## SUPPLEMENTARY MATERIAL

Supplementary Table S1. SadC (TM) mutants defective for interaction with MotC isolated in a bacterial two-hybrid screen.

Plasmid isolate	Point mutation*	Predicted location of mutation in full length SadC
1	L82P	TM3
2	L94P	TM3
3	L134R	TM5
4	L29P L148M	TM1 Between TM5 and TM6
5	L32Q V59E	TM1 TM2
6	W36R L152P	TM1 TM5
7	L66R F136S	TM2 TM5
8	W67R F139L	TM2 TM5
9	P85A S140P	TM3 TM5
10	L116P F <b>136Y</b>	TM4 TM5
11	G22D L57Q I91T	TM1 TM2 TM3
12	L37Q A133V	TM1 TM5
13	L66P	TM2
14	Q86H	TM3
15	L94Q F166Y	TM3 TM6
16	Y112C	TM4
17	L117P	TM4
18	L30P W171G	TM1 TM6

19	R20Q	Before TM1
	L63P	TM2
20	F121L	TM4
	S140P	TM5
21	L98P	TM3
22	L13P	Before TM1
	A133D	TM5
- 22	N/ 4 AF	
23	Y 44F	
	L134P	TM5
24	D70C	Detween TM2 and TM2
24	D790	Between 11012 and 11015
25	Y44N	TM1
20	T49S	Between TM1 and TM2
	A90V	TM3
	I142F	TM5
26	L104Q	TM4
	R130H	TM5
	I142T	TM5
27	Y77F	Between TM2 and TM3
	L114Q	TM4
	L134Q	TM5
	S140P	TM5

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\*Bolded alleles were retested for interaction with MotC and other phenotypes as described in the text.

## 10 Supplementary Table S2. Strains used in this study.

Strain Name	Genotype/Description	Source
<i>E. coli</i> strains		
S17-1 λpir	<i>thi pro hsdR- hsdM+</i> $\Delta recA$ RP4-2::TcMu-Km::Tn7	(1)
BTH101	F, cya <sup>-99</sup> , araD139, galE15, galK16, rpsL1 (Str <sup>r</sup> ), hsdR2, mcrA1, mcrB1	Euromedex
S17-1 λpir	pmq30, SadC-3xFLAG KI construct, Gm <sup>R</sup>	This study
S17-1 λpir	pmq30, SadC-3xFLAG L94P KI construct, Gm <sup>R</sup>	This study
<i>P. aeruginosa</i> strains		
SMC 232	PA14 wild type (WT)	(Rahme et al., 1995)
SMC 6365	WT (gfp-motD)	(2)
SMC 6366	$\Delta bifA (gfp-motD)$	(3)
SMC 7562	$\Delta bifA \ \Delta motAB \ (gfp-motD)$	This study
SMC 3351	$\Delta bifA$	(4)
SMC 5770	$\Delta bifA \ \Delta motAB$	(3)
SMC 5769	$\Delta motAB$	(3)
SMC 5684	ΔmotCD	(3)
SMC 7563	$\Delta bifA \ \Delta motCD$	This study
SMC 8238	SadC-3xFLAG	This study
SMC 8239	SadC-3xFLAG L94P	This study
SMC 8240	WT pMotAB	This study
SMC 7659	$\Delta motCD$ pmq72 empty vector	This study
SMC 8241	Δ <i>motCD</i> pMotAB	This study
SMC 4045	$\Delta sadC$ pmq72 empty vector	(5)
SMC 8242	$\Delta sadC  pMotAB$	This study

Supplementary Table S3. Plasmids used in this study. 15

Plasmid	Description	Source
Name		
pMQ30	Shuttle vector for yeast cloning and Gram-negative allelic	(6)
	replacement, Gm <sup>r</sup>	
pmq30	SadC-3xFLAG cloned into pmq30, Gm <sup>R</sup>	This study
pmq30	SadC-3xFLAG L94P cloned into pmq30, Gm <sup>R</sup>	This study
pMQ72	Shuttle vector for yeast cloning and arabinose-inducible	(6)
	gene expression, Gm <sup>r</sup>	
pKT25	BACTH vector allowing fusion to the C-terminus of the	Euromedex
	<i>cyaA</i> T25 fragment, Kan <sup>r</sup>	
pKNT25	BACTH vector allowing fusion to the N-terminus of the	Euromedex
	<i>cyaA</i> T25 fragment, Kan <sup>r</sup>	
pUT18	BACTH vector allowing fusion to the N-terminus of the	Euromedex
	<i>cyaA</i> T18 fragment, Amp <sup>r</sup>	
pUT18C	BACTH vector allowing fusion to the C-terminus of the	Euromedex
	<i>cyaA</i> T18 fragment, Amp <sup>r</sup>	
pKT25-zip	Leucine zipper of GCN4 fused to T25 in pKT25, Kan <sup>r</sup>	Euromedex
pUT18C-zip	Leucine zipper of GCN4 fused to T18 in pUT18C, Amp <sup>r</sup>	Euromedex
pUT18C-sadC	Full length sadC cloned into pUT18C, Amp <sup>r</sup>	This study
pUT18C-roeA	Full length <i>roeA</i> cloned into pUT18C, Amp <sup>r</sup>	This study
pKT25-motA	Full length <i>motA</i> with a C-terminal 6xHis tag cloned into	(7)
	pKT25, Kan <sup>r</sup>	
pKT25-motC	Full length <i>motC</i> cloned into pKT25, Kan	(7)
pUT18C-	Transmembrane domain of <i>sadC</i> (amino acids 1-187)	This study

sadC(TM)	cloned into pUT18C, Amp <sup>r</sup>	
pUT18C-	Cytoplasmic domain of <i>sadC</i> (amino acids 188-375) cloned	This study
sadC(cyto)	into pUT18C, Amp <sup>r</sup>	
pUT18C-	Transmembrane domain of <i>roeA</i> (amino acids 1-197)	This study
roeA(TM)	cloned into pUT18C, Amp <sup>r</sup>	
pUT18C-	Cytoplasmic domain of <i>roeA</i> (amino acids 1-187) cloned	This study
<i>roeA</i> (cyto)	into pUT18C, Amp <sup>r</sup>	
pUT18C-	sadC(L82P) cloned into pUT18C, Amp <sup>r</sup>	This study
sadC(L82P)		
pUT18C-	sadC(L94P) cloned into pUT18C, Amp <sup>r</sup>	This study
sadC(L94P)		
pUT18C-	sadC(L134R) cloned into pUT18C, Amp <sup>r</sup>	This study
sadC(L134R)		
рКТ25-	<i>sadC</i> (L82P) cloned into pKT25, Kan <sup>r</sup>	This study
sadC(L82P)		
pKT25-	<i>sadC</i> (L94P) cloned into pKT25, Kan <sup>r</sup>	This study
sadC(L94P)		
рКТ25-	sadC(L134R) cloned into pKT25, Kan <sup>r</sup>	This study
sadC(L134R)		

## Supplementary Table S4. Oligonucleotide primers used in this study. 20

Primer Name	Primer Sequence*
B2H-sadC-F	NNNNNTCTAGAGATGCGCACAGACAAGCCTC
B2H-sadC-R	NNNNNGGATCCTCGGCACTGGTGACCTCCCA
SadC 1-187 BTH R	GGCGGGATCCTCGCGCATGCGTTGCCGCATC
SadC 188-375 BTH F	GGCGTCTAGAGCAGCGCCGCTATGCCTTG
SadC 188-375 BTH R	GGCGGGATCCTCGGCACTGGTGACCTCCCAG
RoeA 1-197 BTH R	GCGGGGATCCTCCCCGTGCCAGAGGATCAG
RoeA 198-398 BTH F	GGCGTCTAGAGCACGTGCGCAACCTGCGC
RoeA 198-398 BTH R	GGCGGGATCCTCCCGCAGGCTTTCCGCGAG
SadC L82P For	GTTACGCCGATCCCAGCC <u>C</u> GACCGAGCCGCAGGTGC
SadC L82P Rev	GCACCTGCGGCTCGGTC <u>G</u> GGCTGGGATCGGCGTAAC
SadC L94P For	GGTGGCGATCGCCTGGC <u>C</u> GACCTATTTCCTCTATCACGTC
SadC L94P Rev	GACGTGATAGAGGAAATAGGTC <u>G</u> GCCAGGCGATCGCCAC C
SadC L134R For	CGCCCGCTGTGCGGCGC <u>G</u> GGCGTTCATCGC
SadC L134R Rev	GCGATGAACGCC <u>C</u> GCGCCGCACAGCGGGCG
SadC F136Y For	CGCTGTGCGGCGCTGGCGT <u>A</u> CATCGCTTTTTCCG
SadC F136Y Rev	CCGGAAAAAGCGATG <u>T</u> ACGCCAGCGCCGCACAGC
B2H-MotA For	GGCGTCTAGAGATGTCAAAAATCATCGGCATCATCG
B2H-MotA Rev (used a His tagged template for this amplification)	GGCG GGATCC TC GTGGTGATGGTGGTGGTG
B2H-MotC For	GGCGTCTAGA G ATGGATGTGCTCAGCCTGGTC

B2H-MotC Rev	GGCGGGATCC TC GTCCATGAAGCCTTGCAGC
SadC native RBS pMQ72 F	caactetetactgtttetecataccegtttttttgggTCTTCAGGCGGGTAATTCG AATG
SadC-His pMQ72 R	taatetgtateaggetgaaaatetteteteateegeeteaGTGGTGATGGTGGTG GTGGGCACTGGTGACCTCCCAGG
Linker + 3x FLAG G- Block sequence	GGCGGCAGCGGCGGCGGCAGCGGCGGCGACTACAAAGACCATGACG GTGATTATAAAGATCATGATATCGACTACAAAGATGACGACGATAAA TAG
SadC-3x FLAG KI F_1 upstream	tgtaaaacgacggccagtgccaagcttgcatgcctgCCGATGCCCAGCTGGTTGACC
SadC-3x FLAG KI R_2	CGCTGTTTCAGAGGAGGCTTGTCTGTGCGCATGGGCTCCGTCC CGTAATGGCACCTGG
SadC-3x FLAG KI F_3	CGACTACAAAGATGACGACGATAAATAGGTGCCTGACATACG GGTCGGCGAGCGACGTC
SadC-3x FLAG KI F_4 downstream	ccatgattacgaattcgagctcggtacccggggatccTCAGGCGTGGGGGCAGGAACA G

22 \* In primer sequences, uppercase boldface letters indicate a 6xHis tag, lowercase letters indicate

23 sequence complementary to pMQ72, and underlined letters indicate point mutations.

## 26 Literature Cited.

- Simon R, Priefer U, Puhler A. 1983. A broad host range mobilization system for in vivo
  genetic engineering: transposon mutagenesis in gram negative bacteria. Nat Biotech
  1:784-791.
- Rahme LG, Stevens EJ, Wolfort SF, Shao J, Tompkins RG, Ausubel FM. 1995. Common
  virulence factors for bacterial pathogenicity in plants and animals. Science 268:1899-902.
- 32 3. Kuchma SL, Delalez NJ, Filkins LM, Snavely EA, Armitage JP, O'Toole GA. 2015.
- 33 Cyclic di-GMP-mediated repression of swarming motility by *Pseudomonas aeruginosa*
- 34 PA14 requires the MotAB stator. J Bacteriol 197:420-30.
- 4. Kuchma SL, Brothers KM, Merritt JH, Liberati NT, Ausubel FM, O'Toole GA. 2007.
- BifA, a cyclic-Di-GMP phosphodiesterase, inversely regulates biofilm formation and
  swarming motility by *Pseudomonas aeruginosa* PA14. J Bacteriol 189:8165-78.
- 38 5. Merritt JH, Brothers KM, Kuchma SL, O'Toole GA. 2007. SadC reciprocally influences
- 39 biofilm formation and swarming motility via modulation of exopolysaccharide
- 40 production and flagellar function. Journal of bacteriology 189:8154-64.
- 41 6. Shanks RM, Caiazza NC, Hinsa SM, Toutain CM, O'Toole GA. 2006. Saccharomyces
- 42 *cerevisiae*-based molecular tool kit for manipulation of genes from gram-negative
- 43 bacteria. Appl Environ Microbiol 72:5027-36.
- 44 7. Baker AE, Diepold A, Kuchma SL, Scott JE, Ha DG, Orazi G, Armitage JP, O'Toole GA.
- 45 2016. PilZ domain protein FlgZ Mediates cyclic di-GMP-dependent swarming motility
- 46 control in *Pseudomonas aeruginosa*. J Bacteriol 198:1837-46.
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