

1 **Neurobiological substrates underlying the effect of genomic risk for**  
2 **depression on conversion of amnesic mild cognitive impairment**

3 **Author names and affiliations:** Jiayuan Xu <sup>1,8</sup>, Qiaojun Li <sup>2,8</sup>, Wen Qin <sup>1, 8</sup>,  
4 Mulin Jun Li <sup>3</sup>, Chuanjun Zhuo <sup>1,4</sup>, Huaigui Liu <sup>1</sup>, Feng Liu <sup>1</sup>, Junping Wang <sup>1</sup>,  
5 Gunter Schumann <sup>5,6</sup>, Chunshui Yu <sup>1, 7\*</sup>, Alzheimer's Disease Neuroimaging  
6 Initiative.

7 <sup>1</sup> Department of Radiology and Tianjin Key Laboratory of Functional Imaging,  
8 Tianjin Medical University General Hospital, Tianjin 300052, P.R. China

9 <sup>2</sup> College of Information Engineering, Tianjin University of Commerce, Tianjin  
10 300052, P.R. China

11 <sup>3</sup> Collaborative Innovation Center of Tianjin for Medical Epigenetics, Tianjin  
12 Key Laboratory of Medical Epigenetics, Department of Pharmacology, Tianjin  
13 Medical University, Tianjin 300052, P.R. China.

14 <sup>4</sup> Department of Psychiatry Functional Neuroimaging Laboratory, Tianjin  
15 Mental Health Center, Tianjin Anding Hospital, Tianjin 300052, P.R. China

16 <sup>5</sup> Institute of Psychiatry, Psychology and Neuroscience, King's College London,  
17 London SE5 8AF, United Kingdom

18 <sup>6</sup> Medical Research Council Social, Genetic and Developmental Psychiatry  
19 Centre, London SE5 8AF, United Kingdom

1 <sup>7</sup> CAS Center for Excellence in Brain Science and Intelligence Technology,  
2 Chinese Academy of Sciences, Shanghai, 200031, P.R. China

3 <sup>8</sup>These authors contributed equally to this work.

4 **\*Correspondence to:** Chunshui Yu, Ph.D., Department of Radiology, Tianjin  
5 Medical University General Hospital, No. 154, Anshan Road, Heping District, Tianjin  
6 300052, China, E-mail: [chunshuiyu@tjmu.edu.cn](mailto:chunshuiyu@tjmu.edu.cn). Phone: +86-22-60362026; Fax:  
7 +86-22-60362290

8

9 **Content:**

- 10 1. Supplementary Method  
11 2. Supplementary Results  
12 3. Supplementary Figures S1-S2  
13 4. Supplementary Tables S1-S11  
14 5. References

15

16

## Supplementary Method

### Discovery sample

This study included two discovery samples. The first discovery sample was used to calculate the  $PRS_{MDD}$ , including 9240 MDD patients and 9519 controls provided by the Psychiatric Genomics Consortium (PGC) (Sullivan, 2010). All patients met the criteria for a lifetime history of MDD based on the Diagnostic and Statistical Manual of Mental Disorders, 4th edition (DSM-IV). The genetic summary data from this sample were used to identify the MDD risk variants, reference allele,  $P$  values, and odd ratios (OR) (<http://www.med.unc.edu/pgc/results-and-downloads>) (Ripke *et al.*, 2013).

The second discovery sample was used for calculating  $PRS_{AD}$ . The International Genomics of Alzheimer's Project (IGAP) (Lambert *et al.*, 2013) is a large, two-stage study based on the GWAS of individuals with European ancestry. In this study, we only used the results of stage 1, which has been used the genotyped and imputed data of 7,055,881 SNPs to meta-analyze four previously published GWAS datasets consisting of 17,008 AD cases and 37,154 controls. The summary meta-GWAS statistics from the IGAP (stage 1) were used to identify the information of SNPs that are associated with the risk for AD ([http://web.pasteur-lille.fr/en/recherche/u744/igap/igap\\_download.php](http://web.pasteur-lille.fr/en/recherche/u744/igap/igap_download.php)).

### Target sample

The target sample included 398 patients with aMCI provided by the first stage of Alzheimer's disease Neuroimaging Initiative (ADNI-1) (<http://www.adni-info.org>).

1 Diagnosis of aMCI was made according to the criteria by Petersen (Petersen *et al.*,  
2 1999). After excluding 14 patients only with the baseline data and 9 patients who  
3 were diagnosed as normal during the follow-up evaluation, the remaindering 375  
4 aMCI patients were included in the following analysis. According to the follow-up  
5 results in July 2015 released by ADNI, aMCI subjects were divided into the  
6 conversion (aMCI-C, N= 205) and stable (aMCI-S, N=170) groups. Here, the final  
7 diagnosis for patients with follow-up loss was based on the last clinical evaluation.  
8 The sample was used to test whether PRS could predict the conversion from aMCI to  
9 AD (up to 108 months follow up).

#### 10 **Quality control for individual level**

11 For the 757 subjects from ADNI-1, the quality control was performed using the  
12 PLINK version 1.90 beta3 (<http://www.cog-genomics.org/plink2/>) (Purcell *et al.*, 2007;  
13 Chang *et al.*, 2015). We firstly removed 1 subject with a missing genotype rate of  
14 greater than 0.05. Then we removed 2 subjects with sex inconsistency based on the X  
15 chromosome information. After that, we identified individuals with possible relative  
16 relationships by using the estimate of pairwise identity-by-descent (IBD) to find pairs  
17 of individuals who had more similar genotypes than we would have expected by  
18 chance in a random sample and removed the one of each pair who had the greater  
19 missing genotype rate (3 subjects were excluded). The resulting SNP sets was then  
20 used to calculate multidimensional scaling (MDS) to assess the population  
21 stratification with HapMap phase 3 genetic data as the reference set, 54 participants  
22 were excluded from the sample as European population outliers. The first 4

1 components of MDS analysis were controlled in the subsequent analysis.

## 2 **Quality control at SNP level**

3 We applied SNP-level filtering to remove SNPs with missing call rates greater than  
4 0.05, a minor allele frequency (MAF) less than 0.01, and significant deviation from  
5 Hardy-Weinberg Equilibrium ( $P < 5.00e-6$ ). Strand ambiguous SNPs were also  
6 removed. After individual- and SNP-level quality control, we retained 697 individuals  
7 with European ancestry (419 males) with a genotyping rate of 99.66% for 521,695  
8 SNPs.

## 9 **Imputation**

10 The MaCH (Li *et al.*, 2010) (<http://www.sph.umich.edu/csg/abecasis/MACH>) was  
11 used for haplotype phasing and the MiniMac (Howie *et al.*, 2012)  
12 (<http://genome.sph.umich.edu/wiki/Minimac>) was used for imputation with the 1000  
13 Genomes Phase 1 version 3 CEU as the reference dataset (hg19). Finally, 7,747,882  
14 autosomal SNPs with imputation quality score greater than 0.8 ( $R^2 > 0.8$ ) were used  
15 for further analysis.

## 16 **GMV calculation**

17 All structural images were visually checked by two experimenters of radiology. In the  
18 697 subjects with qualified genetic data, we removed 19 subjects because of poor  
19 image quality. Finally, a total of 322 aMCI patients were finally included in the  
20 voxel-based morphometry (VBM) analysis. The GMV maps were calculated using the  
21 VBM8 implemented in Statistical Parametric Mapping software package (SPM8,  
22 <http://www.fil.ion.ucl.ac.uk/spm>). In the segmentation of VBM8, an adaptive

1 Maximum A Posterior technique (Rajapakse *et al.*, 1997) and a Partial Volume  
2 Estimation (Tohka *et al.*, 2004) were used to estimate the fraction of each pure tissue  
3 type present in every voxel. After the structural images were segmented into gray  
4 matter (GM), white matter and cerebrospinal fluid, the individual's GM concentration  
5 map was normalized into the Dartel template in Montreal Neurological Institute (MNI)  
6 space (<http://www.mni.mcgill.ca/>). This template was derived from 550 healthy  
7 control subjects of the IXI-database (<http://www.brain-development.org>). In the  
8 modulated normalized process, we multiplied the individual's GM concentration map  
9 only by the non-linear determinants derived from the spatial normalization procedure.  
10 This step resulted in normalized GM density or relative GMV map for each subject.  
11 Here, the GMV of each voxel represents the fraction of GM present in each voxel,  
12 which preserves the local GM density while removing the confounding effect of  
13 variance in individual brain sizes. After that, we resliced the normalized GMV to a  
14 1.5-mm cubic voxel. Finally, the GMV images were smoothed with a kernel of  $8 \times 8$   
15  $\times 8 \text{ mm}^3$  full width at half maximum. Then, the spatial pre-processing, normalized,  
16 modulated, and smoothed GMV maps were used for further analysis.

### 17 **LD score regression and colocalization analysis**

18 To validate the specificity of these index SNPs to MDD but not to AD, we performed  
19 LD score regression (Bulik-Sullivan *et al.*, 2015) and colocalization analyses  
20 (Giambartolomei *et al.*, 2014; Pickrell *et al.*, 2016). LD score regression was applied  
21 to quantify genetic correlation pattern ( $r_g$ ) between GWAS summary statistics of the  
22 1,806 MDD-specific index SNPs from PGC-MDD dataset and GWAS summary

1 statistics from IGAP-AD datasets. By estimating the Bayesian posterior probability,  
2 the colocalization analysis was used to test the hypothesis that the 1,806  
3 MDD-specific index SNPs are associated with MDD, but not with AD. The Bayesian  
4 posterior probability  $> 0.90$  was used as a cutoff threshold.

### 5 **Mediation analysis and Mendelian Randomization (MR)**

6 The SPSS macro (<http://www.afhayes.com/spss-sas-and-mplus-macros-and-code.html>)  
7 (Preacher and Hayes, 2008; Hayes, 2013) was used to perform the mediation analysis  
8 to test whether the GMV of each significant brain region mediates the association  
9 between the  $PRS_{sMDD}$  and the status of aMCI-C. The  $PRS_{sMDD}$  was defined as an  
10 independent variable, the GMV of each significant brain region as a mediator variable,  
11 and the aMCI group assignment (aMCI-S vs aMCI-C) as a binary dependent variable.  
12 The first step was to confirm that the independent variable ( $PRS_{sMDD}$ ) was a predictor  
13 of the dependent variable (aMCI-S vs aMCI-C), which is known as the direct effect.  
14 The second step was to confirm that the independent variable ( $PRS_{sMDD}$ ) was a  
15 predictor of the mediator (GMV). The third step was to confirm that the mediator  
16 (GMV) was a predictor of the dependent variable (aMCI-S vs aMCI-C), while  
17 controlling for the independent variable ( $PRS_{sMDD}$ ). The indirect effect is the product  
18 of path coefficients of the last two steps. Then the bootstrapping method was used to  
19 assess the significance of the mediation effect. After 5000 bias-corrected  
20 bootstrapping, we could estimate the distribution of the indirect effect and calculate  
21 its 95% confidence intervals (CI). If zero does not fall between the resulting 95%  
22 confidence interval of the bootstrapping method, we could confirm the existence of a  
23 significant mediation effect ( $P < 0.05$ ).

24

1 Mendelian randomization (MR) is a statistical technique that uses genetic variants  
2 associated with modifiable exposure as instrumental variables to infer the causal  
3 effect of the exposure on outcome under several assumptions, which can overcome  
4 confounders and reverse causality (Smith and Ebrahim, 2003; Davey Smith and  
5 Hemani, 2014). The pleiotropy may influence on the validity of causal estimates  
6 derived from MR methods. It has been suggested that the conventional MR analysis is  
7 valid when there is vertical pleiotropy or balanced horizontal pleiotropy. However,  
8 when there is unbalanced horizontal pleiotropy, the conventional MR analysis may  
9 generate false positive or false negative estimate. In this situation, MR-Egger method  
10 could correct for unbalanced horizontal pleiotropy and yield a valid estimate (see  
11 Figure 1 in (White *et al.*, 2016) for a recent pictorial description of vertical and  
12 balanced/unbalanced horizontal pleiotropy) (Bowden *et al.*, 2015).

13 Here, the conventional two-stage method of MR analysis was applied using  
14  $PRS_{sMDD}$  as instrumental variable (G) to make causal inference between left  
15 hippocampal volume (X) and aMCI conversion (Y) (Burgess and Thompson, 2013;  
16 Burgess, 2014; Burgess and Thompson, 2015)  
17 (<http://www.mendelianrandomization.com/index.php/software-code>). A total of 1806  
18 index SNPs specific to MDD ( $r^2 < 0.25$  within 250 kb window) were included in  
19 calculation of the  $PRS_{sMDD}$  after excluding genetic variants common to  $PRS_{MDD}$  ( $P_T =$   
20 0.009) and  $PRS_{AD}$  ( $P_T = 0.352$ ). In the first stage, the regression of X on G is fit,  
21 which creates a predicted value ( $\hat{X}$ ) of X for each G using equation (1). In the second  
22 stage, a logistic regression is fit in which the binary outcome is Y (aMCI-S and



1 aMCI-C) and the independent variable is  $\hat{X}$  using equation (2).

$$2 \quad X = \beta_{XG}G + e_X \quad (1)$$

$$3 \quad \ln(P(Y = 1)) = \beta_1 \hat{X} + e_Y \quad (2)$$

4 To investigate potential bias due to unbalanced horizontal pleiotropy of PRS<sub>sMDD</sub>  
5 on the conversion of aMCI, MR Egger regression method was applied in sensitivity  
6 analysis (Bowden *et al.*, 2015)  
7 (<http://www.mendelianrandomization.com/index.php/software-code>). If the estimated  
8 intercept of this method differs from zero, this provides evidence that there is  
9 unbalanced horizontal pleiotropy. And the slope coefficient from MR Egger regression  
10 ( $\beta_{\text{Egger}}$ ) estimated the causal effect even the presence of unbalanced horizontal  
11 pleiotropy (Burgess and Thompson, 2017).

12

## Supplementary Results

### **PRS<sub>sMDD</sub> and PRS<sub>tsAD</sub> predict the conversion from aMCI to AD**

After excluding the overlapping SNPs ( $n=4,711$ ) between the PRS<sub>MDD</sub> ( $P_T = 0.009$ ) and PRS<sub>AD</sub> ( $P_T = 0.009$ ), the PRS<sub>sMDD</sub> and PRS<sub>sAD</sub> could significantly predict the aMCI-C ( $P = 0.002$  and  $P = 1.05e-9$ ). To balance the number of index SNPs used to construct the PRS, we also created the PRS<sub>AD</sub> for the top 1,806 AD-specific index SNPs (PRS<sub>tsAD</sub>). Although the PRS<sub>tsAD</sub> could significantly predict the status of aMCI-C ( $P = 0.013$ ), it could only explain 2.63% of variance for the aMCI-C (Table S5). In addition, under the same  $P_T$  ( $P_T = 0.009$ ) as the PRS<sub>MDD</sub>, the PRS<sub>AD</sub> could also predict the aMCI-C ( $P = 0.046$ ) and explained 1.80% of variance for the aMCI-C based on 2,554 index SNPs (Table S5). After excluding the overlapping SNPs ( $n=184$ ) between the PRS<sub>MDD</sub> and PRS<sub>AD</sub> under the same  $P_T$  ( $P_T = 0.009$ ), only the PRS<sub>sMDD</sub> could significantly predict the aMCI-C ( $P = 7.51e-5$ ), and explained 6.86% of variance for the aMCI-C (Table S5). These results indicate that the conversion from aMCI to AD is related to only a small number of MDD-specific genetic variants but to a large number of AD-specific genetic variants.

### **Ten-fold cross validation**

For each PRS, we used ten-fold cross validation to test the unbiased prediction accuracy of the PRS on the conversion of aMCI. That is, we randomly divided the 322 aMCI patients into ten disjoint sets (eight sets: each included 32 patients; two set: each included 33 patients), and then used nine of the sets as training data ( $N =$

1 289 or 290) and the remaining one set as test data ( $N = 33$  or  $32$ ). We repeated this  
2 process ten times using different possible combinations of training and test sets.  
3 For each validation test, we calculated the accuracy of the test set (32 or 33 aMCI  
4 patients). For each PRS, the mean accuracy and coefficient of variance (CV) of  
5 validation tests are shown in Table S2.

### 6 **Conversion rates of aMCI in the trisected PRS groups**

7 When  $PRS_{sMDD}$  and  $PRS_{sAD}$  were trisected into the low, middle and high risk. There  
8 were significant differences in conversion rate among the 9 hierarchical PRS groups  
9 ( $P = 3.23e-6$ ). In the middle  $PRS_{sAD}$  group, aMCI patients with high  $PRS_{sMDD}$  showed  
10 significantly higher conversion rate than those with low  $PRS_{sMDD}$  (59.38% vs 31.58%,  
11  $P = 0.022$ ). In the high  $PRS_{sAD}$  group, the aMCI patients with high  $PRS_{sMDD}$  showed  
12 higher conversion rate than those with low  $PRS_{sMDD}$  (89.65% vs 72.97%,  $P = 0.047$ )  
13 (Table S6).

### 14 **Enrichment analyses using genes fine-mapped based on physical position**

15 We remapped the 8,762 SNPs calculated for  $PRS_{sMDD}$  into 1,608 genes only based on  
16 the physical position of each variant (within 5kb window). The specific enrichment  
17 results for the 1,608 genes are as the follows: in the annotation of gene ontology,  
18 540/1,608 genes were enriched in the development process, 910/1,608 in the protein  
19 binding, and 783/1,608 in membrane part (Figure S2A and Table S7). Specifically,  
20 these genes mainly over-represented in biological processes of the anatomical  
21 structure morphogenesis ( $q_c = 5.27e-5$ , FDR-BH correction), development process ( $q_c$   
22  $= 1.83e-4$ , FDR-BH correction) and cellular developmental process ( $q_c = 7.48e-4$ ,

1 FDR-BH correction) (Figure S2B), in the molecular function of the amyloid-beta  
2 binding ( $q_c = 9.40e-5$ , FDR-BH correction) (Figure S2C), and in the neuron part ( $q_c =$   
3  $8.13e-11$ , FDR-BH correction) and neuron projection ( $q_c = 2.96e-5$ , FDR-BH  
4 correction) (Figure S2C). And in the KEGG pathway analysis, these genes were also  
5 significantly enriched in neuronal development-related axon guidance ( $q_c = 7.95e-3$ ,  
6 FDR-BH correction) (Figure S2C and Table S9).

7 In the PPI network analysis, we mapped the PRS<sub>sMDD</sub> fine-mapping 1,608 genes  
8 to the PPI network of the BIOGRID (315 unmatched genes were excluded). A tightly  
9 connected PPI sub-network was instituted by 928 genes from the remaining 1,293  
10 genes. Using NTA, 928 seed genes and top ten neighboring genes were included in  
11 the construction of the final PPI network consisting of 938 genes. APP was also  
12 identified as the most functionally neighboring genes (Figure S2D). Table S10  
13 showed that the 938 genes of the final PPI network were significantly enriched in  
14 various nervous system related development processes (Figure S2D), such as nervous  
15 system development ( $q_c = 9.91e-6$ , FDR-BH correction), neuron projection  
16 development ( $q_c = 3.78e-5$ , FDR-BH correction), neuron development ( $q_c = 1.01e-4$ ,  
17 FDR-BH correction) and generation of neurons ( $q_c = 4.11e-4$ , FDR-BH correction).  
18 The PPI-based enrichment analysis further confirmed that the PRS<sub>sMDD</sub> fine-mapping  
19 1,608 genes also involved in the developmental process and amyloid beta binding.

20 We further explored in which the developmental periods these 1,608 genes were  
21 over-represented in the hippocampus. Under a pSI threshold of 0.05, 65 genes showed  
22 temporal-specific expression in the hippocampus in the middle-late fetal

1 developmental period ( $q_c = 0.002$ , FDR-BH correction) (Figure S2E). Under the most  
2 stringent threshold ( $pSI = 0.001$ ), CDC20B also showed temporal-specific high  
3 expression in the hippocampus in the middle-late fetal stage, which is well consistent  
4 with the finding (CDC20B) from the 1,860 genes obtained based on expression  
5 patterns in the hippocampus (Figure S2E).

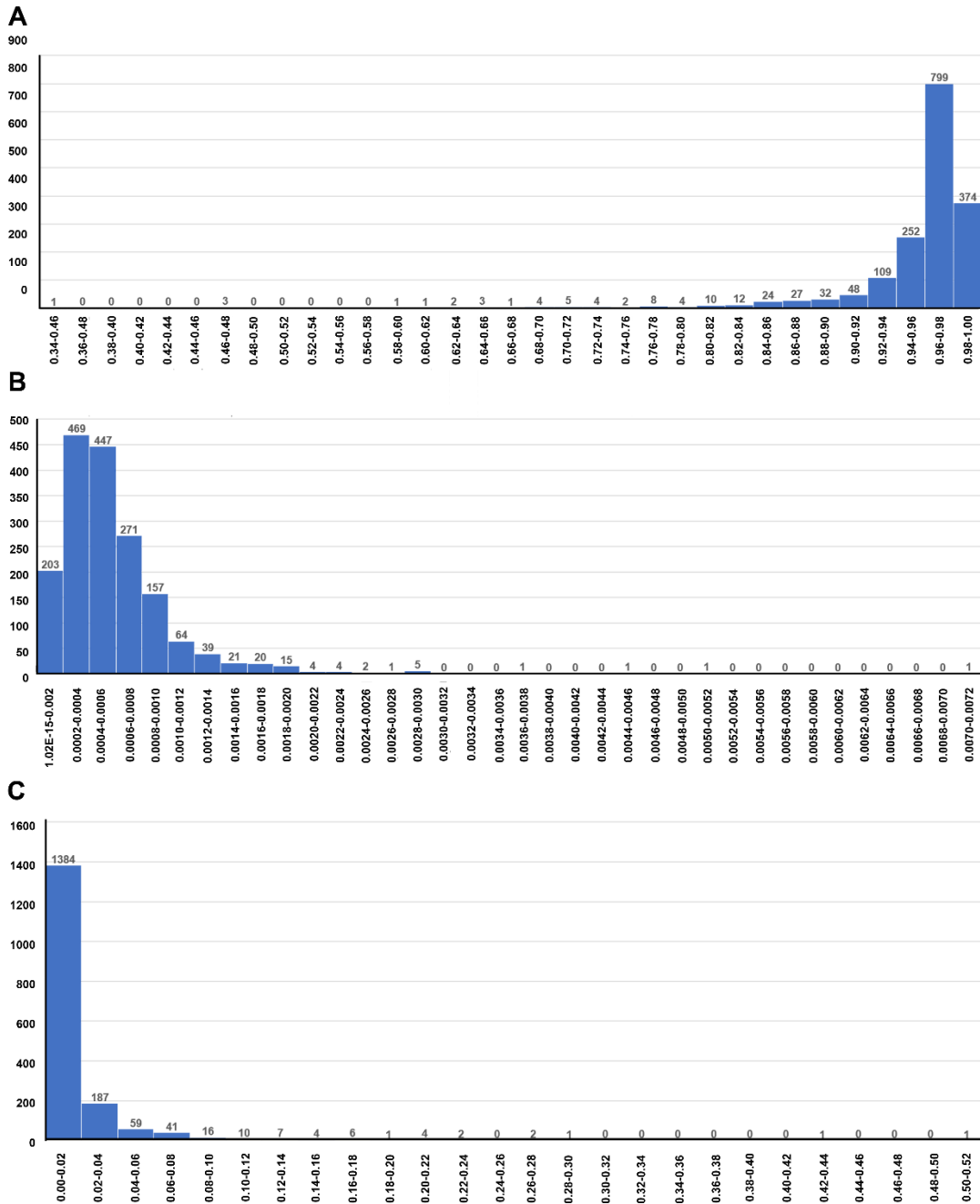
6

7

8

1

### Supplementary Figures



2

3 **Figure S1. Histograms of Bayesian posterior probability for 1,806 index SNPs.**

4 Using PP = 0.90 as a cutoff threshold, there are 1582/1,806 (87.5%) index SNPs are

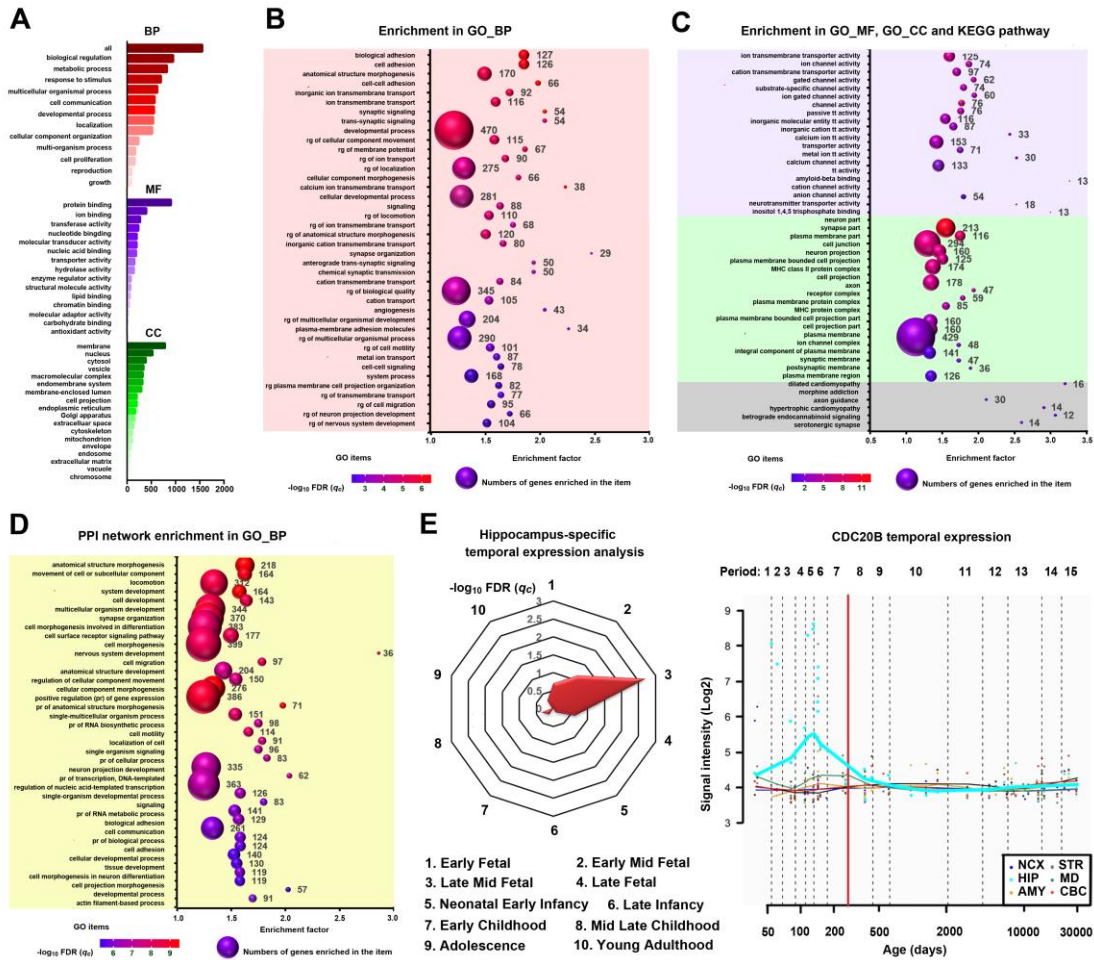
5 highly associated with MDD (A), but none SNPs are associated with AD (B), and

6 none SNPs show colocalization between MDD and AD (C). The X-axis represents the

7 intervals of Bayesian posterior probability and Y-axis denotes the number of index

8 SNPs within each interval.

9



**Figure S2. Gene enrichment of the PRS<sub>MDD</sub> fine-mapping 1,608 genes.** (A) Enrichment of the PRS<sub>MDD</sub> genes in GO items. The X-axis shows the numbers of genes enriched in each item (Y-axis). The red, purple, green bars denote the biological process, molecular function and cellular component, respectively. (B) Top 40 significant enriched GO BP items of the PRS<sub>MDD</sub> genes. The X-axis shows enrichment factor of each GO item (Y-axis). The size of balls shows the numbers of genes enriched in each item (numbers of genes are labeled beside the balls). The color of balls demonstrates the significance of the enrichment analysis. (C) Top 20 significant enriched GO MF and CC items, and all significant KEGG pathway items of the PRS<sub>MDD</sub> genes. The purple, green and grey ground colors show the MF, CC and KEGG pathway, respectively. (D) Top 40 significant enriched GO BP items of PPI network. (E) Left, PRS<sub>MDD</sub> genes were highly expressed in the middle-late fetal developmental period in the hippocampus; right, CDC20B was highly expressed in the early and late mid-fetal developmental periods in the hippocampus. Period 1-15

1 have been described in Table S11. AMY, amygdala; BP, biological process; CBC,  
2 cerebellar cortex; CC, cellular component; CDC20B, cell division cycle 20B; cp, cell  
3 projection; GO, gene ontology; HIP, hippocampus; MF, molecular function; MD,  
4 mediodorsal nucleus of the thalamus; NCX, neocortex; PPI, protein-protein  
5 interaction; pr, positive regulation; rg, regulation; STR, striatum; tt, transmembrane  
6 transporter.

7



1  
2  
3  
4  
5

## Supplementary Tables

**Table S1. Quality control of aMCI patients**

Steps	Exclusion No.	Reasons for exclusion	Remainder No.
0	0	The total number of aMCI patients	398
1	14	Only with the baseline data	384
2	9	With a normal diagnosis at the follow-up	375
3	9	Without genotyping data	366
4	1	Sex inconsistency	365
5	30	European population outliers	335
6	13	Poor image quality	322

1 **Table S2. The predictive accuracies of PRS measures on aMCI conversion in**  
2 **ten-fold cross validation**

Fold No.	PRS <sub>AD</sub> (N = 322)		PRS <sub>sAD</sub> (N = 322)		PRS <sub>MDD</sub> (N = 322)		PRS <sub>sMDD</sub> (N = 322)		PRS <sub>sMDD+AD</sub> (N = 322)	
	Training*	Test	Training*	Test	Training*	Test	Training*	Test	Training*	Test
1	71.25%	66.26%	68.91%	67.24%	65.69%	64.38%	61.38%	60.75%	71.72%	70.63%
2	71.72%	67.10%	68.07%	65.63%	63.45%	60.42%	62.76%	56.88%	78.13%	70.24%
3	70.02%	70.15%	72.45%	63.13%	65.34%	64.34%	61.72%	60.62%	74.24%	70.13%
4	68.17%	68.56%	72.79%	66.67%	65.17%	62.21%	60.55%	59.23%	74.17%	68.97%
5	71.88%	70.36%	72.79%	68.97%	65.34%	60.34%	61.03%	62.88%	69.97%	68.63%
6	70.13%	66.34%	72.38%	68.97%	65.34%	59.75%	62.76%	62.52%	69.97%	69.34%
7	69.31%	65.62%	68.41%	65.63%	65.34%	64.34%	62.41%	59.13%	79.31%	70.63%
8	70.69%	63.56%	68.14%	65.63%	62.41%	60.13%	62.41%	60.13%	74.69%	69.34%
9	70.21%	67.65%	68.14%	68.28%	62.07%	59.98%	62.76%	60.13%	72.28%	69.70%
10	70.24%	66.89%	68.36%	68.70%	64.59%	60.58%	62.63%	60.61%	74.24%	72.63%
M	70.36%	67.25%	70.04%	66.89%	64.47%	61.65%	62.04%	60.29%	73.87%	70.02%
CV	0.02	0.02	0.03	0.03	0.02	0.03	0.01	0.03	0.04	0.02

3 \* The 322 aMCI patients are divided into ten sets, 8 sets include 32 patients for each and 2  
4 sets include 33 patients for each. Thus, the training data include 289 or 290 aMCI patients and  
5 the corresponding test data include 33 or 32 patients.

6 AD, Alzheimer's disease; aMCI, amnesic mild cognitive impairment; CV, coefficient of  
7 variance; MDD, major depressive disorder; PRS, polygenic risk scores; PRS<sub>AD</sub>, PRS for  
8 AD-related genetic variants; PRS<sub>sAD</sub>, PRS for AD-specific genetic variants; PRS<sub>MDD</sub>, PRS for  
9 MDD-related genetic variants; PRS<sub>sMDD</sub>, PRS for MDD-specific genetic variants; PRS<sub>sMDD+AD</sub>,  
10 PRS for MDD-specific and AD-related genetic variants.

11

1 **Table S3. Top ten known AD locus from AD GWAS meta-analysis <sup>a</sup>**

Rank	Gene	SNP	Ethnicity	OR (95% CI)	P value	Sample size
1	APOE	APOE_ e2/3/4	All	3.685 (3.30-4.12)	<1.00E-50	4,167
2	BIN1	rs744373	All	1.166 (1.13-1.20)	1.59E-26	49,650
3	CLU	rs11136000	Caucasian	0.879 (0.86-0.90)	3.37E-23	72,432
4	ABCA7	rs3764650	All	1.229 (1.18-1.28)	8.17E-22	60,569
5	CR1	rs3818361	Caucasian	1.174 (1.14-1.21)	4.72E-21	47,052
6	PICALM	rs3851179	Caucasian	0.879 (0.86-0.92)	2.85E-20	44,358
7	MS4A6A	rs610932	All	0.904 (0.88-0.93)	1.81E-11	63,026
8	CD33	rs3865444	All	0.893 (0.86-0.93)	2.04E-10	37,767
9	MS4A4E	rs670139	All	1.079 (1.05-1.11)	9.51E-10	64,577
10	CD2AP	rs9349407	All	1.117 (1.08-1.16)	2.75E-09	35,840

2 AD, Alzheimer's disease; CI, confidence interval; SNP, single-nucleotide polymorphisms;  
3 OR, odd ratios; GWAS, genome-wide association analysis.

4 <sup>a</sup> These data are from <http://www.alzgene.org/TopResults.asp>.

5

6

1 **Table S4. The predictive effects of PRS on the conversion of aMCI (N=322) after**  
2 **removing SNPs located in the top ten AD loci**

	$P_T$	SNPs	iSNPs	$P$	$R^2$	Sp	Se	Ac	AUC
<b>PRS<sub>AD</sub></b>	0.352	2,622,829	49,829	7.93e-10	18.44%	71.84%	87.17%	80.75%	0.72
<b>PRS<sub>MDD</sub></b>	0.009	13,462	2,559	7.49e-5	6.86%	68.89%	83.43%	77.33%	0.65
<b>PRS<sub>sAD</sub></b>	NA	2,618,124	49,504	1.05e-9	18.09%	70.24%	85.63%	80.12%	0.70
<b>PRS<sub>sMDD</sub></b>	NA	8,756	1,806	1.74e-3	4.19%	65.11%	81.74%	76.01%	0.62
<b>PRS<sub>sMDD+AD</sub></b>	NA	2,631,585	50,523	2.26e-10	18.71%	70.37%	88.22%	80.86%	0.75

3 Ac, accuracy; AD, Alzheimer's disease; aMCI, amnesic mild cognitive impairment; AUC,  
4 area under curve of receiver operating characteristic curve; iSNPs, numbers of index  
5 single-nucleotide polymorphisms that constitute PRS; MDD, major depressive disorder; NA,  
6 not applicable; PRS, polygenic risk scores; PRS<sub>AD</sub>, PRS for AD-related genetic variants;  
7 PRS<sub>MDD</sub>, PRS for MDD-related genetic variants; PRS<sub>sMDD</sub>, PRS for MDD-specific genetic  
8 variants; PRS<sub>sMDD+AD</sub>, PRS for MDD-specific and AD-related genetic variants;  $P_T$ ,  $P$  values  
9 threshold of genome-wide association studies;  $R^2$ , Nagelkerke's pseudo  $R^2$  of logistic  
10 regression; Se, sensitivity; SNPs, numbers of single-nucleotide polymorphisms that constitute  
11 PRS; Sp, specificity.

12

1 **Table S5. The predictive effects of PRS measures on the conversion of aMCI**

	$P_T$	No. of SNPs	No. of iSNPs	$P$	$R^2$
PRS <sub>MDD</sub>	0.009	13,472	2,559	<b>7.49e-5</b>	<b>6.86%</b>
PRS <sub>AD</sub>	0.009	100,088	2,554	<b>4.62e-2</b>	<b>1.80%</b>
PRS <sub>tsAD</sub>	NA	NA	1,806	<b>1.30e-2</b>	<b>2.63%</b>
PRS <sub>sMDD</sub>	NA	13,286	2,535	<b>7.51e-5</b>	<b>6.86%</b>
PRS <sub>sAD</sub>	NA	99,904	3,254	1.05e-1	1.10%

2 aMCI, amnesic mild cognitive impairment; AD, Alzheimer's disease; iSNPs, index SNPs;  
3 MDD, major depressive disorder; NA, not applicable; PRS, polygenic risk scores; PRS<sub>AD</sub>, PRS  
4 for AD-related genetic variants; PRS<sub>MDD</sub>, PRS for MDD-related genetic variants; PRS<sub>sAD</sub>, PRS  
5 for AD-specific genetic variants; PRS<sub>sMDD</sub>, PRS for MDD-specific genetic variants. PRS<sub>tsAD</sub>,  
6 PRS for the top 1,806 AD-specific genetic variants;  $P_T$ ,  $P$  values threshold of genome-wide  
7 association studies;  $R^2$ , Nagelkerke's pseudo  $R^2$  of logistic regression; and SNP,  
8 single-nucleotide polymorphism.

9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24

1 **Table S6. Conversion rates of aMCI in the trisected PRS groups**

PRS groups	aMCI-S (N)	aMCI-C (N)	Conversion rate
<b>Low PRS<sub>sAD</sub> and low PRS<sub>sMDD</sub></b>	22	11	33.33%
<b>Low PRS<sub>sAD</sub> and middle PRS<sub>sMDD</sub></b>	19	14	42.42%
<b>Low PRS<sub>sAD</sub> and high PRS<sub>sMDD</sub></b>	19	21	52.50%
<b>Middle PRS<sub>sAD</sub> and low PRS<sub>sMDD</sub></b>	26	12	31.58%
<b>Middle PRS<sub>sAD</sub> and middle PRS<sub>sMDD</sub></b>	16	21	56.76%
<b>Middle PRS<sub>sAD</sub> and high PRS<sub>sMDD</sub></b>	13	19	59.38%
<b>High PRS<sub>sAD</sub> and low PRS<sub>sMDD</sub></b>	10	27	72.97%
<b>High PRS<sub>sAD</sub> and middle PRS<sub>sMDD</sub></b>	7	36	83.72%
<b>High PRS<sub>sAD</sub> and high PRS<sub>sMDD</sub></b>	3	26	89.65%
<b>All</b>	135	187	58.07%

2 AD, Alzheimer’s disease; aMCI, amnesic mild cognitive impairment; aMCI-C, conversion  
3 from aMCI to AD; aMCI-S, aMCI patients with a stable diagnosis; PRS, polygenic risk scores;  
4 PRS<sub>sAD</sub>, PRS for AD-specific genetic variants; and PRS<sub>sMDD</sub>, PRS for MDD-specific genetic  
5 variants.

6

7

1 **Table S7. Numbers of PRS<sub>sMDD</sub> fine-mapping 1,860 genes and PRS<sub>sMDD</sub>**  
 2 **fine-mapping 1,608 genes observed in the GO category**

<b>Items</b>	<b>Description</b>	<b>PRS<sub>sMDD</sub> fine-mapping 1,860 genes<sup>a</sup></b>	<b>PRS<sub>sMDD</sub> fine-mapping 1,608 genes<sup>b</sup></b>	
GO:	biological regulation	901	954	
biological process	metabolic process	777	825	
	response to stimulus	661	700	
	multicellular organismal process	590	627	
	cell communication	542	571	
	developmental process	505	540	
	localization	523	558	
	cellular component organization	480	522	
	multi-organism process	148	158	
	cell proliferation	160	168	
	reproduction	83	92	
	growth	80	85	
	unclassified	561	237	
	GO:	protein binding	855	910
	molecular function	ion binding	381	400
transferase activity		192	204	
nucleotide binding		189	202	
molecular transducer activity		140	143	
nucleic acid binding		257	273	
transporter activity		144	157	
hydrolase activity		181	189	
enzyme regulator activity		77	79	
structural molecule activity		61	71	
lipid binding		58	58	
chromatin binding		38	41	
molecular adaptor activity		23	24	
carbohydrate binding		19	17	
antioxidant activity		10	11	
unclassified		560	238	
GO:		membrane	738	783
cellular component		nucleus	491	525
		cytosol	266	295
		vesicle	302	327
	macromolecular complex	362	388	
	endomembrane system	320	341	

membrane-enclosed lumen	300	319
cell projection	194	200
endoplasmic reticulum	146	155
Golgi apparatus	127	134
extracellular space	97	104
cytoskeleton	154	172
mitochondrion	100	103
envelope	69	74
endosome	58	62
extracellular matrix	56	56
vacuole	55	60
chromosome	45	51
unclassified	528	201

1 GO, gene ontology; PRS<sub>sMDD</sub>, PRS for MDD-specific genetic variants.

2 <sup>a</sup> PRS<sub>sMDD</sub> genetic variants were fine-mapped into 1,860 genes based on the  
3 hippocampal-specific regulatory probability between eQTLs and epigenomic features (within  
4 a 5kb window)

5 <sup>b</sup> PRS<sub>sMDD</sub> genetic variants were fine-mapped into 1,608 genes based on physical position of  
6 each variant (within a 5kb window)

7

8 **Table S8. Gene enrichment analysis of PRS<sub>sMDD</sub> fine-mapping 1,860 genes**

9 **(See accompanying Excel file)**

10

11 **Table S9. Gene enrichment analysis of PRS<sub>sMDD</sub> fine-mapping 1,608 genes**

12 **(See accompanying Excel file)**

13

14 **Table S10. Gene enrichment analysis of the PPI network from PRS<sub>sMDD</sub>**  
15 **fine-mapping 1,860 genes and PRS<sub>sMDD</sub> fine-mapping 1,608 genes**

16 **(See accompanying Excel file)**

17

18

19



1 **Table S11. Period 1-15 in the temporal expression analysis**

Period	Description	Age
1	Embryonic	4PCW-8PCW
2	Early fetal	8PCM-10PCW
3	Early fetal	10PCM-13PCW
4	Early mid-fetal	13PCW-16PCW
5	Early mid-fetal	16PCW-19PCW
6	Late mid-fetal	19PCW-24PCW
7	Late fetal	24PCW-38PCW
8	Neonatal and early infancy	0M-6M
9	Late infancy	6M-12M
10	Early childhood	1Y-6Y
11	Middle and late childhood	6Y-12Y
12	Adolescence	12Y-20Y
13	Young adulthood	20Y-40Y
14	Middle adulthood	40Y-60Y
15	Late adulthood	60Y-

2 M, postnatal months; PCW, post-conceptional weeks; Y, postnatal years.

3



## 1 **References**

- 2 Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid  
3 instruments: effect estimation and bias detection through Egger regression. *Int J*  
4 *Epidemiol* 2015; 44(2): 512-25.  
5
- 6 Bulik-Sullivan BK, Loh PR, Finucane HK, Ripke S, Yang J. LD Score regression  
7 distinguishes confounding from polygenicity in genome-wide association studies.  
8 *Nat Genet* 2015; 47(3): 291-5.  
9
- 10 Burgess S. Sample size and power calculations in Mendelian randomization with a  
11 single instrumental variable and a binary outcome. *Int J Epidemiol* 2014; 43(3):  
12 922-9.  
13
- 14 Burgess S, Thompson SG. Use of allele scores as instrumental variables for Mendelian  
15 randomization. *Int J Epidemiol* 2013; 42(4): 1134-44.  
16
- 17 Burgess S, Thompson SG. *Mendelian Randomization: Methods for Using Genetic*  
18 *Variants in Causal Estimation*. Routledge Chapman & Hall 2015.  
19
- 20 Burgess S, Thompson SG. Interpreting findings from Mendelian randomization using  
21 the MR-Egger method. *Eur J Epidemiol* 2017; 32(5): 377-89.  
22
- 23 Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation  
24 PLINK: rising to the challenge of larger and richer datasets. *Gigascience* 2015; 4: 7.  
25
- 26 Davey Smith G, Hemani G. Mendelian randomization: genetic anchors for causal  
27 inference in epidemiological studies. *Hum Mol Genet* 2014; 23(R1): R89-98.  
28
- 29 Giambartolomei C, Vukcevic D, Schadt EE, Franke L, Hingorani AD, Wallace C, *et al.*  
30 Bayesian test for colocalisation between pairs of genetic association studies using  
31 summary statistics. *PLoS Genet* 2014; 10(5): e1004383.  
32
- 33 Hayes AF. *Introduction to mediation, moderation, and conditional process analysis: A*  
34 *regression-based approach*: Guilford Press; 2013.  
35
- 36 Howie B, Fuchsberger C, Stephens M, Marchini J, Abecasis GR. Fast and accurate  
37 genotype imputation in genome-wide association studies through pre-phasing. *Nat*  
38 *Genet* 2012; 44(8): 955-9.  
39
- 40 Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, *et al.*  
41 Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for  
42 Alzheimer's disease. *Nat Genet* 2013; 45(12): 1452-8.  
43
- 44 Li Y, Willer CJ, Ding J, Scheet P, Abecasis GR. MaCH: using sequence and genotype  
45 data to estimate haplotypes and unobserved genotypes. *Genet Epidemiol* 2010; 34(8):  
46 816-34.  
47
- 48 Petersen RC, Smith GE, Waring SC, Ivnik RJ, Tangalos EG, Kokmen E. Mild  
49 cognitive impairment: clinical characterization and outcome. *Arch Neurol* 1999; 56(3):

1 303-8.

2

3 Pickrell JK, Berisa T, Liu JZ. Detection and interpretation of shared genetic influences  
4 on 42 human traits. *Nat Genet* 2016; 48(7): 709-17.

5

6 Preacher KJ, Hayes AF. Asymptotic and resampling strategies for assessing and  
7 comparing indirect effects in multiple mediator models. *Behav Res Methods* 2008;  
8 40(3): 879-91.

9

10 Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, *et al.* PLINK:  
11 a tool set for whole-genome association and population-based linkage analyses. *Am J*  
12 *Hum Genet* 2007; 81(3): 559-75.

13

14 Rajapakse JC, Giedd JN, Rapoport JL. Statistical approach to segmentation of  
15 single-channel cerebral MR images. *IEEE Trans Med Imaging* 1997; 16(2): 176-86.

16

17 Ripke S, Wray NR, Lewis CM, Hamilton SP, Weissman MM, Breen G, *et al.* A  
18 mega-analysis of genome-wide association studies for major depressive disorder. *Mol*  
19 *Psychiatry* 2013; 18(4): 497-511.

20

21 Smith GD, Ebrahim S. 'Mendelian randomization': can genetic epidemiology  
22 contribute to understanding environmental determinants of disease? *Int J Epidemiol*  
23 2003; 32(1): 1-22.

24

25 Sullivan PF. The psychiatric GWAS consortium: big science comes to psychiatry.  
26 *Neuron* 2010; 68(2): 182-6.

27

28 Tohka J, Zijdenbos A, Evans A. Fast and robust parameter estimation for statistical  
29 partial volume models in brain MRI. *Neuroimage* 2004; 23(1): 84-97.

30

31 White J, Swerdlow DI, Preiss D, Fairhurst-Hunter Z, Keating BJ, Asselbergs FW, *et*  
32 *al.* Association of Lipid Fractions With Risks for Coronary Artery Disease and  
33 Diabetes. *JAMA Cardiol* 2016; 1(6): 692-9.

34

35