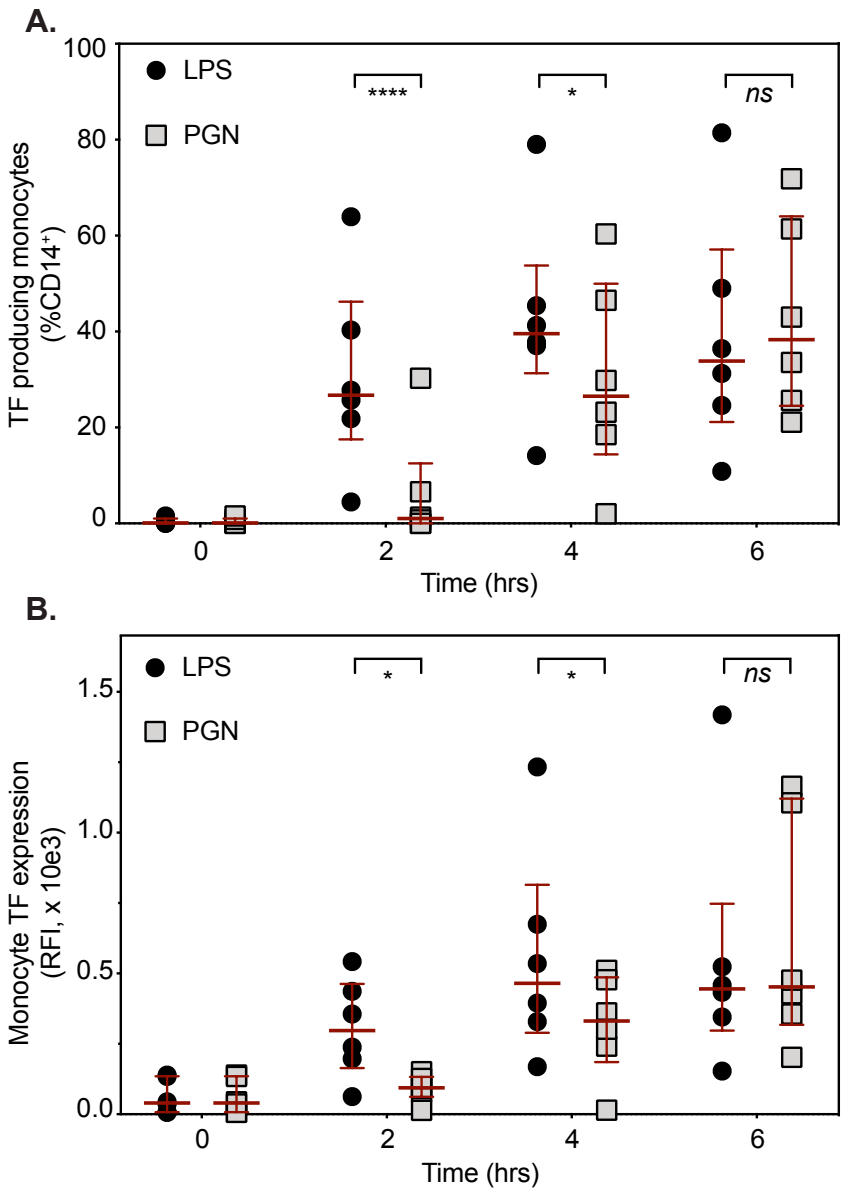
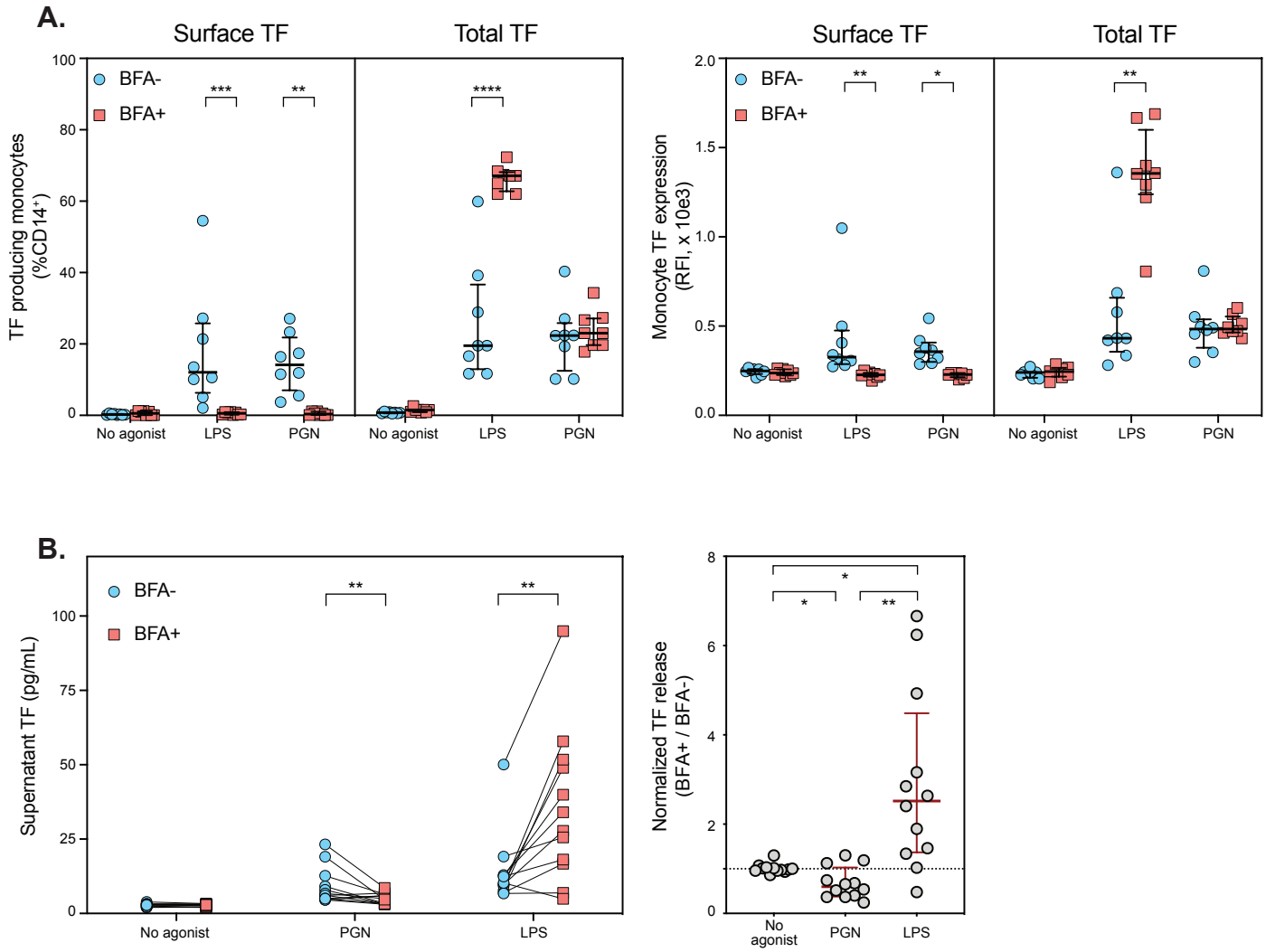
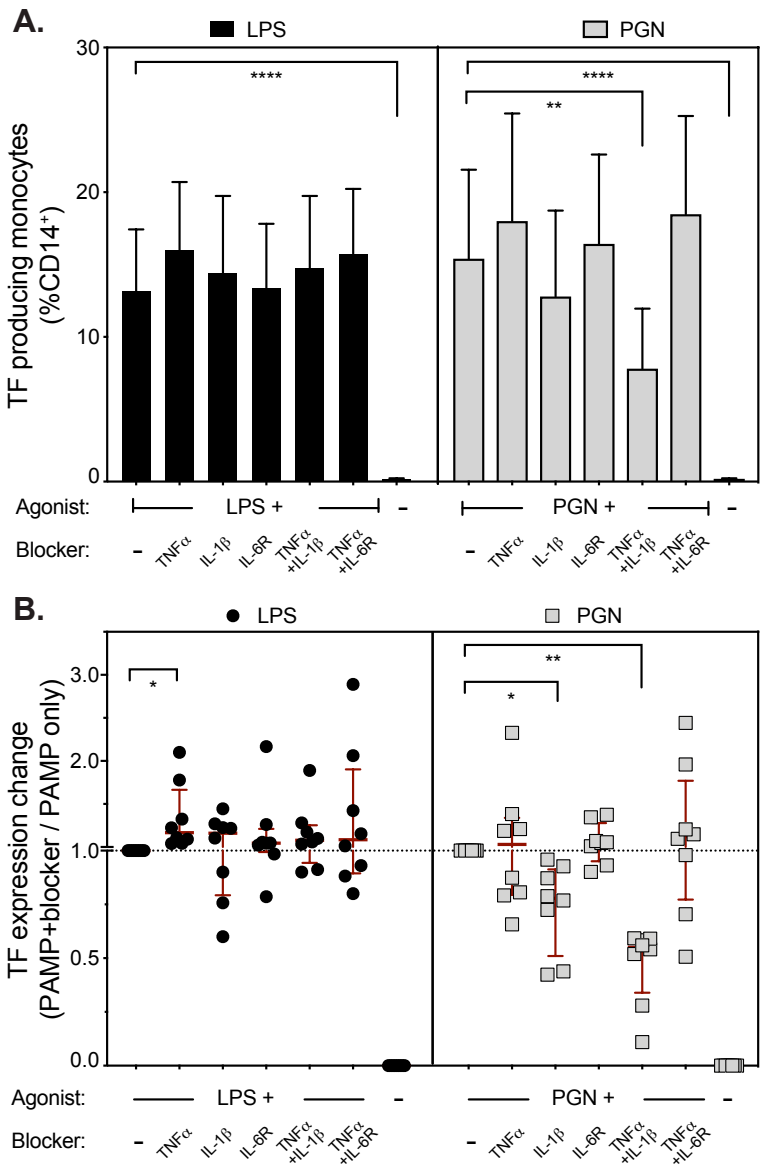


Supplemental figure S1. Pairwise comparison of kinetics of LPS- and PGN-induced monocyte procoagulant responses. Freshly isolated PBMCs were stimulated in parallel with either LPS (10 µg/mL, black circles) or PGN (10 µg/mL, grey squares) for up to 6 hours, and TF expression was quantified by flow cytometry. Individual procoagulant responses are depicted as (A) frequency of procoagulant monocytes and (B) monocyte TF fluorescence intensity (geometric mean). Horizontal bars depict median responses and interquartile range. Differences between groups were assessed by RM two-way ANOVA with Sidak's multiple comparison test. (*P < .05, ****P < .0001, *ns* not significant)



Supplemental figure 2. Differential effect of brefeldin A (BFA) on TF cellular localization and release from PAMP-stimulated monocytes. (A) Freshly isolated PBMCs were stimulated in parallel for 6 hours with either LPS (1 µg/mL) or PGN (10 µg/mL) in the absence (blue circles) or presence of brefeldin A (3 µg/mL, red squares). Cell associated TF was detected before (surface TF) or after cell permeabilization (total TF). Individual procoagulant responses, either frequency of procoagulant monocytes (left) or TF fluorescence intensity (right) are shown, while horizontal bars represent median responses and interquartile range. Differences between groups were assessed by RM two-way ANOVA with Sidak's multiple comparison test. (*P < .05,**P < .01, ***P < .001, ****P < .0001). (B) Freshly isolated PBMCs were stimulated for 6 hours with either PGN (10 µg/mL) or LPS (1 µg/mL) in the absence (blue circles) or presence of brefeldin A (3 µg/mL, red squares). After stimulation, cells were spun and supernatants were transferred to new microcentrifuge tubes and kept frozen (-20ºC) until analysis. TF in cell supernatants was quantified using commercial ELISA kits (Quantikine TF ELISA, R&D Systems) according to manufacturer protocol. Pairwise representation of the differential effect of BFA on TF release (left panel) and normalized changes in TF secretion after BFA treatment (right panel) are shown. Differences between groups were assessed by ratio-paired *t* tests or RM one-way ANOVA with Holm-Sidak's multiple comparison test. (*P < .05,**P < .01)



Supplemental figure 3. Pairwise comparison of cytokine neutralization effect on LPS- and PGN-induced monocyte procoagulant responses. Freshly isolated PBMCs were stimulated in parrallel with either LPS (1 μg/mL) or PGN (10 μg/mL) in the absence or presence of the stated neutralizing antibodies, and TF expression was quantified by flow cytometry. (A) Pairwise representation of cytokine neutralization effect on the frequency of procoagulant monocytes induced by LPS (left panel, black bars) or PGN (right panel, grey bars). (B) Normalized changes in monocyte TF antigen expression induced by LPS (left panel, black circles) or PGN (right panel, grey squares) in the presence of neutralizing antibodies are shown. Differences between paired groups were analyzed by ratio-paired *t* tests. (*P < .05, **P < .01, ****P < .0001)