Supporting Information

Variant Bacterial Riboswitches Associated with Nucleotide Hydrolase Genes Sense Nucleoside Diphosphates

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the spontaneous RNA cleavage products generated during in-line probing of 5'-³²P-labeled 129 *nudix* RNA. In-line probing experiments contained either no ligand (–), 1 mM, or 100 μ M of the nucleoside di- or triphosphate indicated. Additional annotations are described in the legend to **Figure 2**.



Figure S2. The *ykkC* subtype 2c RNA 126 *nudix* responds differently to ADP and CDP. (A) Sequence and secondary structure of the 126 nucleotide RNA derived from a *nudix* gene form *B. cellulosilyticus*. In-line probing data used to infer structural modulation with ADP were derived from the in-line probing reactions depicted in C. Other annotations are as described for **Figure 2A**. (B) Sequence and secondary structure model depicted in A, but representing the in-line probing data for CDP. Note that CDP is added to the in-line probing assay depicted in C at a concentration below its K_D value (**Figure 4C**), and thus the extent of structural modulation is likely to be poorer than if it were saturated with ligand. Annotations are as described for A. (C) PAGE analysis of the products of in-line probing of 5′ ³²P-labeled 126 *nudix* RNA, as depicted in panels A and B, in the presence of ADP or CDP. In-line probing experiments contained either no

ligand (-) or 2 mM of the compound indicated. Annotations are described in the legend to Figure 2B.

Sherlock et al.



Figure S3. Nucleoside diphosphate binding by the 130 *had* RNA from *L. mesenteroides*. (A, B) PAGE analysis of the products of in-line probing of 5' ³²P-labeled 130 *had* RNA (**Figure 4B**) in the absence of ligand (–), or in the presence of either 1 mM or 100 μ M of the nucleoside di- or triphosphate indicated. Annotations are described in the legend to **Figure 2B**. (C) PAGE analysis of the products of in-line probing of 5' ³²P-labeled 130 *had* RNA in the absence of ligand (–) or in the presence of 1 mM of the compound indicated. Annotations are described in the legend to **Figure 2B**.



Figure S4. Ligand binding by the 126 *nudix* RNA from *B. cellulosilyticus*. (A, B) PAGE analysis of the products of in-line probing of 5' 32 P-labeled 126 *nudix* RNA (**Figure S1**) in the absence of ligand (–), or in the presence of 1 mM or 100 µM of the nucleoside di- or triphosphate compound indicated. Annotations are described in the legend to **Figure 2B**. (C) PAGE analysis of the products of in-line probing of 5' 32 P-labeled 126 *nudix* RNA in the presence of either no ligand (–) or 1 mM of the compound indicated. Annotations are described in the legend to **Figure 2B**.



Figure S5. Nucleoside diphosphate binding by the 126 *nudix* RNA from *B. cellulosilyticus*. PAGE analysis of the products of in-line probing of 5' 32 P-labeled 126 *nudix* RNA (**Figure 4A**) in the presence of increasing CDP (A) or ADP (B) concentrations. In-line probing experiments contained either no ligand (–) or ligand ranging from 5 M to 100 nM. Annotations are described in the legend to **Figure 2B**.



Figure S6. Nucleoside diphosphate binding by the 131 *nudix* RNA from *C. botulinum*. (A, B, C) PAGE analysis of the products of in-line probing of 5′ ³²P-labeled 126 *nudix* RNA (**Figure 3A**) in the presence of various concentrations of ADP, CDP, or dADP, respectively. In-line probing

experiments contained either no ligand (–) or ligand ranging from 5 M to 100 nM. Annotations are described in the legend to **Figure 2B**.



Figure S7. Nucleoside diphosphate binding by the 129 *nudix* RNA from *S. kocurii*. (A, B, C) PAGE analysis of the products of in-line probing of 5' ³²P-labeled 129 *nudix* RNA (**Figure 2A**) in the presence of various concentrations ADP, CDP, or dADP, respectively. In-line probing

experiments contained either no ligand (–) or each compound ranging from 5 mM to 100 nM. Annotations are described in the legend to **Figure 2B**.



Figure S8. Nucleoside diphosphate binding by the 130 *had* RNA from *L. mesenteroides*. (A, B, C) PAGE analysis of the products of in-line probing of 5'-³²P-labeled 130 *had* RNA (**Figure 4B**) in the presence of various concentrations of ADP, CDP, or dADP, respectively. In-line probing experiments contained either no ligand (–) or each compound ranging from 5 mM to 100 nM. Annotations are described in the legend to **Figure 2B**.



Figure S9. Nucleoside diphosphates are selectively recognized by only subtype 2c *ykkC* RNAs. (A) Sequence and secondary structure of the 112 nucleotide ppGpp riboswitch RNA derived from the *ilvE* gene of *T. oceani*. Data collected in B were used to determine regions of constant or reduced scission upon the addition of ppGpp to in-line probing reactions. (B) PAGE analysis of the products of in-line probing of 5' ³²P-labeled 112 *ilvE* as depicted in A in the presence of ligands for previously-validated riboswitches represented by *ykkC* motif RNAs. (C) PAGE analysis of the products of in-line probing of 5' ³²P-labeled 126 *nudix* as depicted in **Figure 4A** in the presence of ligands for previously-validated riboswitches represented by *ykkC* motif RNAs. In-line probing experiments contained either no ligand (–) or 1 mM of the compound indicated. Annotations are described in the legend to **Figure 2B**.

Table S1.	Sequences	of Synthetic	DNAs used	in this Study.
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Name	Sequence	Annotation	
	TAATACGACTCACTATAGGTATTGAA	Forward template for	
HS005	CTGGGACTAGATAGCTAGGGTTCCGC	transcription of the WT, M2	
	CTTAATATATTAAGGGTTTGTGACCA	and M3 C. botulinum 131	
	AGCGCTATAGCTCTTGTTTTG	nudix RNAs	
	AACACGCTGAACCTTCCCCCTCGGGC	Reverse template for	
MES140	ATTTTATTCCAATAGGTGCCTCTTGC	transcription of the WT C.	
	AAAACAAGAGCTATAGCGC	botulinum 131 nudix RNA	
	TAATACGACTCACTATAGGATAGGA	Forward tomplata for	
45006	ACTGGAAAATTGCAGCTAGGGTTCCG	transcription of the WT S	
115000	CTATCTAGGTAGGTCAGTGACCGAGG	kogurii 120 nudir D NA	
	GCTGCATCTTTTAC		
	ATACGCTGAACCTTTCCCCTCGGACC	Reverse template for	
MES142	TTTTATGTCATTAGGTGCCTTTTACCT	transcription of the WT S.	
	ATATGTAAAAGATGCAGCCCTCGG	kocurii 129 nudix RNA	
HS015	TAATACGACTCACTATAGGTAATGAA	Forward template for	
	CTGGAAAACTGTAGCTAGGGTTCCGC	transcription of the WT B	
	TAACGATAGGTCTGTGACCGAGGGCT	cellulosilyticus 126 nudix RNA	
	ACAT		
HS016	ATACGCTGAACCTTTCCCCTCGGACC	Pavarsa tamplata for	
	TTTTATGTCAATAGGTGCCTTTTACTA	transcription of the WT B	
	TCGTAAAAGATGTAGCCCTCGGTCAC	cellulosilyticus 126 nudix RNA	
	AGACCT	centrostryneus 120 main 1141	
MES289	TAATACGACTCACTATAGGAATTACA	Forward template for	
	AACGAACTGGAAAACTGGAGCTAGG	transcription of the WT L.	
	GTTCCGCAATATGGTCAGTGACCGAG	mesenteroides 130 had RNA	

	GGCTCCATC	
MES266	AAAACGCTAAACCTTTCCCCTCGGAT TTTTTATATCAATAGGTGCCTTTTGA ATATTTCAAAAGATGGAGCCCTCGGT CACTG	Reverse template for transcription of the WT <i>L.</i> <i>mesenteroides</i> 130 <i>had</i> RNA
MES137	TAATACGACTCACTATAGGAAGTGTA CCTTAGGGTTCCGGCCATAAGGCGTC AGCGACCGAGCGGTACAATCCGGGG	Forward template for transcription of the WT <i>T.</i> <i>oceanii</i> 112 <i>ilvE</i> RNA (ppGpp aptamer)
MES138	TCTTTTAGGAACTTGCCGCTGGAGCC TTTTATGCTCACGGTGTTCCGGGTTTC CCCGGATTGTACCGCTCGG	ReversetemplatefortranscriptionoftheWToceanii112ilvERNA (ppGppaptamer)
HS017	TAATACGACTCACTATAGGTATTGAA CTGGGACTAGATAGCTAGGGTTCCGC CTTAATATATTAAGGGTTTGTGACCA AACGCTATAGCTCTTGTTTTG	Forward template for transcription of the M1 <i>C</i> . <i>botulinum</i> 131 <i>nudix</i> RNA
MES224	AACACGCTGAACCTTCCCCCTCGGGC ATTTTATTCCAATAGGTGCCTCTTGC AAAACAAGAGCTATAGCGT	ReversetemplatefortranscriptionoftheM1 <i>botulinum</i> 131 <i>nudix</i> RNA
MES225	AACACGCTGAACCTTCCCCCTCGGGC ATTTTATTCCAATAGGCGCCTCTTGC AAAACAAGAGCTATAGCGC	ReversetemplatefortranscriptionoftheM2botulinum131nudixRNA
MES226	AACACGCTGAACCTTCCCCCGCGGGC ATTTTATTCCAATAGGTGCCTCTTGC AAAACAAGAGCTATAGCGC	ReversetemplatefortranscriptionoftheM3 <i>botulinum</i> 131 <i>nudix</i> RNA