

1 **Supplemental materials**

2 **Continuous PTH in Male Mice Causes Bone Loss Because It Induces Serum Amyloid A**
3 **(SAA)**

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11 **Supplemental Figure legends (Fig. 1 to Fig. 4); Supplemental Table 1 and 2; and**
12 **Supplemental Figure 1 to Figure 4.**

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14 **Legends for Supplemental figures**

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16 **Supplemental Figure 1.** Generation of SAA3 KO mice. (A) SAA3 mice were generated by
17 CRISPR/Cas9. The *Saa3* single-guided RNA (*Saa3*gRNA), Cas9 mRNA and the targeting
18 single-stranded oligo-directed nucleotides (ssODN) bearing two in-frame tandem termination
19 codons (TAA and TGA) and a *Sal1* restriction enzyme digestion site were microinjected into the
20 pro-nucleus of CD-1 one-cell embryos. Through specific CRISPR/Cas9 editing and homology-
21 directed repair, the termination codons and the *Sal1* point mutation were knockin into the first
22 coding exon of *Saa3* gene, which resulted into a founder mouse with one WT allele and one
23 knockin allele. (B and C) The heterozygous (HET) founder mouse was first identified by PCR

24 followed by SalI digestion and then confirmed by sequencing of PCR product. (D) The HET
25 founder mouse was bred with WT CD-1 mice to generate more HET mice, which were
26 genotyped by PCR, using allele specific primers as listed in the Methods section. For
27 experiments, WT and SAA3 KO mice were generated by HET x HET mating.

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29 **Supplemental Figure 2.** RANKL increased SAA3 protein expression and its secretion from WT
30 BMMs. Continuous PTH increased gene expression for *Tnfsf11* (RANKL) and *Tnfrsf11b* (OPG)
31 similarly in WT and SAA3 KO BMSC cultures. Continuous PTH increased osteoclast-like cells
32 similarly in WT and SAA3 KO bone marrow cultures. (A) Immunofluorescence for SAA3
33 protein in BMMs and ELISA of SAA3 protein in the medium of BMMs. WT and SAA3 KO
34 BMMs were treated with M-CSF+RANKL (30 ng/ml each) for 1 day. BMMs were labeled for
35 SAA3 (red) and DAPI (blue) proteins. Representative 200x magnification (scale bars: 50 μ m)
36 merged microscopic images. Und= Undetectable. (B) Osteoclast-like cells in PTH-stimulated
37 WT and SAA3 KO BMSCs cultured with vehicle (VEH) or PTH (10 nM) for 7 days.
38 Representative image of TRAP⁺ cells, marked by blue arrow, showing partial coverage by
39 canopy of cells, at 40x magnification (scale bars: 500 μ m). Expression of genes for *Tnfsf11* and
40 *Tnfrsf11b* measured on day 7 by qPCR and reported as relative quantification (RQ) values. (C)
41 Bone marrow cultures were treated with PTH (10 nM). Representative TRAP stained 100x
42 magnification (scale bars: 200 μ m) microscopic images. Data are means \pm SEM for $n=3$
43 independent samples. $**P < 0.01$, determined by 2-way ANOVA, post-hoc Bonferroni pairwise
44 multiple comparisons.

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46 **Supplemental Figure 3.** Continuous PTH stimulated prostaglandinE₂ (PGE₂) in hBMSCs.
47 Human BMSCs were cultured in osteoblast differentiation medium in the presence of vehicle
48 (VEH) or PTH (10 nM) ± NS398 (100 nM), a selective inhibitor of COX2 activity. At each
49 medium change, the medium was collected and assessed for PGE₂ by ELISA. Nd=Not
50 detectable. In this dataset, the data are from the same experiment as Figure 6A but expanded to
51 show that NS398 blocked all PGE₂ production. Bars are means ± SEM for *n*=3 independent
52 samples. ***P* < 0.01, determined by 1-way ANOVA, post-hoc Bonferroni pairwise multiple
53 comparisons.

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55 **Supplemental Figure 4.** Addition of conditioned medium (CM) from RANKL-treated hBMMs,
56 but not from RANKL + NS398 treated hBMMs, inhibited PTH-stimulated osteoblast
57 differentiation in hBMSC cultures. *BGLAP* mRNA expression in hBMSC cultures at day 21.
58 hBMSCs were cultured in the presence of OPG (100 ng/ml), vehicle (VEH) or PTH (10 nM) ±
59 CM from hBMMs. hBMMs were treated with M-CSF+RANKL (30 ng/ml each) in the absence
60 or presence of NS398 (100 nM) for 8 days. CM from days 4-6 of hBMM cultures were pooled
61 and added to hBMSCs as 3 parts CM and 1 part osteoblast differentiation medium. Bars are
62 means ± SEM for *n*=3 independent samples. ***P* < 0.01, determined by 1-way ANOVA, post-
63 hoc Bonferroni pairwise multiple comparisons.

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68 **Supplemental Table 1. Primers used for quantitative real-time PCR (qPCR)**

Gene Name	Taqman[®] probe number	Gene Name	Taqman[®] probe number
<i>Bmp2</i>	Mm01340178_m1	<i>BMP2</i>	Hs00154192_m1
<i>Dkk1</i>	Mm00438422_m1	<i>DKK1</i>	Hs00183740_m1
<i>Gapdh</i>	Mm99999915_g1	<i>GAPDH</i>	Hs02758991_g1
<i>Igf1</i>	Mm00439560_m1	<i>IGF1</i>	Hs01547656_m1
<i>Runx2</i>	Mm00501584_m1	<i>RUNX2</i>	Hs01047973_m1
<i>Saa1</i>	Mm00656927_g1	<i>SAA1</i>	Hs00761940_s1
<i>Saa2/Saa1</i> *	Mm04208126_mH	<i>SAA2</i>	Hs01667582_m1
<i>Tnfsf11</i> (RANKL)	Mm00441908_m1	<i>TNFSF11</i> (RANKL)	Hs00243522_m1
<i>Wnt10b</i>	Mm00442104_m1	<i>WNT10B</i>	Hs00559664_m1
<i>Ctnnb1</i> (catenin, beta 1)	Mm00483039_m1	<i>ALPL</i> (alkaline phosphatase)	Hs01029144_m1
<i>Ptgs2</i> (COX2)	Mm00478374_m1	<i>BGLAP</i> (osteocalcin)	Hs01587814_g1
<i>Pthlh</i> (Pthrp)	Mm00436057_m1		
<i>Ramp3</i>	Mm00840142_m1		
<i>Saa3</i>	Mm00441203_m1		
<i>Sost</i>	Mm00470479_m1		
<i>Tnfrsf11b</i> (OPG)	Mm01205928_m1		
<i>Wnt4</i>	Mm01194003_m1		
<i>Wnt7b</i>	Mm01301717_m1		
Gene Name	SYBR Green validated primer sequence		
<i>Bglap</i> (osteocalcin)	Reverse TGG TCT GAT AGC TCG TCA CAA G Forward CTG ACC TCA CAG ATC CCA AGC		
<i>Actb</i> (actin, beta)	Reverse CCA GTT GGT AAC AAT GCC ATG T Forward GGC TGT ATT CCC CTC CAT CG		

69 *The gene assay ID Mm04208126_mH for *Saa2* detected both *Saa2* (RefSeq NM_011314.2) and
70 *Saa1* (RefSeq NM_009117.3) transcripts.
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72 **Supplemental Table 2. Mendelian ratios of WT and SAA3 KO mice**

Genotype	Males (368)		Females (360)	
	Number	Ratio	Number	Ratio
Wild type (WT)	88	0.96	90	1.00
SAA3 knockout (KO)	90	0.98	92	1.02
SAA3 heterozygous (HET)	190	2.06	178	1.98

73 All experimental SAA3 KO and WT mice were obtained by HET x HET mating. All three
74 genotypes were obtained in the expected Mendelian ratios.

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