

S6 Fig. Both RavA and RavS have autokinase activity and transfer the phosphoryl group to RavR in a RavR<sup>D496</sup> dependent manner. (a) RavA phosphorylates RavR in vitro. Autophosphorylation of the histidine kinase RavA and its recombinant derivative RavA<sup>H164A</sup> in the presence of [y-<sup>32</sup>P]ATP at room temperature for 10 min. RavA phosphotransfer to RavR was then carried out for 30 s. Assays contained 10 µM of soluble protein RavA or RavA<sup>H164A</sup>. Ten micromolar RavR or RavR<sup>D496A</sup> was added into the mixtures as indicated. The experiment was repeated three times. (b) RavA phosphorylated the recombinant protein RavR<sup>ΔEAL</sup> in vitro. Assays contained 15 μM of soluble protein RavA or RavA<sup>H164A</sup>. Five micromolar RavR<sup>ΔEAL</sup> or RavR<sup>ΔEAL</sup> or RavR<sup>ΔEAL</sup> was added as indicated. (c) RavS<sup>ΔN</sup> but not RavS<sup>ΔTrM</sup> possesses robust autophosphorylation activity. Five micromolar of RavS<sup>ΔTrM</sup> or RavS<sup>ΔN</sup> was added into the in vitro autophosphorylation mixture in the presence of [y-32P]ATP at room temperature for 10 min. (d-f) Detection phosphorylated HK and RR using Phos-tag acrylamide gel. (d) RavA phosphorylates RavR or RavR<sup>ΔEAL</sup> in vivo. RavA or RavA<sup>H164A</sup> was incubated with 2 mM ATP at 28 °C for 15 min, respectively. RavR (or RavR<sup>D496A</sup>) or RavR<sup>ΔEAL</sup> (or RavR<sup>Δ</sup> into mixtures for 2 min. (e – f) RavS $^{\Delta N}$  phosphorylates RavR $^{\Delta EAL}$  or RavR $^{EAL-AAA}$  in vitro. RavS $^{\Delta N}$  was autophosphorylated with 2 mM ATP at 28 °C for 15 min. A total of 20 μM various recombinant RavR proteins, including RavR<sup>ΔEAL</sup>, RavR<sup>ΔEAL</sup>(D496A),  $RavR^{\text{EAL-AAA}} or \ RavR^{\text{EAL-AAA}} or \ RavR^{\text{EAL-AAAA}} or \ RavR^{\text{EAL-AAA}} or \ RavR^{\text{EAL-AAA}} or \ RavR^{\text{EAL-AAA}} or \ RavR^{\text{EAL-AAA}} or \ RavR^{\text{EAL-AAAA}} or \ RavR^{\text{EAL-AAA}} or \ RavR^{\text{EAL-AAAA}} or \ RavR^{\text{EAL-A$ as indicated. The reactions were stopped with 3 × SDS loading buffer and the products were separated by 8% or 12% acrylamide gels at 4 °C, the gels were stained with Coomassie brilliant blue (d-f). Each experiment was repeated three times.