SUPPLEMENTAL TO

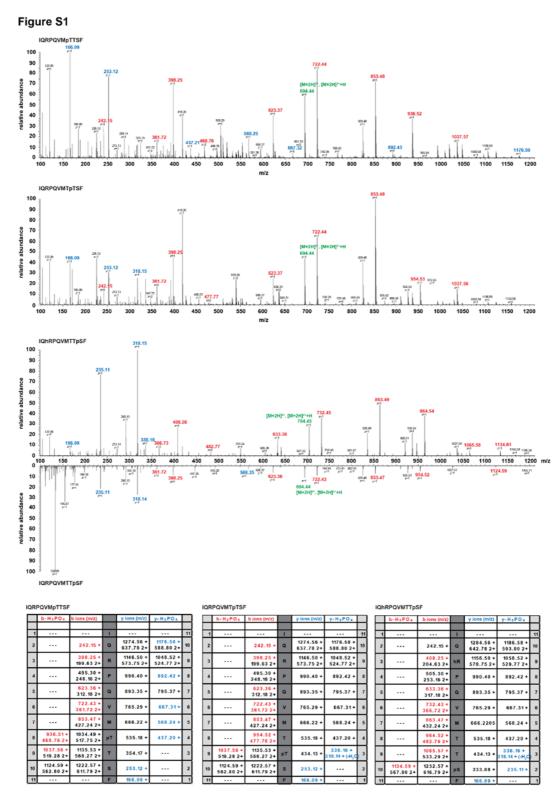


Fig. S1. Identification of the MUC17 C-terminal phosphorylation site S4492 within peptide IQRPQVMTTSF.

Caco-2 cells overexpressing MUC17-3TR were harvested and subjected to immunoprecipitation, followed by SDS-PAGE, trypsinization of bands and mass spectrometry analysis. MS/MS spectra of the sample and standard peptides (average

of 50 subsequent fragmentation spectra) for all three potential phosphorylation sites (T4490, T4491, S4492) are presented. A mirror plot of MS/MS spectra comparing the MUC17-3TR sample with standard peptide IQhRPQVMTTpSF demonstrates phosphorylation at S4492. Tables show identified fragments for all standard peptides with major identified fragments of the b/y ion series labeled in red/blue and theoretical mass to charge values in black.

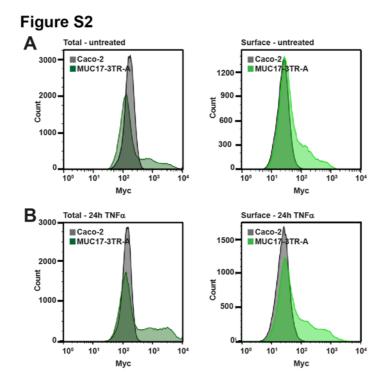


Fig. S2. Surface and total MUC17-3TR expressing Caco-2 can be clearly distinguished from control cells.

MUC17-3TR-A was overexpressed in Caco-2. Cells were left untreated (**A**) or stimulated with TNF α for 24 h (**B**) followed by FACS analysis using anti-Myc mAb. Surface localized MUC17-3TR was detected in unpermeabilized cells and total MUC17-3TR was analyzed following permeabilization. MUC17-3TR overexpressing Caco-2 showed a distinct signal for MYC-positive cells under all tested conditions. Histograms of a representative experiment are shown.

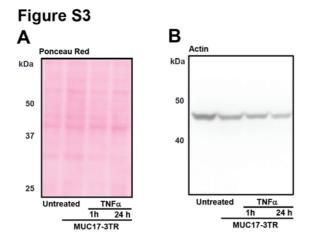


Fig. S3. Overexpression of MUC17-3TR in Caco-2 cells has no significant effect on actin and total protein amounts.

Caco-2 cells were transduced with MUC17-3TR adenovirus and stimulated with TNF α for 1 h or 24 h. Untreated Caco-2 cells served as controls. Whole cell lysates were analyzed by immunoblot. **A**) Ponceau Red staining confirmed equal protein amounts in all samples. **B**) Actin was detected using an anti-actin-C4 antibody. A small decrease was observed in adenovirus transduced cells in comparison to control cells.

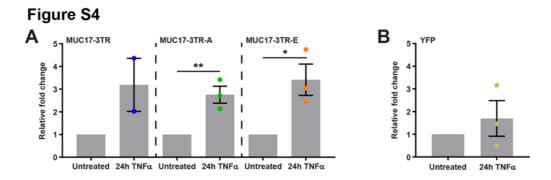


Fig. S4. MUC17-3TR mRNA quantities are increased following 24 h TNF α -treatment.

A) MUC17-3TR, MUC17-3TR-A and MUC17-3TR-E were overexpressed in Caco-2 and cells stimulated with TNFα for 24 h. Transcripts of MUC17 were subjected to quantitative real-time PCR analysis. Expression of MUC17-3TR-A and MUC17-3TR-E was significantly augmented following TNFα-stimulation. MUC17-3TR demonstrated a trended increase. (n=2-3; *P≤0.05 or **P≤0.01) **B)** YFP adenovirus was used in a control experiment to transduce Caco-2 cells, followed by TNFα-treatment for 24 h. Quantitative real-time PCR analysis showed no significant change of YFP mRNA quantities in stimulated samples when compared to controls. (n=3)

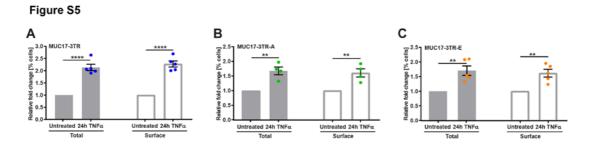


Fig. S5. 24 h TNF α -stimulation increases the number of surface and total MUC17-3TR positive cells.

Caco-2 cells overexpressing MUC17-3TR (**A**), MUC17-3TR-A (**B**) or MUC17-3TR-E (**C**) were stimulated with TNFα for 24 h and compared to untreated controls using FACS analysis. Surface localized MUC17-3TR was detected in unpermeabilized Caco-2 using anti-Myc mAb and total MUC17-3TR expression was quantified in permeabilized cells using the same antibody. Following TNFα treatment, an increased number of cells expressing total and surface localized MUC17-3TR was determined.

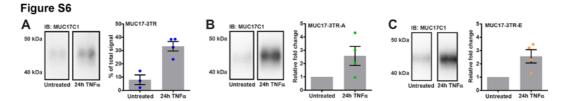


Fig. S6. MUC17-3TR is detected in spent cell culture medium following 24 h TNF α -stimulation.

A) Elevated levels of MUC17-3TR protein were detected in cell culture medium following incubation with TNF α for 24 h. MUC17-3TR in cell culture medium from Caco-2 was analyzed by immunoblot with anti-MUC17C1 pAb. Non-stimulated Caco-2 cells expressing MUC17-3TR served as control. Values represent the proportion of the sample signal compared to the total signal of all samples in an individual experiment. (n=3-4) **B, C**) Protein amounts of MUC17-3TR-A and MUC17-3TR-E in vesicles isolated from Caco-2 medium were increased following 24 h TNFα-stimulation. Caco-2 cells overexpressing MUC17-3TR-A and MUC17-3TR-E were treated with TNF α for 24 h and spent cell culture medium was harvested. Vesicles were isolated from the spent cell culture medium by ultracentrifugation and samples analyzed by gel electrophoresis and immunoblot using anti-MUC17C1 pAb. (n=4)

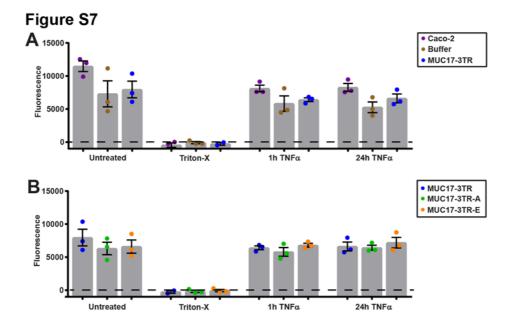


Fig. S7. Caco-2 cell viability is not impaired by transduction with MUC17-3TR adenovirus or TNF α -treatment.

Caco-2 cells were transduced with MUC17-3TR, MUC17-3TR-A or MUC17-3TR-E adenovirus and stimulated with TNF α . Alamar Blue® solution was added to the cell culture medium to analyze changes in metabolic activity using fluorescence measurements. Triton-X treated cells served as negative control. **A**) Caco-2 cells transduced with MUC17-3TR adenovirus or mock-treated with adenovirus storage buffer showed a small decrease in metabolic activity compared to controls. Treatment with TNF α had no significant effect on cell viability in comparison to untreated controls. (n=3) **B**) No differences in metabolic activity could be detected in Caco-2 cells overexpressing MUC17-3TR, MUC17-3TR-A or MUC17-3TR-E. Stimulation with TNF α had likewise no effect on cell viability. (n=3)

Figure S8

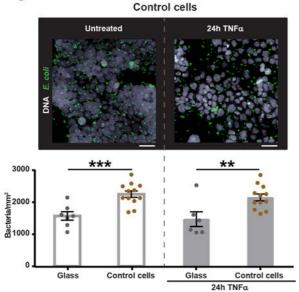


Fig. S8. Enteropathogenic *E. coli* specifically bind to Caco-2 cells.

Caco-2 cells were left untreated or stimulated with TNF α for 24 h, incubated with EPEC for 1 h and stained for *E. coli* (green) and DNA (gray). Samples were analyzed by confocal microscopy to quantify the number of bacteria adhering to glass cover slips and Caco-2 cells. Significantly more bacteria bound to the surface of Caco-2 cells than to glass. Scale bars, 50 μ m. (n=6-12, cells: 3 technical replicates per data point, glass: 1-3 technical replicates per data point, **P<0.001 or ***P<0.001)