

Supplementary Figures

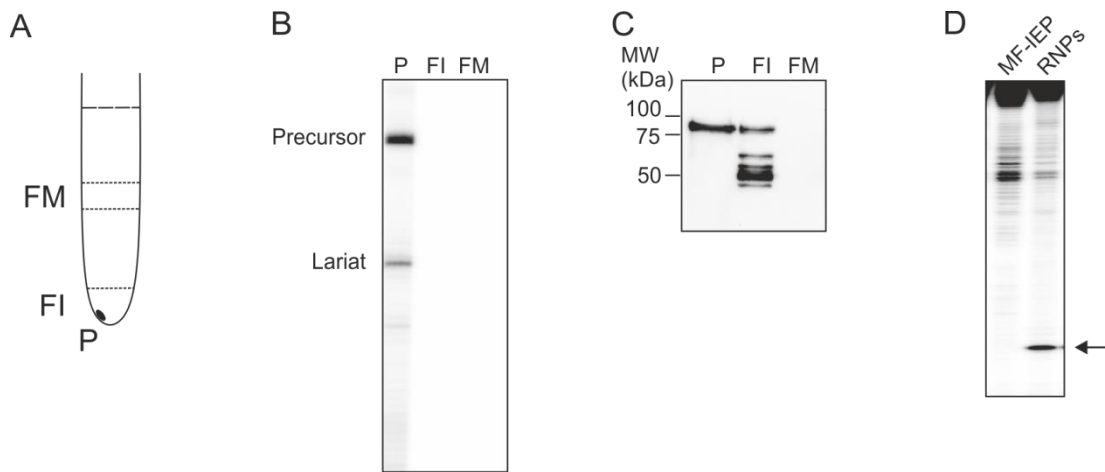


Figure S1. Detection of intron RNA and MF-IEP protein during RNP preparation. (A) Fractions taken along the tube during RNP preparations. FM (in the middle), FI (at the bottom) and P (pellet). (B) Primer extension to determine the presence of intron RNA in the different fractions during the RNP preparations. (C) Detection of MF-IEP protein in the fractions was carried out by western blot using antibodies anti-Flag. (D) Endonuclease assay of reconstituted RNPs. Cleavage of substrate was observed only in lane when RNPs were used (RNPs lane arrow) and not when the protein was used.

RmInt1-IEP	MTSE-----STTDKPF-----RIEKRRVYEA	21
5HHL-Eu.re.I2	MDT-----SNLME-----QILSSDNLNRA	19
5HHJ-Ro.in.IEP	MVKSSGTERKERMDTSSLME-----QILSNDNLNRA	31
6AR1-Ge.st.IEP	M-----ALLE-----RILARDNLITA	16
5G2X-Ll.LtrA	MKPT-----MAILERISKNSQENIDEVFTRLYRYLLRPDIYYVA	39
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RmInt1-IEP	YKAVKANRGAAGVDGQTLEIFEKDLAANLYKIWNRMSSGTYFPPPVRAVS	71
5HHL-Eu.re.I2	YLQVVRNKGAEQVDGMKYTELKEHLAKNGETIKGQLRTRKYKQPARRVE	69
5HHJ-Ro.in.IEP	YLQVVRNKGAEQVDGMKYTELKEYLAKNGEIIKEQLRIRKYKQPARRVE	81
6AR1-Ge.st.IEP	LKRVEANQGAPGIDGVSTDQLRDYIRAHWSTIHAQLLAGTYRPAPVRRVE	66
5G2X-Ll.LtrA	YQNLYSNKGAST-KGILDDTADGFSEEKIKKIIQSLKDGTYYPQPVRRMY	88
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RmInt1-IEP	IPKKAG-GERVLGVPTVSDRIAQMVKQMIEPDLDSLFLPDSYGYRPGKS	120
5HHL-Eu.re.I2	IPKPDG-GVRNLGVPTVTDRFIQQAIQVLTPIYEEQFHDHSYGFRPNRC	118
5HHJ-Ro.in.IEP	IPKPDG-GVRNLGVPTVTDRFIQQAIQVLTPIYEEQFHDHSYGFRPNRC	130
6AR1-Ge.st.IEP	IPKPGG-GTRQLGIPTVVDRLIQQAILQELTPIFDPDFSSSSFGFRPGRN	115
5G2X-Ll.LtrA	IAKKNSKMRPLGIPTFTDKLIQEAVRIILESIIYEPVFEVDSHGFRPQRS	138
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RmInt1-IEP	ALDAVGVTRQR-CWKYDWWLEFDIKGLFDNLPHDLLLLKAVRKDVKCNWAL	169
5HHL-Eu.re.I2	AQQAILTALNIMNDGNDWIVDIDLEKFFDTVNHDKLMTLIGRTIKDGDVI	168
5HHJ-Ro.in.IEP	AQQAILTALDMMNDGNDWIVDIDLEKFFDTVNHDKLMTLIIGRTIKDGDVI	180
6AR1-Ge.st.IEP	AHDAVRQAQGYIQEGYRYVVDMDLEKFFDRVNHDILMSRVARKVKDKRVL	165
5G2X-Ll.LtrA	CHTALKTIKRE-FGGARWFVEGDIKGCFDNIDHVTLIGLINLKIKDMKMS	187
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RmInt1-IEP	LYIERWLTAPMEKNGEVIERSRGTPQGGVSPILANLFLHYAFDLWM---	216
5HHL-Eu.re.I2	SIVRKYLVSIMIDDEYEDSIVGTPQGGNLSPLLANIMLNE-LDKEM---	214
5HHJ-Ro.in.IEP	SIVRKYLVSIMIDDEYEDSIVGTPQGGNLSPLLANIMLNE-LDKEM---	226
6AR1-Ge.st.IEP	KLIRAYLQAGVMIEGVKQVTEEGTPQGGPLSPLLANILLDD-LDKEL---	211
5G2X-Ll.LtrA	QLIYKFLKAGYLENQYHKTYSGTPQGGILSPLLANIYLHE-LDKFVLQL	236
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RmInt1-IEP	-----	216
5HHL-Eu.re.I2	-----	214
5HHJ-Ro.in.IEP	-----	226
6AR1-Ge.st.IEP	-----	211
5G2X-Ll.LtrA	KMKFDRESPERITPEYRELHNEIKRISHRLKLEGEKAKVLLLEYQEKRK	286
RmInt1-IEP	-----TRTHPDLPCRYADDGLVHCQ-SEQQAEALRVELSSRLA-AC	256
5HHL-Eu.re.I2	-----EKRGLNFVRYADDCIIMVG-SEMSANRVMRNISRFIEEKL	253
5HHJ-Ro.in.IEP	-----EKRGLNFVRYADDCIIMVG-SEMSANRVMRNISRFIEEKL	265
6AR1-Ge.st.IEP	-----EKRGLKFCRYADDCNIYVK-SLRAGQVRKQSIQRFLEKTL	250
5G2X-Ll.LtrA	RLPTLPCTSQTNKVLKYVRYADDFIISVKGSKEDCQWIKEQLKLFHKNKL	336
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RmInt1-IEP	GLQMHPKTKIVYCKDQRRREAYPNVTFDFLGYQFRPRRVANTQRDEFFC	306
5HHL-Eu.re.I2	GLKVNMTKSKVDR-----PS-GLKYLGFYFDYDPRAH-----	284
5HHJ-Ro.in.IEP	GLKVNMTKSKVDR-----PR-GIKYLGFYFYDTSAQ-----	296
6AR1-Ge.st.IEP	KLKVNEEKSAVDR-----PW-KRAFLGFSFTPERKARIRLAPRS-	288
5G2X-Ll.LtrA	KMELSEKTLITH-----SSQPARFLGYDIRVRRSGTIKRSKGVK	376
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RmInt1-IEP	GYTPAVSPTALKSMRATIKSLNIPRQT-----PGTLA	338
5HHL-Eu.re.I2	-----	284
5HHJ-Ro.in.IEP	-----	296
6AR1-Ge.st.IEP	-----IQRLKQIRIRQLTNPNS-----ISMP	309
5G2X-Ll.LtrA	KRTLNGSVELLIPLQDKIRQFIFDKKIAIQKKDSSWFPVHRKYLIRSTDL	426
RmInt1-IEP	EIAKQLNPLLRGWIAYYGRYS-----RSALSTLADYVNQKLR	375
5HHL-Eu.re.I2	-----	284
5HHJ-Ro.in.IEP	-----	296
6AR1-Ge.st.IEP	ERIHRVNQYVMGWIGYFRLVET-----PSVLQTIIEGWIRRLR	347
5G2X-Ll.LtrA	EIITIYNSELRGICNYYGLASNFNQLNYFAYLMEYSCLKTIASKHKGTLS	476
RmInt1-IEP	A-----WIRRKFRFQSHK-----	389
5HHL-Eu.re.I2	-----	284
5HHJ-Ro.in.IEP	-----	296
6AR1-Ge.st.IEP	LCQ-----WLQW--KRVRT-----	359
5G2X-Ll.LtrA	KTISMFKDGSWSWGIPIYEIKQKQRRYFANFSECKSPYQFTDEISQAPVL	526
RmInt1-IEP	-----	389
5HHL-Eu.re.I2	-----	284
5HHJ-Ro.in.IEP	-----	296
6AR1-Ge.st.IEP	-----RIRELRALGLKETAVMEIANTRKGAWRTTKTPQLHQ	395
5G2X-Ll.LtrA	YGYARNTLENRLKAKCCELCGTSDENTS-YEI-----HHVNKVKVN	565
RmInt1-IEP	--TRASLFLRKLARENPGLEFV--HWKAFGTNT-FT	419
5HHL-Eu.re.I2	-----QFK-----AKP-HA	292
5HHJ-Ro.in.IEP	-----QFK-----AKPHAK	305
6AR1-Ge.st.IEP	ALGKTYWTAQGLKSLTQRYFEL-RQGHHHHHH-HH	428
5G2X-Ll.LtrA	LKGKEKWEMAMIAKQRKTLVVCFHCHRHVVIHK-HK	599

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Figure S2. Structure-based sequence alignment of IEPs. The sequence of RmInt1 IEP is aligned to the sequences of Eu.re.I2 (from PDB: 5HHL), Ro.in.IEP (from PDB: 5HHJ), Ge.st.IEP (from PDB: 6AR1) and LtrA (from PDB: 5G2X). The QMxxxxLxxLFL motif is highlighted in yellow

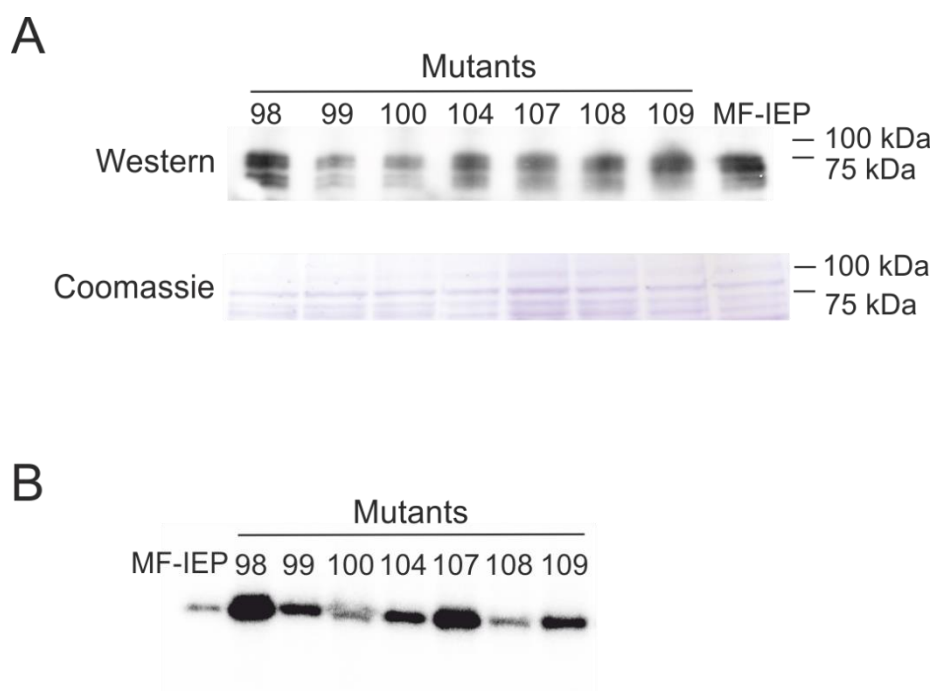


Figure S3. (A). Western analysis using anti-Flag antibodies of mutants done in MF-IEP. Protein loaded in each lane is shown in the panel labelled as Coomassie (B) Primer extension of mutants of MF-IEP protein.

Supplementary material and methods

Primer extension

15 µg RNA samples were annealed with 300,000 c.p.m. 5'-labeled P primer (5'-TGAAAGCCGATCCCGGAG-3') in 10 mM Pipes (pH 7.5) and 400 mM NaCl by heating at 85°C for 5 minutes and fast cooling to 60°C followed by slow chilling to 45°C. The cDNA was synthesized using AMV Reverse Transcriptase (Roche Diagnostics) in the presence of 50 mM Tris-HCl pH 8.0; 60 mM NaCl; 10 mM DTT; 6 mM MgOAc; 1 mM dNTPs; 60 µg/ml actinomycin D (Sigma-Aldrich) and 0.4 U RNase Inhibitor (NEB) at 42°C for 60 minutes. The reactions were ethanol precipitated and the products were separated by denaturing 6% polyacrylamide gel electrophoresis

Endonuclease assays.

For DNA cleavage assays, 5' end-labeled gel-purified 70-mer single-stranded DNA oligonucleotide (300,000 cpm) was incubated with reconstituted RNP s (1.5 µg) or MF-IEP protein (1.5 µg) at 37°C for 2 h in reaction buffer [50 mM Tris-HCl pH 7.5, 10 mM KCl, 25 mM MgCl₂, 5 mM DTT]. The reactions were extracted with phenol-chloroform-isoamyl alcohol (25:24:1) followed by ethanol precipitation, and the products were separated by denaturing 7 M urea- 6% (w/v) polyacrylamide gel electrophoresis. Bands were revealed by dry gel exposition to digital imaging plates (Fuji), which were scanned using Quantity One software (Bio-Rad).