

1 **Supplementary Material for Sulkowski *et al***

2 A multi-protein complex anchors adhesive holdfast at the outer membrane of *Caulobacter*
3 *crescentus*.

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5 **Authors**

6 Nina I. Sulkowski,^{a,b,*} Gail G. Hardy,^{c,*,#} Yves V. Brun,^{c,d} Tanmay A.M. Bharat^{a,b,#}

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8 **Affiliations**

9 ^a Sir William Dunn School of Pathology, University of Oxford, Oxford OX1 3RE, UK

10 ^b Central Oxford Structural and Molecular Imaging Centre, Oxford OX1 3RE, UK

11 ^c Department of Biology, Indiana University, Bloomington, Indiana 47405, USA.

12 ^d Département de microbiologie, infectiologie et immunologie, Université de Montréal, C.P.

13 6128, succ. Centre-ville, Montréal (Québec) H3C 3J7, Canada

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16 **Running Title**

17 *Caulobacter crescentus* outer membrane holdfast anchor

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19 **Correspondence**

20 # Address correspondence to Gail Hardy, gahardy@indiana.edu or

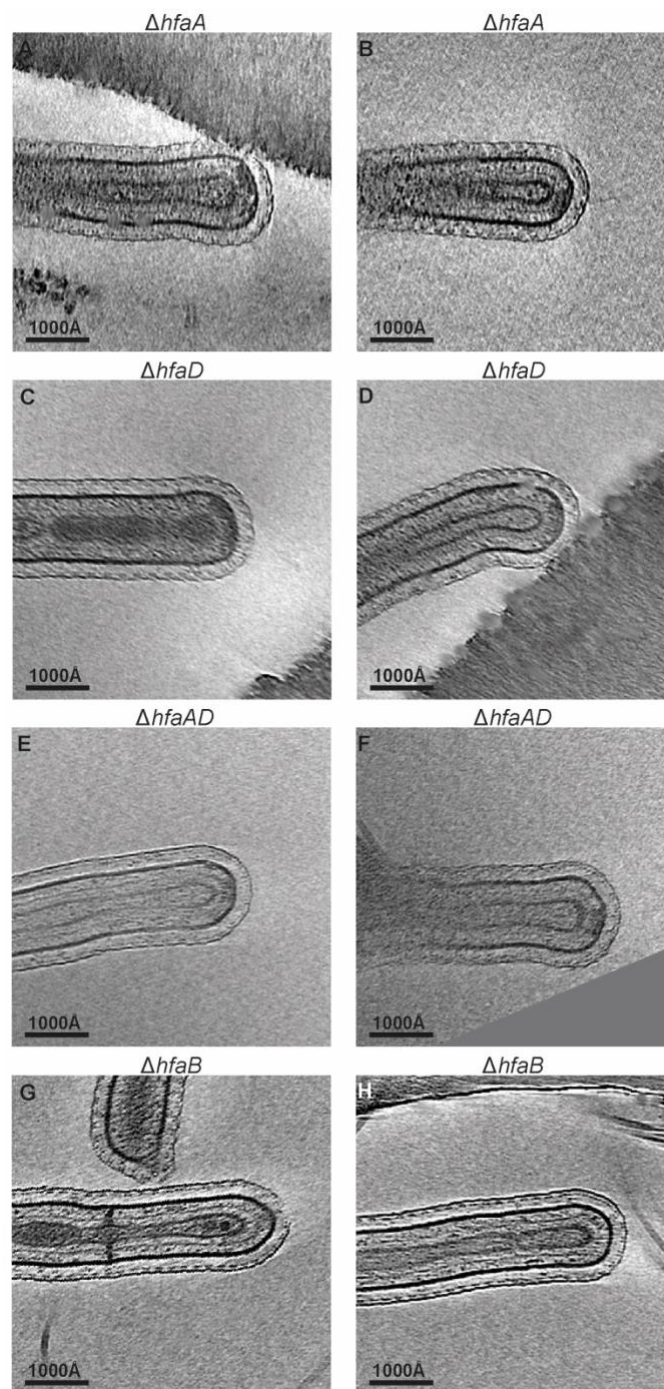
21 Tanmay A.M. Bharat, tanmay.bharat@path.ox.ac.uk

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23 * N.I.S. and G.G.H. contributed equally to this work.

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25 SUPPLEMENTARY FIGURES



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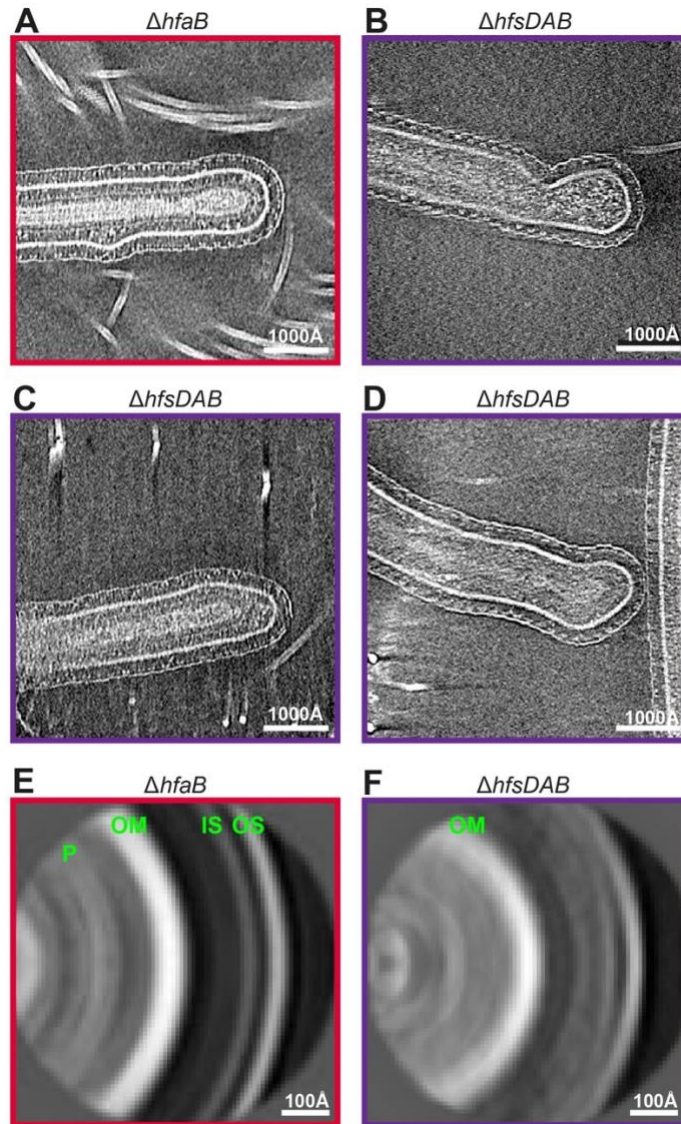
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29 **Figure S1. Cryo-ET gallery of *C. crescentus* stalk tips.**

30 Collection of holdfast anchor mutants imaged in this study. (A-B) $\Delta hfaA$ (C-D) $\Delta hfaD$ (E-F)

31 $\Delta hfaAD$ (G-H) $\Delta hfaB$.



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33 **Figure S2. Sub-tomogram averaging of the holdfast secretion complex mutant**

34 **($\Delta hfsDAB$).** (A) $\Delta hfaB$ strain displays uniform curvature along the stalk tip. Panels A is same

35 as Figure 4A, shown here for comparison with B-D panels. (B-D) $\Delta hfsDAB$ mutant displays

36 aberrations in stalk morphology, with visible narrowing at various regions near the tip. (E) Sub-

37 tomogram averaging analysis of $\Delta hfaB$ stalk tips. Panel E is same as in Figure 4C. The density

38 layers corresponding to the outer S-layer (OS), inner S-layer (IS), outer membrane (OM) and

39 the peptidoglycan (P) have been marked. (F) Sub-tomogram averaging analysis of $\Delta hfsDAB$

40 stalks reveals increased OM curvature in the final average.

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42 SUPPLEMENTARY TABLES

43 TABLE S1. Bacterial strains and plasmids used in this study.

Strain or plasmid	Description or construction	Source or reference
Strains		
<i>E. coli</i>		
S17-1	<i>E. coli</i> 294::RP4-2(Tc::Mu)(KM::Tn7)	(1)
Alpha-select TM	<i>deoR endA1 recA1 relA1 gyrA96 hsdR17</i> (τ_k^- m_k^+) <i>supE44 thi-1</i> Δ (<i>lacZYA-argFV169</i>) Φ 80 δ <i>lacZ</i> Δ M15 F ⁻	Bioline
<i>C. crescentus</i>		
CB15	Wild-type	(2)
YB1109	CB15 NY111d1 is a clone that has the Abs phenotype (stalk abscission Abs2)	(3)
YB2811	CB15 Abs2 <i>pstS</i> ::miniTn5	(4)
YB767	CB15N <i>pstS</i> ::miniTn5	(5)
YB7793	CB15 Abs2 <i>pstS</i> ::miniTn5, Δ <i>hfsG</i>	This study
YB7795	CB15 Abs2 <i>pstS</i> ::miniTn5, Δ <i>hfsDAB</i>	This study
YB7797	CB15 Abs2 <i>pstS</i> ::miniTn5, Δ <i>hfaB</i>	This study
YB8679	CB15 Abs2 <i>pstS</i> ::miniTn5, Δ <i>hfaA</i>	This study
YB8680	CB15 Abs2 <i>pstS</i> ::miniTn5, Δ <i>hfaD</i>	This study
YB8681	CB15 Abs2 <i>pstS</i> ::miniTn5, Δ <i>hfaA</i> , Δ <i>hfaD</i>	This study
Plasmids		
pNPTS138/9	pLitmus 39 derivative; with <i>nptI</i> , <i>sacB</i> and RK2 <i>oriT</i> sequences, and deleted <i>bla</i> gene; Km ^R	M.R.K. Alley, unpublished
pNPTS138 Δ <i>hfaA</i>	pNPTS138 parent vector containing 500 bp fragments upstream and downstream of <i>hfaA</i>	(6)
pNPTS138 Δ <i>hfaB</i>	pNPTS138 parent vector containing 500 bp fragments upstream and downstream of <i>hfaB</i>	(6)
pNPTS138 Δ <i>hfaD</i>	pNPTS138 parent vector containing 500 bp fragments upstream and downstream of <i>hfaD</i>	(6)
pNPTS138 Δ <i>hfsDAB</i>	pNPTS138 parent vector containing 500 bp fragments downstream of <i>hfsD</i> and downstream of <i>hfsB</i>	(6)
pNPTS138 Δ <i>hfsG</i>	pNPTS138 parent vector containing 500 bp fragments upstream and downstream of <i>hfsG</i>	(7)

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45 Table S1. Bacterial strains and plasmids used in this study.

46 Description of bacterial strains and plasmids used in this study is provided in this table.

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49 **TABLE S2. Oligonucleotides used in this study**

Oligo Name	Sequence (5' - 3')	Application
FhfaupA	GCGATCACGAAGCTTACGAGGTAG	<i>hfaA</i> in-frame deletion
RmhfadownA	GGCGTCAATGAATTCCGAGCGGAT	<i>hfaA</i> in-frame deletion
FhfaupD	GGCATCACCAAGCTTAACTACAAC	<i>hfaD</i> in-frame deletion
RmhfadownD	GCGGCCTGGGAATTCTAGTCCTGA	<i>hfaD</i> in-frame deletion
FupSphhfsG	CTGCCCGCATGCGTTCGGCCTC	<i>hfsG</i> in-frame deletion
RdwnBamHihfsG	GTTGAGATCGGATCCGTAGGTGAT	<i>hfsG</i> in-frame deletion
hfsDHindend	GATCAGCTTAAGCTTCTCCTCAGG	<i>hfsDAB</i> in-frame deletion
hfsBHindend	TCCATAGCCAAGCTTAGGCGCCGG	<i>hfsDAB</i> in-frame deletion
FhfaBup	GCCTTCACGCCGGGATCCAACAATCTGGGA	<i>hfaB</i> in-frame deletion
RmhfadownB	CACGTTGGCGAATTCCGACTGGCT	<i>hfaB</i> in-frame deletion

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52 **Table S2. Oligonucleotides used in this study**

53 Oligonucleotide sequences used in this study are listed in this table.

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56 **SUPPLEMENTARY MOVIE LEGENDS**

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58 **Movie S1. Tomogram of CB15 cell stalks.**

59 A movie containing sequential z-slices of a reconstructed tomogram of *C. crescentus* CB15
60 cell stalk tips is presented. This tomogram shows the presence of the holdfast anchor complex
61 at the stalk tip.

62

63 **Movie S2. Tomogram of a cell stalk tip from the *hfsG* deletion strain.**

64 A movie containing sequential z-slices of a reconstructed tomogram of *C. crescentus* $\Delta hfsG$
65 cell stalk tips is presented. This tomogram also shows the presence of the holdfast anchor
66 complex at the stalk tip.

67

68 **Movie S3. Tomogram of a cell stalk tip from the *hfaB* deletion strain**

69 A movie containing sequential z-slices of a reconstructed tomogram of *C. crescentus* $\Delta hfaB$
70 cell stalk tips is presented. This tomogram shows the absence of the holdfast anchor complex
71 at the stalk tip.

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