

Supplementary Information

Impaired mitochondrial calcium efflux contributes to disease progression in models of Alzheimer's disease

Jadiya et al. Nature Communications. 2019.

Supplementary Methods

Membrane Rupture and cell viability assay. Equal numbers of N2a, APP^{swe} and APP^{swe} infected with Ad-NCLX for 48h were treated with Ionomycin, (1-5 μ M) for 24h and an oxidizing agent *tert*-Butyl hydroperoxide (TBH) (10- 30 μ M) for 14h and glutamate (NDMAR-agonist, neuroexcitotoxicity agent) (10- 50 μ M) for 24 h. To measure number of viable cells, CellTiter-Blue Reagent (10 μ l/well in 96 well plate) is added directly to each well, incubated at 37°C for 2 hrs and the fluorescent signal at (560(20)_{Ex}/590(10)_{Em}).was measured using a Tecan Infinite M1000 Pro plate reader. Data is normalized to vehicle control to avoid any differences in cell numbers between the groups.

Citrate synthase activity. Citrate synthase activity was measured according to the method described in manual (BioVision Catalog #: K318). Briefly, mitochondria were isolated from brain cortex of mice, transferred to a 96-well plate (4 μ g/well) and brought to a volume of 50 μ l with CS assay buffer provided in the kit. The reaction was initiated by adding of 50 μ l of reaction mix (CS assay buffer, developer and substrate mix) to each well containing samples. Changes in absorbance at 412 nm in 25°C were measured in kinetic mode for 20 min. The results were presented as changes in activity within the groups (activity/ μ g protein/min).

Supplementary Table 1. Demographics of brain samples examined in this study.

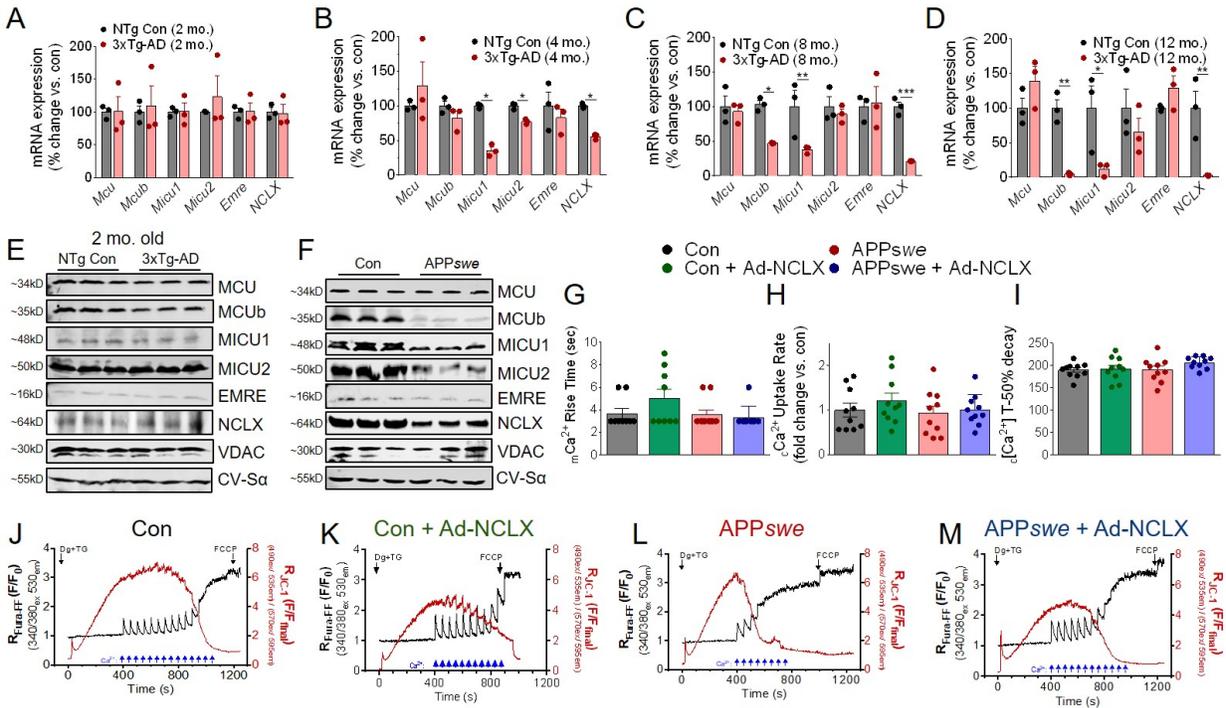
Diagnosis	Gender	Age	Source
No dementia	Male	81	ASU Brain bank
No dementia	Male	78	ASU Brain bank
No dementia	Female	86	ASU Brain bank
No dementia	Female	82	ASU Brain bank
No dementia	Male	71	ASU Brain bank
No dementia	Male	80	ASU Brain bank
No dementia	Female	85	ASU Brain bank
Sporadic, non-familial AD	Female	86	ASU Brain bank
Sporadic, non-familial AD	Female	82	ASU Brain bank
Sporadic, non-familial AD	Female	80	ASU Brain bank
Sporadic, non-familial AD	Male	72	ASU Brain bank
Sporadic, non-familial AD	Male	75	ASU Brain bank
Sporadic, non-familial AD	Male	81	ASU Brain bank
Sporadic, non-familial AD	Female	77	ASU Brain bank

ASU: Arizona State University

Supplementary Table 2. List of primer sequences used in this study.

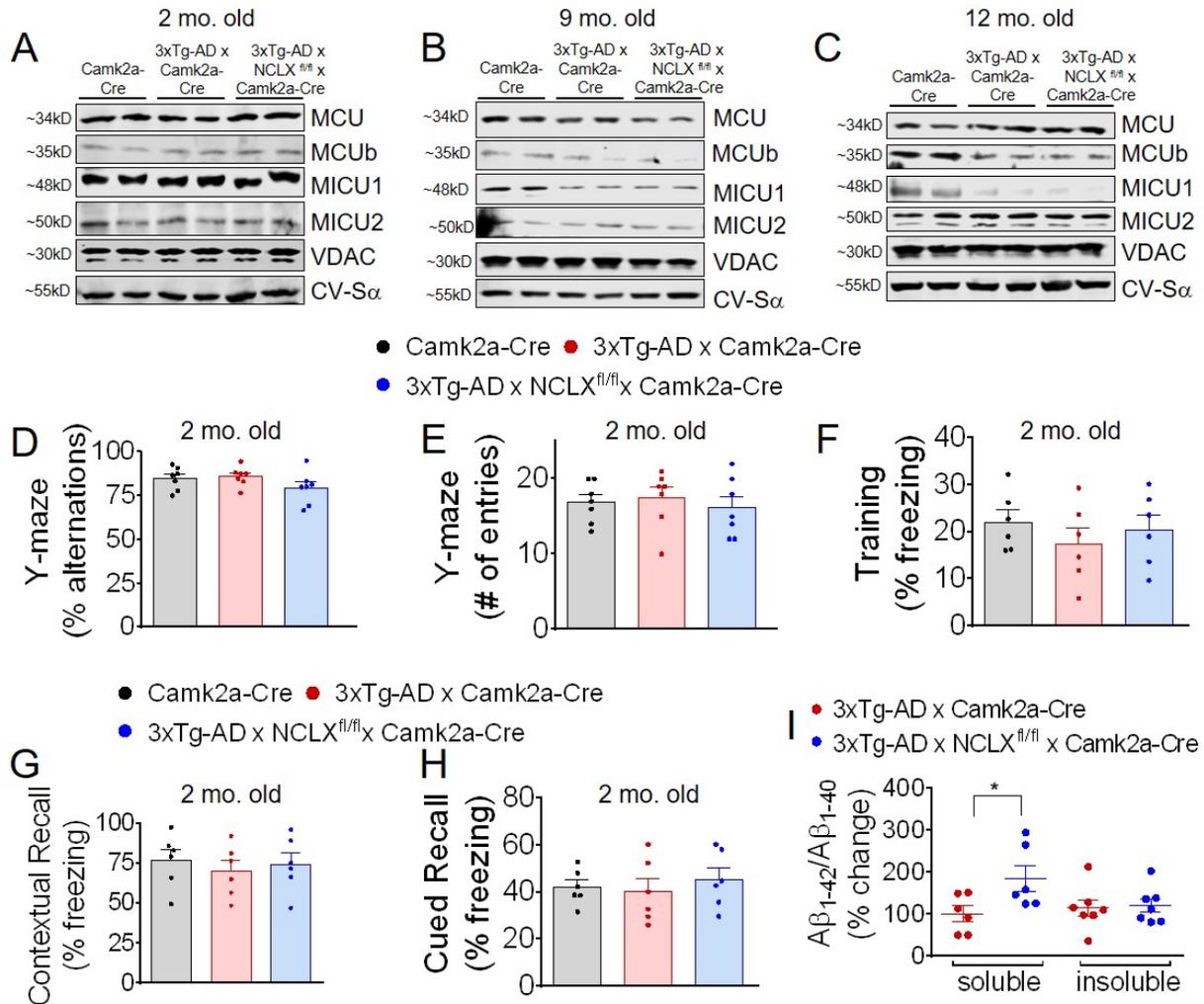
Primer sequences for Jadiya et al.		
Gene (mouse or human)	Forward primer	Reverse primer
<i>Rps13</i>	GCACCTTGAGAGGAACAGAA	GAGCACCCGCTTAGTCTTATAG
<i>Mcu</i>	GATGACGTGACGGTGGTTTA	GTCAGAGATAGGCTTGAGTGTG
<i>Mcub</i>	CGACAACATCGGCTTGACTA	GTGGAGCCACAGGATGAAATA
<i>Micu1</i>	AAGAACACTCCCTGCCATTT	GCCAGGGTCATCTGCATTAT
<i>Micu2</i>	GTGCTTTCTGGAGGGCTAAA	CTGCAAGTATTCCCTAAGCTATCA
<i>Emre</i>	GGGACACTCATCAGCAAGAA	CTCCCTGTGCCCTGTTAATC
<i>NCLX (Slc8b1)</i>	GCCATCTCCACTAACCTCAA	GGGTCTGAGAAAGCCACTAAA
<i>Hbb-bt genomic</i>	GAAGCGATTCTAGGGAGCAG	GGAGCAGCGATTCTGAGTAGA
<i>RPS13</i>	CCTTCACAGATCGGTGTAATCC	TCAGGAGCAAGTCCCTTAGA
<i>MCU</i>	GGGGTACCCACCAAACAGCTATTC	CCCAAGCTTGGGTTCCGATCTGTCGG
<i>MCUB</i>	GTGTGAAGCTGTGTGGAAATG	CAAGGGAAGGCCATGTCTATAA
<i>MICU1</i>	AACATTCCTTGACTTCCCTCC	TCTGAATCTTGCTGTGTTCCC
<i>MICU2</i>	TGGAAGGATAAAACAATACATATGGG	GGAACATGAAGATGTCTGGAATTG
<i>EMRE</i>	CTGAGATGTGCTGTCCACTAAG	GAGACAACCTGCACCAACTAGAA
<i>NCLX (SLC8B1)</i>	CAGAAAGGGAAGTGGAGAGTAAG	GCCATTAGCAGCACACAAAG
<i>PGC-1α</i>	CCGAGAATTCATGGAGCAAT	GTGTGAGGAGGGTCATCGTT
<i>COXII</i>	GCCGACTAAATCAAGCAACA	CAATGGGCATAAAGCTATGG

Supplementary Figures



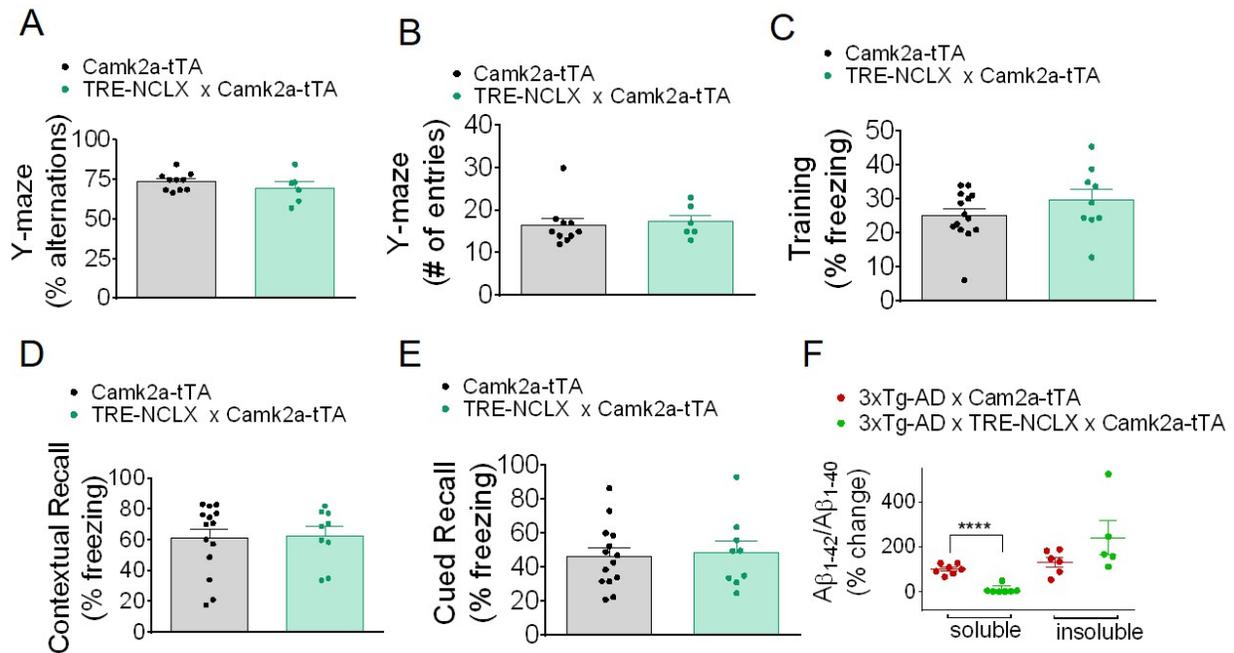
Supplementary Figure 1: mCa²⁺ exchanger expression and mCa²⁺ handling in AD.

(A) mRNA expression of mCa²⁺ exchanger in brain tissue isolated from the frontal cortex of 2 mo. old 3xTg-AD mutant mice and age-matched outbred non-transgenic controls (NTg). n = 3 for both groups. (B) mRNA expression of mCa²⁺ exchanger in brain tissue isolated from the frontal cortex of 4 month old 3xTg-AD mutant mice and age-matched outbred non-transgenic controls (NTg). n = 3 for both groups. (C) mRNA expression of mCa²⁺ exchanger in brain tissue isolated from the frontal cortex of 8 month old 3xTg-AD mutant mice and age-matched outbred non-transgenic controls (NTg). n = 3 for both groups. (D) mRNA expression of mCa²⁺ exchanger in brain tissue isolated from the frontal cortex of aged (12 mo.) 3xTg-AD mutant mice and outbred non-transgenic controls (NTg). n = 3 for both groups. (E) Western blots for mCa²⁺ exchanger protein in 3xTg-AD mutant mice (2 mo.) and age-matched outbred non-transgenic controls (NTg). n = 3 for both groups. (F) Western blots for mCa²⁺ exchanger proteins in neuroblastoma control cell line (N2a) vs. cells stably expressing cDNA encoding the APP Swedish mutant (K670N, M671L, APPswe). n = 3 for both groups. (G) Quantification of mCa²⁺ rise time. (H) Fold change in cCa²⁺ uptake rate of con + Ad-NCLX, APPswe and APPswe + Ad-NCLX vs. con (N2a) cells. n=10. (I) Time to 50% cCa²⁺ transient decay (T-50%). n = 10 for all groups. (J-M) Representative traces for mCa²⁺ retention capacity in con, con + Ad-NCLX, APPswe and APPswe cells infected with adenovirus encoding mitochondrial Na⁺/Ca²⁺ exchanger (NCLX). Data was presented as mean ± SEM, ***p<0.001, **p<0.01, *p<0.05, one-way ANOVA, Sidak's multiple comparisons test.



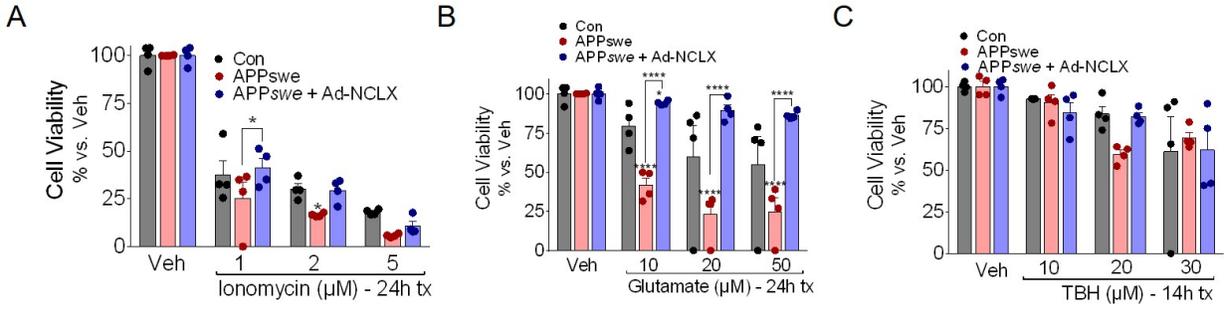
Supplementary Figure 2: NCLX deletion effect on the memory at the age of 2 mo.

(A-C) Western blots for NCLX expression and proteins associated with mCa^{2+} exchange in tissue isolated from the hippocampus of 2, 9 and 12 mo. old 3xTg-AD x NCLX^{fl/fl} x Camk2a-Cre mutant mice compared to age-matched control. MCU, mitochondrial calcium uniporter; MCUB, mitochondrial calcium uniporter β subunit; MICU1, mitochondrial calcium uptake 1; MICU2, mitochondrial calcium uptake 2; EMRE, essential MCU regulator; NCLX, Na^+/Ca^{2+} exchanger. Voltage dependent anion channel (VDAC) and oxidative phosphorylation component CV-S α , complex V α subunit; were used as mitochondrial loading controls and has been shown in Fig. 2. (D-E) Working memory was assessed in the Y-maze spontaneous alternation test in mice at the age of 2 mo. in Camk2a-Cre, 3xTg-AD x Camk2a-Cre and 3xTg-AD x NCLX^{fl/fl} x Camk2a-Cre mice (D) Percentage spontaneous alternations. (E) Number of total arm entries. n = number of dots as shown for all groups. (F-H) Hippocampus and amygdala associated memory was assessed in the fear conditioning test in mice at the age of 2 mo. in Camk2a-Cre, 3xTg-AD x Camk2a-Cre and 3xTg-AD x NCLX^{fl/fl} x Camk2a-Cre mice (F) Freezing responses in the training phase (G) Contextual recall freezing responses (H) Cued recall freezing responses. n= number of dots as shown for all groups. (I) Soluble (RIPA) and insoluble (formic acid extractable) A β_{1-42} /A β_{1-40} ratio in brain cortex of 3xTg-AD x Camk2a-Cre and 3xTg-AD x NCLX^{fl/fl} x Camk2a-Cre mice at the age of 12 mo. were measured by sandwich ELISA. Data was presented as mean \pm SEM, ***p<0.001, **p<0.01, *p<0.05, one-way ANOVA, Sidak's multiple comparisons test.



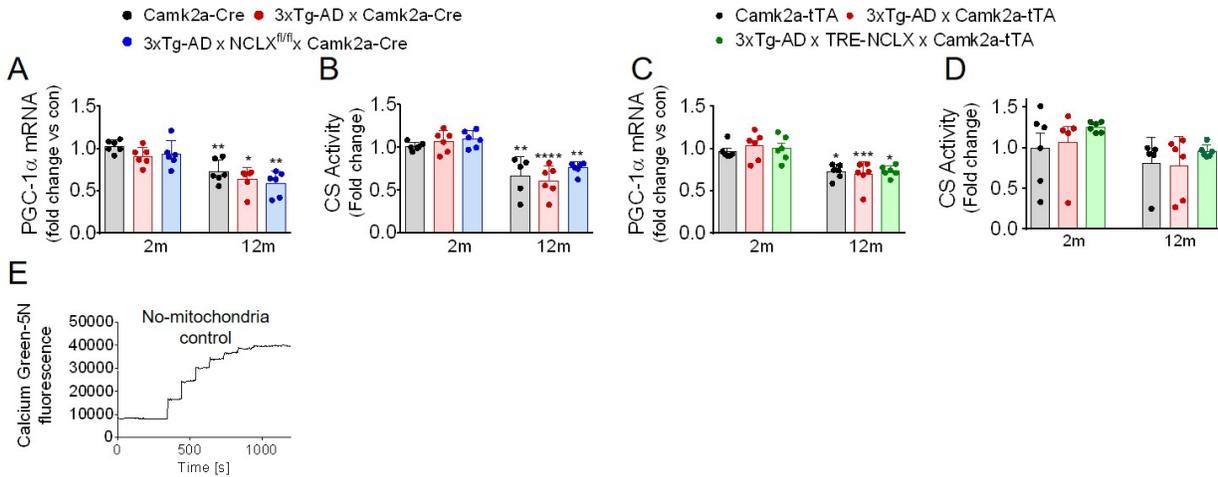
Supplementary Figure 3: NCLX overexpression effect on the memory in wild type mice

(A-B) Working memory was assessed in the Y-maze spontaneous alternation test in mice at the age of 6 mo. in Camk2a-tTA and TRE-NCLX x Camk2a-tTA mice **(A)** Percentage spontaneous alternations. **(B)** Number of total arm entries. $n =$ number of dots shown in Fig. **(C-E)** Hippocampus and amygdala associated memory was assessed in the fear conditioning test in mice at the age of 6 mo. in Camk2a-tTA and TRE-NCLX x Camk2a-tTA mice **(C)** Freezing responses in the training phase **(D)** Contextual recall freezing responses **(E)** Cued recall freezing responses. **(F)** Soluble (RIPA) and insoluble (formic acid extractable) $A\beta_{1-42}/A\beta_{1-40}$ ratio in brain cortex of 3xTg-AD x Camk2a-tTA and 3xTg-AD x TRE-NCLX x Camk2a-tTA mice at the age of 12 mo. were measured by sandwich ELISA. All data shown as mean \pm SEM; one-way ANOVA, Sidak's multiple comparisons test, $n =$ number of dots shown in Fig. for all groups. *** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$.



Supplementary Figure 4: Enhancing mCa^{2+} efflux effect on cell viability

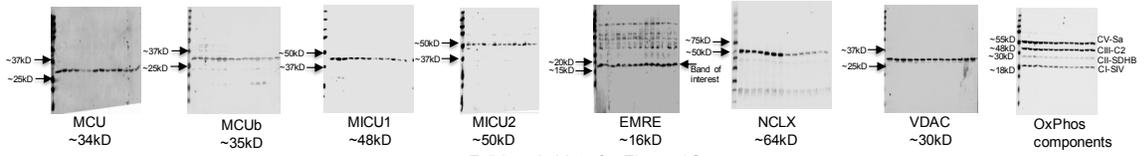
(A-C) N2a, APP_{swe} and APP_{swe} infected with Ad-NCLX for 48h were assessed for cell viability using Cell Titer Blue after treatment with (A) Ionomycin (Ca^{2+} overload, 1-5 μ M), (B) glutamate (NDMAR-agonist, neuroexcitotoxicity agent, 10-50 μ M). (C) *tert*-Butyl hydroperoxide (TBH, oxidizing agent, 10-30 μ M), n = individual dots shown for each group. Data was presented as mean \pm SEM, ***p<0.001, **p<0.01, *p<0.05, one-way ANOVA, Sidak's multiple comparisons test.



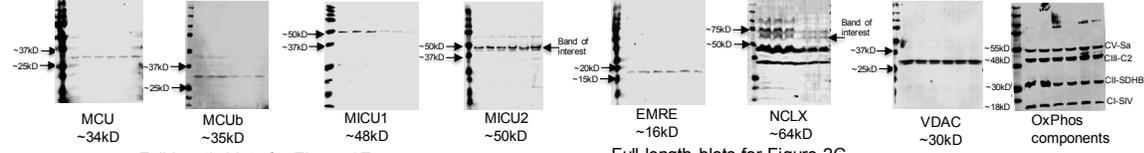
Supplementary Figure 5: Mitochondrial mass in deletion and overexpression of NCLX

(A) PGC-1 α mRNA expression in tissue isolated from the brain cortex of 2 & 12 mo. old Camk2a-Cre, 3xTg-AD x Camk2a-Cre and 3xTg-AD x NCLX^{fl/fl} x Camk2a-Cre mice expressed as fold-change vs. 2m old Camk2a-Cre con. n = 6 for all groups. **(B)** Citrate synthase activity in mitochondria isolated from the brain cortex of 2 & 12 mo. old Camk2a-Cre, 3xTg-AD x Camk2a-Cre and 3xTg-AD x NCLX^{fl/fl} x Camk2a-Cre mice expressed as fold-change vs. 2m old Camk2a-Cre con. n = 6 for all groups. **(C)** PGC-1 α mRNA expression in tissue isolated from the brain cortex of 2 & 12 mo. old Camk2a-tTA, 3xTg-AD x Camk2a-tTA and 3xTg-AD x TRE-NCLX x Camk2a-tTA mice expressed as fold-change vs. 2m old Camk2a-tTA con. n = 6 for all groups. **(D)** Citrate synthase activity in mitochondria isolated from the brain cortex of 2 & 12 mo. old Camk2a-tTA, 3xTg-AD x Camk2a-tTA and 3xTg-AD x TRE-NCLX x Camk2a-tTA mice expressed as fold-change vs. 2m old Camk2a-tTA con. n = 6 for all groups. **(E)** A standard curve of Ca²⁺ (5-50 μ m) in experimental intracellular buffer to quantify actual Ca²⁺ content as shown in Fig. 5A-D. Data was presented as mean \pm SEM, ***p<0.001, **p<0.01, *p<0.05, one-way ANOVA, Sidak's multiple comparisons test.

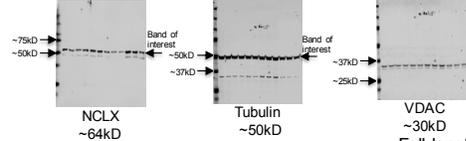
Full length blots for Figure 1A



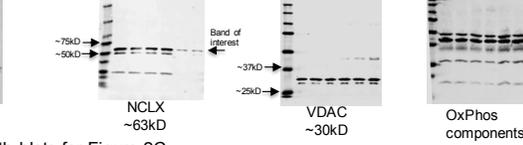
Full length blots for Figure 1C



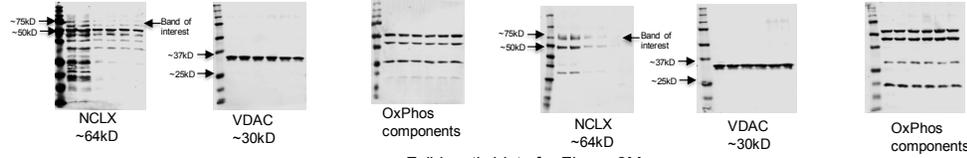
Full length blots for Figure 1E



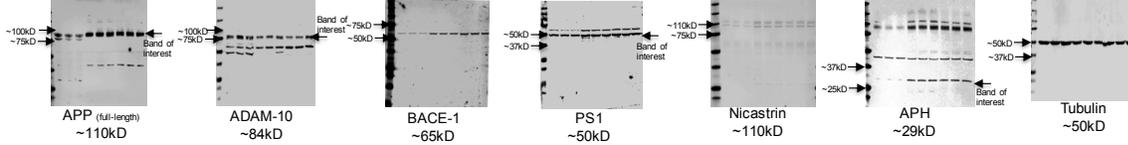
Full length blots for Figure 2C



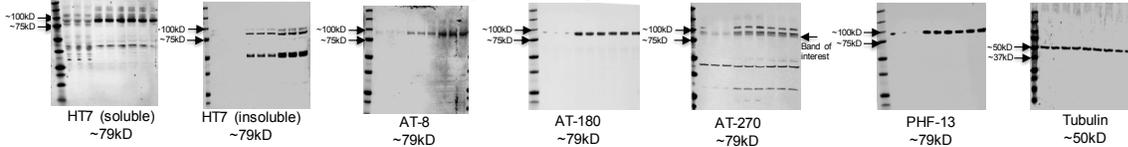
Full length blots for Figure 2C



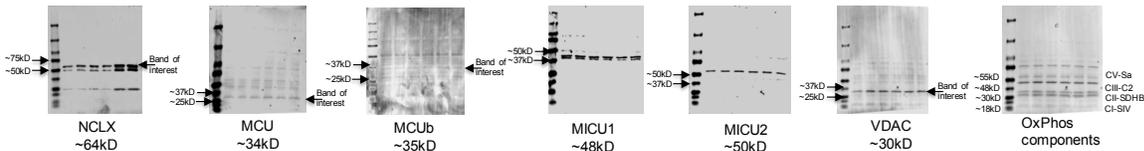
Full length blots for Figure 2M



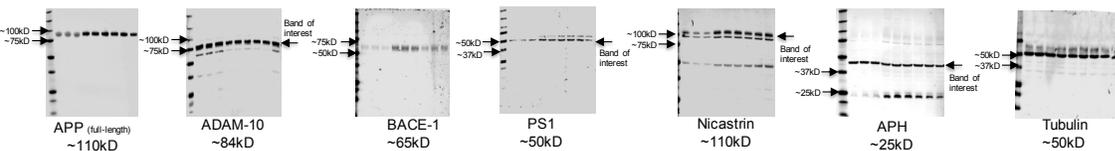
Full length blots for Figure 2N



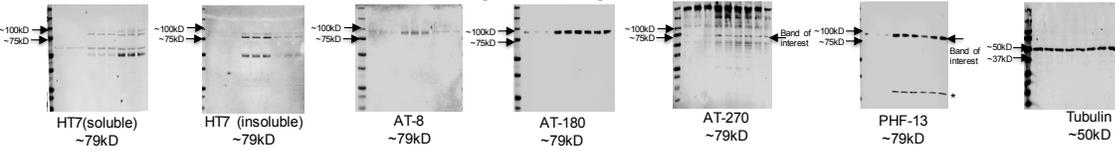
Full length blots for Figure 3C

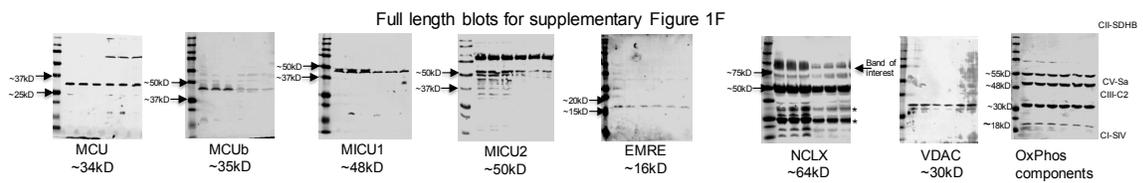
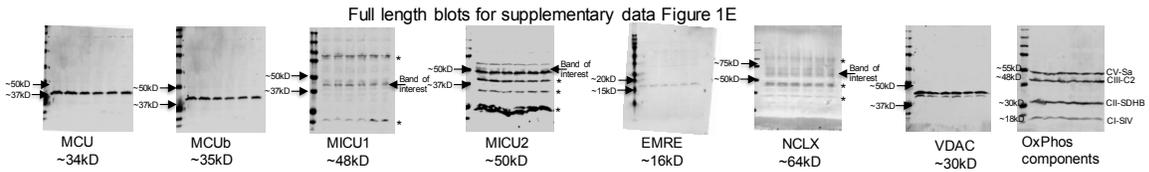
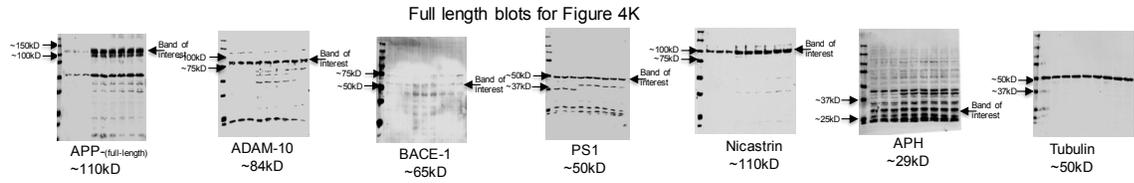


Full length blots for Figure 3M

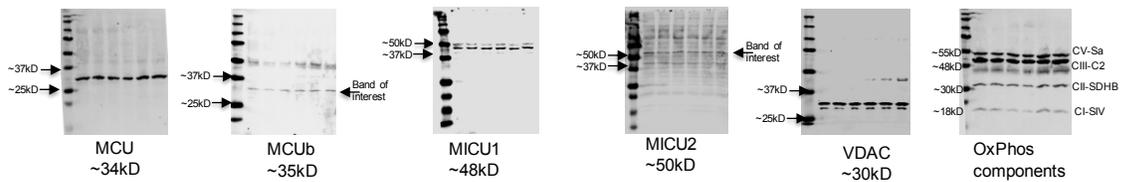


Full length blots for Figure 3N

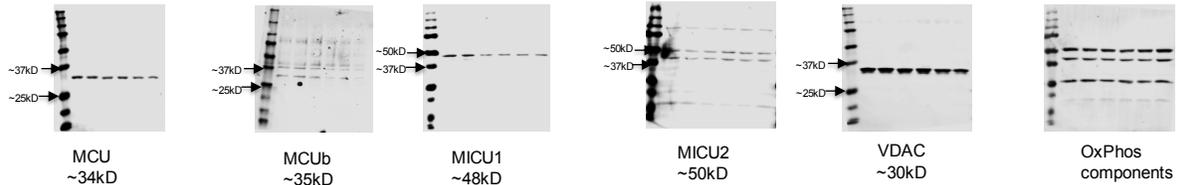




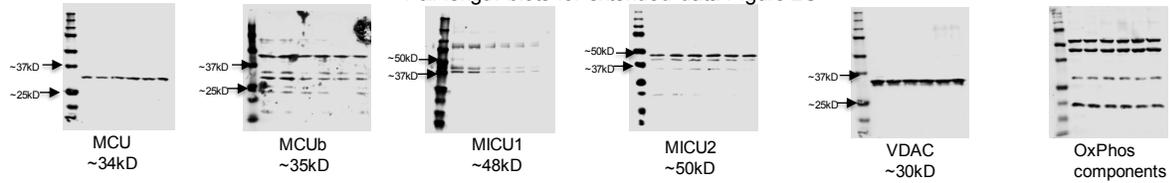
Full length blots for supplementary data Figure 2A



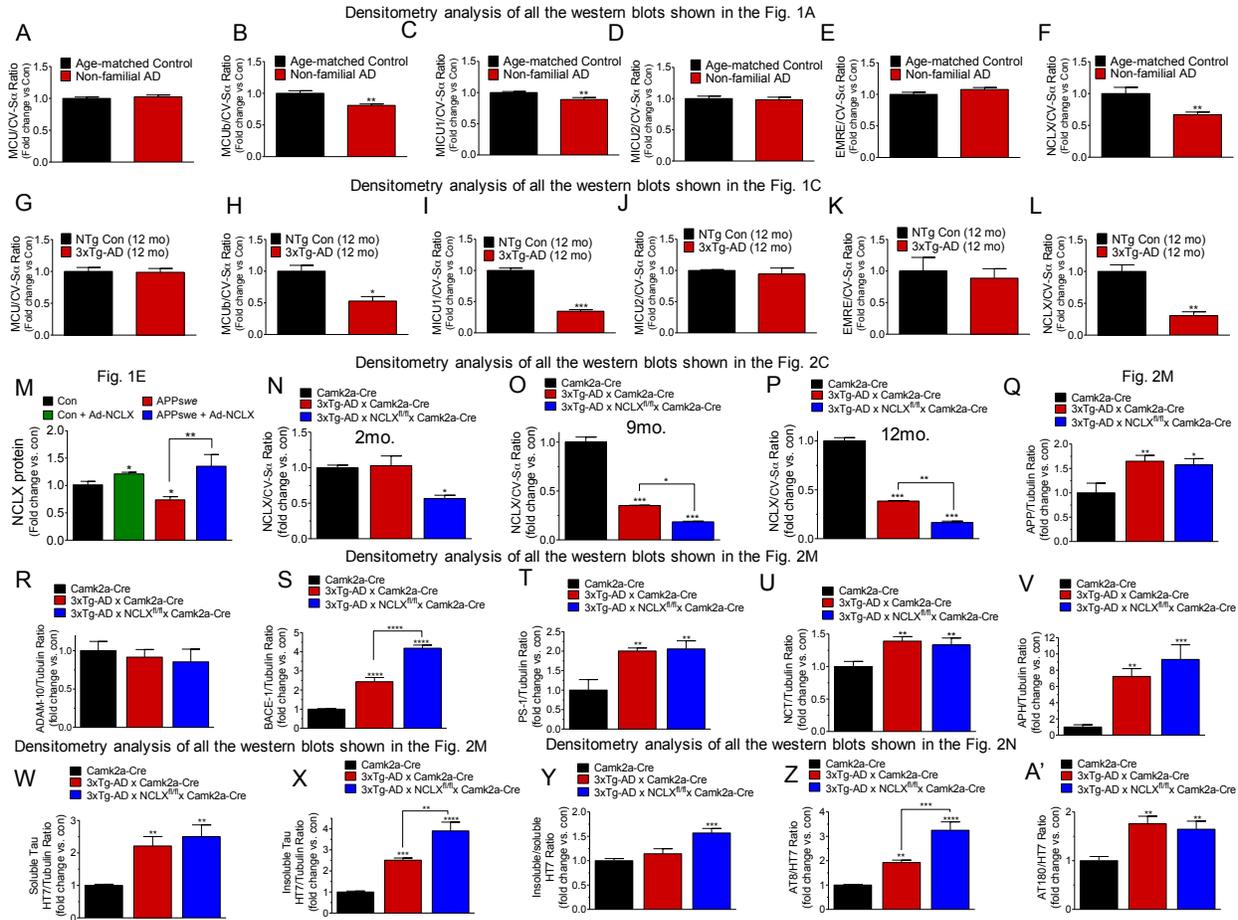
Full length blots for supplementary data Figure 2B



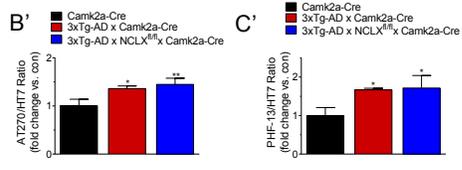
Full length blots for extended data Figure 2C



Supplementary Figure 6: Full-length Western blots.



Densitometry analysis of all the western blots shown in the Fig. 2N



Densitometry analysis of all the western blots shown in the Fig. 3C

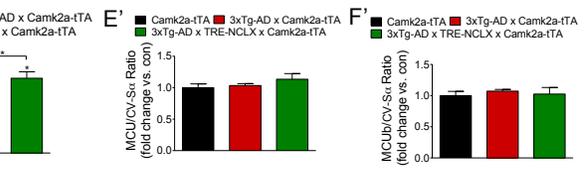
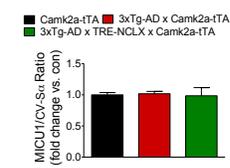


Fig. 3C



Densitometry analysis of all the western blots shown in the Fig. 3M

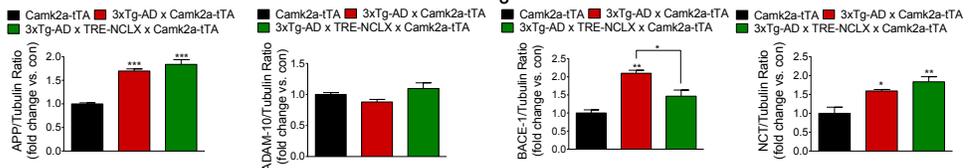


Fig. 3M

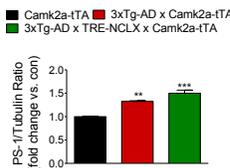
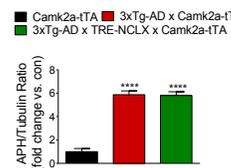
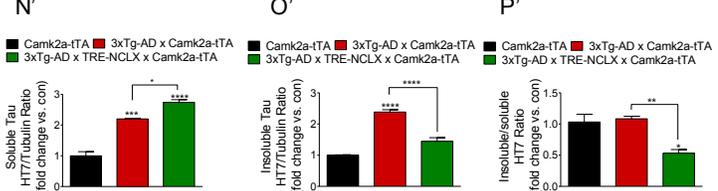


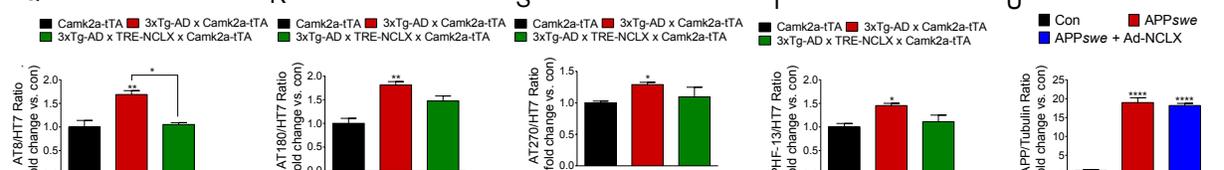
Fig. 3M



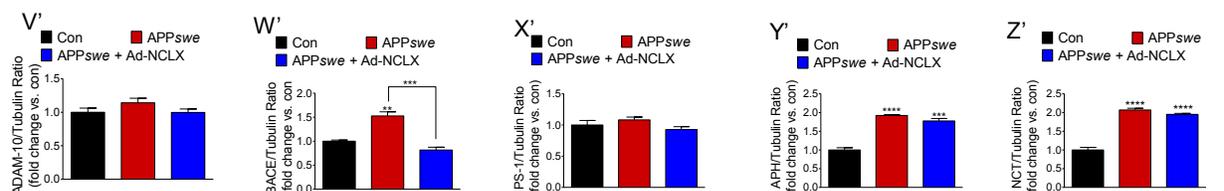
Densitometry analysis of all the western blots shown in the Fig. 3N



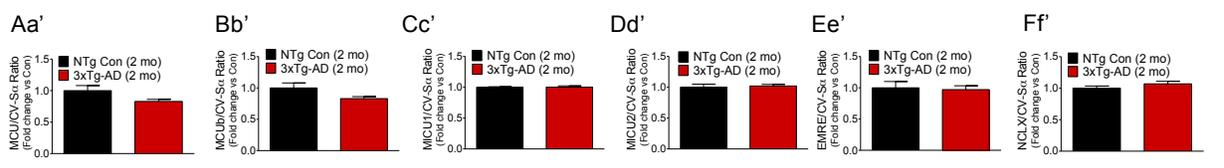
Densitometry analysis of all the western blots shown in the Fig. 3N



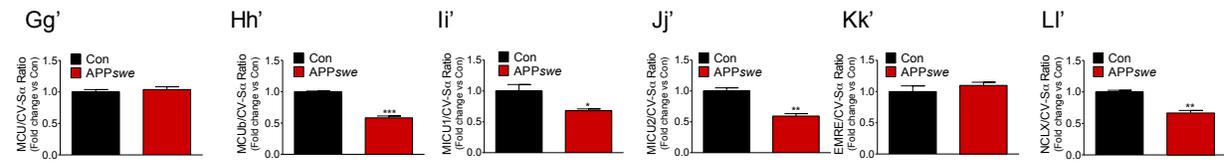
Densitometry analysis of all the western blots shown in the Fig. 4K



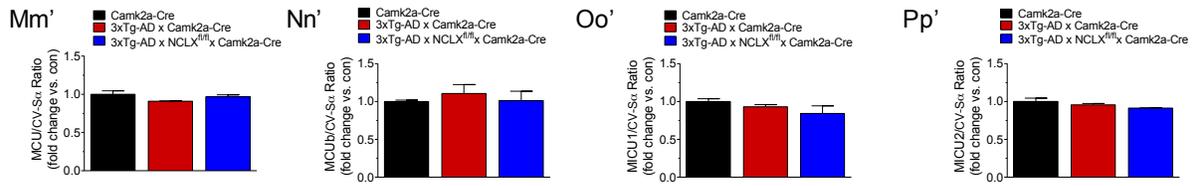
Densitometry analysis of all the western blots shown in the supplementary Fig. 1E



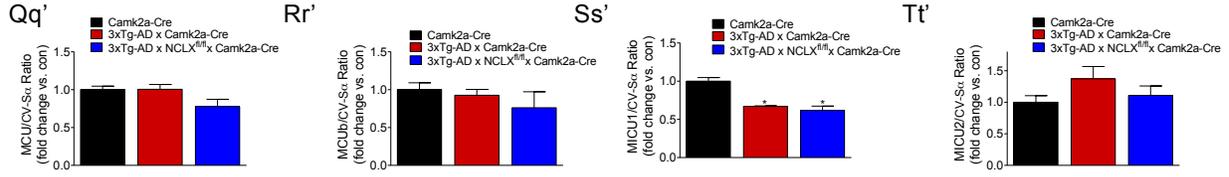
Densitometry analysis of all the western blots shown in the supplementary Fig. 1F



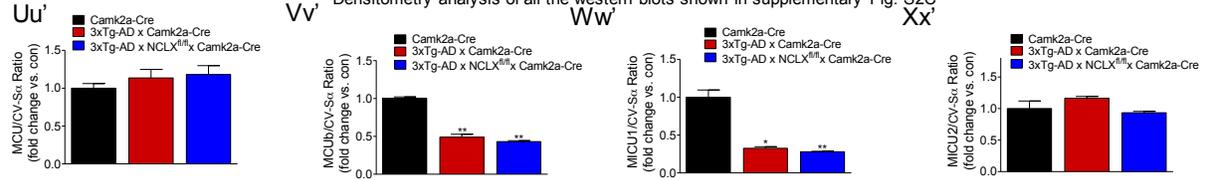
Densitometry analysis of all the western blots shown in supplementary Fig. S2A



Densitometry analysis of all the western blots shown in supplementary Fig. S2B



Densitometry analysis of all the western blots shown in supplementary Fig. S2C



Supplementary Figure 7: Densitometry analysis of Western blots.