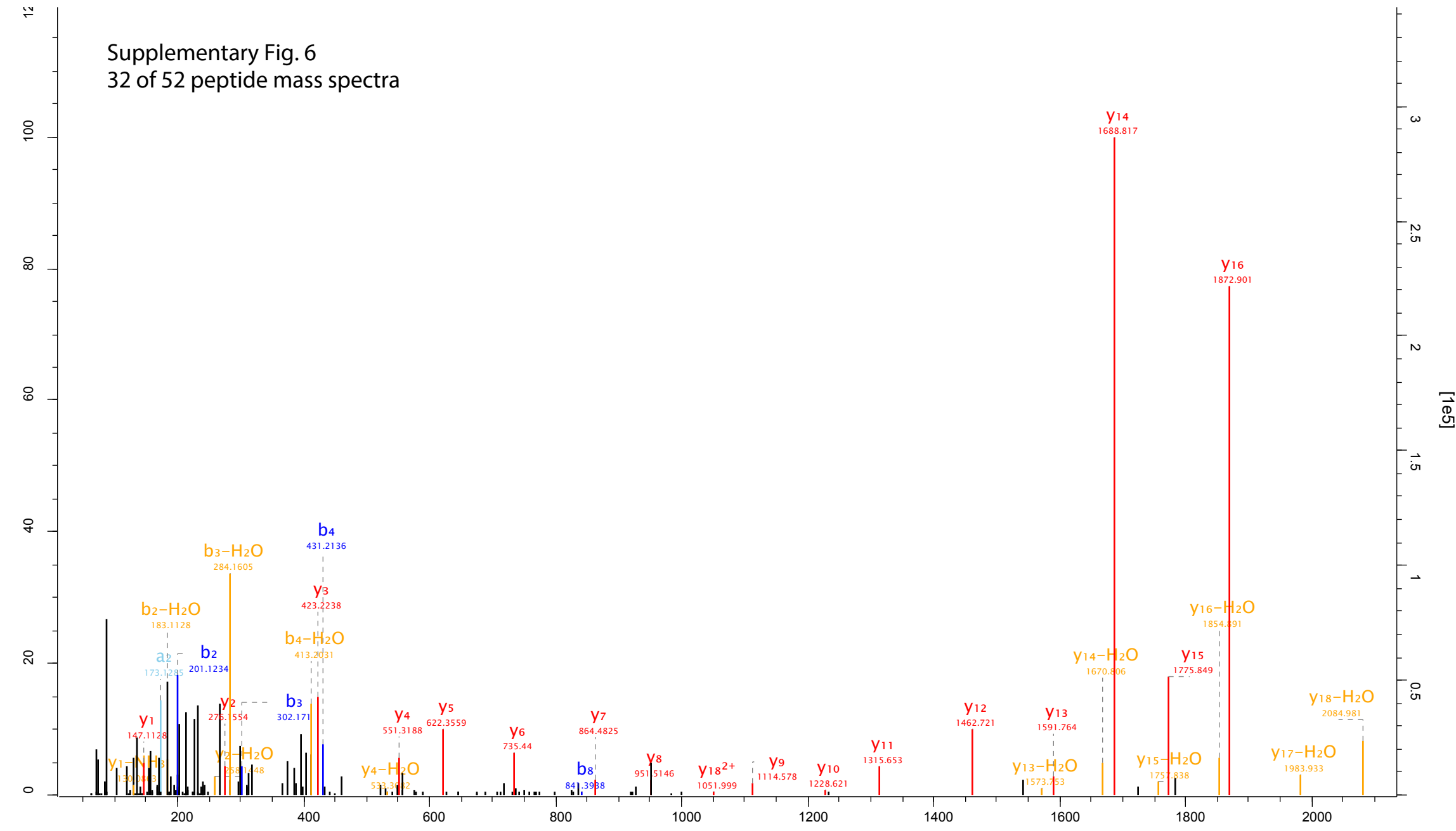
Supplementary Fig. 6
30 of 52 peptide mass spectra



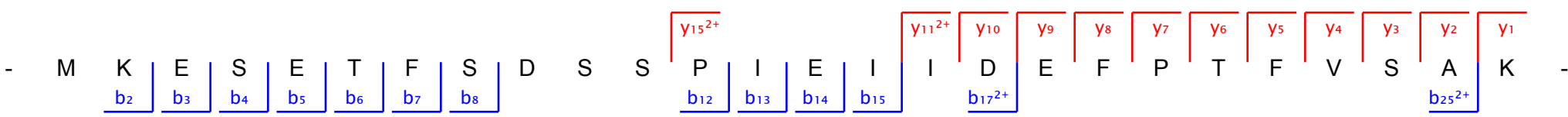
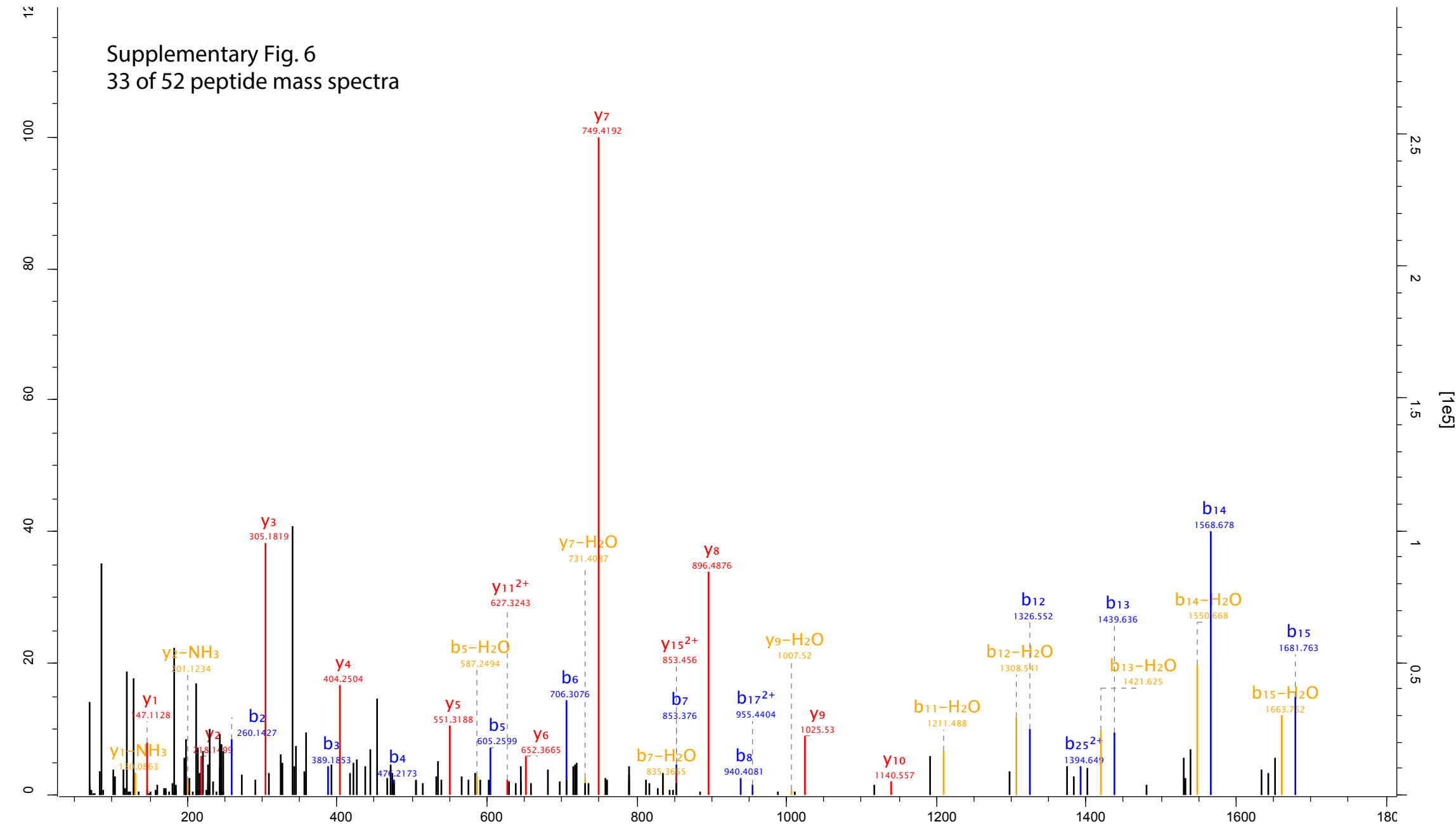
Supplementary Fig. 6
31 of 52 peptide mass spectra



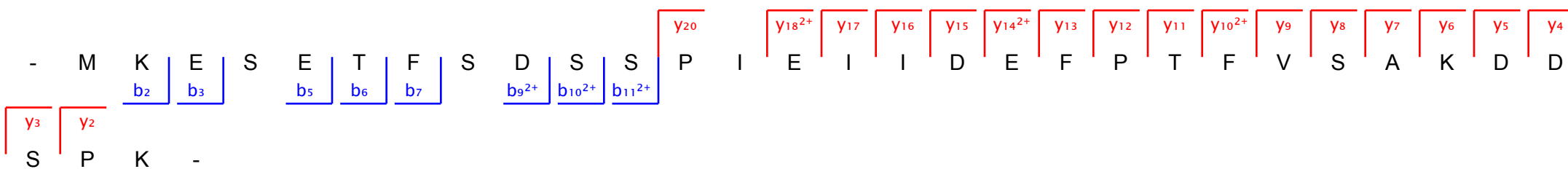
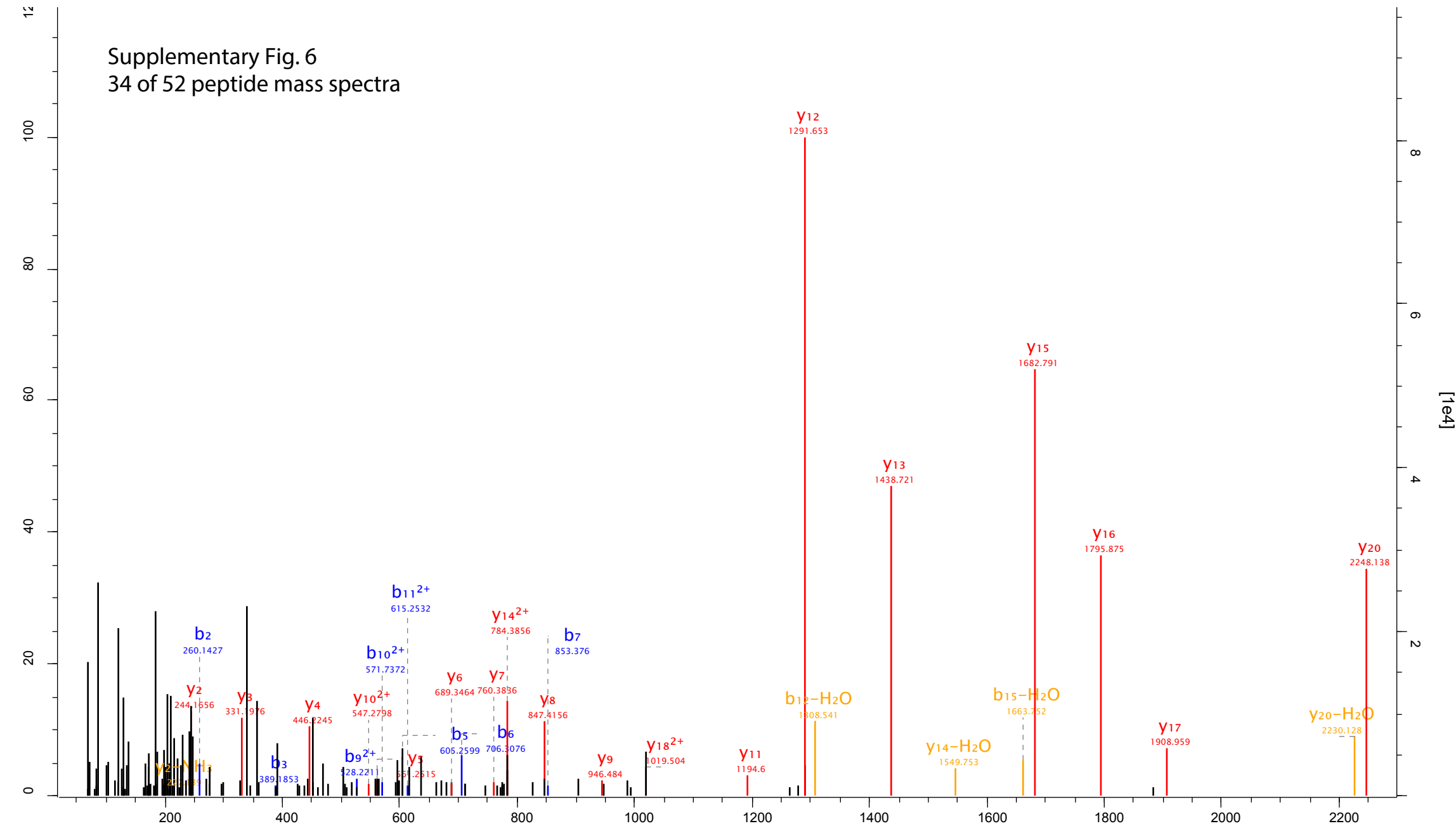
Supplementary Fig. 6
32 of 52 peptide mass spectra



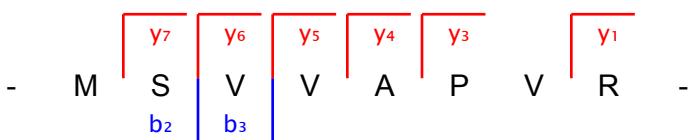
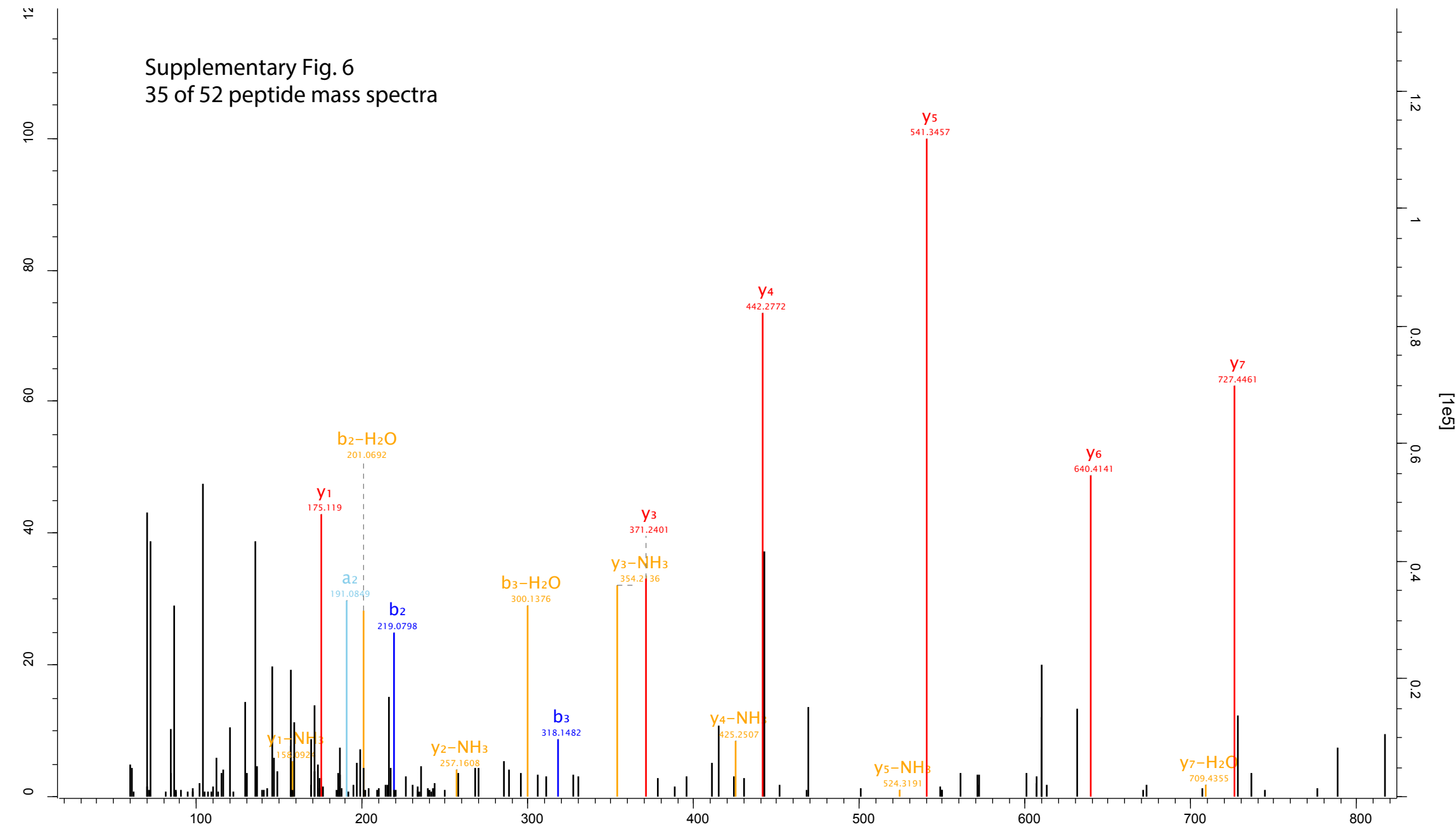
Supplementary Fig. 6
33 of 52 peptide mass spectra



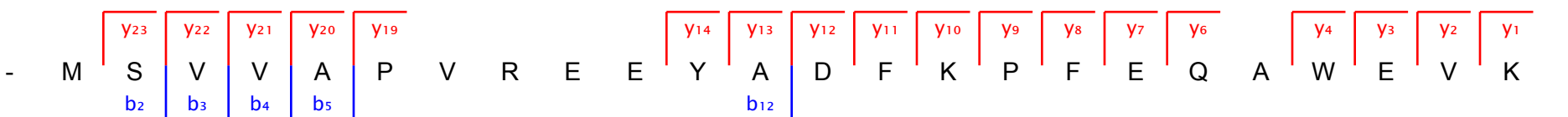
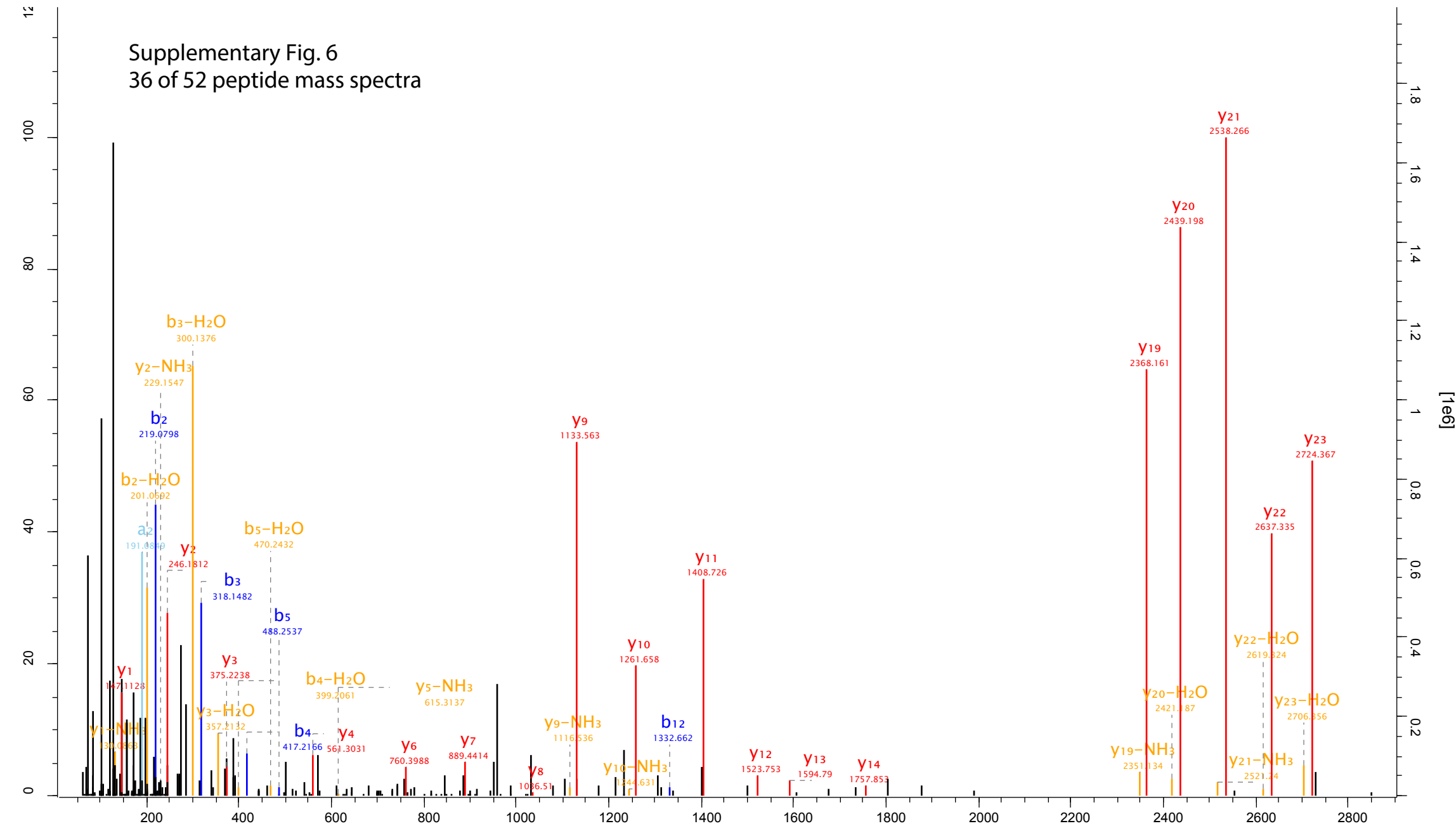
Supplementary Fig. 6
34 of 52 peptide mass spectra



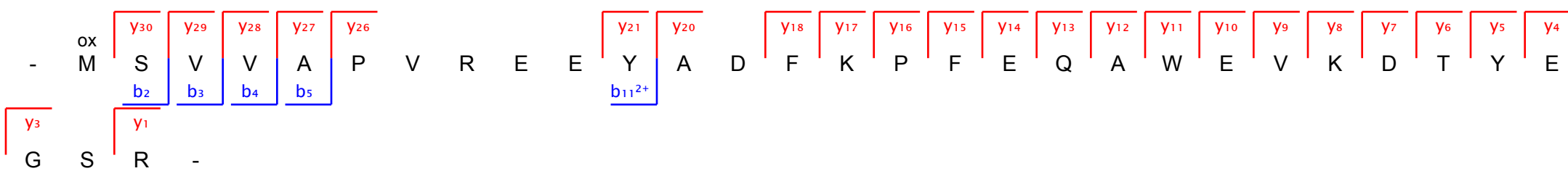
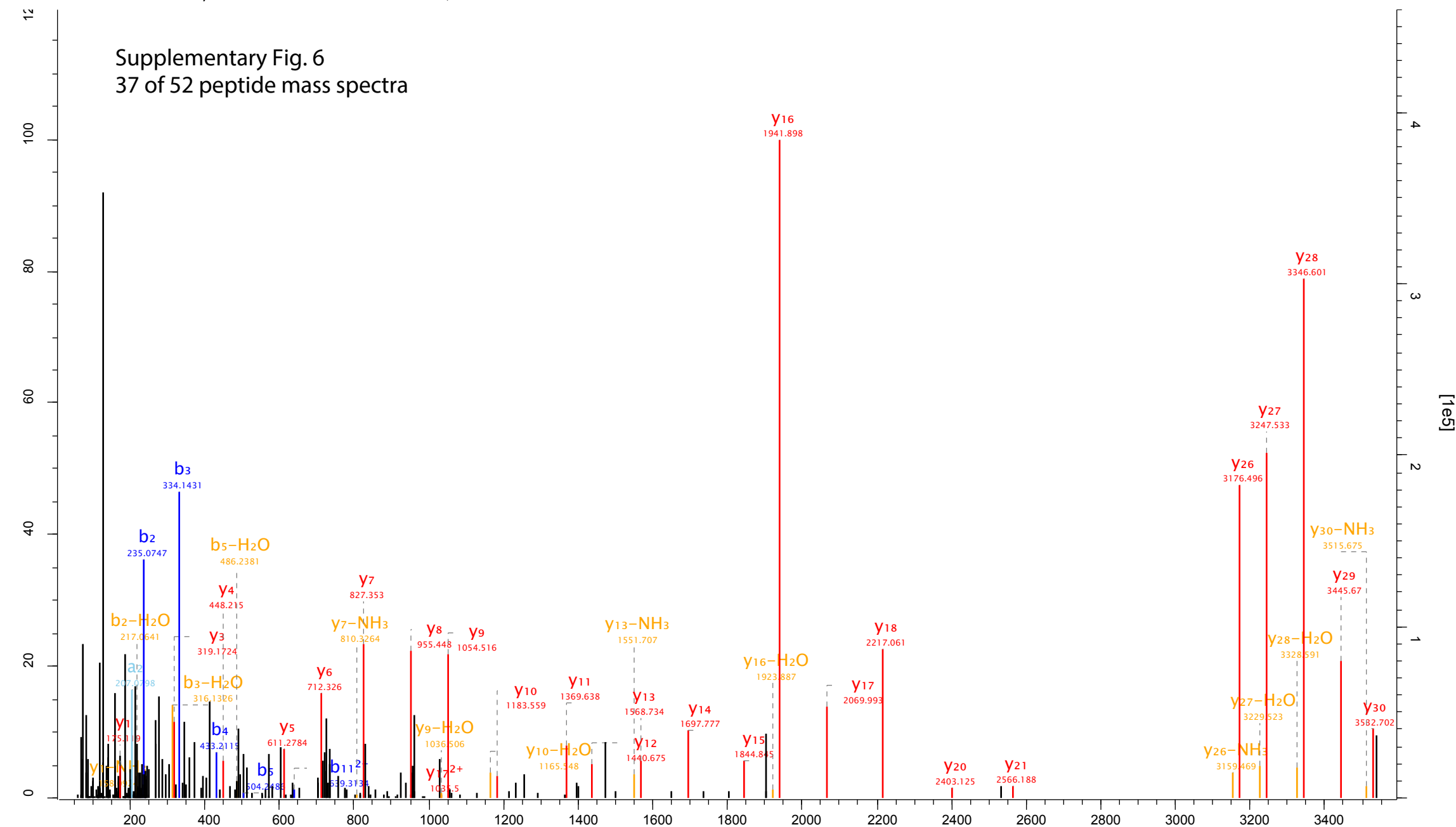
Supplementary Fig. 6 35 of 52 peptide mass spectra



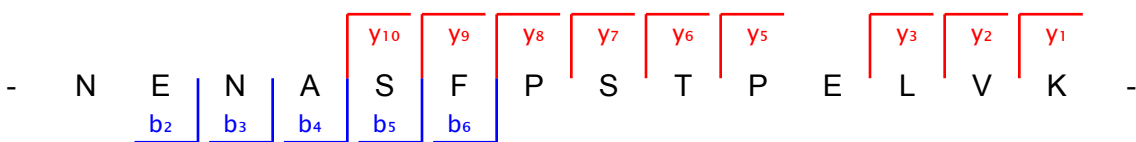
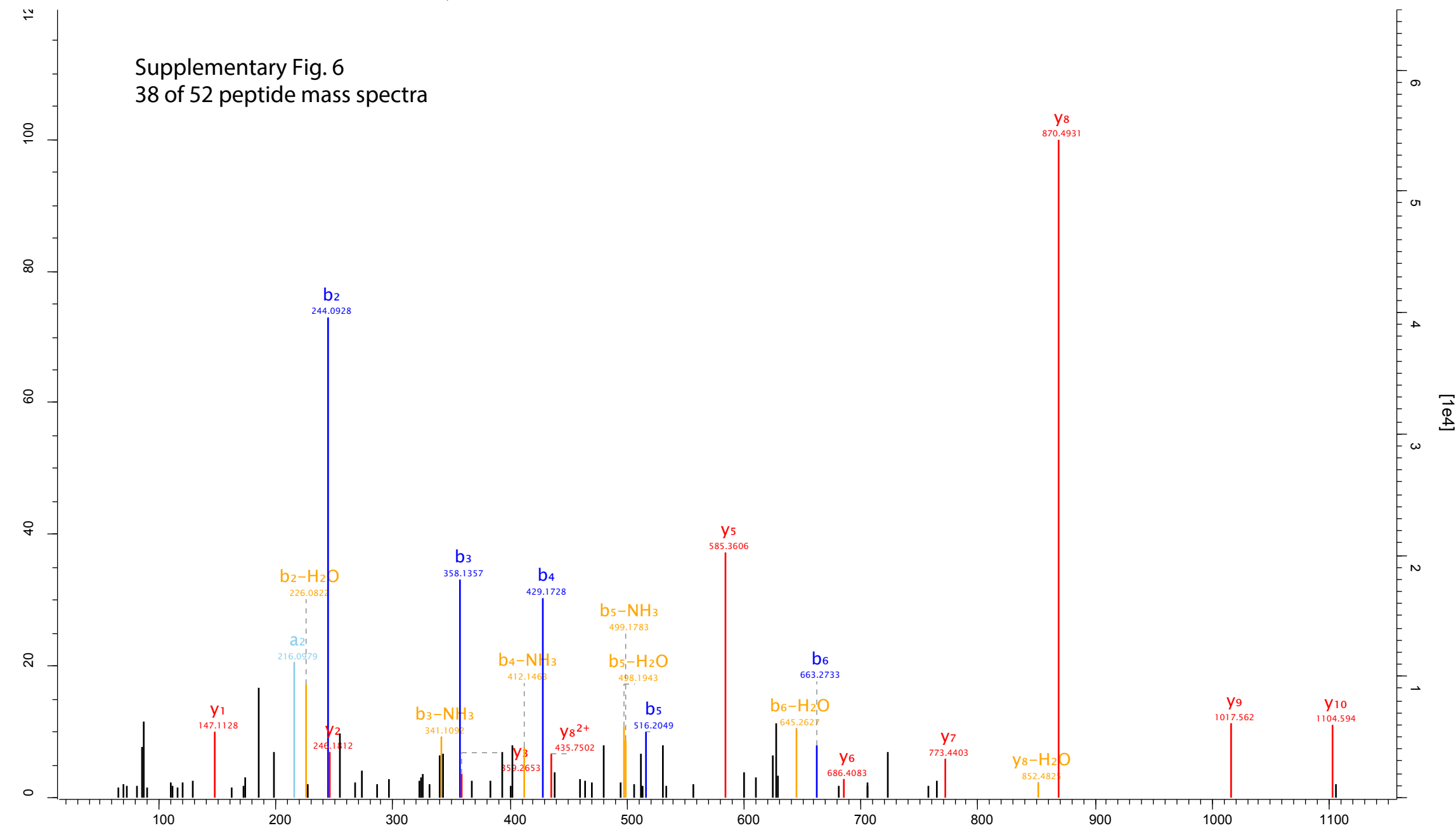
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36 of 52 peptide mass spectra



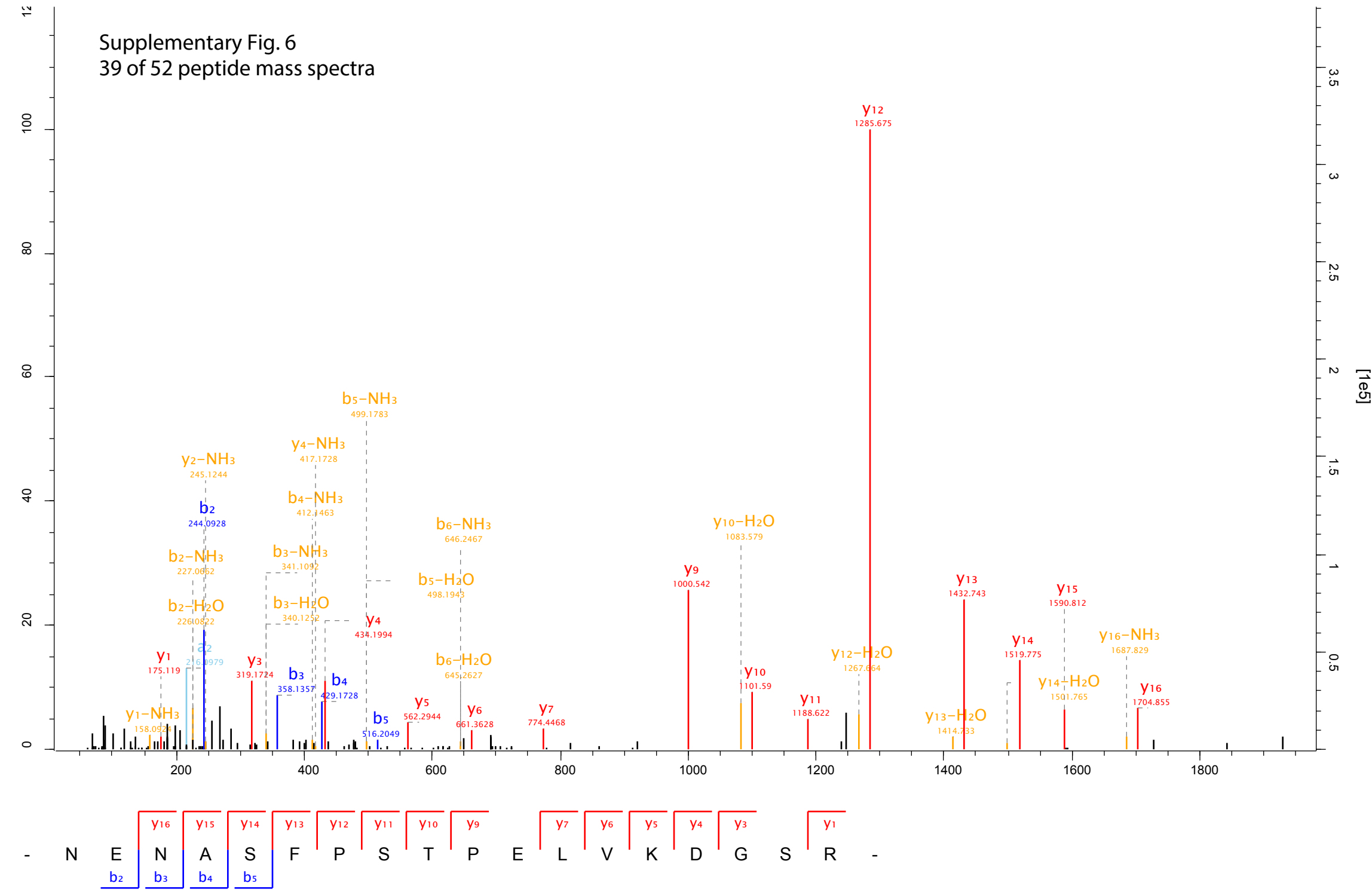
Supplementary Fig. 6
37 of 52 peptide mass spectra



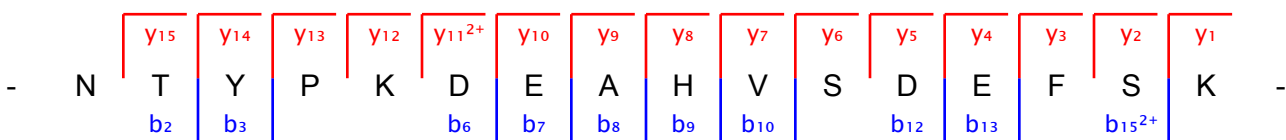
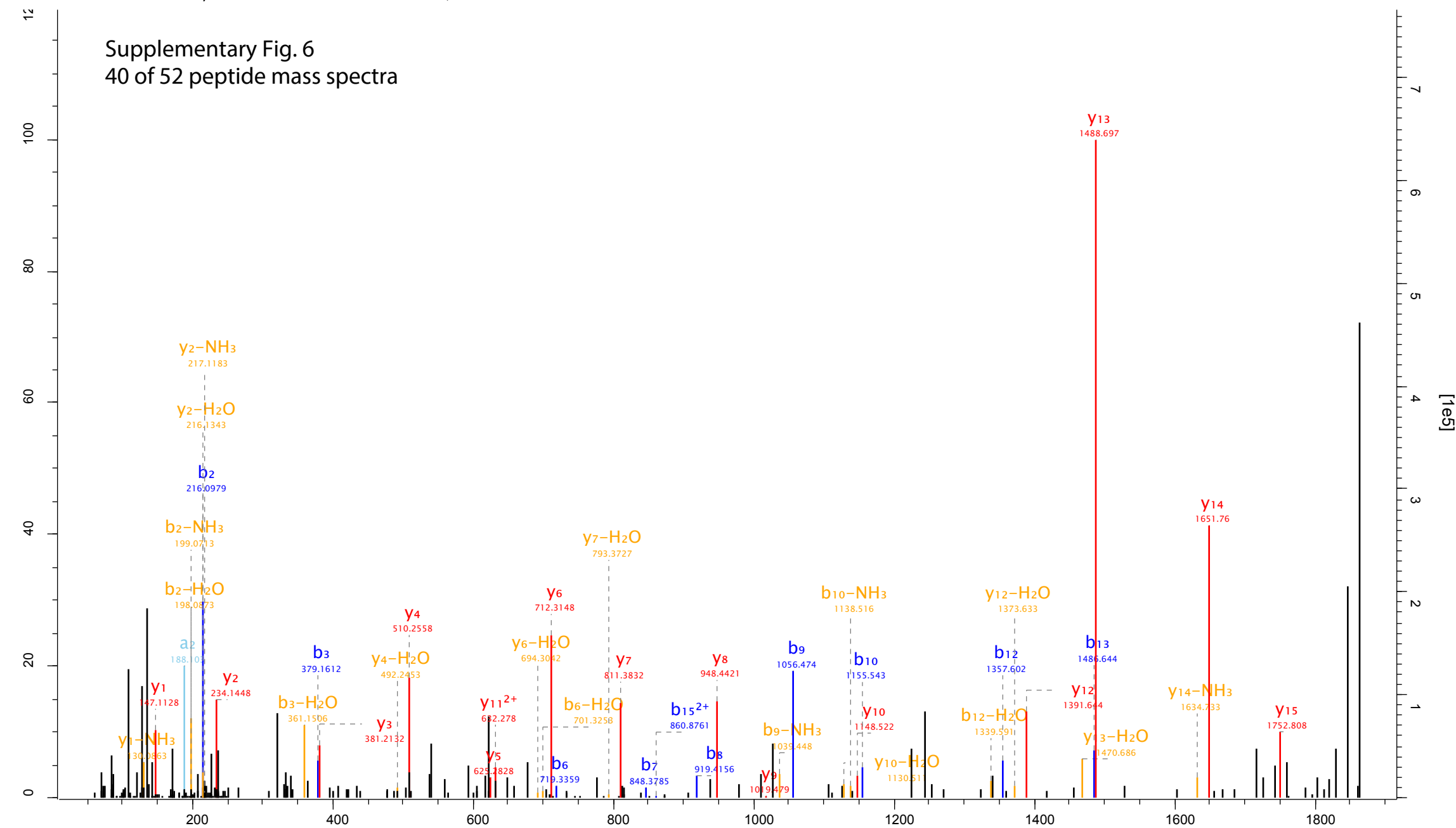
Supplementary Fig. 6
38 of 52 peptide mass spectra



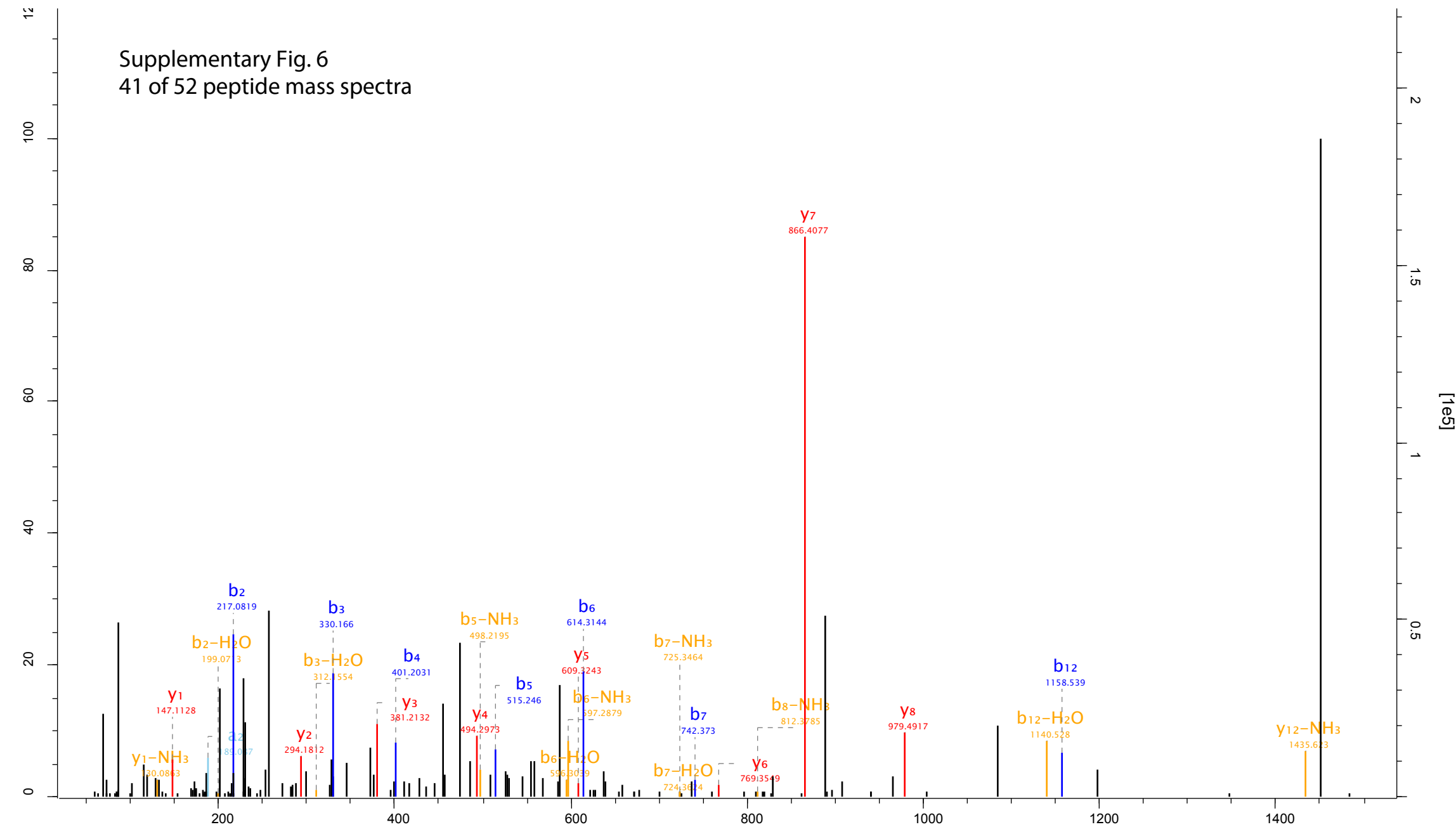
Supplementary Fig. 6
39 of 52 peptide mass spectra



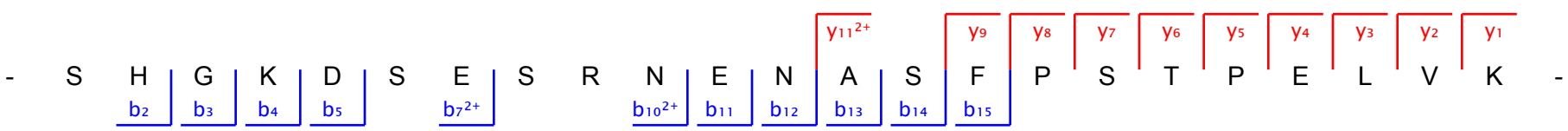
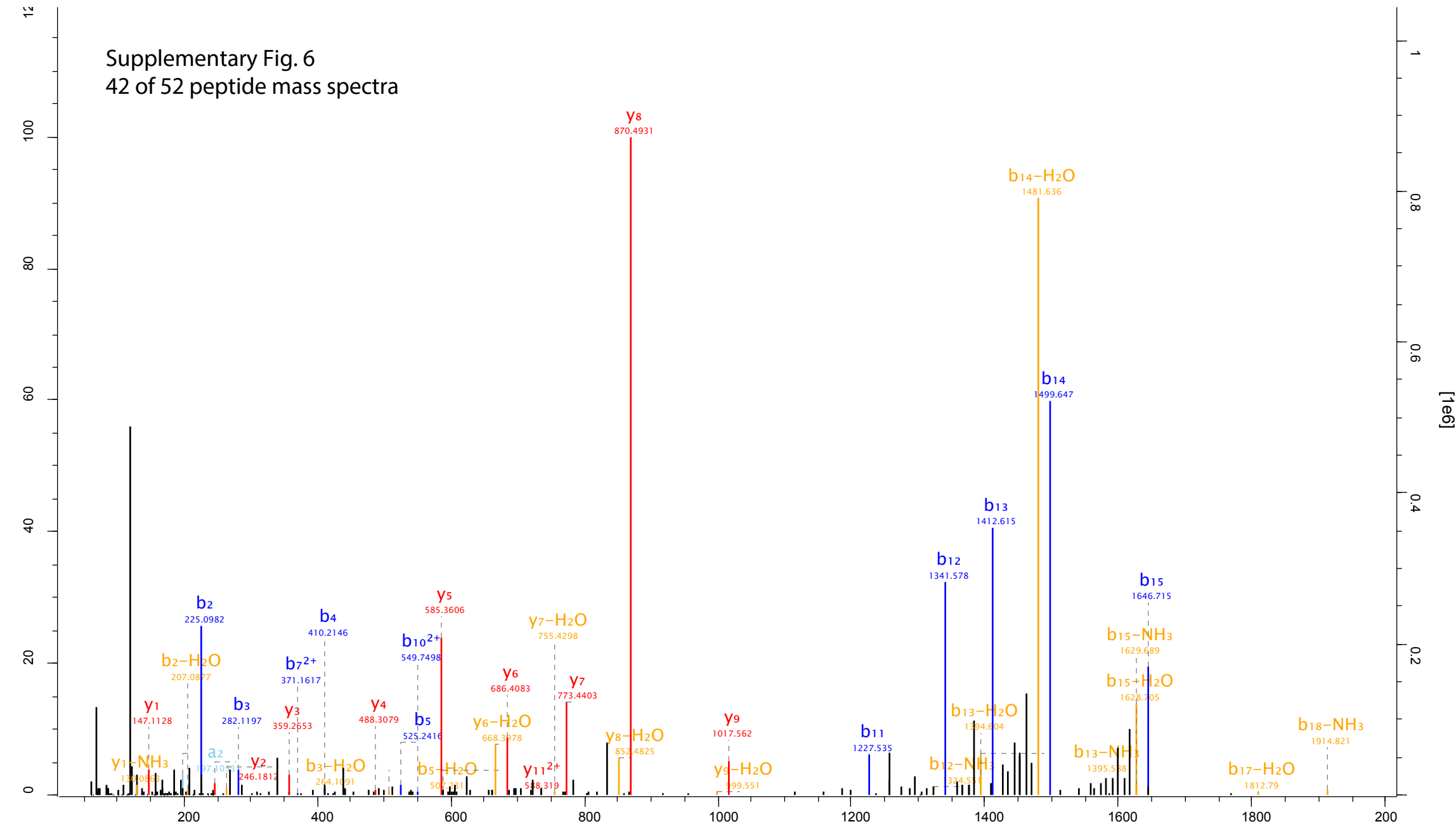
Supplementary Fig. 6
40 of 52 peptide mass spectra



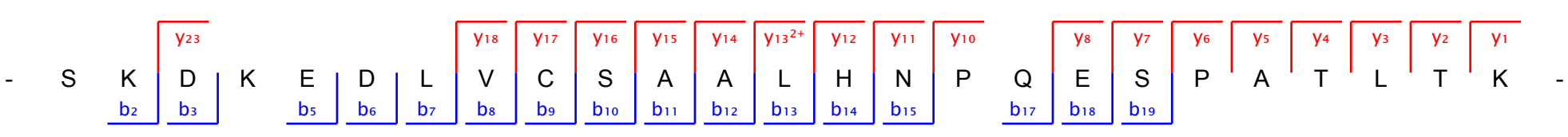
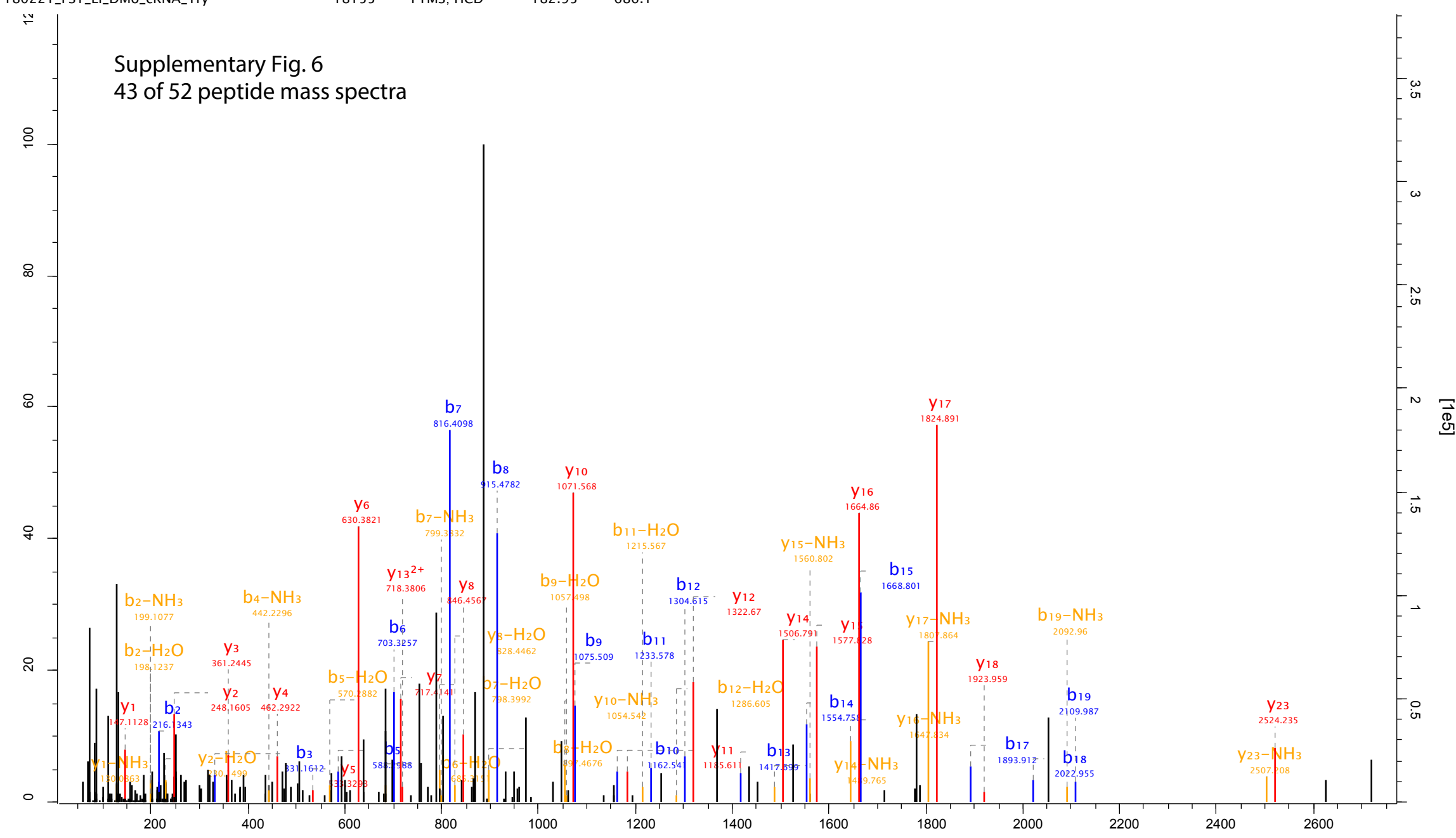
Supplementary Fig. 6
41 of 52 peptide mass spectra



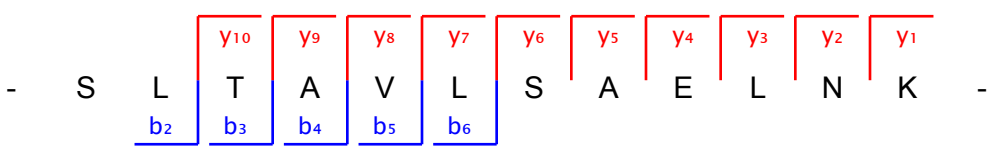
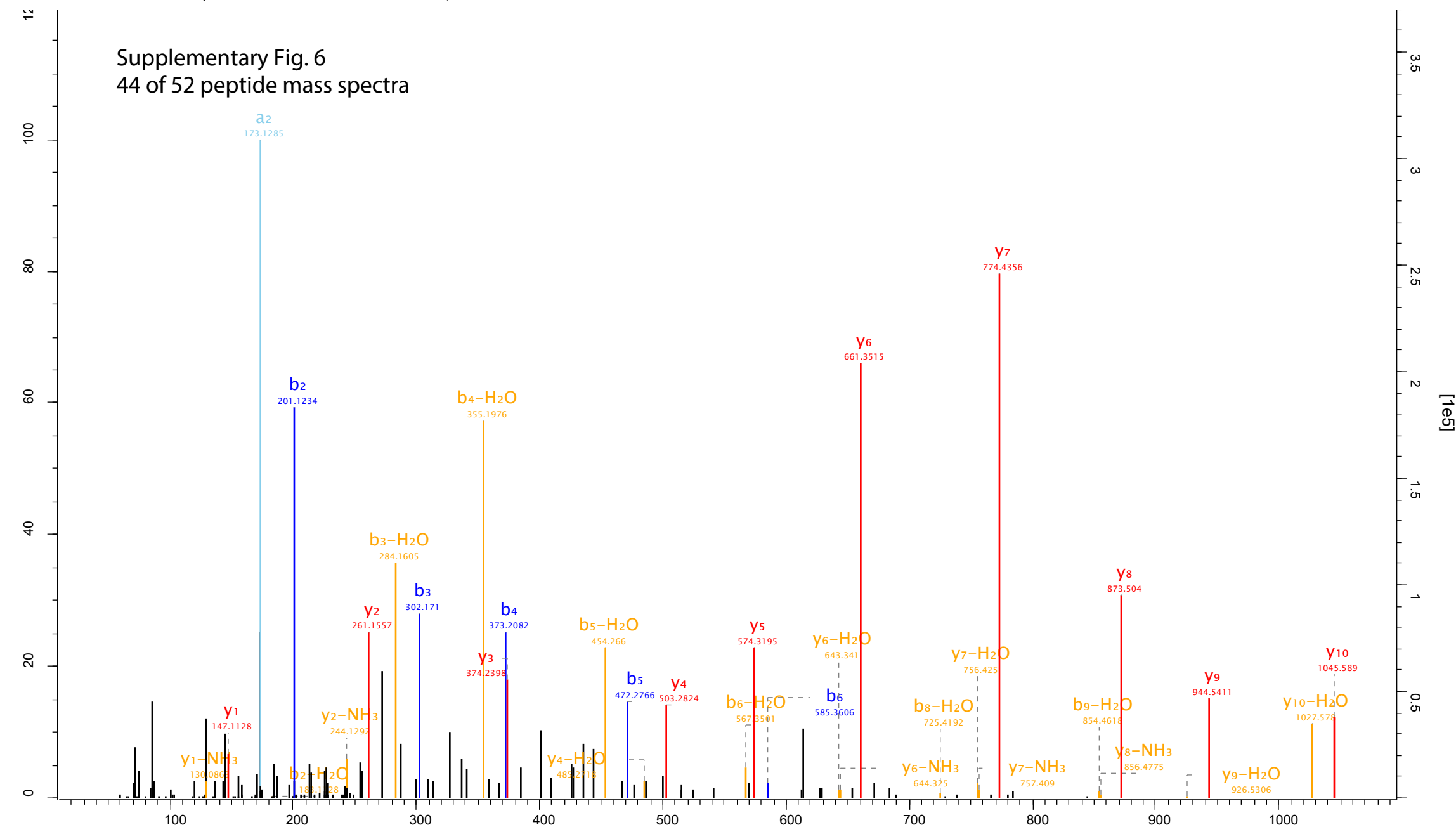
Supplementary Fig. 6 42 of 52 peptide mass spectra



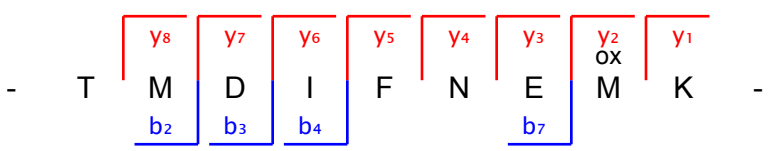
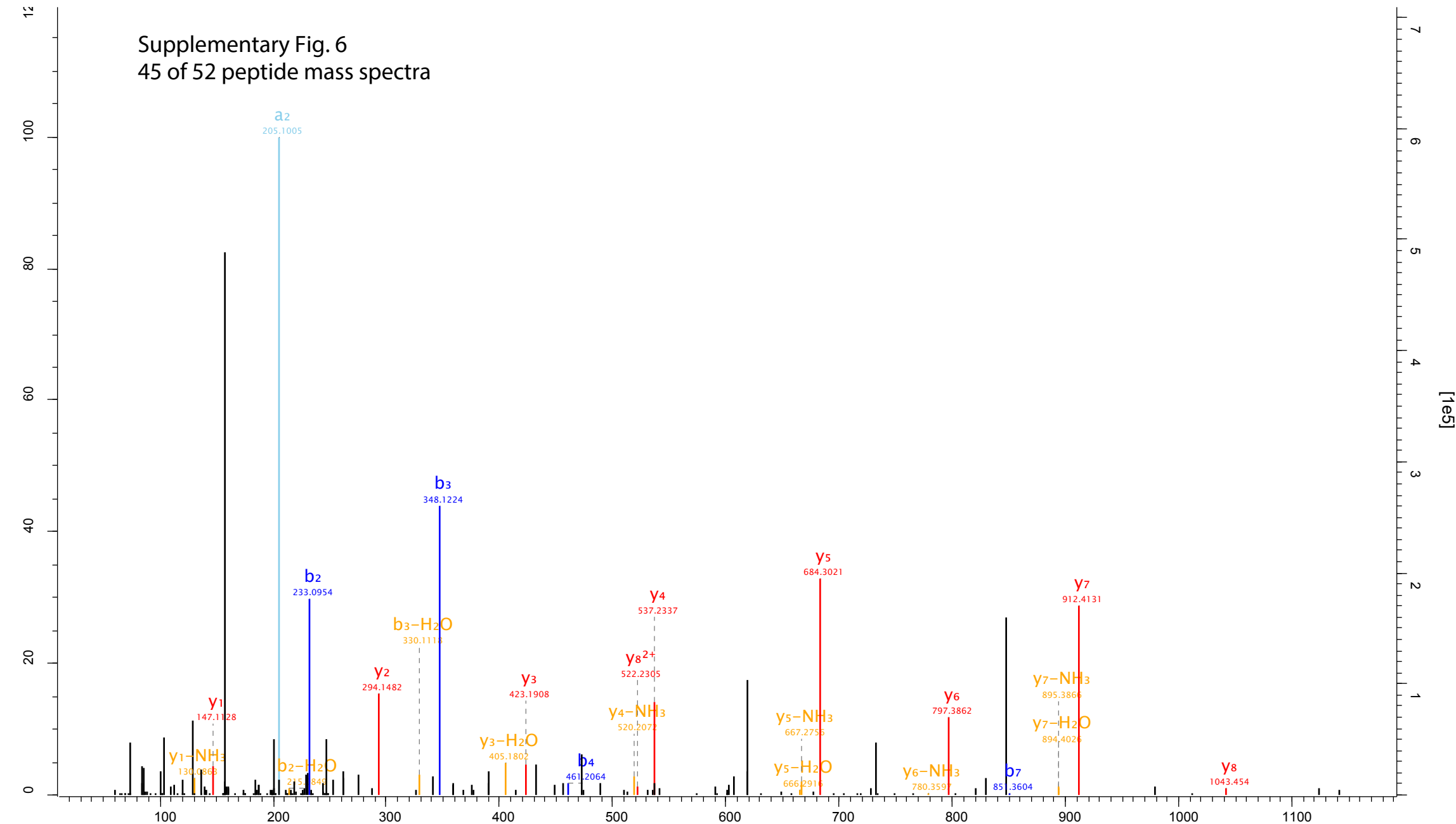
Supplementary Fig. 6
43 of 52 peptide mass spectra



Supplementary Fig. 6
44 of 52 peptide mass spectra

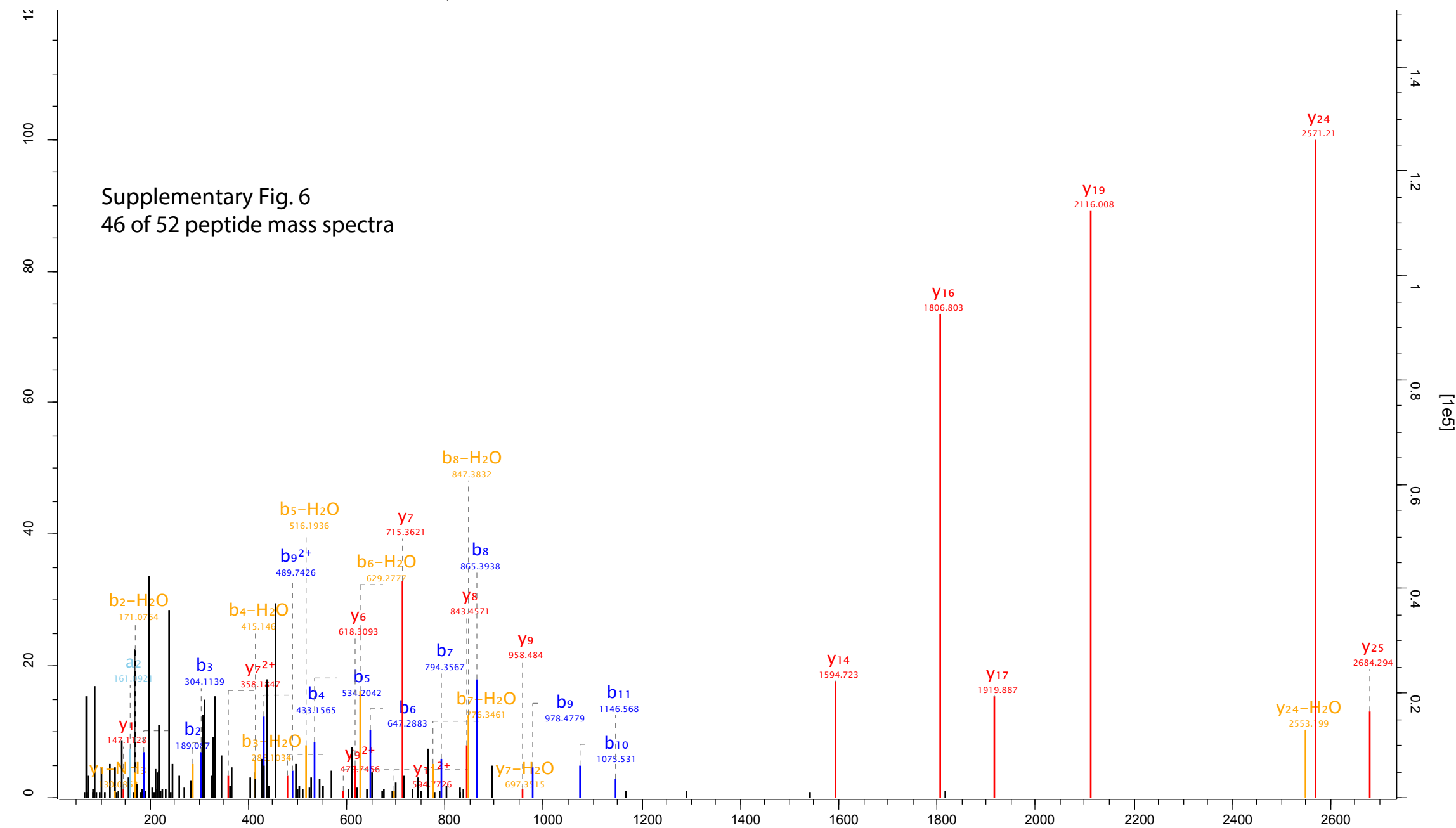


Supplementary Fig. 6
45 of 52 peptide mass spectra

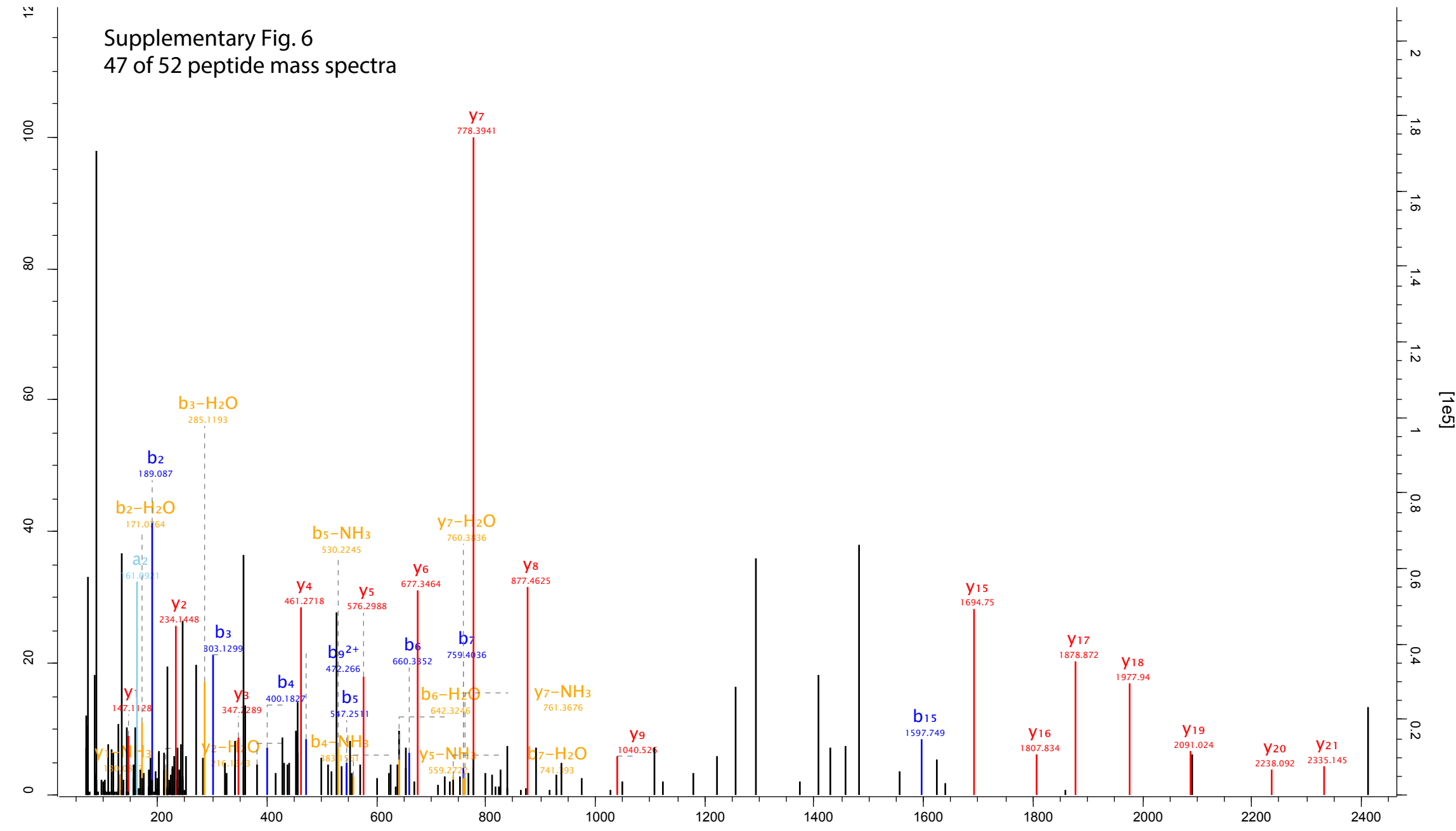


Raw file 180221_FS1_Li_DMo_cRNA_F_P Scan 32825 Method FTMS; HCD Score 69.52 m/z 888.44

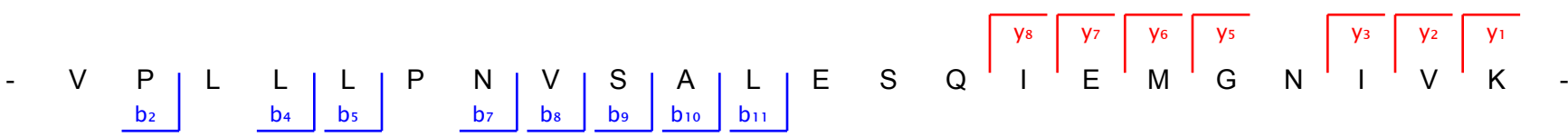
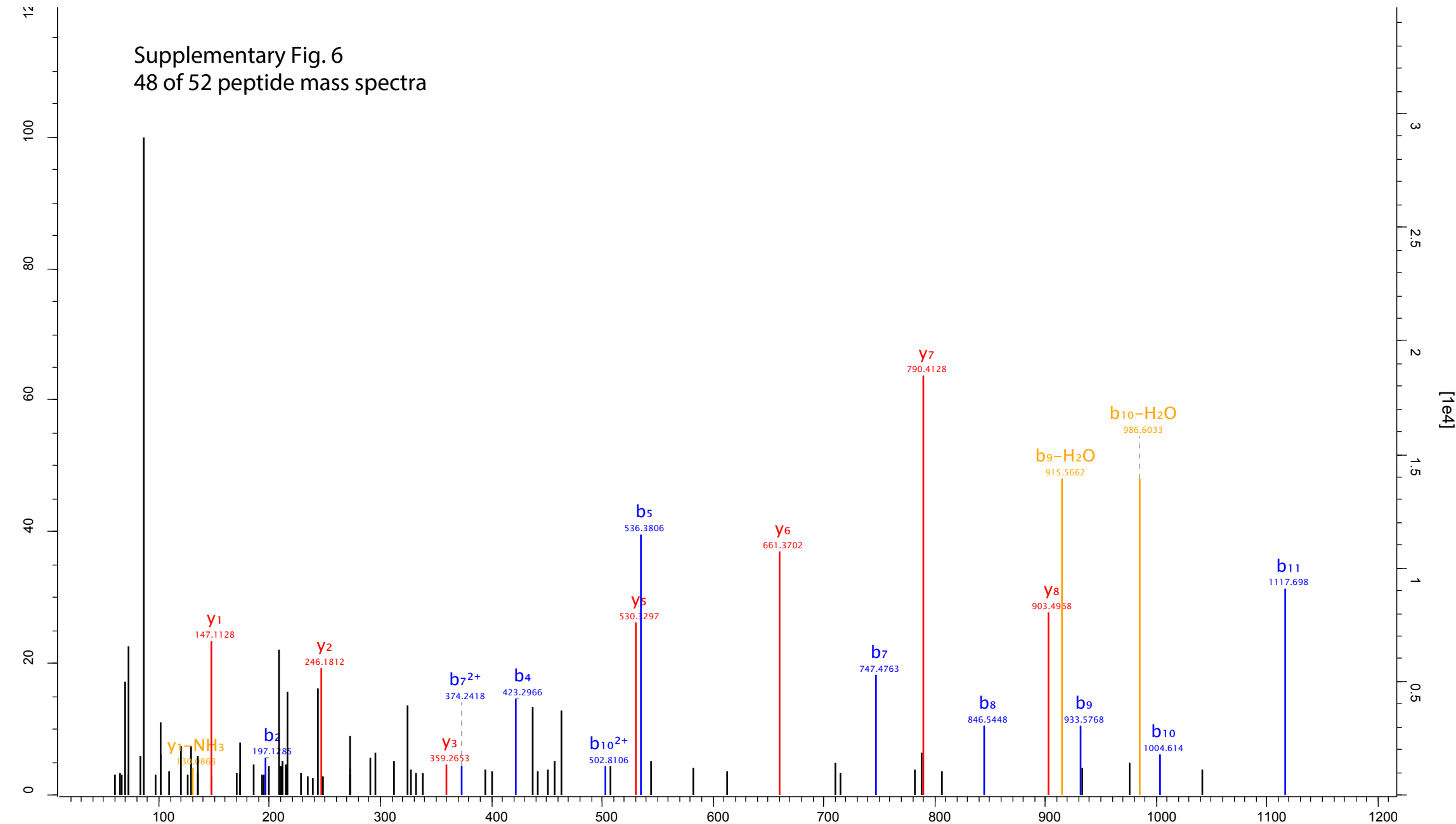
Supplementary Fig. 6
46 of 52 peptide mass spectra



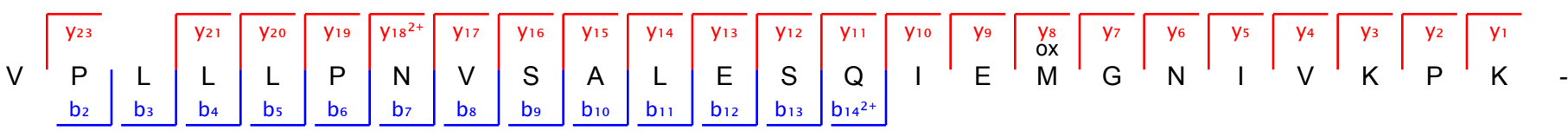
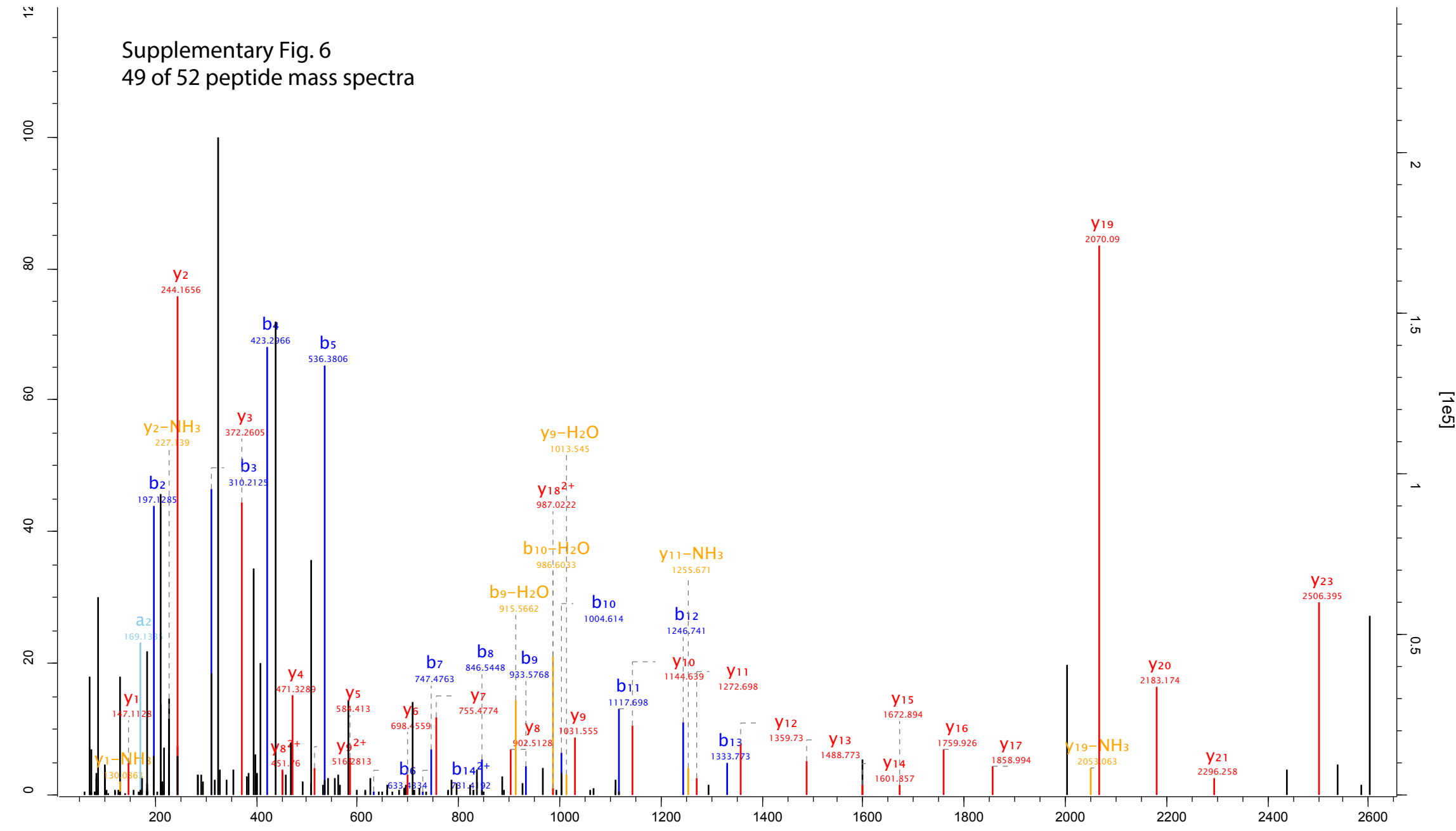
Supplementary Fig. 6
47 of 52 peptide mass spectra



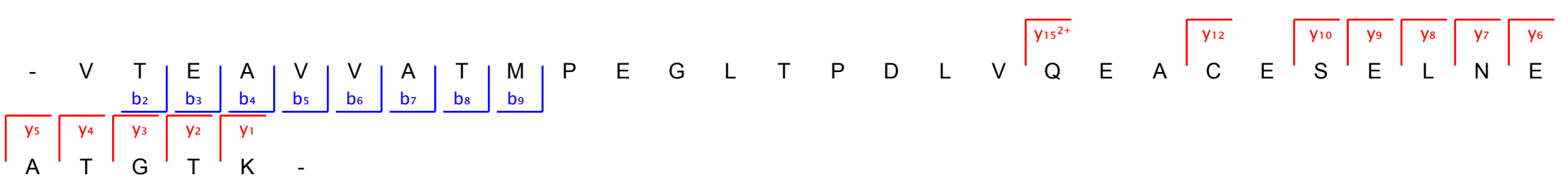
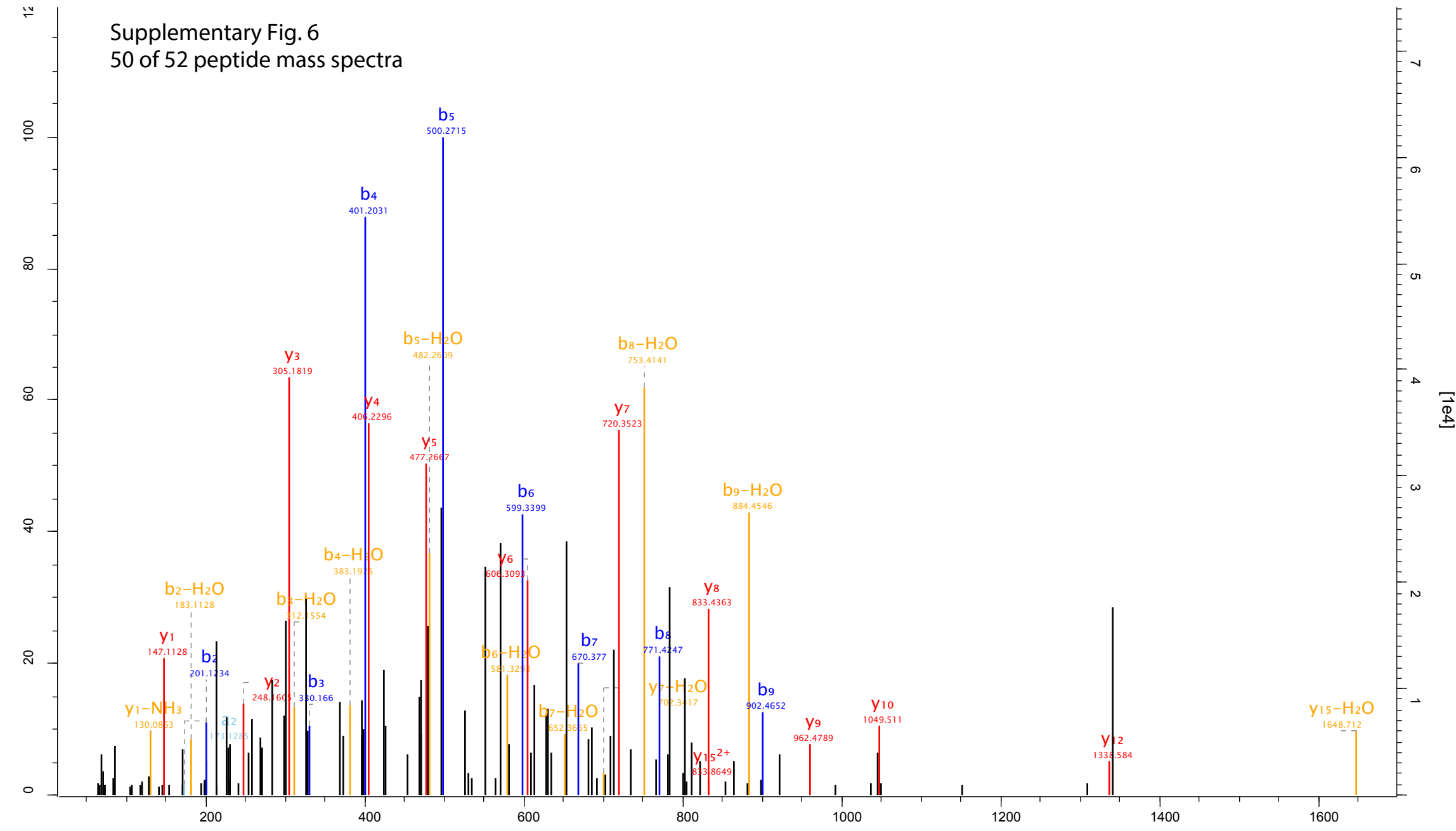
Supplementary Fig. 6
48 of 52 peptide mass spectra



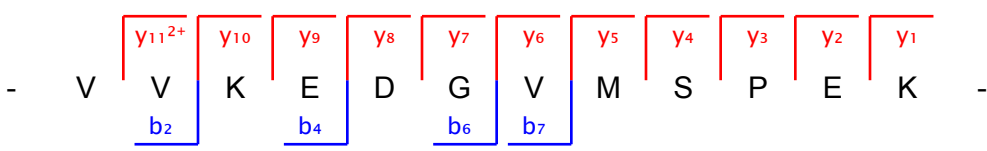
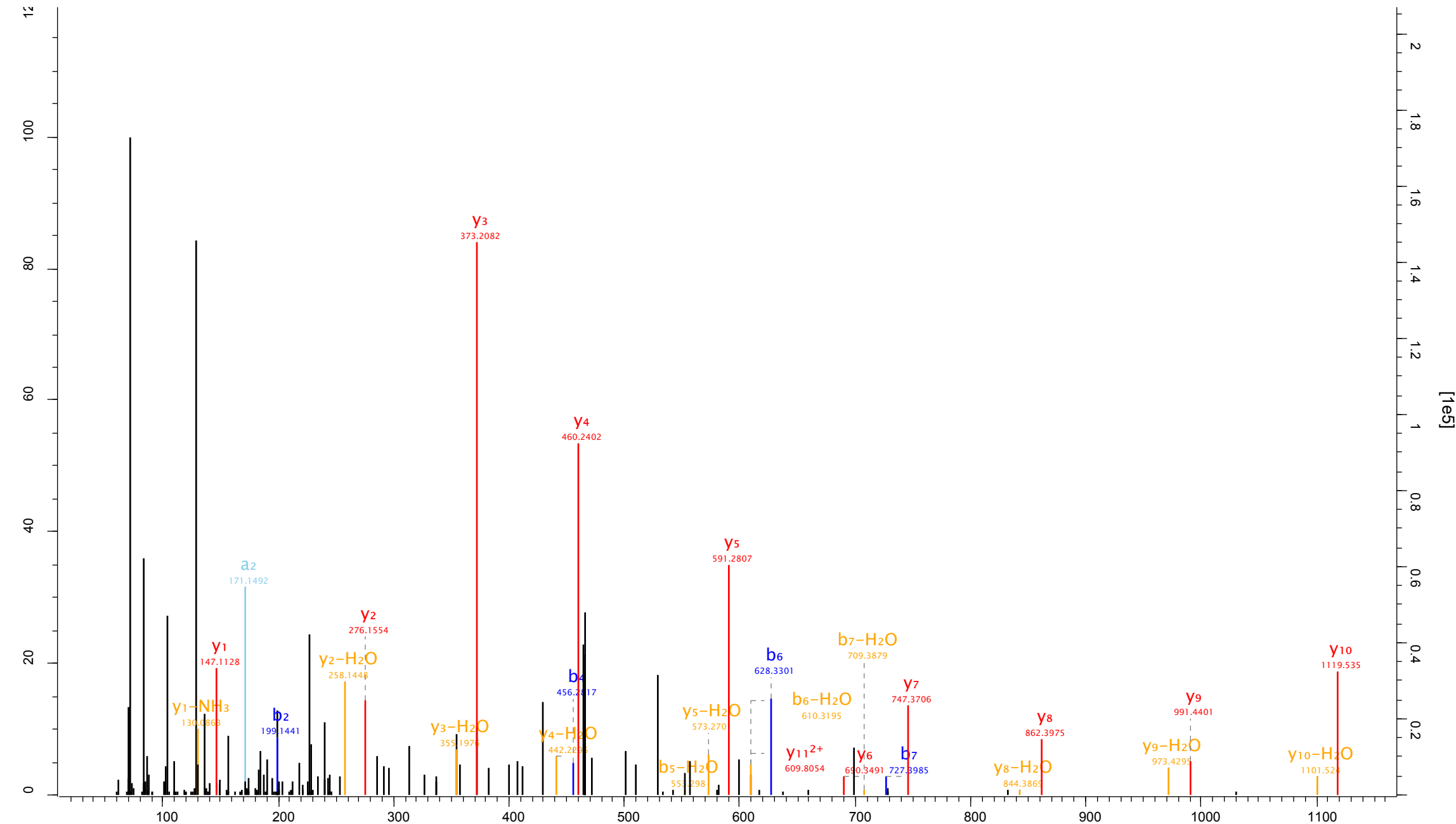
Supplementary Fig. 6
49 of 52 peptide mass spectra



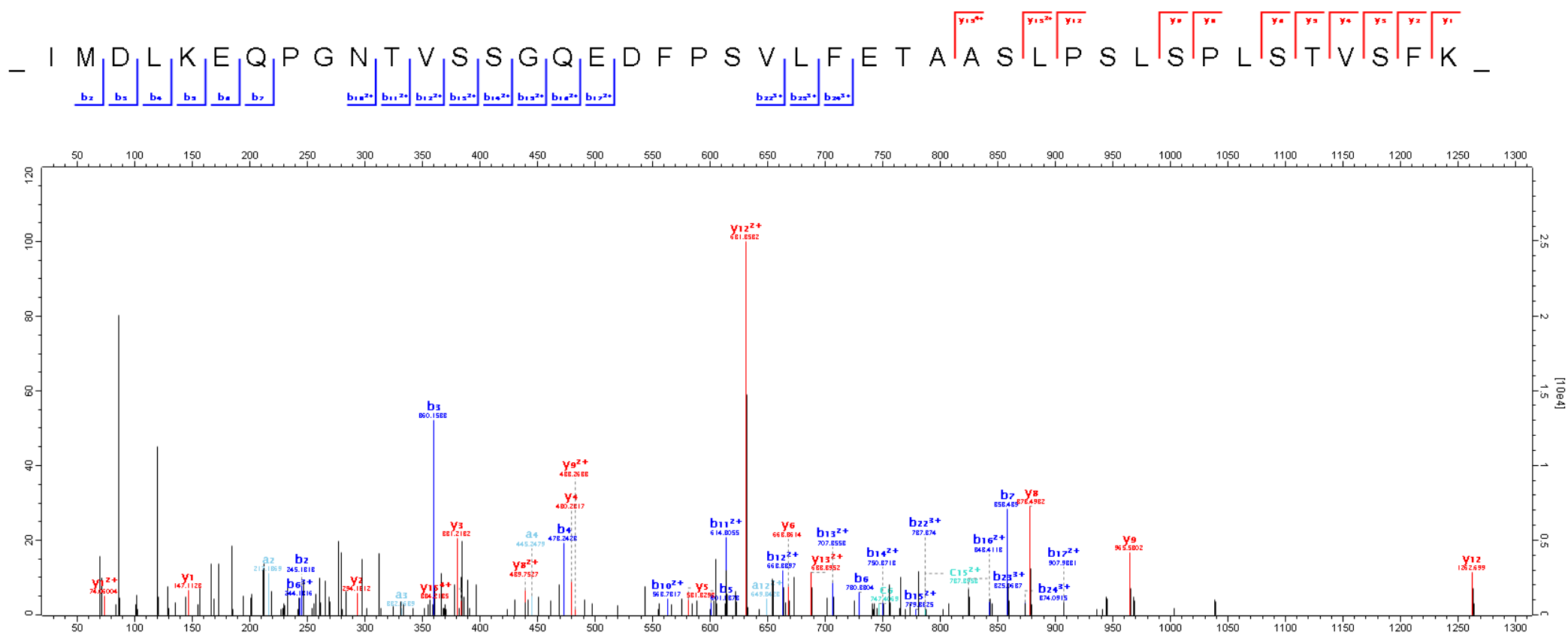
Supplementary Fig. 6
50 of 52 peptide mass spectra

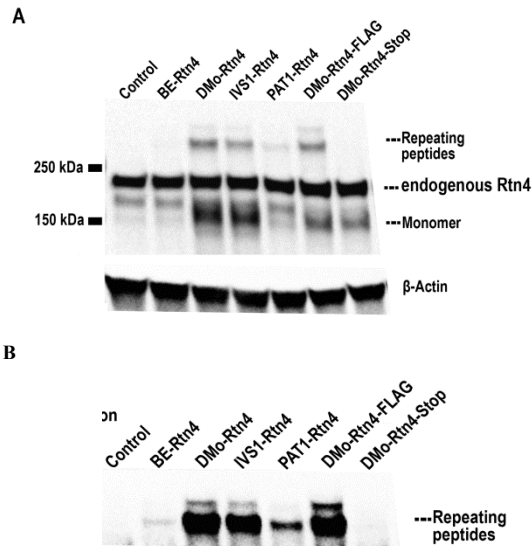


Raw file 180221_FS1_Li_DMo_cRNA_F_P Scan 5595 Method FTMS; HCD Score 112.02 m/z 439.9



Supplementary Fig. 6
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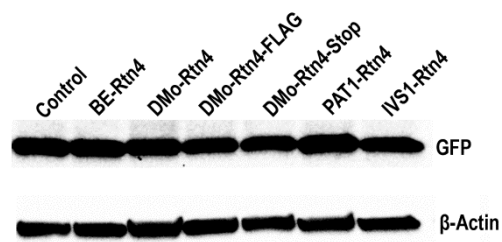




Supplementary Fig. 7.

Several IMEs promote circRtn4 translation in N2a cells.

A. Western blot with Anti-Nogo A antibody of protein isolated from N2a cells with circRtn4 overexpression. Control, empty vector; BE-Rtn4, pCircRNA-BE-Rtn4; DMO-Rtn4, pCircRNA-DMo-Rtn4; IVS1-Rtn4, pCircRNA-IVS1-Rtn4; PAT1-Rtn4, pCircRNA-PAT1-Rtn4; DMO-Rtn4-FLAG, pCircRNA-DMo-Rtn4-FLAG; DMO-Rtn4-Stop, pCircRNA-DMo-Rtn4-Stop; the high molecular weight bands larger than 250 kDa represent polypeptides from circRtn4 continuous translation; monomer designates the putative product from single round of circRtn4 translation; actin is used as loading control. B. long exposure of the high molecular weight bands larger than 250 kDa.



Supplementary Fig. 9 Equal transfection efficiency from circRNA vectors.

GFP expression from the SV40 promoter (Supplementary Fig. 1) in each circRNA expression plasmid DNA transfection in HEK293 cells was measured by Western blot analysis with antibody against GFP. β -Actin was used as loading control. Control, pCircRNA-DMo; BE-Rtn4, pCircRNA-BE-Rtn4; DMO-Rtn4, pCircRNA-DMo-Rtn4; DMO-Rtn4-FLAG, pCircRNA-DMo-Rtn4-FLAG; DMO-Rtn4-Stop, pCircRNA-DMo-Rtn4-Stop; PAT1-Rtn4, pCircRNA- PAT1-Rtn4; IVS1-Rtn4, pCircRNA-IVS1-Rtn4.