# Supplementary Information for:

# Divergent engagements between adeno-associated viruses with their cellular receptor AAVR

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#### **Supplementary Figure 1**

**Supplementary Figure 1. Resolution assessment.** Fourier shell correlation (FSC) of the final 3D reconstruction following gold standard refinement using RELION and THUNDER. The resolutions corresponding to an FSC of 0.143 are shown for unbound AAV1 (a), the AAV1-AAVR complex (b), unbound AAV5 (c) and the AAV5-AAVR complex (d). FSC curves are plotted before (red) and after (green) masking in addition to postcorrection (blue), accounting for the effect of the mask using phase randomization.



**Supplementary Figure 2. Density maps of the AAV5-AAVR and AAV1-AAVR complexes.** Shaded surface representation of the density maps for the trimeric AAV5 (a) or AAV1 (b) capsomers in complex with three molecules of PKD1 (in gold) (a) or PKD2 (in magenta) (b). The densities for three AAV1 and AAV5 capsomers are colored green, blue and cyan, respectively, in both panels. In the surrounding boxes, atomic models shown as sticks are superimposed to indicate the representative regions in wire frames. In the stick models, the residue numbers are indicated. The AAVR, AAV1 and AAV5 residues are labeled with a subscript.



**Supplementary Figure 3. (a)** Sequence alignment of AAVR PKD1-5. Residues with red or yellow backgrounds are identical or conserved, respectively. Residues in PKD1 and PKD2 at the AAV5-AAVR and AAV1-AAVR interfaces are

indicated with blue and red triangles, respectively. Residues in PKD1 and PKD2 spanning the density in panel **b** and panel **c** are highlighted in blue and red boxes, respectively. (**b**) The residues in a featured fragment from PKD1 ( $_{AAVR}H351-_{AAVR}S356$ ) are shown as colored sticks and surrounded by density. The large densities of the side chains of  $_{AAVR}H351$  and  $_{AAVR}Y355$ , and short density of the side chain at position  $_{AAVR}S356$  are different from those for their counterparts in other PKDs. (**c**) The residues in a featured fragment from PKD2 ( $_{AAVR}G430-_{AAVR}T435$ ) are shown as colored sticks and surrounded by density. The short density of the side chain at position  $_{AAVR}S356$  are different from those for their counterparts in other PKDs. (**c**) The residues in a featured fragment from PKD2 ( $_{AAVR}G430-_{AAVR}T435$ ) are shown as colored sticks and surrounded by density. The short density of the side chain at position  $_{AAVR}S433$  distinguishes it from an asparagine residue in PKD3 and a leucine residue in PKD1.



Supplementary Figure 4. Structure of AAVR PKD1 bound to AAV5. (a) PKD1 adopts an Ig-like fold and contains nine  $\beta$ -strands labeled A-G and colored in orange. The loops between each  $\beta$ -strand are shown in green. (b) Topological secondary structures of PKD1. The  $\beta$ -strands are alphabetically labeled and shown as orange arrows. The regions that interact with AAV5 are highlighted by red frames. The N-terminal residue, <sub>AAVR</sub>V305, is also involved in the interaction with AAV5 and is highlighted with a red dot.



Supplementary Figure 5. Structural alignment of unbound and AAVRbound AAV5 and AAV1. The structures of the AAV5 capsomer in its unbound form (blue) and in an AAVR-bound state (red) (a) and the AAV1 capsomer in its unbound form (blue) and in an AAVR-bound state (red) (b) are shown in cartoon representations and aligned individually. The region showing a significant conformational shift in the AAVR-bound AAV1 capsomer is highlighted by a red circle and enlarged in the right panel of (b).





Supplementary Figure 6. Mutagenesis study of AAVR for AAV5 and AAV1 binding assays. (a-j) A total of nine PKD1 mutants were tested for their ability to bind to AAV5 by BIAcore sensorgrams. (k-x) A total of thirteen PKD2 mutants were tested for their ability to bind to AAV5 by BIAcore sensorgrams. The concentrations of the analytes are indicated in each panel. The analytes with RU

values under 20 for the highest concentration tested are denoted as nondetectable (N.D.). Each panel is a representative for triplicate experiments.



Supplementary Figure 7. Different impacts of the interacting residues on AAV5 and AAV1 transduction. The interacting regions at the virus-receptor interface on the AAV5 capsid (a) and AAVR PKD1 (b), the AAV1 capsid (c) and AAVR PKD2 (d) are framed with dashed lines. Three AAV capsomers of AAV5 (a) and AAV1 (c) are covered with blue, green and cyan surfaces. AAVR PKD1 (b) and PKD2 (d) bound to AAVs are colored orange and magenta, respectively. The interacting residues with the greatest impact, a mild impact, or a negligible impact on viral transduction are colored red, light orange and white, respectively. Residues that increased viral transduction are colored green. Residues whose side chains were not involved in the interaction with AAVR are colored white.



**Supplementary Figure 8. Virus overlay assays.** Equal amount  $(6 \ \mu g)$  of wt AAVR was loaded onto Bis-Tris gels, and virus overlay assays with AAV5 **(a)** or AAV1 **(b)** and the related mutations were performed. The molecular weights for

standard protein makers are 180 kDa, 130 kDa, 95 kDa, 55 kDa, 43 kDa, 34 kDa, 26 kDa, 17 kDa and 10 kDa from top to bottom, respectively.



**Supplementary Figure 9. (a)** A total of nine AAVR PKD1 mutants were tested for their ability to bind AAV1 by BIAcore sensorgrams in triplicate experiments. **(b)** A total of thirteen AAVR PKD2 mutants were tested for their ability to bind AAV5 by BIAcore sensorgrams in triplicate experiments. The calculated KD values for the binding of each mutant to virus are summarized as the mean values of three experiments with standard errors. **(c)** The impact of AAVR PKD1 mutants overexpressed in AAVR-silenced HEK293T cells on AAV1 transduction. Cells were transfected with wt AAVR or different AAVR mutants as indicated followed by infection with AAV1-mCherry at an MOI of  $5 \times 10^5$  vg per cell. **(d)** The overexpression of AAVR PKD2 mutants in AAVR-silenced HEK293T cells impacted on AAV5 transduction. Cells were transfected with wt AAVR or different AAVR mutants as indicated followed by infection with AAV5-mCherry at an MOI of  $3 \times 10^6$  vg per cell. The percentage of mCherry positive cells are plotted as means +/- standard errors (n=3). Source data are provided as a Source Data file.



Supplementary Figure 10. Sequence alignment of the VP3 region of the AAV1-AAV12 capsids. The labeled numbers correspond to the amino acid

residues in the AAV1 VP1 coding region. The variable regions between different AAV serotypes are colored green and labeled. Residues that interact with HPSG analog in AAV2, PKD2 in AAV1, and PKD1 in AAV5 are highlighted in blue, magenta and orange frames, respectively. Residues that contact PKD2 from AAV2 are marked with red stars. Identical or conserved residues are shown as letters with red or yellow backgrounds, while the nonconserved residues have a white background.



**Supplementary Figure 11. Three positions are involved in the PKD2-AAV2 interaction but not the PKD2-AAV1 interaction.** The AAV1-AAVR PKD2 and AAV2-AAVR PKD2 complex structures are aligned referenced by the polypeptides of the AAV1 and AAV2 capsomers. Two AAV1 capsomer polypeptides are shown as blue and green cartoons, while two AAV2 capsomer polypeptides are displayed as white cartoons. The PKD2 molecules bound to AAV1 and AAV2 are represented as magenta and while cartoons, respectively. Three positions in the AAV1 and AAV2 capsids with distinct interactions with AAVR PKD2, as well as the interacting residues in AAVR PKD2 are shown as colored sticks with labels. Dashed lines denote bonds with a distance less than 3.5 Å.



**Supplementary Figure 12. Glycosylated sites in AAVR.** Among the five reported glycosylated sites in AAVR, three are found in the structure of PKD2 bound to AAV1 (a), and one is located in a PKD1 molecule bound to AAV5 (b). The two AAV1 or AAV5 capsomers are shown as blue and green surfaces, while the bound PKD1 and PKD2 are shown as orange and magenta cartoons, respectively. The reported glycosylated residues are highlighted as colored spheres.



Supplementary Figure 13. Full-length gels and immunoblots. (a, c, e) Whole gels showing the expression of wt AAVR or AAVR mutants in HEK293T cells with shRNA. After blocking, membranes were cut to 2 halves along the 72 kDa protein marker. AAVR was immunoblotted with anti-AAVR antibodies in the upper half. The corresponding samples were loaded onto another gel, and  $\beta$ -actin was immunoblotted as a control in the lower half (b, d, f). The molecular weights of AAVR and  $\beta$ -actin are 150 kDa and 43 kDa, respectively. The molecular weights of standard protein markers are labeled in each panel. (g) Illustration of gating strategy of flow cytometry.

# Supplementary Tables

# Supplementary Table 1. Cryo-EM data statistics

	AAV1 alone EMD-9795, PDB 6JCR	AAV1-AAVR EMD-9794, PDB 6JCQ	AAV5 alone EMD-9797, PDB 6JCT	AAV5- AAVR EMD-9796, PDB 6JCS
Data collection and processin	g			
Magnification Voltage (kV) Electron exposure (e–/Ų)	110,000 200 25.04	110,000 200 25.04	110,000 200 25.04	110,000 200 25.04
Defocus range (µm)	-2.5 to -1.2	-2.5 to -1.2	-2.5 to -1.2	-2.5 to -1.2
Pixel size (Å)	0.93	0.93	0.93	0.93
Symmetry imposed	11	l1	11	11
Initial particle images (no.)	3,499	3,831	3,545	14,221
Final particle images (no.)	2,642	2,926	2,900	12,590
Map resolution (Å)	3.07	3.30	3.18	3.18
FSC threshold	0.143	0.143	0.143	0.143
Map resolution range (Å)	2.6-3.07	2.7-3.3	2.7-3.18	2.7-3.18
Refinement				
Initial model used (PDB	5EG3	5EG3,	3NTT	3NTT,
code)		6IHB		2E7M
Model resolution (Å)	3.07	3.30	3.18	3.18
FSC threshold	0.143	0.143	0.143	0.143
Model resolution range (A)	∞ to 3.07	∞ to 3.30	∞ to 3.18	co to 3.18
Map sharpening <i>B</i> factor $(Å^2)$	-132.4	-159.3	-148.00	-152.8
Correlation coefficient	0 824	0.815	0 825	0.811
between map and model	0.024	0.010	0.020	0.011
Model composition				
Non-hydrogen atoms	4,112	4,830	4,110	4,881
Protein residues	517	611	515	611
Ligands	0	0	0	0
<i>B</i> factors (A <sup>2</sup> )				
Protein	32.9	36.7	35.1	33.8
Ligand				
R.m.s. deviations				
Bond lengths (A)	0.009	0.009	0.008	0.010
Bond angles (°)	1.030	0.971	0.970	1.008
Validation				
MolProbity score	1.68	1.97	1.69	1.79
Clashscore	4.88	6.27	4.39	5.90
Poor rotamers (%)	0.22	0.19	0.44	0.56
Ramachandran plot				
Favored (%)	93.60	89.46	92.61	92.45

Allowed (%)	6.40	10.21	7.20	7.22	
Disallowed (%)	0	0.33	0.19	0.33	

No.	Secondary Structure	Range of Residues	No.	Secondary Structure	Range of Residues
1	βA	221-225	22	βGH5	446-451
2	βΒ	228-239	23	βGH6	469-470
3	αBC1	241-245	24	βGH7	474-475
4	βBC2	248-252	25	αGH8	488-492
5	αBC3	258-262	26	βGH9	494-497
6	βC	263-272	27	βGH10	500-503
7	αCD1	277-281	28	βGH11	511-512
8	αΑ	283-293	29	βGH12	521-522
9	βD	294-315	30	βGH13	528-530
10	βDE1	322-324	31	βGH14	547-549
11	βE	331-336	32	αGH15	552-556
12	αEF1	344-348	33	βGH16	560-561
13	βEF2	361-362	34	βGH17	567-570
14	βEF3	365-370	35	βGH18	582-585
15	αEF4	387-391	36	βGH19	608-610
16	βF	394-396	37	βGH20	627-629
17	βG	402-407	38	βН	635-640
18	βGH1	414-415	39	βΙ	661-677
19	βGH2	418-419	40	βΙ1	702-703
20	αGH3	423-427	41	βl2	709-710
21	βGH4	436-442	42	βΙ3	721-722

Supplementary Table 2. Secondary structure assignment of the AAV5 capsid

The secondary structures of AAV5 capsid are assigned as a previously reported crystallographic AAV5 structure (PDB code: 3NTT).

AAVR Residues	Contacts <sup>1</sup>	AAV5 Residues
V305	1	S319
1349	1	N535
T350	4	Q532
H351	4, 2, 4, 8	Q532, N546, E708, R710
P352	5, 4	G545, N546
R353	2, 6, 2, 1, 4	S531, G545, M547, F698, T712
D354	1	T712
Y355	2, 4	F698, R710
S356	3	Q697
P374	1	G545
G375	3	L543
L376	1, 2, 1, 6	A540, T541, Y542, N546
E378	1	A540
T397	1	L543
V398	1	L543
K399	3	N443

#### Supplementary Table 3. Interaction between AAV5 and PKD1

<sup>1</sup>Numbers represent the number of atom-to-atom contacts between the AAVR residues and the AAV5 residues, analyzed by the Contact program in the CCP4 suite (with a distance cutoff of 4 Å ).

No.	Secondary Structure	Range of Residues	No.	Secondary Structure	Range of Residues
1	βA	232-235	17	βGH2	443-449
2	βΒ	238-249	18	βGH3	460-465
3	αBC1	253-255	19	βGH4	487-490
4	βBC2	258-262	20	αGH5	501-504
5	βC	273-282	21	βGH6	508-511
6	αCD1	288-290	22	βGH7	514-516
7	αΑ	294-302	23	βGH8	533-535
8	βD	304-325	24	βGH9	542-544
9	βDE1	330-334	25	αGH10	563-567
10	βE	342-346	26	βGH11	578-581
11	βEF1	371-373	27	βGH12	593-596
12	βEF2	375-381	28	βGH13	619-621
13	αEF3	396-398	29	βGH14	638-640
14	βF	401-405	30	βΗ	647-651
15	βG	409-416	31	βΙ	673-689
16	βGH1	426-428	32	βI1	732-734

Supplementary Table 4. Secondary structure assignment of the AAV1 capsid

The secondary structures of AAV1 capsid are referenced with the structure of AAV2 (PDB code: 1LP3).

AAVR Residues	Contacts <sup>1</sup>	AAV1 Residues	
S425	9	D590	
T426	1	D590	
V427	2	T504	
D429	1, 5	T504, W503	
S431	1	W503	
Q432	2, 1	S268, W503	
S433	1, 1	S268, N269	
T434	1, 6, 5	G266, A267, S268	
D435	3, 5, 3	G266, A267, H272	
D436	3, 2, 3	A263, H272, S385	
D437	6, 1, 5	H272, S385, Q386	
K438	1, 2, 1	N269, N383, G384	
1439	1	N269	
Y442	2	N269	
K464	1	T593	

#### Supplementary Table 5. Interaction between AAV1 and PKD2

Numbers represent the number of atom-to-atom contacts between the AAVR residues and the AAV1 residues, analyzed by the Contact program in the CCP4 suite (with a distance cutoff of 4 Å ).

No.	Name of variable region	Corresponding secondary structure	Range of residues	in AAVR interaction?
1	VR-I	loop βBC2-aBC3	253-258	No
2	VR-II	loop βD-βDE1	315-321	Yes
3	VR-III	loop βEF2-aEF4	370-381	No
4	VR-IV	loop βGH4-βGH5	442-446	No
5	VR-V	loop βGH9-βGH10	497-500	No
6	VR-VI	loop βGH11-βGH12	514-520	No
7	VR-VII	loop βGH13-βGH14	536-546	Yes
8	VR-VIII	loop βGH17-βGH18	572-581	No
9	VR-IX	Ιοορ βΙ-βΙ1	693-698	Yes

#### Supplementary Table 6. Assignment of variable regions in the AAV5 capsid

#### Supplementary Table 7. Assignment of variable regions in the AAV1 capsid

No.	Name of variable region	Corresponding secondary structure	Range of residues	in AAVR interaction?
1	VR-I	Ιοορ βΒC2-βC	263-265	Yes
2	VR-II	loop βD-βDE1	326-330	No
3	VR-III	loop βEF2-aEF3	381-388	Yes
4	VR-IV	loop βGH2-βGH3	449-457	No
5	VR-V	loop βGH6-βGH7	511-513	No
6	VR-VI	loop βGH7-βGH8	525-530	No
7	VR-VII	loop βGH9-aGH10	544-554	No
8	VR-VIII	loop βGH11-βGH12	582-591	Yes
9	VR-IX	Ιοορ βΙ-βΙ1	704-709	No

# Supplementary Table 8. Primers for AAV capsid mutations cloning

Primer Name	Sequence (5'-3')
cap5-F	gcgcagccatcgacgtcagac
cap5-319A-R	gatggtggtggtggcgtcctgcaccg
cap5-443A-R	ctggactccgccagtggcatttgtgctcac
cap5-531A-R	gggttcgccggctgggcgttgaagatcatag
cap5-532A-R	gcccgggttcgccggggcgctgttgaagatcat
cap5-535A-R	gtggtgcccgggggccgccggctggctgttg
cap5-540A-R	gccctcgaggtacgtgccggtggtgcccgggt
cap5-542A-R	atgttgccctcgagggccgtggcggtggtg
cap5-543A-R	agcatgttgccctcggcgtacgtggcggtg
cap5-546A-R	ctggtgatgagcatggcgccctcgaggtac
cap5-697A-R	ggcaaagtccacaaaggcggggtcgttgtagtt
cap5-698A-R	cggggcaaagtccacggcctgggggtcgttgta
cap5-708A-R	ctggtggttctgtaggccccggtgctgtccg
cap5-710A-R	gataggtctggtggtggcgtattccccggtgct
cap5-712A-R	cgataggtctggcggttctgtattcccc
cap5-R	gaattccagcacactggcggccgttactagtggatcctagagcatggaaactagataag
cap5-319A-F	cggtgcaggacgccaccaccatc
cap5-443A-F	gtgagcacaaatgccactggcggagtccag
cap5-531A-F	ctatgatcttcaacgcccagccggcgaaccc
cap5-532A-F	atgatcttcaacagcgccccggcgaacccgggc
cap5-535A-F	cagccagccggcggccccgggcaccaccgc
cap5-540A-F	acccgggcaccaccggcacgtacctcgagggc
cap5-542A-F	caccaccgccacggccctcgagggcaacat
cap5-543A-F	caccgccacgtacgccgagggcaacatgct
cap5-546A-F	gtacctcgagggcgccatgctcatcaccag
cap5-697A-F	aactacaacgaccccgcctttgtggactttgcc
cap5-698A-F	tacaacgacccccaggccgtggactttgccccg
cap5-708A-F	acagcaccgggggcctacagaaccaccagac
cap5-710A-F	agcaccggggaatacgccaccaccagacctatc
cap5-712A-F	gaatacagaaccgccagacctatcggaa
cap1-F	gcgcagccatcgacgtcagac
cap1-263G-R	ctggccccgttgagccactggagatttg
cap1-268A-R	gtggttgtcgttgGCggcccccgttg
cap1-269A-R	gtagtggttgtcgGCgctggcccccg
cap1-272A-R	gtagccgaagtagGCgttgtcgttgc
cap1-383A-R	cacggcttggctgccGGCgttgagcgtcaggtag
cap1-385A-R	cccacggcttggGCgccattgttga
cap1-386A-R	tgaacgtcccacggcGGCgctgccattgttgag
cap1-503A-R	gttatattttgaagcaccagtGGCggtaaaattgctgttg
cap1-504A-R	gttatattttgaagcaccGGCccaggtaaaattgctg
cap1-590A-R	ccggtcgcagggGctgtgctgctgc

cap1-R	cttgagtccaaagccgcccat
cap1-263G-F	caaatctccagtggctcaacgggggccag
cap1-268A-F	caacgggggccGCcaacgacaaccac
cap1-269A-F	cgggggccagcGCcgacaaccactac
cap1-272A-F	gcaacgacaacGCctacttcggctac
cap1-383A-F	ctacctgacgctcaacGCCggcagccaagccgtg
cap1-385A-F	tcaacaatggcGCccaagccgtggg
cap1-386A-F	ctcaacaatggcagcGCCgccgtgggacgttca
cap1-503A-F	caacagcaattttaccGCCactggtgcttcaaaatatAAC
cap1-504A-F	cagcaattttacctggGCCggtgcttcaaaatataac
cap1-590A-F	gcagcagcacagCccctgcgaccgg

Primer Name	Sequence (5'-3')
AAVR-349A-F	ACACCTACGACTGGCAGCTGGCTACTCATCCTA
AAVR-349A-R	GCCAGCTGCCAGTCGTAGGTGTAGGTTTCTCC
AAVR-350A-F	ACCTACGACTGGCAGCTGATTGCTCATCCTAGA
AAVR-350A-R	CAATCAGCTGCCAGTCGTAGGTGTAGGTTTCT
AAVR-353A-F	GGCAGCTGATTACTCATCCTGCAGACTACAGTG
AAVR-353A-R	GCAGGATGAGTAATCAGCTGCCAGTCGTAGGTG
AAVR-354A-F	GCTGATTACTCATCCTAGAGCCTACAGTGGA
AAVR-354A-R	GCTCTAGGATGAGTAATCAGCTGCCAGTCGTA
AAVR-356A-F	TACTCATCCTAGAGACTACGCTGGAGAAATGG
AAVR-356A-R	GCGTAGTCTCTAGGATGAGTAATCAGCTGCC
AAVR-376A-F	ATCGAAGCTCACTCCAGGCGC GTATGAATTCA
AAVR-376A-R	GCGCCTGGAGTGAGCTTCGATAGTTTGAGGA
AAVR-378A-F	AGCTCACTCCAGGCCTGTATGCATTCAAAGTGA
AAVR-378A-R	GCATACAGGCCTGGAGTGAGCTTCGATAGTT
AAVR-395A-F	CCCATGGGGAAGGCTATGTGGCCGTGACAGTCA
AAVR-395A-R	GCCACATAGCCTTCCCCATGGGCATTTTGACCC
AAVR-397A-F	GGGAAGGCTATGTGAACGTGGCAGTCAAGCCAG
AAVR-397A-R	CCACGTTCACATAGCCTTCCCCATGGGCATTT
AAVR-399A-F	GCTATGTGAACGTGACAGTCGCGCCAGAGCCCC
AAVR-399A-R	GCGACTGTCACGTTCACATAGCCTTCCCCATG
AAVR-425A-F	GATCTCTTTGCCAACCACTGCTACAGTCAT
AAVR-425A-R	CAGTGGTTGGCAAAGAGATCTCCTGGAACT
AAVR-426A-F	CTCTTTGCCAACCACTTCTGCAGTCATTGA
AAVR-426A-R	GAGAAGTGGTTGGCAAAGAGATCTCCTGGAAC
AAVR-427A-F	TTGCCAACCACTTCTACAGCCATTGATGGCAG
AAVR-427A-R	GCTGTAGAAGTGGTTGGCAAAGAGATCTCCT
AAVR-429A-F	AACCACTTCTACAGTCATTGCTGGCAGTCAAAG
AAVR-429A-R	GCAATGACTGTAGAAGTGGTTGGCAAAGAGAT
AAVR-431A-F	ACTTCTACAGTCATTGATGGCGCTCAAAGCACTGAT
AAVR-431A-R	GCGCCATCAATGACTGTAGAAGTGGTTGGCAA
AAVR-433A-F	
AAVR-433A-R	GCTTGACTGCCATCAATGACTGTAGAAGTGGT
AAVR-434A-F	
AAVR-434A-R	
AAVR-435A-F	TTGATGGCAGTCAAAGCACTGCTGATGATAAAATC
AAVR-435A-R	GCAGIGCTIIGACIGCCAICAAIGACIGIAGA
AAVR-436A-F	
AAVR-436A-R	GCATCAGIGCTTIGACIGCCATCAATGACTG
AAVR-437A-F	GGCAGICAAAGCACIGATGATGCTAAAATCGTTCA
AAVR-437A-R	GCATCATCAGIGCTTIGACIGCCATCAATGAC
AAVR-438A-F	AGTCAAAGCACTGATGATGATGCAATCGTTCAGT

# Supplementary Table 9. Primers for AAVR mutations cloning

AAVR-438A-R	GCATCATCAGTGCTTTGACTGCCATCAAT
AAVR-442A-F	GATGATGATAAAATCGTTCAGGCCCATTGGGAAG
AAVR-442A-R	GCCTGAACGATTTTATCATCATCAGTGCTTTG
AAVR-464A-F	CTGAAGATACAGCCATATTAGCACTAAGTAAACTCG
AAVR-464A-R	GCTAATATGGCTGTATCTTCAGAAATCTTCTC