## Both Ligands and Macromolecular Crowders Preferentially Bind to Closed Conformations of Maltose Binding Protein

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Supporting Information

Two-state model for wild-type MBP <sup>a</sup>				
Titrant	$K_{d;app} (\mu M) \text{ or} \ K_{d}^{C} (mM)^{b}$	$\lambda_{f} \text{ or } \lambda_{P}$ (nm)	$\lambda_b \text{ or } \lambda_{CP}$ (nm)	
Maltose ([C] <sub>T</sub> =0)	$1.2 \pm 0.1^{\circ}$	$345.8\pm0.1$	$350.0 \pm 0.1$	
Maltose ([C] <sub>T</sub> =100 g/L)	$1.9\pm0.3$	$344.0\pm0.1$	$349.8\pm0.2$	
Maltose ( $[C]_T$ =200 g/L)	$3.9 \pm 0.2$	$343.0\pm0.1$	$349.9\pm0.1$	
Maltose ( $[C]_T$ =300 g/L)	$8 \pm 1$	$343.0 \pm 0.1$	$350.3 \pm 0.1$	
Ficoll ( $[L]_T=0$ )	$2.0 \pm 0.5$	$345.6 \pm 0.1$	$341.5 \pm 0.4$	

Three-state model for wild-type MBP<sup>a</sup>

	$K_{\rm d}(\mu { m M})$ or	$\lambda_{P}(nm)$	$\lambda_b$ or $\lambda_{CP}$
	$K_{d}^{C}$ (mM)		(nm)
Maltose and Ficoll	$1.3 \pm 0.1$	$345.8\pm0.1$	$350.0 \pm 0.1$
	$1.5 \pm 0.2$		$\textbf{341.8} \pm \textbf{0.2}$

Two-state model for A96F				
Maltose ( $[C]_T=0$ )	$0.33 \pm 0.05$	$345.9 \pm 0.1$	$349.3 \pm 0.1$	
Maltose ( $[C]_T$ =50 g/L)	$0.67 \pm 0.05$	$344.0\pm0.1$	$349.4\pm0.1$	
Maltose ([C] <sub>T</sub> =100 g/L)	$0.91 \pm 0.09$	$343.5\pm0.1$	$349.2\pm0.1$	
Maltose ([C] <sub>T</sub> =200 g/L)	$1.6 \pm 0.2$	$342.6\pm0.1$	$349.7\pm0.2$	
Maltose ( $[C]_T$ =300 g/L)	$2.2 \pm 0.1$	$341.8\pm0.1$	$349.6\pm0.1$	
Ficoll ([L] <sub>T</sub> =0)	$0.54 \pm 0.09$	$346.1\pm0.3$	$342.3\pm0.2$	

Three-state model for A96F			
Maltose and Ficoll	$0.42 \pm 0.04$	$346.1 \pm 0.1$	$349.4 \pm 0.1$
	0.91 ± 0.09		$343.2 \pm 0.1$

Two-state model for A96W				
Maltose ([C] <sub>T</sub> =0)	$0.036 \pm 0.006$	$346.5 \pm 0.1$	$349.3\pm0.1$	
Maltose ( $[C]_T$ =100 g/L)	$0.053 \pm 0.003$	$344.0\pm0.04$	$349.3\pm0.04$	
Maltose ([C] <sub>T</sub> = $200 \text{ g/L}$ )	$0.097 \pm 0.010$	$343.5\pm0.1$	$349.1\pm0.1$	
Maltose ([C] <sub>T</sub> =300 g/L)	$0.22 \pm 0.02$	$343.3\pm0.1$	$349.3\pm0.1$	
Ficoll ([L] <sub>T</sub> =0)	$0.35\pm0.04$	$346.1 \pm 0.2$	$343.6 \pm 0.1$	

Three-state	model	for	A96W
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The estate model for 11900			
Maltose and Ficoll	$0.019 \pm 0.002$	$346.4 \pm 0.1$	$349.3 \pm 0.1$
	$0.47 \pm 0.05$		$343.2 \pm 0.1$

<i>Two-state model for 1329W</i>				
Maltose ( $[C]_T=0$ )	$0.014 \pm 0.009$	$346.4 \pm 0.04$	$349.7 \pm 0.03$	
Maltose ([C] <sub>T</sub> =100 g/L)	$0.030 \pm 0.006$	$344.4\pm0.1$	$350.1\pm0.1$	
Maltose ( $[C]_T$ =200 g/L)	$0.056 \pm 0.011$	$344.4\pm0.1$	$350.6\pm0.1$	
Maltose ( $[C]_T$ =300 g/L)	$0.12 \pm 0.01$	$343.6\pm0.1$	$350.8\pm0.1$	
Ficoll ( $[L]_T=0$ )	$0.65 \pm 0.11$	$346.4 \pm 0.2$	$343.1 \pm 0.2$	

## Two-state model for I329W

Three-state model for I329W				
Maltaga and Eisall	$0.012 \pm 0.002$	$346.4 \pm 0.1$	$349.7 \pm 0.1$	
Manose and Picon	$0.70 \pm 0.11$		$343.2 \pm 0.1$	
<i>Tw</i>	vo-state model for A	<u>96W/I329W</u>		
Ficoll ( $[L]_T=0$ )	$0.45\pm0.11$	$348.9\pm0.3$	$345.9\pm0.3$	
Two-st	ate model for MBP-	NBD in Ficoll70		
	$K_{d;app}$ ( $\mu$ M)			
	Peak wavelength	Fluo. intensity at		
	with 280 nm	550 nm with 500		
	excitation	nm excitation		
Maltose ([C] <sub>T</sub> =0)	$5.8 \pm 0.4$	29 ± 1		
Maltose ( $[C]_T$ =50 g/L)	$14 \pm 1$	$30 \pm 1$		
Maltose ( $[C]_T$ =100 g/L)	$30 \pm 2$	61 ± 1		
Maltose ([C] <sub>T</sub> = $200 \text{ g/L}$ )	$61 \pm 3$	$145 \pm 3$		
Two-state model for MBP-NBD in BSA				
Maltose ( $[C]_T=0$ )		$26 \pm 0.7$		

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Maltose ([C] <sub>T</sub> =0)		$26 \pm 0.7$	
Maltose ( $[C]_T$ =50 g/L)		$219 \pm 14$	
Maltose ( $[C]_T$ =100 g/L)		$469 \pm 22$	

<sup>a</sup>Taken from Miklos et al. (2013).

 ${}^{b}K_{d;app}$  (µM) applies to rows where maltose is the titrant;  $K_{d}^{C}$  (mM) applies to rows where Ficol70 is the titrant.

<sup>c</sup>Bold entries are obtained in both two-state and three-state fits. Whereas the two-state model was used to independently fit five or six titration curves, the three-model in essence provides a constrained, simultaneous fit of all these titration curves. The degree of agreement between the corresponding parameters in the two- and three-state fits is thus a measure of the soundness of the three-state competitive model.



**Figure S1.** (A) Two-state fit of binding isotherms from titrating maltose into the A96F mutant in the absence or presence of fixed concentrations of Ficoll70. (B) Two-state fit of the binding isotherm from titrating Ficoll70. (C) Three-state fit of all the binding isotherms.



**Figure S2.** (A) Two-state fit of binding isotherms from titrating maltose into the I329W mutant in the absence or presence of fixed concentrations of Ficoll70. (B) Two-state fit of the binding isotherm from titrating Ficoll70. (C) Three-state fit of all the binding isotherms.



**Figure S3.** Linear dependence of the apparent maltose dissociation constants on Ficoll70 concentration, expected for a three-state competitive model. (A) Wild-type MBP. (B) A96F mutant. (C) A96W mutant. (D) I329W mutant.



**Figure S4.** Two-state fit of the binding isotherm from titrating Ficoll70 into the A96W/I329W mutant.



**Figure S5.** FMAP calculation of the protein-crowder interaction energies. An MBP molecule (green) is fictitiously placed into many locations inside a cubic box with side length of 200 Å containing eight BSA molecules (gray), representing a concentration of 110 g/L. The interaction energies within a slice of the crowder solution are displayed as colors according to a scale (in kcal/mol) shown on the right. The particular MBP molecule displayed is within a "hot" region (center of blue rectangle). An enlarged view of the pose with this MBP molecule docked to a neighboring BSA molecule is shown at lower bottom.



**Figure S6.** Histograms of MBP-BSA pairwise interaction energies. MBP is either in the closed form (blue curve) or open form (red curve). By the FMAP method,  $2 \times 10^6$  MBP placements in a box of BSA molecules with the lowest interaction energies were obtained. Out of these, the interaction energies were further calculated by an atom-based method and the results were collected for  $1.52 \times 10^5$  MBP placements (the remaining placements were newly found to have clashes with the crowders). The latter results were grouped into bins with 0.02 kcal/mol width, and the count in each bin is displayed.