

Table S1. Sequence description of the Kv channel genes used in the study, related to Figures 3, 4, 9, S7 and S12.

Nucleotides variations with respect to the original refSeq are listed.

Synonymous modifications introduced in the cloning primers to improve primer quality are labeled as “modified nt”. Unreported missense variations shown in bold, have been corrected by mutagenesis and tested for functionality. These variations do not affect the activity of the corresponding channels (data not shown).

¹ *KCND3*-isoform 2 is the only isoform we detected in the rat brain. This isoform has a 19aa shorter C-tail compared to isoform 1.

² The amplified sequence of *KCNQ3* corresponds to a former sequence version (NM_031597.3). The new version (NM_031597.4) contains 117 supplementary nucleotides at the 5'-end that we did not detect in the rat brain.

³ This deletion was previously reported as XM_001071249, the only isoform we detected in the rat brain.

⁴ We amplified two sequences for rat *KCNH1* and *KCNQ2*. As these Refseqs are provisional in NCBI, we chose sequences homologous to human and mouse isoform 1: NM_031742 with an 81nt insertion at position nt952 for *KCNH1*; NM_133322 with a 54nt insertion at position nt1248 and a few nucleotide differences for *KCNQ2*.

Gene	Protein	NCBI Ref Seq	Nucleotide modifications compared to RefSeq
Rat Kv genes			
<i>KCNA1</i>	Kv1.1	NM_173095	
<i>KCNA2</i>	Kv1.2	NM_012970	nt396>C (synonymous, reported in Ensembl)
<i>KCNA3</i>	Kv1.3	NM_019270	modified nt15>A; nt297>C (synonymous, rs199378897)
<i>KCNA4</i>	Kv1.4	NM_012971	Codons nt126>GCT and nt925>GGC (missense, reported in Ensembl)
<i>KCNA5</i>	Kv1.5	NM_012972	modified nt15>T
<i>KCNA6</i>	Kv1.6	NM_023954	
<i>KCNA7</i>	Kv1.7	NM_001108914	modified nt15>T
<i>KCNA10</i>	Kv1.8	NM_0001191713	
<i>KCNB1</i>	Kv2.1	NM_013186	
<i>KCNB2</i>	Kv2.2	NM_054000	nt2292>A (synonymous, not reported)
<i>KCNC1</i>	Kv3.1	NM_012856	
<i>KCNC2</i>	Kv3.2	NM_139216	
<i>KCNC3</i>	Kv3.3	NM_053997	out of frame TAG insertion nt1981 (reported in Ensembl: Kcnc3-203)
<i>KCNC4</i>	Kv3.4	NM_001122776	modified nt15>T; nt657>T (synonymous), nt1817>G (missense) (both reported in Ensembl), nt1492>G (missense, not reported)
<i>KCND1</i>	Kv4.1	NM_001105748	modified nt12>A
<i>KCND2</i>	Kv4.2	NM_031730	nt1173>T (synonymous, not reported)
<i>KCND3</i>	Kv4.3	NM_001270963	(= isoform 2) ¹
<i>KCNF1</i>	Kv5.1	NM_001169104	nt31>G (missense, rs198924451)
<i>KCNG1</i>	Kv6.1	NM_001106545	
<i>KCNG2</i>	Kv6.2	NM_001107372	
<i>KCNG3</i>	Kv6.3	NM_133426	
<i>KCNG4</i>	Kv6.4	NM_001107435	
<i>KCNQ1</i>	Kv7.1	NM_032073	modified nt12>T, nt1402>G (missense, not reported), nt1912>G (missense, reported in Ensembl)
<i>KCNQ2</i>	Kv7.2	Kcnq2_201 ⁴	Δ(1526-1528), nt 2447>G (missense, not reported), 7 others synonymous variations (reported in NM_133322).
<i>KCNQ3</i>	Kv7.3	NM_031597 ²	nt1656>T, nt2127>C, nt2505>T (all synonymous: rs105980670, rs198097269, rs105394238)
<i>KCNQ4</i>	Kv7.4	XM_008764109	
<i>KCNQ5</i>	Kv7.5	NM_001134643	Δ(1246-1302) ³ , nt1770>A, nt1815>C, nt2232>C, nt2748>A (all synonymous, not reported)
<i>KCNV1</i>	Kv8.1	NM_021697	
<i>KCNV2</i>	Kv8.2	NM_001106370	nt306>A (synonymous, not reported)
<i>KCNS1</i>	Kv9.1	NM_053954	modified nt18>A
<i>KCNS2</i>	Kv9.2	NM_023966	
<i>KCNS3</i>	Kv9.3	NM_031778	modified nt12>C; nt327>T, nt900>T (synonymous, not reported)

<i>KCNH1</i>	Kv10.1	kcnh1_003 ⁴	
<i>KCNH5</i>	Kv10.2	NM_133610	
<i>KCNH2</i>	Kv11.1	NM_053949	nt156>T (synonymous, not reported), codon nt900>CCT (missense, reported in Ensembl)
<i>KCNH6</i>	Kv11.2	NM_053937	nt405>T, nt414>G, nt786>C (synonymous, rs107171774, rs106162519, rs105164542), nt2744>A (missense, rs105334634), nt1538>A (missense, not reported)
<i>KCNH7</i>	Kv11.3	NM_131912	
<i>KCNH8</i>	Kv12.1	NM_145095	
<i>KCNH3</i>	Kv12.2	NM_017108	
<i>KCNH4</i>	Kv12.3	NM_053630	modified nt15>A; nt1640>G (missense, not reported)
Mouse Kv genes			
<i>KCNA1</i>	Kv1.1	NM_010595	modified nt15>A
<i>KCNA4</i>	Kv1.4	NM_021275	modified nt6>G and nt12>A
<i>KCNA5</i>	Kv1.5	NM_145983	modified nt15>T and nt1794>T, and nt2557>A
<i>KCNA6</i>	Kv1.6	NM_013568	
<i>KCNB1</i>	Kv2.1	NM_008420	modified nt12>T and nt2557>A
Human Kv genes			
<i>KCNA1</i>	Kv1.1	NM_000217	
<i>KCNA4</i>	Kv1.4	NM_002233	modified nt9>G
<i>KCNA5</i>	Kv1.5	NM_002234	nt1149>C (synonymous, rs2359641)
<i>KCNA6</i>	Kv1.6	NM_002235	
<i>KCNB1</i>	Kv2.1	NM_004975	modified nt12>T

Table S2. New variants cloned from rat brain, related to Table S1

Brief description of seven new variants of the Kv genes cloned from rat brain.

Gene	Variant	Variation	Effect
<i>KCNA5</i>	A	$\Delta(779-859)$	change in reading frame leading to a premature STOP codon at nt846
	B	$\Delta(490-868)$	no change in reading frame but shorter protein (477 AA)
	C	$\Delta(792-858)$	change in reading frame leading to a premature STOP codon at nt900
<i>KCNC3</i>	A		premature STOP codon at nt1518 leading to a shorter isoform (506 AA)
<i>KCND1</i>	A	Different Exon 3	premature stop codon and shorter protein
<i>KCNQ1</i>	A	$\Delta(1733-1794)$	change in reading frame leading to a new C-terminus sequence
<i>KCNQ2</i>	A	$\Delta(1525-1527)$	1 missing codon

Table S3. Transcriptome analysis of the three host cell lines, related to Figures S4 and 9

Absolute count values (in FPKM) of the expression of ion channel-related genes in CHO-FT, HEK-FT or CV1-FT cell lines, obtained by transcriptome sequencing. Alignment was done on CHO genome for CHO-FT cell line, and on human genome for HEK-FT and CV1-FT cell lines. See attached xlsx file.

Table S4. Absolute count values of the multiplexed gene expression assay, related to Figure S3

See attached xlsx file.

Table S5. Cell counts, related to Figures 3, 5, 6, S7 and S9

Cell counts (N) for each ion channel group used for comparative chart of the Kv kinetic properties used in Figure 2, 3, 4, 5, 9, S7, S9, S13 and for overlay plots in Figure 6. See attached xlsx file.

Table S6. Cloning primers list, related to Method “IC gene cloning”

List of primers used for PCR amplification and cloning in pENTR/TOPO vector of rat, mouse and human Kv genes.

Rat genes

Gene	Accession n.	Forward primer (5' to 3')	Reverse primer (5' to 3')
<i>KCNA1</i>	NM_173095	CACCATGACGGTGATGTCAGGGGAGA	TTAAACATCGGTCAGGAGC
<i>KCNA2</i>	NM_012970	CACCATGACAGTGGCTACCGGAGACCCAGTGGAT	TCAGACATCAGTTAACATTTTGGTAATATTCAC
<i>KCNA3</i>	NM_019270	CACCATGACCGTGGTCCAGGGGACC	TTAGACATCAGTGAATATCTTTTGA
<i>KCNA4</i>	NM_012971	CACCATGGAGGTGGCAATGGTGAG	TCACACATCAGTCTCCACAGCCT
<i>KCNA5</i>	NM_012972	CACCATGGAGATCTCCCTTGTGCC	TTACAAATCTGTTTCAGCGCTAGT
<i>KCNA6</i>	NM_023954	CACCATGAGATCGGAGAAATCCCTGA	TCAAACCTCGGTGAGCATCCTT
<i>KCNA7</i>	NM_001108914	CACCATGCTATTTTACGTGCGGACACC	TCACACCTCGGTACCATGTGTT
<i>KCNA10</i>	NM_0001191713	CACCATGGATGTGTGTAGCTGGAGAG	TCACTTCTGGACTTCTCTGCAG
<i>KCNB1</i>	NM_013186	CACCATGACGAAGCATGGCTCGCG	TCAGATACTCTGATCCCTAGTGC
<i>KCNB2</i>	NM_054000	CACCATGGCAGAAAAGGCACCTCC	TCACATGCTGGTCTCAGAGTGG
<i>KCNC1</i>	NM_012856	CACCATGGGCAAGGGGACGAGA	TCAAGTCACTCTCACAGCCTCT
<i>KCNC2</i>	NM_139216	CACCATGGGCAAGATCGAGAACAACG	TTAAGTAAGAGATCTGGCCTCGG
<i>KCNC3</i>	NM_053997	CACCATGCTCAGCTCAGTGTGCGTCT	CTAGAGGATGGAGGGCAGGGTGC
<i>KCNC4</i>	NM_001122776	CACCATGATCAGCTCGGTTTGTGTC	TTAAAGAGCACAGGTGTGAGACA
<i>KCND1</i>	NM_001105748	CACCATGGCGGCAGGAGTGGC	TCACAGGAAGAGATCTTGACAG
<i>KCND2</i>	NM_031730	CACCATGGCAGCCGGTGTTC	TTACAAAGCAGACACCCTGACGA
<i>KCND3</i>	NM_001270963	CACCATGGCGGCAGGAGTTGC	TTAAATACACAGTGTATAATTTCTTCC
<i>KCNF1</i>	NM_001169104	CACCATGGAACCGGGGCTTGC	TCACTTGCAGCTCTGTAGCC
<i>KCNG1</i>	NM_001106545	CACCATGACCTGTTACCAGGAGACAA	TCAGTTGTTGCCCTAGTGTCTG
<i>KCNG2</i>	NM_001107372	CACCATGGCCCGGCTCCCGGGGCAC	TCAGGGTCCCACCGGGCCTG
<i>KCNG3</i>	NM_133426	CACCATGACCTTCGGGCGCG	TTAATTCAGGAACCTCAGCGGAG
<i>KCNG4</i>	NM_001107435	CACCATGCCATGTCTTCCCGAG	CTACATGTAGCGGCCTGGGGAG
<i>KCNQ1</i>	NM_032073	CACCATGGACACGGCTTCCCTCCC	TCAGGAGCCTTCATCAGGGCCTG
<i>KCNQ2</i>	NM_133322	CACCATGGTGCAAAAGTCGCGCAAC	TCACTTCTAGGCCCTGCCCAA
<i>KCNQ3</i>	NM_031597	CACCATGGGGCTCAAGGCTCGCA	CTAAGTGGGCTTGTGGAAGGGG
<i>KCNQ4</i>	XM_008764109	CACCATGGCCGAGGCCCCCGG	TCAGTCCATGTTGGTGTGACTG
<i>KCNQ5</i>	NM_001134643	F1-GCCGCCCCCTTACCATGCCCCGCCACCACGC	R1-AACTTCCGCTTTGCAACATGAAACTTCATGATTCTGA
		F2-TGCAAAGCGGAAGTTTAAAGAAACATTACGCCATAT	R2-GGCGCGCCACCCTTTTAGTTCAGTTTGACGTGAGGCA
<i>KCNV1</i>	NM_021697	CACCATGGATCTGTCACCCGCAACC	TTAAAACCAGAAATCATCTCCACC
<i>KCNV2</i>	NM_001106370	CACCATGCTGAAACAGAGCACCGAGA	TTAATTCCTTGCCTTGTGGCGGACTG
<i>KCNS1</i>	NM_053954	CACCATGGTGAGCGAGTTCCGGGTCC	TTAATAACTGTGAGATTTTGCAGGC
<i>KCNS2</i>	NM_023966	CACCATGACCCGCCAGAGCCTGTG	CTAATGTAGAGAGTCTGCTAAAC
<i>KCNS3</i>	NM_031778	CACCATGGTGTGTTGGCGAGTTTTC	TCATTTTGCAGTACAGTTCTCCA
<i>KCNH1</i>	Not available	CACCATGACCATGGCTGGCGG	TCAGCTTGGCCAAAAATGTCTC
<i>KCNH5</i>	NM_133610	F1-GCCGCCCCCTTACCATGCCGGGGGCAAGA GAG	R1-CTAAGGCCTTTTCGGGACCTGATATAAATTTCA GGA
		F2-CCCGAAAGGCCTTAGTGAACGAGTCATGGAT TAC	R2-GGCGCGCCACCCTTTTAAAGTTGATTTCATCTTTG
<i>KCNH2</i>	NM_053949	CACCATGCCGGTGGGAGGGGC	CTAACTGCCTGGATCTGAGCC
<i>KCNH6</i>	NM_053937	CACCATGCCAGTCCGAGGGG	CTAACTCCTTGTGAATCCAGGATCGGA
<i>KCNH7</i>	NM_131912	CACCATGCCTGTTTCGAGGGG	TTACTTTCCCGGAAGACCAGGAT
<i>KCNH8</i>	NM_145095	CACCATGCCGGTTATGAAAGGATTGCT	TTATACGTTTATGGCTTTGCTGT

<i>KCNH3</i>	NM_017108	F1- GCCGCCCCCTTCACCATGCCGGCCATGCCGGGG CTC	R1-TCGAGCATGCCGCTGCTTGAGGGGCTTGGGA TGC
		F2-GCAGCGCATGCTCGAGTACTTCCAAGCCACC TGG	R2-GGCGCGCCACCCTTTCAGACTCCTGTGCCCT CCT
<i>KCNH4</i>	NM_053630	F1- GCCGCCCCCTTCACCATGCCGGTCATGAAAGGG TTGCTG	R1-GACGATGCACTCGGATGAAGTCCTTGAGATC
		F2-CATCCGAGTGCATCGTCTGCCCGCCCACT	R2-GGCGCGCCACCCTTTCAGTGAATGTGTCTG ACCCAG

Mouse genes

Gene	Accession n.	Forward primer (5' to 3')	Reverse primer (5' to 3')
<i>KCNA1</i>	NM_010595	CACCATGACGGTGATGTCAGGGGAGA	TTAAACATCGGTCAGGAGC
<i>KCNA4</i>	NM_021275	CACCATGGAGGTGGCAATGGTGAG	TCACACATCAGTCTCCACAGCCT
<i>KCNA5</i>	NM_145983	CACCATGGAGATCTCCCTTGTGCC	TTACAAATCTGTTTCACGGCTAGT
<i>KCNA6</i>	NM_013568	CACCATGAGATCGGAGAAATCCCTGA	TCAAACCTCGGTGAGCATCCTT
<i>KCNB1</i>	NM_008420	CACCATGCCGGCGGGTATGACGAA	TCAGATACTCTGATCCTAGTGC

Human genes

Gene	Accession n.	Forward primer (5' to 3')	Reverse primer (5' to 3')
<i>KCNA1</i>	NM_000217	CACCATGACGGTGATGCTGCGGGAGAA	TTAAACATCGGTCAGTAGCTTGC
<i>KCNA4</i>	NM_002233	CACCATGGAGGTGGCAATGGTGAG	TCACACATCAGTCTCCACAGCCT
<i>KCNA5</i>	NM_002234	CACCATGGAGATCGCCCTAGTGC	TCACAAATCTGTTTCCCGGCTGG
<i>KCNA6</i>	NM_002235	CACCATGAGATCGGAGAAATCCCTTAC	TCAGACCTCCGTGAGCATTCTTT
<i>KCNB1</i>	NM_004975	CACCATGCCGGCGGGTATGACGAA	TCAGATGCTCTGATCTCGTGTGC

Table S7. Screening primers list, related to Method “Cell line validation by RT-PCR”

List of primers used for the amplification of the Kv genes by RT-PCR for cell lines validation, with expected amplification size.

Gene	Species	Forward primer (5' to 3')	Reverse primer (5' to 3')	Size (bp)
<i>KCNA1</i>	rat, mouse, human	CATGACCACTGTGGGATACG	AGCAACTGAGCCTGCTCTTC	177
<i>KCNA2</i>	rat	TTATCACCTGTGGGACAGAG	TGACCCCAATGAAGAGGAAG	219
<i>KCNA3</i>	rat	GGAGACCCTGTGCATCATCT	TCAGGATGGCCAGTGACATA	197
<i>KCNA4</i>	rat, mouse, human	CGGGTGTCTTAACCATTGCT	TGACTTGTCCCCAGGAAG	188
<i>KCNA5</i>	rat, mouse	CTCCGACGTCTGGACTCAAT	CCCGGAGATGTTTATGAGGA	248
	human	CGAGGATGAGGGCTTCATTA	CTGAACCTCAGGCAGGGTCTC	186
<i>KCNA6</i>	rat, mouse	CCGGTACCGGTCAATTGTCTC	TCTGCATAAGCTCGATGTGG	208
	human	GCGGCTGGTGATCAATATCT	CTCCTCCAGGAAAATGTCCA	238
<i>KCNA7</i>	rat	TTCTTGTGGTGGAGACCCCT	TAGGATGGCCAGGGACATAG	205
<i>KCNA10</i>	rat	ATCTTCAAGCTCTCCGACA	GTTGTGATGGTGACCACAGC	212
<i>KCNB1</i>	rat	AAGGAGCAGATGAACGAGGA	GCGATGGTGGACAGGACGAT	194
	mouse	AAGGAGCAGATGAACGAGGA	GCAATGGTGGAGAGGACAAT	194
	human	AAAGAGCAGATGAACGAGGA	GCAATGGTGGAGAGGACGAT	194
<i>KCNB2</i>	rat	CCCTTCTATCCCATCATT	TTGATATTGGCCGACTCTCC	208
<i>KCNC1</i>	rat	GAAGACTGCCCCACATAGA	CACTCTCACAGCCTCTGTCC	204
<i>KCNC2</i>	rat	GTACCCCAAAACATGGTCAG	GTAGGTGAGCTTGCCAGAGG	192
<i>KCNC3</i>	rat	TTATGGGATTGCCCTCTCTG	TGATCGGGCTTGTCTTCT	183
<i>KCNC4</i>	rat	CAGAAGCTTCCCAAGAAACG	TAGCGTCACCATTCTGCTTG	184
<i>KCND1</i>	rat	CCTACGTGTGTTTCGGGTCT	GGGATGCTTGTGAAGTTGGT	201
<i>KCND2</i>	rat	TGAGTGCATCTCGGCTTATG	TCTGCCTAGCAGTCATGGTG	182
<i>KCND3</i>	rat	GGCTACACCCTGAAGAGCTG	GCCAAATATCTTCCCAGCAA	222
<i>KCNF1</i>	rat	CCTGCAGACCCTCACCTACG	CACTGTGGTCATGGTATGA	186
<i>KCNG1</i>	rat	CGAGAATGAGATGGCAGACA	ATCAGCACCCGTTCTTGTTT	243
<i>KCNG2</i>	rat	CATCCTGCCCTTCTACGTGT	GAGGAAGAGCAGCAGCAGTC	214
<i>KCNG3</i>	rat	CGATGCTACCGAGAGATGGT	ACGATCCCACTGACAACACA	245
<i>KCNG4</i>	rat	CTGGAGCACCTGGATGCCT	ATGCCCCAGTAGCTCAGCTC	215
<i>KCNQ1</i>	rat	GGCGCATGCAGTACTTTGTA	TGTCACCTTGTCTTCCACCC	245
<i>KCNQ2</i>	rat	TCGACATCATGGTGTGATT	CCAATGTACCAGCAGTCAC	205
<i>KCNQ3</i>	rat	ATCGGGTTTCGCTTCTAAT	TCTTCTGTCTATGGGGTCTCC	214
<i>KCNQ4</i>	rat	AGTATGCGCTTCTCGAGAT	GAGTTGGCATCCTTCTCAGC	182
<i>KCNQ5</i>	rat	TTGTTGCCTCCAAGGAAAAC	GAGCCACTTGACTCACTGCC	208
<i>KCNV1</i>	rat	TCCGACAAAAGCTCTGGGAT	AGGAAGTGGCACCTGTCTTT	238
<i>KCNV2</i>	rat	ACTCCAACCCAGCTGTCTAC	CTTTGCCTCGTAGTCATCGC	162
<i>KCNS1</i>	rat	ACAAGTGTTCGCTCATGC	GTCGAAGCCCAGTTCTTTT	193
<i>KCNS2</i>	rat	CTGAGCTCTTCCCCTACGTG	TCTCCTGCTCAGTTTCCACT	177
<i>KCNS3</i>	rat	GAAGTGGATGACCCGTGTGCT	ACGTGGCATAGAAGGGAATG	163
<i>KCNH1</i>	rat	TGGTCCCTTACAACGTCTCC	ACTGAACAGGCTGCTGATGC	271
<i>KCNH5</i>	rat	ATGCCAACCAACCGTTAC	TCAGCTCTCATGTCTTGGG	181
<i>KCNH2</i>	rat	TGGATGCACTCCAGAGACAG	AACCTGAGAAAGCGAGTCCA	209
<i>KCNH6</i>	rat	GAAGCTGGACCGCTATTCTG	GGCGGTGACATACTTGTCTT	235
<i>KCNH7</i>	rat	ATCCCGCAACTCACTCTGAA	CACTGCTTTGAAAGGGCTGT	234
<i>KCNH8</i>	rat	AAAACCAAGCAGGAAGCTGA	ACGCAGCTCCAATCTCTGAT	238
<i>KCNH3</i>	rat	GAGTACCTCATTACCAAGG	GGGGCAACTCAGGGTACAG	250
<i>KCNH4</i>	rat	AATGTACAGCCATCATCCA	GTTGACGGCCCATGTAGTCT	153
<i>GAPDH</i>	rat	TTGTGATGGGTGTGAACCAC	GGATGCAGGGATGATGTCTT	238
<i>GAPDH</i>	human	CGAGATCCCTCCAAAATCAA	TGACGATCTTGAGGCTGTTG	206

Table S8. APC automation steps, related to Method “APC automation”

Steps	Sub-steps	Comment
Initialize	Initialize amplifier and Patchliner	Specify stimulus protocol file. Location of IC/EC/SE solutions
	Initialize log file with meta data	User, experiment details, cell line, passage, induction, temperature, cell count
Prepare chips	Fill IC and EC	Volume 10 μ l, Speed 40
Catch cells	Set pressure	-100 mBar
	Set Vhold -20 mV	
	Get cells from cell hotel	
	Wait 30 s	
	Set Vhold -80 mV	
Make seal	Add seal enhancer	
	Wait for 30 s	
Whole cell patch	Apply atmospheric pressure	
	Wait for whole cell	Suction is applied to the cells until the whole cell configuration is reached
	Wash with EC	To remove seal enhancer
Whole cell setting	Auto C-slow and R-series	Apply auto compensation for C-slow and R-series
	Seal quality control	If seal resistance is less than 100 M, then the channel gets turned off
Current clamp	Record resting potential of cells	
Voltage clamp	Apply Activation protocol	
Protocol loop	Auto C-slow	Apply auto compensation for C-slow
	Activation protocol	
	Ramp	
	Deactivation	
	AP	
	Inactivation	
	Wash with EC	
End loop		End loop if repetition >2
	Recovery	
Save files	Save files in format YYMMDD_00X	Where YY is year, MM is month, DD is date and X is the experiment number for a given day
Clean up	Clean pipettes and move to next recordings	