Supplementary Tables

Supplementary Table S1. List of mitocans and non-mitocans used for bioinformatics analysis.

Mitocans	*Non-mitocans*
Cytarabine	Hydroxyurea
Doxorubicin	Fludarabine
Paclitaxel	BN-2629
Etoposide	E-7820
Rapamycin	Fulvestrant
Vinorelbine	Dasatinib
Temsirolimus	Pazopanib
Arsenic trioxide	Afatinib
Obatoclax	Dolastatin 10
Salinomycin	Estramustine
Staurosporine	Midostaurin
Simvastatin	Testolactone
Mitoxanthrone	Valrubicin
L-Asparaginase	Fostamatinib

Supplementary Table S2. List of cell lines included in the study.

Cell line	Cell line origin	Disease	Age	Sex	Prior treatment	Ploidy	Genetic mutations / rearrangements
MV-4-11	Peripheral blood	Acute myeloid leukemia	10	M	NA	2n=48 (hyperdiploid)	FLT3-ITD[1], t(4;11)[2] [PMID: 12529668]
OCI-AML2	Peripheral blood	Acute myeloid leukemia	65	M	NA	Hyperdiploid 48(43-49)	*DNMT3A* [PMID: 21904384]
THP-1	Peripheral blood	Acute myeloid leukemia	1	M	NA	Near-diploid (46, XY)	*MLL-AF9* fusion, deletions in *CDKN2A, PTEN* [PMID: 11066077, 19635138]
MOLM-13	Peripheral blood	Acute myeloid leukemia	20	M	NA	Hyperdiploid 51(48-52)	FLT3-ITD, CBL deltaExon8 mutant, *MLL-AF9* fusion [PMID: 12529668; 9305600]
U251	Central nervous system	Glioblastoma	75	M	NA	2n=46	*CDKN2A, C9orf53, PTEN, TP53* [PMID: 24810477; 17088437]
SKOV3	Ovary	Adeno-carcinoma	64	F	Thiotepa	Hypodiploid (42-45)	*TP53, CDKN2A, PIK3CA, APC* [PMID: 26585234; 17088437]
OVCAR3	Ovary	Adeno-carcinoma	60	F	CyPh/CsPt/ Adr	Aneuploid, near-triploid 69+/-	*TP53* [PMID: 17088437]

[1] internal tandem duplication
[2] chromosomal translocation

Supplementary Table S3. Basic characteristics of the patients, enrolled in the study.

N	PB/BM[1]	age	sex	WBC	% blast	Pretreatment	Mutation status: gene mutations/genetic rearrangements	Karyotype
1.	PB	29	F	4.6	36	Decitabine, Venetoclax, Dasatinib	PTPN11, CSF3R	48,XX,-11,del(17)(p12),+r,+2mar[2]/49,idem,+8[12]/48~50,XX, t(7;11)(q22;q24),+8,del(9)(q13q22),-11,del(17)(p12),+r,+2~3mar[cp6]
2.	PB	83	M	15.6	30	ONC201, Cytarabine	TERT, CSF3R, TP53, GATA2	45,XY,t(3;8)(q25;q22),-5,del(7)(q22q34),add(8)(q24.1),del(12)(p11.2),16,+mar[18]/46~49,idem,+del(7)(q22q34)[cp2]
3.	PB	35	F	23.2	62	Decitabine	ASXL1, ASXL2, NRAS, RUNX1, FLT3 ITD, PHF6	46, XX
4.	PB	53	M	2.1	63	DCLL9718S	KMT2A, U2AF1	46,XY,add(1)(p36.1),del(7)(q22q34),del(20)(q11.2q13.3)[19]/47,idem,+del(7)(q22q34)[1]
5.	PB	41	F	13.5	92	Hydroxyurea, Crenolanib, Decitabine, Venetoclax, Gilteritinib	TET2, FLT3 ITD	49,XX,+4,t(6;9)(p22;q34),+8,+15,del(15)(q11.2q21)x2[14]/48~50,idem,+13[cp6]
6.	PB	77	M	4.9	57	Azacitidine, Cytarabine, Cladribine, Venetoclax	TET2, SRSF2, RUNX1, CBL, IL7R	46, XY
7.	PB	32	F	12.7	5.5	No treatment	FLT3 ITD, NPM1, DNMT3A	46, XX
8.	PB	57	M	37.2	15	Hydroxyurea. Cytarabine	ASXL1, NRAS, TET2, RUNX1T1	47,XY,+19[1]/47,idem,der(22)t(8;22)(q21.2;p11.2)[11]/46,XY[8]
9.	PB	79	F	5.4	6	IMGN622	ASXL, BCOR, DNMT3A	46,XX,t(2;17)(p14;q12),add(10)(q24),del(21)(q22q22)[19]/46,XX[1]
10.	BM	80	M	128	89	Hydroxyurea	FLT3 ITD, RUNX1 ISH2, PIGA, DNMT3A, ZRSR2	46, XY
11.	BM	57	M	9.3	86	No treatment	SMC1A, SRSF2, IDH1, ASXL2, FLT3 ITD	47,XY,+8[3]/46,XY,add(5)(q35)[2]/48,XY,+8,+mar[7]/47,XY,add(5)(q35),+8[6]/48,XY,add(5)(q35),+8,+mar[1]/49,XY,+8,+2mar[1]
12.	PB	64	M	3.1	67	Decitabine	TET2, TP53, EZH2, CBL	41~45,Y,der(X)t(X;1)(q24;p13),-2,add(3)(p13),del(5)(q13q35),-8,psudic(9;10)(p22;p11.2),-15,-17,add(17)(p11.2),add(21)(p11.2),del(22)(q11.2),+2~6mar[cp19]/46,XY[1]
13	PB	58	M	38	31	No treatment	ETV6, EZH2, NRAS, SETBP1, DNMT3A	46, XY
14.	PB	70	M	8.2	48	No treatment	KDM6A, RUNX1, TP53	46~47,XY,+1,der(1;10)(q10;q10),der(1;12)(p10;q10),del(5)(q11.2),del(6)(q15q25),-14,-16,add(16)(p11.2),-17,-21,+3~6mar[cp15]/46,XY[4]
15.	BM	89	M	95.8	65	No treatment	ASXL1, CBL, CREBBP, PHF6, TET2	50,XY,+der(1;11)(q10;q10),+8,+8,+8[10]/47~51,XY,+add(1)(p13),+8,+8,+8,der(9)t(1;9)(q21;p24)[cp10]
16.	BM	58	M	38	31	No treatment	ETV6, EZH2, NRAS, SETBP1, DNMT3A	46, XY
17.	PB	64	M	9.2	23	Dasatinib, Decitabine, Venetoclax	ASXL1	46,XY,add(2)(q12),t(4;12)(q12;q24.1),ins(12;?)(p13;?)[18]/46,XY[2]

18.	PB	72	M	5.2	90	AMG 330, Tocilizumab, Hydrea	BCOR, U2AF1, DNMT3A	46, XY
19.	PB	21	M	4.8	86	Decitabine, Venetoclax,	NPM1, IDH2	46,XY,t(2;4)(p23;q21)[3]/46,XY,t(5;17)(q13;q25)[2]/46,XY[15]
20.	PB	30	M	2.3	61	Mitoxantrone, Etoposide, Cytarabine	BCOR, PTPN11	45,XX,t(3;3)(q21;q26.2),del(6)(q12),-7[16]
21.	PB	72	M	1.4	38	Hydroxyurea, Quizartinib, Decitabine	CEBPA, FLT3 ITD, KRAS, PTPN11, WT1	46, XY

[1] Source of primary AML cells: PB (peripheral blood) / BM (bone marrow).

Supplementary Table S4. Primer sequences used for qPCR studies.

Locus	Forward (5'-3')	Reverse (5'-3')	PCR protocol
Primers used for estimation of relative mtDNA content (mtDNA/gDNA ratio)			I. 95 ^{0}C for 30 s; II. 1. 95 ^{0}C for 10 s; 2. 55 ^{0}C for 45 s; +Plate read (40 cycles) III. Melt Curve 65 to 95 ^{0}C, increment 0.5 ^{0}C for 5 s +Plate read END
GAPDH	TCTGACTTCAACAGCGACAC	CAGCCACATACCAGGAAATG	
B2M	TGTAAGCAGCATCATGGAGG	CTCCACATAGTGAGGGTTATC	
TBP	GAGAGTTCTGGGATTGTACC	CCATCTGAAAACAGAGCAGG	
ACTB	AAAGACCTGTACGCCAACAC	GGAAAGACACCCACCTTGAT	
ND1	GTATTATACCCACACCCACC	GTACAATGAGGAGTAGGAGG	
ND4	CACATATGGCCTAGACTACG	GTGGGTGGTTGTGTTGATTC	
Primers used for gene expression studies			
MFN1	TCTCCGCCTTTAACTTCTCG	TTATGCTAAGTCTCCGCTCC	
MFN2	GCAGGACATGATAGATGGCT	ATAGTTGAGGGAGAAGCACTG	
OPA1	GTCAGTCAAATGGACCCTCA	ATCTGCTGAATCCTGCTTGG	
DNML1	GAACAGCGAGATTGTGAGGTT	TGCATTACTGCCTTTGGCAC	
FIS1	ATCCGTAAAGGCATCGTGCT	AGAAGACGTAATCCCGCTGT	
MFF	ACCTGACTGTTGTAGATGCAG	AGCACGTTCTTTGTTCTCCTC	
MID49	GTGAAGCGGTTCATTGACAG	TTGAGCAGGCTCAGTTCCTT	
MID51	ATGTACTTGAGTGGCAGCCT	GGGCACAATGAGTTGGATGT	
ACTB	GATCAAGATCATTGCTCCTCCTG	ATACTCCTGCTTGCTGATCCA	

Supplementary Table S5. Results of multiple linear regression: association of cancer type with the sensitivity to mitocan treatment.

Dependent variable[1]	B-coefficient	SE(B)	*p*-value
After adjustment for age, gender and prior treatment of the cell line donor			
Mitocan activity Z-scores	0.382	0.168	**0.031**
Non-mitocan activity Z-scores	0.346	0.171	0.054

[1] With cancer type as an independent predictor

$p < 0.05$ are highlighted in bold

Supplementary Table S6. Fold changes (positive – increase; negative – decrease) of mitochondrial and bioenergetic parameters after doxorubicin treatment (DOX/untreated).

Cell line	mtDNA content	MTG fluore-scence	JC-1, red/ green	ATP content	Basal OCR	ATP-linked OCR	Proton leak	Spare capacity	Coupling efficiency	ECAR	WB (ATP syn ß)	LD50 (DOX), µM
MV-4-11	**-1.49**			**-19.71**	**-2.19**	**-2.84**	-1.94	-1.10	-1.43	1.22		1.634
OCI-AML2	**-1.44**			**-26.80**	**-2.02**	**-3.26**	-1.28	**-4.26**	**-2.22**	-1.03		21.67
MOLM-13	**-1.77**	**-32.03**	**-63.96**	**-9.57**	**-3.11**	**-6.29**	-1.23	**-5.95**	**-2.16**	-1.10	1.118	26.75
THP-1	**-1.40**			**-5.66**	**-5.32**	**-9.26**	**-3.30**	**-3.80**	**-2.36**	**-1.92**		68.70
PBMC	1.40	**-13.97**	**-4.71**	**-8.76**	**-1.55**	**-1.61**	-1.41	**-2.23**	-1.20	**-1.52**	**3.772**	40.85
U251	**5.51**			-1.05								122.22
SKOV3	**2.37**	**-29.19**	**-15.34**	1.07	-1.28	-1.26	-1.03	**-1.90**	-1.07	1.09	1.002	110.82
OVCAR3	**5.59**			1.05								93.87

Cases with $p<0.05$ are in **bold**.

Supplementary Table S7. Fold change differences in expression of fusion and fission genes in DOX-treated (LD50, 8 h) cells compared to untreated cells.

Gene / Cell line	Fusion			Fission				
	MFN1	*MFN2*	*OPA1*	*DNM1L*	*FIS1*	*MFF*	*MID49*	*MID51*
MV-4-11	0.83 (0.50-1.37)*	2.13 (1.34-3.39)	1.35 (0.84-2.18)	2.06 (1.26-3.36)	3.06 (1.88-4.99)	1.99 (1.19-3.34)	5.71 (3.52-9.27)	2.86 (1.69-4.85)
MOLM-13	1.09 (0.53-2.26)	2.19 (0.76-6.32)	0.51 (0.25-1.03)	0.58 (0.24-1.38)	0.80 (0.30-2.12)	0.53 (0.28-1.01)	16.82 (4.96-56.95)	1.41 (0.42-4.71)
THP-1	5.43 (3.71-7.95)	6.95 (3.61-13.37)	0.71 (0.52-0.98)	0.36 (0.13-0.99)	1.84 (1.26-2.69)	0.66 (0.48-0.92)	32.98 (15.96-68.15)	3.14 (1.78-5.54)
PBMC	3.09 (1.11-8.63)	4.82 (1.18-19.67)	1.29 (0.47-3.59)	0.91 (0.45-1.84)	1.52 (0.74-3.10)	2.24 (1.04-4.84)	13.67 (2.79-67.09)	4.72 (1.08-20.68)
SKOV3	0.33 (0.11-0.95)	0.67 (0.51-0.86)	0.46 (0.17-1.21)	0.52 (0.18-1.51)	0.93 (0.72-1.21)	0.63 (0.35-1.12)	0.58 (0.37-0.89)	0.25 (0.21-0.30)

* Fold changes were calculated using the comparative $\Delta\Delta C_t$ method

Supplementary Table S8. Relative potency of drugs used for combination studies (single-ray design) in cell lines and primary AML samples.

Cell line/Patient sample	$LD25_{1st\ drug}$ / $LD25_{2nd\ drug}$ = Relative potency	Fixed ratio (50:50) concentrations of the 1st drug, 2nd drug
2-DG (1st drug) / CCCP (2nd drug)		
MV-4-11	7500 µM / 2.07 µM = 3623	10.7 mM, 3 µM; 7.5 mM, 2.1 µM; 3.6 mM, 1 µM
OCI-AML2	16240 µM / 15.63 µM = 1039	16.2 mM, 15.6 µM; 8.1 mM, 7.8 µM; 6.55 mM, 6.3 µM
MOLM-13	51600 µM / 6.3 µM = 8190	51.6 mM, 6.3 µM; 41 mM, 5 µM.
THP-1	74800 µM / 4.06 µM = 18424	74.8 mM, 4.1 µM; 36.8 mM, 2 µM; 18.4 mM, 1 µM
PBMC	106000 µM / 45.41 µM = 2334	106 mM, 45.41 µM; 70 mM, 30 µM; 46.7 mM, 20 µM; 23.3 mM, 10 µM
U251	616150 µM / 92.3 µM = 6676	616.15 mM, 92.3 µM; 467.3 mM, 70 µM; 333.8 mM, 50 µM
SKOV3	737000 µM / 133.4 µM = 5525	800 mM, 139 µM; 737 mM, 133.4 µM; 688.9 mM, 120 µM
OVCAR3	390748 µM / 130.3 µM = 2999	450 mM, 150 µM; 390.7 mM, 130.3 µM; 300 mM, 100 µM; 150 mM, 50 µM; 30 mM, 10 µM
AML 1	8660 µM / 12.33 µM = 702	8.66 mM, 12.33 µM; 5.77 mM, 8.22 µM; 2.87 mM, 4.11 µM
AML 2	79100 µM / 16.07 µM = 4922	79 mM, 16.07 µM; 50 mM, 10 µM; 24.5 mM, 5 µM
AML 3	80700 µM / 11.4 µM = 7079	80.7 mM, 11.4 µM; 53 mM, 7.5 µM; 25 mM, 3.5 µM
AML 4	66200 µM / 7.7 µM = 8597	66.2 mM, 7.7 µM; 43 mM, 5 µM; 21.5 mM, 2.5 µM
AML 5	80900 µM / 8.3 µM = 9747	80.9 mM, 8.3 µM; 57.8 mM, 6 µM; 40.5 mM, 4 µM; 19.5 mM, 2 µM
AML 6	3700 µM / 2.6 µM = 1423	3.7 mM, 2.6 µM; 2.9 mM, 2 µM; 1.4 mM, 1 µM
AML 7	23800 µM / 17.4 µM = 1368	23.8 mM, 17.4 µM; 13.7 mM, 10 µM; 6.8 mM, 5 µM
AML 8	58900 µM / 23.8 µM = 2475	58.9 mM, 23.8 µM; 36.8 mM, 15 µM; 24.5 mM, 10 µM; 12.3 mM, 5 µM
AML 9	268000 µM / 13.5 µM = 19852	268 mM, 13.5 µM; 198.5 mM, 10 µM; 99.3 mM, 5 µM; 19.9 mM, 1 µM
AML 10	58800 µM / 2.23 µM = 26368	58.8 mM, 2.23 µM; 26.4 mM, 1 µM; 13.2 mM, 0.5 µM
AML 11	4800 µM / 14.5 µM = 331	4.8 mM, 14.5 µM; 2.4 mM, 7.25 µM; 1.2 mM, 3.6 µM
AML 12	56100 µM / 10.33 µM = 5431	56.1 mM, 10.3 µM; 28.1 mM, 5.2 µM; 14 mM, 2.6 µM; 7 mM, 1.3 µM
AML 13	200700 µM / 55.1 µM = 3642	200 mM, 55.1 µM; 100 mM, 27.6 µM; 50 mM, 13.8 µM
AML 14	43200 µM / 39.6 µM = 1091	43.2 mM, 39.6 µM; 21.6 mM, 19.8 µM;

		10.8 mM, 9.9 µM
AML 15	7950 µM / 70.2 µM = 113	8 mM, 70 µM; 4 mM, 35 µM; 2 mM, 17.5 µM; 1 mM, 8.75 µM
AML 16	95700 µM / 17 µM = 5629	95.7 mM, 17 µM; 47.9 mM, 8.5 µM; 23.9 mM, 4.25 µM; 12 mM, 2.13 µM
AML 17	81000 µM / 16.7 µM =4850	81 mM, 16.7 µM; 40.5 mM, 8.35 µM; 20.25 mM, 4.18 µM; 10.1 mM, 2.1 µM
AML 18	10300 µM / 4 µM = 2575	10.3 mM, 4 µM; 2.6 mM, 1 µM; 1.3 mM, 0.5 µM
AML 19	18200 µM / 0.55 µM = 33091	18.2 mM, 0.55 µM; 12.1 mM, 0.37 µM; 9.1 mM, 0.28 µM; 4.55 mM, 0.13 µM; 2.28 mM, 0.07 µM
AML 20	850 µM / 2.18 µM = 390	0.85 mM, 2.2 µM; 0.425 mM, 1.1 µM; 0.2 mM, 0.55 µM; 0.1 mM, 0.27 µM
AML 21	105300 µM / 22.8 µM = 4618	105.3 mM, 22.8 µM; 52.7 mM, 11.4 µM; 26.3 mM, 5.7 µM
CCCP (1st drug) / ABT-199 (2nd drug)		
OCI-AML2	15.63 µM / 0.177 µM = 88	15.63 µM, 0.177 µM; 10 µM, 0.113 µM; 5 µM, 0.06 µM; 1 µM, 0.0113 µM
PBMC	45.41 µM / 0.6 µM = 76	45.1 µM, 0.6 µM; 30 µM, 0.4 µM; 20 µM, 0.26 µM; 10 µM, 0.13 µM
2-DG (1st drug) / ABT-199 (2nd drug)		
OCI-AML2	16240 µM / 0.177 µM = 91751	16.24 mM, 0.177 µM; 13.8 mM, 0.15 µM; 9.2 mM, 0.1 µM; 1.8 mM, 0.02 µM
PBMC	106000 µM / 0.6 µM = 176667	106 mM, 0.6 µM; 53 mM, 0.3 µM; 26.5 mM, 0.15 µM; 10.6 mM, 0.06 µM
2-DG (1st drug) / ara-C (2nd drug)		
OCI-AML2	16240 µM / 77.8 µM = 209	16.2 mM, 77.8 µM; 8.1 mM, 38.9 µM; 4.1 mM, 19.5 µM; 2 mM, 9.7 µM
AML 11[1]	4800 µM / 53.4 µM = 90	4.8 mM, 53.4 µM; 2.4 mM, 26.7 µM; 1.2 mM, 13.4 µM
AML 13	200700 µM / 131.8 µM = 1523	200 mM, 131.8 µM; 100 mM, 69.1 µM; 50 mM, 34.5 µM
AML 14	43150 µM / 400 µM = 108	43.2 mM, 400 µM; 21.6 mM, 200 µM; 10.8 mM, 100 µM
AML 15	7950 µM / 50.4 µM = 158	8 mM, 50 µM; 4 mM, 25 µM; 2 mM, 12.5 µM; 1 mM, 6.25 µM
AML 20	850 µM / 141.2 µM = 6	0.85 mM, 141.2 µM; 0.425 mM, 70.6 µM; 0.2 mM, 35.3 µM; 0.1 mM, 17.7 µM

[1] Only primary AML samples, not resistant to ara-C (LD25 < 400 µM corresponding to 0.4% DMSO), have been treated with combination of ara-C and 2-DG.

Supplementary Table S9. Results of multiple linear regression: no association of pretreatment of the patients (n = 21) with mean combination indices of CCCP/2-DG.

Dependent variable[1]	B-coefficient	SE(B)	*p*-value
Before adjustment for age, gender, source of primary cells and % blast cells			
Mean combination index based on LD25, 50, 75 of the mixture CCCP/2DG	-0.085	0.118	0.481
After adjustment for age, gender, source of primary cells and % blast cells			
Mean combination index based on LD25, 50, 75 of the mixture CCCP/2DG	0.002	0.153	0.988

[1] With pretreatment (0 / 1) as an independent predictor

Supplementary Table S10. Correlations between mitochondrial parameters across untreated AML cells, healthy blood cells and SKOV3 cell line[1].

		Max OCR	ATP-linked OCR	Spare capacity	Coupling efficiency	LD25, CCCP	LD25, 2-DG	CI based on LD50 of the mixture	Mean CI based on LD25,50,75 of the mixture
Basal OCR	*r*	**0.915**	**0.975**	0.790	0.189	0.345	0.565	0.521	0.283
	p	**0.011**	**0.001**	0.061	0.719	0.500	0.243	0.289	0.587
Max OCR	*r*		**0.966**	**0.969**	0.477	0.463	0.644	0.579	0.397
	p		**0.002**	**0.001**	0.338	0.355	0.168	0.229	0.436
ATP-linked OCR	*r*			**0.888**	0.376	0.500	0.694	0.639	0.424
	p			**0.018**	0.463	0.312	0.126	0.172	0.402
Spare capacity	*r*				0.651	0.550	0.687	0.61	0.481
	p				0.161	0.258	0.132	0.199	0.335
Coupling efficiency	*r*					**0.840**	0.784	0.749	**0.821**
	p					**0.036**	0.065	0.087	**0.045**
LD25, CCCP	*r*						**0.969**	**0.907**	**0.893**
	p						**0.001**	**0.012**	**0.017**
LD25, 2-DG	*r*							**0.943**	**0.869**
	p							**0.005**	**0.025**
CI based on LD50 of the mixture	*r*								**0.960**
	p								**0.002**

[1] Correlations defined by two-tailed Pierson correlation coefficient *r*; *p* defines *p*-value. Significant correlations ($p<0.05$) are in bold.

Supplementary Table S11. Changes in mitochondrial bioenergetic parameters after treatment with mitocans (CCCP, DOX, LD50), glycolysis inhibitor (2-DG, LD50), or their combination (CCCP and 2-DG, LD25) in a panel of cells.

Drug	Bioenergetic parameter	MV-4-11	OCI-AML2	MOLM-13	THP-1	PBMC	SKOV3
CCCP, LD50	Basal OCR	$p < 0.0001$ **-74%**	$p < 0.0001$ **-92%**	$p < 0.0001$ **-89%**	$p < 0.0001$ **-96%**	NS	$p < 0.0001$ **-79%**
	Maximal OCR	$p < 0.0001$ **-67%**	$p < 0.0001$ **-96%**	$p < 0.0001$ **-96%**	$p < 0.0001$ **-98%**	$p < 0.0001$ **-88%**	$p < 0.0001$ **-88%**
	ATP-linked OCR	$p < 0.0001$ **-101%**	$p < 0.0001$ **-97%**	$p < 0.0001$ **-96%**	$p < 0.0001$ **-95%**	$p = 0.0020$ **-71%**	$p < 0.0001$ **-100%**
	Proton leak	NS	$p < 0.0001$ **-85%**	$p < 0.0001$ **-68%**	$p < 0.0001$ **-100%**	$p = 0.0034$ **+147%**	NS
	Spare capacity	$p = 0.0211$ **-64%**	$p < 0.0001$ **-100%**	$p < 0.0001$ **-100%**	$p < 0.0001$ **-100%**	$p < 0.0001$ **-105%**	$p < 0.0001$ **-96%**
	Coupling efficiency	$p < 0.0001$ **-116%**	$p = 0.0161$ **-31%**	$p = 0.0017$ **-19%**	$p < 0.0001$ **-45%**	$p = 0.0002$ **-105%**	$p < 0.0001$ **-129%**
	ECAR	NS	$p < 0.0047$ **-19%**	$p = 0.0001$ **-49%**	$p = 0.0004$ **-57%**	NS	NS
DOX, LD50	Basal OCR	$p < 0.0001$ **-54%**	$p = 0.0006$ **-51%**	$p < 0.0001$ **-68%**	$p = 0.0002$ **-81%**	$p = 0.0021$ **-35%**	NS
	Maximal OCR	$p = 0.0108$ **-30%**	$p = 0.0007$ **-62%**	$p < 0.0001$ **-77%**	$p < 0.0001$ **-77%**	$p = 0.0097$ **-52%**	NS
	ATP-linked OCR	$p < 0.0001$ **-65%**	$p = 0.0012$ **-69%**	$p = 0.0002$ **-84%**	$p = 0.0010$ **-89%**	$p = 0.0340$ **-38%**	NS
	Proton leak	NS	NS	NS	$p = 0.0090$ **-70%**	NS	NS
	Spare capacity	NS	$p = 0.0051$ **-77%**	$p = 0.0034$ **-83%**	$p = 0.0074$ **-74%**	$p = 0.0194$ **-55%**	$p = 0.0175$ **-47%**
	Coupling efficiency	$p = 0.0062$ **-30%**	$p = 0.0119$ **-55%**	NS	$p = 0.0157$ **-58%**	NS	NS
	ECAR	NS	NS	NS	$p = 0.0087$ **-48%**	$p = 0.0020$ **-34%**	NS
2-DG, LD50	Basal OCR	$p < 0.0001$ **-62%**	$p = 0.0047$ **-38%**	$p < 0.0001$ **-52%**	$p = 0.0020$ **-76%**	NS	$p < 0.0001$ **-47%**
	Maximal OCR	NS	NS	$p = 0.0017$ **-27%**	$p < 0.0001$ **-53%**	NS	$p = 0.0242$ **-52%**
	ATP-linked OCR	$p < 0.0001$ **-63%**	$p = 0.0270$ **-50%**	NS	NS	NS	$p < 0.0001$ **-71%**
	Proton leak	$p = 0.0076$ **-65%**	NS	$p = 0.0188$ **-35%**	$p = 0.0068$ **-63%**	NS	$p = 0.0004$ **+97%**
	Spare capacity	$p = 0.0017$ **+86%**	NS	NS	NS	NS	NS
	Coupling efficiency	NS	NS	NS	NS	NS	$p < 0.0001$ **-57%**
	ECAR	NS	$p < 0.0001$ **-37%**	$p = 0.0017$ **-47%**	$p = 0.0014$ **-56%**	$p < 0.0001$ **-60%**	NS
CCCP, LD25 + 2-DG, LD25	Basal OCR	$p < 0.0001$ **-81%**	$p = 0.0066$ **-42%**	$p < 0.0001$ **-83%**	NS	$p < 0.0001$ **-70%**	$p < 0.0001$ **-95%**
	Maximal OCR	$p < 0.0001$ **-52%**	$p < 0.0001$ **-72%**	$p < 0.0001$ **-89%**	$p < 0.0001$ **-61%**	$p < 0.0001$ **-94%**	$p < 0.0001$ **-96%**
	ATP-linked OCR	$p < 0.0001$ **-95%**	$p < 0.0001$ **-88%**	$p < 0.0001$ **-96%**	$p = 0.0103$ **-87%**	$p < 0.0001$ **-100%**	$p < 0.0001$ **-101%**

	Proton leak	$p = 0.0056$ **-67%**	NS	$p = 0.0038$ **-42%**	NS	NS	NS
	Spare capacity	NS	$p < 0.0001$ **-107%**	$p < 0.0001$ **-92%**	NS	$p < 0.0001$ **-99%**	$p < 0.0001$ **-98%**
	Coupling efficiency	$p = 0.0003$ **-72%**	$p < 0.0001$ **-80%**	$p < 0.0001$ **-67%**	$p = 0.0005$ **-64%**	$p = 0.0016$ **-85%**	$p < 0.0001$ **-121%**
	ECAR	NS	NS	$p < 0.0001$ **-58%**	NS	$p < 0.0001$ **-88%**	$p = 0.0095$ **-41%**

ANOVA with subsequent Dunn's or Fisher LSD test was used. Significant changes ($p<0.05$) are in bold.