

S9 Fig. Overexpression ravS (OE-ravS) but not ravS^{H500A} rescues the swimming motility of the ravR^{ΔEAL} mutant.(a) Overexpression of ravS in bacterial cells. RavS was detected by Western blotting, RNAP was used as loading control. The experiment was repeated three times. (b-i) Phenotypic characterization of the effect of ravS overexpression. WT-EV (harboring a medium copy, broad-host pBBR1MCS2 vector), ravR^{ΔEAL}ΔravS-EV, ravR^{ΔEAL}ΔravS::OE-ravS, ravR^{ΔEAL}ΔravS::OE-ravS^{H500A} strains of *X. campestris* pv. campestris. (**b–c**) Bacterial virulence. Bacterial strains were inoculated onto plant leaves of Brassica olercaeae cv Zhonggan 11. Lesion length was recorded 10 days after inoculation (n = 30). (d) Production of extracellular polysaccharides (EPS). Bacterial strains were grown in TGM medium at 28 °C for 72 hours before EPS guantification, which was calculated as the dry weight of EPS vs. the dry weight of bacterial cells (n = 3). (e) Swimming motility. Bacterial strains were inoculated in NYG plates containing 0.15% agar and grew under 28 °C for 28 h. Average diameters of the migration zones were measured (n = 10). (f) Flagella of bacterial strains. Bacterial flagella were observed by transmission electron microscopy (TEM) after negative staining. Representative images of each strain are shown. Upper panel: bacteria population profile; Lower panel: a single bacterium. (g) Ratio of bacterial cells with flagella. For each strain, cells with flagella were counted (n = 100). (h) Flagellar length of bacterial strains (n = 30). (i) fliC mRNA level in bacterial strains. The amount of fliC mRNA was measured by gRT-PCR. Amplification of cDNA of tmRNA was used as internal control. In (c-e), (g-i), vertical bar indicates standard deviation; asterisk: significant difference, tested by Student's t-test ($P \le 0.05$) were measured. The experiment was repeated three times and a representative result is shown.