

1 Supplemental Information

2 **Early Treatment With Empagliflozin And GABA Improves β -Cell Mass And Glucose**
3 **Tolerance In Streptozotocin-Treated Mice**

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6 **Supplemental Information inventory:**

7 **Figure S1** shows insulin level during the IPGTT assay (A). This figure also shows insulin
8 dose injected to the diabetic mice through the protocol (B and C).

9 **Figure S2** shows immunofluorescence with a Ki-67 and insulin staining.

10 **Table S1** shows primers used for real-time RT-PCR analysis. This table supports data
11 presented in main Figure 7.

12 **Table S2** shows antibodies used for immunofluorescence staining. This table supports data
13 presented in main Figure 6 and supplemental Figure 2.

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15 **Supplemental Materials & Methods**

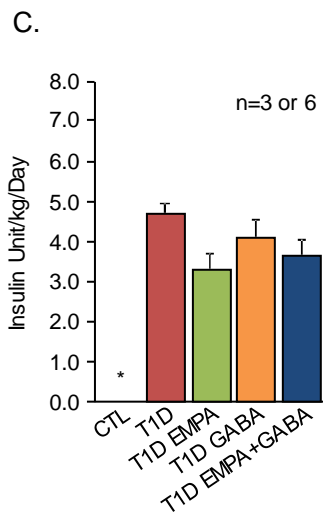
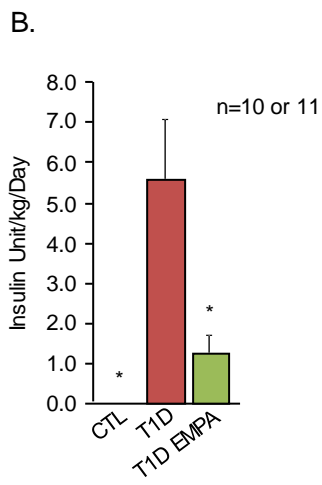
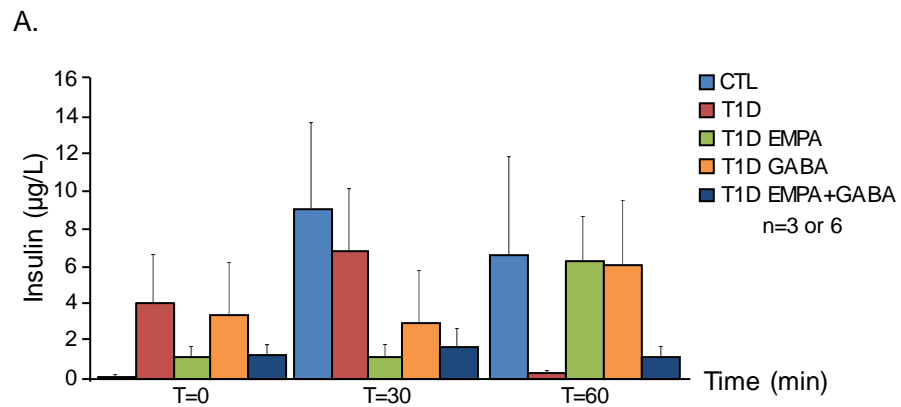
16 **Intraperitoneal glucose tolerance test (IPGTT)**

17 IPGTTs were performed at the end of the treatment protocols (day 12 or 28). Blood samples
18 were collected at 0, 30 and 60 min during IPGTT and were used to measure insulin levels
19 using a commercial ELISA kit (Mercodia), following the manufacturer's instructions. Results
20 were acquired using a spectrophotometer (Victor™ X4, 2030 Multilabel Reader, Perkin
21 Elmer) at 450 nm.

22 **Immunofluorescence**

23 Pancreas tissues were fixed overnight in 4% paraformaldehyde (PFA) and embedded in
24 paraffin. Then, the samples were sectioned into 5 µm thick slices, blocked in 10% bovine
25 serum albumin (Sigma-Aldrich), 3% no fat milk and 0.3% Triton X-100 PBS. For the
26 detection of Ki67, tissues were incubated overnight at 4°C with antibody (listed in **Table S2**)
27 diluted in PBS/0.3% Triton X-100/10% BSA/3% milk. Then tissues were incubated the next
28 day for 2 h at RT in the dark with respective secondary antibody (listed in **Table S2**) diluted
29 in PBS/0.3% Triton X-100/10% BSA. After washing, tissues were incubated for 5 min with
30 DAPI 397-412 nm diluted to 1:10,000 in distilled water. Then, sections were mounted with
31 Faramount mounting medium (Dako, # S3025) and observed under a fluorescence
32 microscope. The digital images were acquired by a digital imaging platform (3DHistech
33 Panoramic P250 Flash III, nicknamed Oyster).

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36 **Figure S1. Daily dose of insulin received by the mice through the study.** (A) At 0, 30 and
37 60 min post glucose injection, blood of control mice (CTL, blue bar), untreated diabetic mice
38 (T1D, red bar), diabetic mice treated with empagliflozin (T1D EMPA, green bar), GABA
39 (T1D GABA, orange bar) or empagliflozin and GABA (T1D EMPA+GABA, navy blue bar)
40 was collected. Insulin level was evaluated by ELISA kit. The bars represent the standard error
41 of the mean. Number of mice per group is indicated (n=3 or 6). The statistics were performed
42 using a ANOVA-Tukey test for the times 0 and 30 min and using a Kruskal-Wallis for the
43 time 60 min. (B and C) Daily, human Lantus insulin was injected in control mice (CTL, blue
44 bar), untreated diabetic mice (T1D, red bar), diabetic mice treated with empagliflozin (T1D
45 EMPA, green bar), GABA (T1D GABA, orange bar) or empagliflozin and GABA (T1D
46 EMPA+GABA, navy blue bar). Diabetic mice received 6 unit of Human Lantus insulin by
47 subcutaneous injection when their glycemia was higher than 400 mg/dL. B and C correspond
48 to the protocol #1 and #2, respectively. (B) The asterisk (*) shows a significant difference
49 ($p=0.004$; ANOVA+Tukey) between T1D group and CTL and T1D EMPA groups. (C) The
50 asterisk (*) shows a significant difference ($p<0.001$; ANOVA-Tukey) between CTL group
51 and T1D, T1D EMPA, T1D GABA and T1D EMPA+GABA groups. The bars represent the
52 standard error of the mean. For each study, number of mice per group is indicated (n=10 or
53 11, n=3 or 6).

54 **Figure S2.**

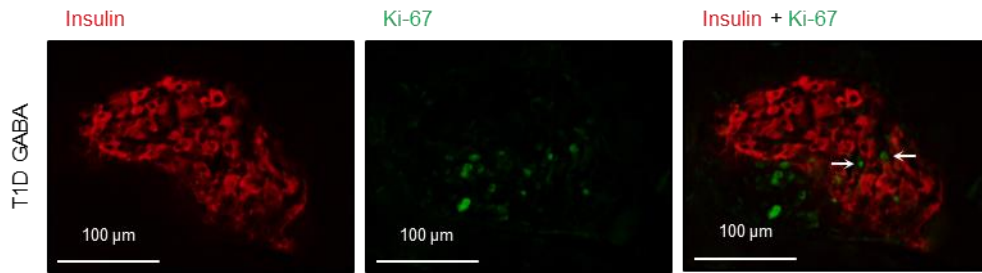
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60 **Figure S2. Effect of GABA on α cells proliferation.** The pancreas of T1D GABA mice was

61 divided into three parts and its tail was used for insulin and Ki-67 staining. The red and green

62 staining corresponds to the insulin and Ki-67, respectively. Arrows indicated Ki-67 positive

63 nucleus. Images were obtained thank the microscope Scope A.1 Zeiss (40x/0.75), the camera

64 Axiocam MRc5 Zeiss (0.63x) and the software Zen (Insulin: 250 ms and Ki-67: 300 ms).

65 Scale bar=100 µm.

66 **Table S1. SYBER Green Primers**

| Function | Gene symbol | Sequence | Ref |
|---------------------------------------|------------------------------|---|-----|
| Oxidative stress | <i>iNOS</i> | Forward: 5'-AATCCCTGGACAAGCTGCAT-3' | |
| | | Reverse: 5'-TCTGGTCAAACCTCTTGGGGTT-3' | |
| Stress ER | <i>sXbp1</i> | Forward: 5'-CTGAGTCCGCAGCAGGTGCAG-3' | [1] |
| | | Reverse: 5'-GTCCATGGGAAGATGTTCTGG-3' | |
| | <i>Atf4</i> | Forward: 5'-GGGTTCTGTCTTCCACTCCA-3' | [1] |
| | | Reverse: 5'-AAGCAGCAGAGTCAGGCTT(TC)-3' | |
| Inflammation | <i>Bip</i> | Forward: 5'-TTCAGCCAATTATCAGCAAACCTCT-3' | [1] |
| | | Reverse: 5'-TTTTCTGATGTATCCTCTTCACCAGT-3' | |
| | <i>Txnip</i> | Forward: 5'-AAGTTACCCGAGTCAAAGCCGT-3' | |
| Reverse: 5'-TTCCAGGCCTCATGATCACCAT-3' | | | |
| Inflammation | <i>IL1β</i> | Forward: 5'-ATCTGGGATCCTCTCCAGCCAA-3' | |
| | | Reverse: 5'-TCATCAGGACAGCCCAGGT-3' | |
| Internal control | <i>IL6</i> | Forward: 5'-AACCGCTATGAAGTTCCTCTCT-3' | |
| | | Reverse: 5'-AGGTCTGTTGGGAGTGGTA-3' | |
| Internal control | <i>RpL19</i> | Forward: 5'-CTGAAGGTCAAAGGGAATGTG-3' | [2] |
| | | Reverse: 5'-GGACAGAGTCTTGATGATCTC-3' | |

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70 **Table S2. Antibodies**

| Antibody | Firm | Reference | Dilution |
|-----------------|-------------------|-----------|----------|
| Insuline | Cell Signaling | 3014S | 1:500 |
| Insuline | Dako | A0564 | 1:500 |
| Glucagon | Santa Cruz | sc-71152 | 1:400 |
| Ki67 | ThermosFisher | MA5-14520 | 1:250 |
| Anti rabbit | Life technologies | A11012 | 1:1,000 |
| Anti mouse | Life technologies | A11001 | 1:1,000 |
| Anti guinea pig | Life technologies | A21450 | 1:1,000 |

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73 **References**

- 74 1. Osowski, C.M. and F. Urano, *Measuring ER stress and the unfolded protein response*
75 *using mammalian tissue culture system*. *Methods Enzymol*, 2011. **490**: p. 71-92.
- 76 2. Guigon, C.J., et al., *Follicular cells acquire sertoli cell characteristics after oocyte*
77 *loss*. *Endocrinology*, 2005. **146**(7): p. 2992-3004.

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