

# SUPPORTING INFORMATION

## Exposure to graphene oxide sheets alters the expression of reference genes used for real-time RT-qPCR normalization

Irene de Lázaro<sup>1,2,\*</sup> and Kostas Kostarelos<sup>1</sup>

<sup>1</sup> Nanomedicine Lab, Faculty Biology, Medicine and Health and National Graphene Institute, AV Hill Building, The University of Manchester, Manchester M13 9PT, United Kingdom

<sup>2</sup> Present address: John A. Paulson School of Engineering and Applied Sciences, Harvard University, Cambridge, MA, 02138, USA; Wyss Institute for Biologically Inspired Engineering, Harvard University, Boston, MA, 02115, USA

---

\* Correspondence to: [idelazaro@g.harvard.edu](mailto:idelazaro@g.harvard.edu)

# Supporting Tables

**Table S1.** Physicochemical characterization of GO material used in this study.

	<b>Technique</b>	<b>Results</b>
<b>Lateral dimensions*</b>	<i>AFM</i>	0.050 – 0.5 $\mu\text{m}$
	<i>TEM</i>	0.1 – 2 $\mu\text{m}$
<b>Thickness*</b>	<i>AFM</i>	1.4 $\pm$ 0.5 nm (1-2 layers)
<b>Optical Properties</b>	<i>Absorbance</i> ( $\lambda = 230$ nm)	$A = 0.050 * C_{\text{GO}}$ ( $\mu\text{g/mL}$ )
	<i>Fluorescence</i> ( $\lambda_{\text{exc}} = 525$ nm)	$F_{600} = 0.823 * C_{\text{GO}}$ ( $\mu\text{g/mL}$ )
<b>Degree of defects (<math>I_D/I_G</math>)</b>	<i>Raman spectroscopy</i>	1.36 $\pm$ 0.03
<b>Surface charge</b>	$\zeta$ -potential	-55.9 $\pm$ 1.4 mV
<b>Functionalisation degree</b>	<i>TGA</i>	41%
<b>Chemical composition (Purity)</b>	<i>XPS</i>	C: 67.6%, O: 32.2%, (99.8%) S: 0.2%
<b>C:O ratio</b>	<i>XPS</i>	2.1
<b><math>\pi</math>-<math>\pi</math>, O=C-O, C=O, C-O-C, C-C &amp; C=C</b>	<i>XPS</i>	3.0%, 15.3%, 31.2%, 11.3%, 39.2%

\*Lateral dimensions and thickness are reported as a range between the minimum and maximum sizes detected. Full characterization of the material is provided in Mukherjee et al<sup>15</sup>.

**Table S2.** MIQE checklist.

See separate XLS file

**Table S3.** RNA yield and integrity assessment, MCF7 samples.

Sample	[RNA] µg/ml	A <sub>260/280</sub>	A <sub>260/230</sub>	28S/18S	RIN
Untreated 1	554	2.01	2.1	3.0	9.2
Untreated 2	583	2.06	2.02	2.4	9.2
Untreated 3	535	1.87	1.93	3.2	9.5
5 µg/ml 1	559	2.02	2.08	3.8	9.3
5 µg/ml 2	639	2.04	2.18	2.5	9.6
5 µg/ml 3	563	2.02	1.82	2.8	9.3
10 µg/ml 1	636	2.02	1.99	3.7	9.4
10 µg/ml 2	597	2.01	2.11	3.3	9.4
10 µg/ml 3	723	2.03	2.19	3.9	9.4
50 µg/ml 1	523	1.79	1.70	3.0	9.6
50 µg/ml 2	594	2.02	1.75	3.0	9.8
50 µg/ml 3	529	1.99	2.09	2.7	9.8

[RNA] µg/ml, A<sub>260/A280</sub> and A<sub>260/230</sub> are calculated by spectroscopy; 28S/18S ratios and RIN numbers are calculated upon RNA denaturation and electrophoretic separation (**Figure S1, a**). The table shows the results corresponding to all four experimental groups, n=3.

**Table S4.** RNA yield and integrity assessment, MEF samples.

Sample	[RNA] µg/ml	A <sub>260/280</sub>	A <sub>260/230</sub>	28S/18S	RIN
Untreated 1	358	1.89	1.88	3.1	9.1
Untreated 2	333	1.96	1.93	2.8	9.3
Untreated 3	324	2.01	2.19	2.7	9.3
5 µg/ml 1	266	2.09	1.71	2.2	9.6
5 µg/ml 2	223	2.03	2.1	2.5	8.9
5 µg/ml 3	246	1.99	2.14	2.4	9.1
10 µg/ml 1	371	1.82	1.95	2.5	9.3
10 µg/ml 2	263	1.77	1.77	2.1	9.3
10 µg/ml 3	252	1.92	1.77	2.3	9.3
50 µg/ml 1	96	1.96	2.04	2.0	9.4
50 µg/ml 2	176	1.99	2.09	2.4	9.2
50 µg/ml 3	110	1.71	1.75	2.6	9.3

[RNA] µg/ml, A<sub>260/A280</sub> and A<sub>260/230</sub> are calculated by spectroscopy; 28S/18S ratios and RIN numbers are calculated upon RNA denaturation and electrophoretic separation (**Figure S1, b**). The table shows the results corresponding to all four experimental groups, n=3.

**Table S5.** Details of primer pairs used in MCF7 study (human).

Gene symbol	Accession number (mRNA)	Primer sequences	Amplicon size (bp)	E	R <sup>2</sup>
<b>RPS13</b>	NM_001017.2	<b>Fwd</b> CGCTCTCCTTTTCGTTGCCT <b>Rv</b> CGCTGCGTCGATAGGGTAAA	96	2.11	0.9956
<b>RPL27</b>	NM_000988.3	<b>Fwd</b> ATCGCCAAGAGATCAAAGATAA <b>Rv</b> TCTGAAGACATCCTTATTGACG	123	2.02	0.9993
<b>RPL30</b>	NM_000989.3	<b>Fwd</b> ACAGCATGCGGAAAATACTAC <b>Rv</b> AAAGGAAAATTTTGCAGGTTT	158	1.99	0.9987
<b>OAZ1</b>	NM_004152.3 NM_001301020.1	<b>Fwd</b> CTCCACTGCTGTAGTAACCCG <b>Rv</b> GATCCCTCTGACTATTCCCTCG	104	1.75	0.9998
<b>ACTB</b>	NM_001101.3	<b>Fwd</b> AGCACAGAGCCTCGCCTTT <b>Rv</b> GAGCGCGGCGATATCATCA	82	2.0	0.9961
<b>GAPDH</b>	NM_001289746.1 NM_001289745.1 NM_001256799.2 NM_002046.5	<b>Fwd</b> CCACATGGCCTCCAAGGAGTAAGAC <b>Rv</b> AGGAGGGGAGATTCAGTGTGGTGGG	131	2.0	0.9984
<b>MAPK1</b>	NM_002745.4 NM_138957.3	<b>Fwd</b> TCCCAAATGCTGACTCCAAAG <b>Rv</b> CATGTCGAACTTGAATGGTGC	164	2.08	0.9993
<b>UBC</b>	NM_021009.6	<b>Fwd</b> GCCTTAGAACCCCAAGTATCAG <b>Rv</b> AAGAAAACCAAGTGCCCTAGAG	74	1.98	0.9907
<b>HMBS</b>	NM_000190.3 NM_001024382.1 NM_001258208.1 NM_001258209.1 XM_017017629.1 XM_005271531.1 XM_005271532.1 XM_005271533.3 XM_011542796.1	<b>Fwd</b> AGCTTGCTCGCATAACAGACG <b>Rv</b> AGCTCCTTGGTAAACAGGCTT	157	2.03	0.9990
<b>TBP</b>	NM_003194.4 NM_001172085.1	<b>Fwd</b> CCACTCACAGACTCTCACAAC <b>Rv</b> CTGCGGTACAATCCCAGAACT	127	2.00	0.9968

E, efficiency of qPCR reaction; R<sup>2</sup>, coefficient of determination from linear regression of Cq values (cDNA serial dilution). Primer pairs amplify all transcription variants with equal amplicon length.

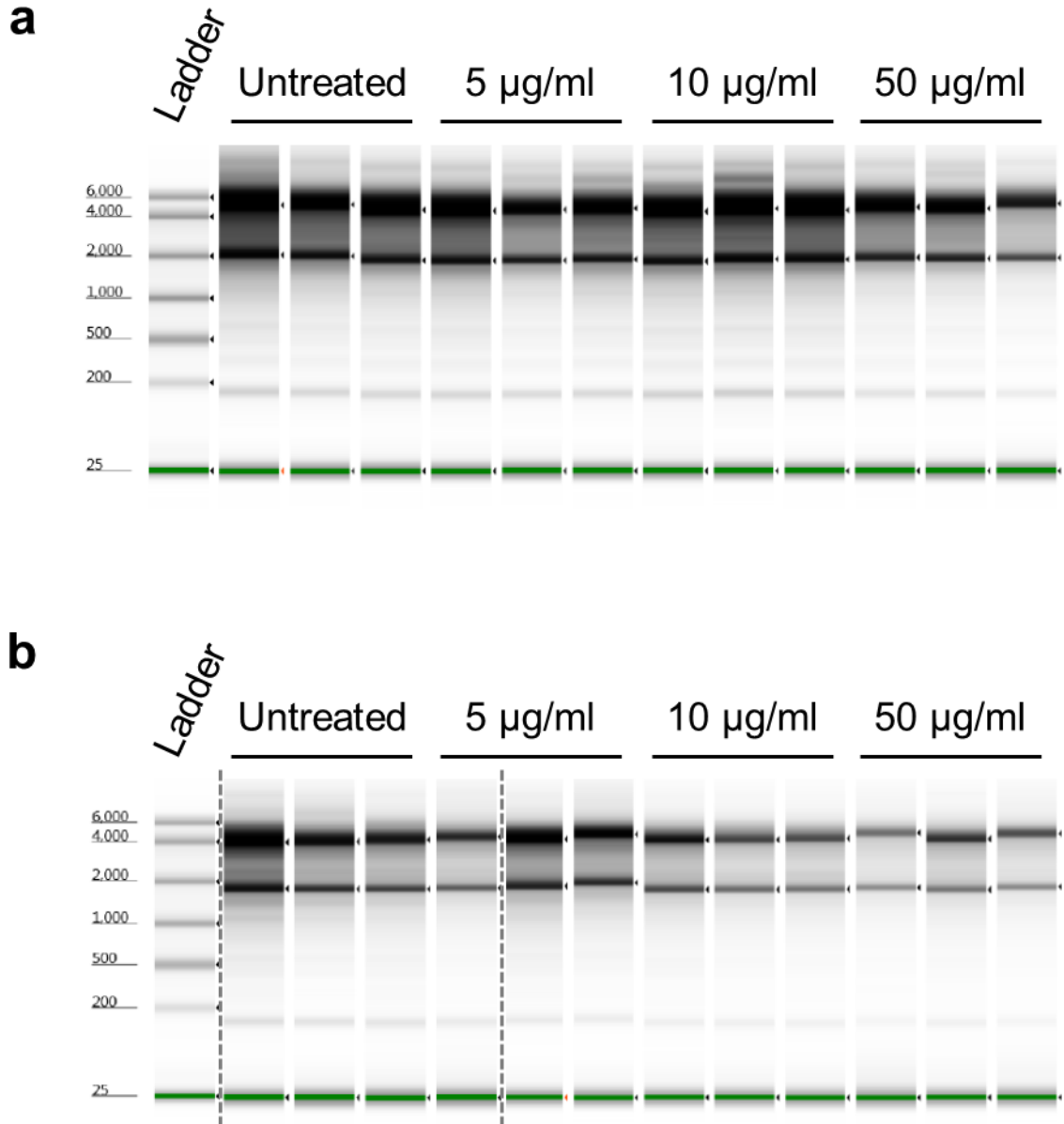
**Table S6.** Details of primer pairs used in MEF study (mouse).

Gene symbol	Accession number (mRNA)	Primer sequences	Amplicon size (bp)	E	R <sup>2</sup>
<b>Rps13</b>	NM_026533.3	<b>Fwd</b> CCCAGGTCCGTTTTGTGACT <b>Rv</b> TCCTCTCAAGGTGCTTTTCGG	132	2.01	0.9972
<b>Rpl27</b>	NM_011289.3	<b>Fwd</b> CAAAAACGCAGTGCCCGA <b>Rv</b> CTTACGGAGAGGTGGCTTCA	120	2.27	0.9907
<b>Rpl30</b>	NM_009083.4 NM_001163485.1	<b>Fwd</b> GAAGAGCTTTGCATTGTGGGAG <b>Rv</b> CCATCTTCCTGCCTTAGGTGC	102	1.95	0.9948
<b>OAZ1</b>	NM_008753.4 NM_001301034.1	<b>Fwd</b> GGGTTGCCCTTAATTGCTGT <b>Rv</b> TCTTGTGCGTTAGACGTCGGC	187	1.99	0.9985
<b>Actb</b>	NM_007393.5	<b>Fwd</b> CTGAGCTGCGTTTTACACCC <b>Rv</b> CGCCTTCACCGTTCCAGTTT	200	1.93	0.9995
<b>Gapdh</b>	NM_001289726.1 NM_008084.3 XM_017321385.1	<b>Fwd</b> AGGTCGGTGTGAACGGATTTG <b>Rv</b> TGTAGACCATGTAGTTGAGGTCA	123	2.00	0.9988
<b>Mapk1</b>	NM_011949.3 NM_001038663.1 XM_006522147.3	<b>Fwd</b> GGTTGTTCCCAAATGCTGACT <b>Rv</b> CAACTTCAATCCTCTTGTGAGGG	84	1.99	0.9979
<b>Ubc</b>	NM_019639.4	<b>Fwd</b> CAAACAGGAAGACAGACGTACC <b>Rv</b> CCCATCACACCCAAGAACAAG	80	2.04	0.9986
<b>Hmbs</b>	NM_013551.2 NM_001110251.1	<b>Fwd</b> ATCTTGACCTAGTGAGTGTGT <b>Rv</b> GTACAGTTGCCCATCTTTCATCA	141	2.01	0.9985
<b>Tbp</b>	NM_013684.3	<b>Fwd</b> TTTGGCTAGGTTTCTGCGGT <b>Rv</b> GCCCTGAGCATAAGGTGGAA	195	2.04	0.9984

E, efficiency of qPCR reaction; R<sup>2</sup>, coefficient of determination from linear regression of C<sub>q</sub> values (cDNA serial dilution). Primer pairs amplify all transcription variants with equal amplicon length.

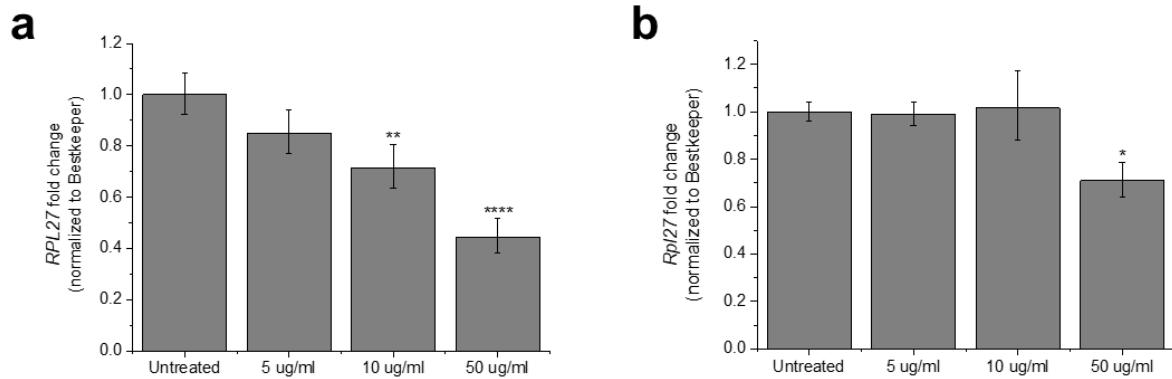
# Supporting Figures

## Supporting Figure 1



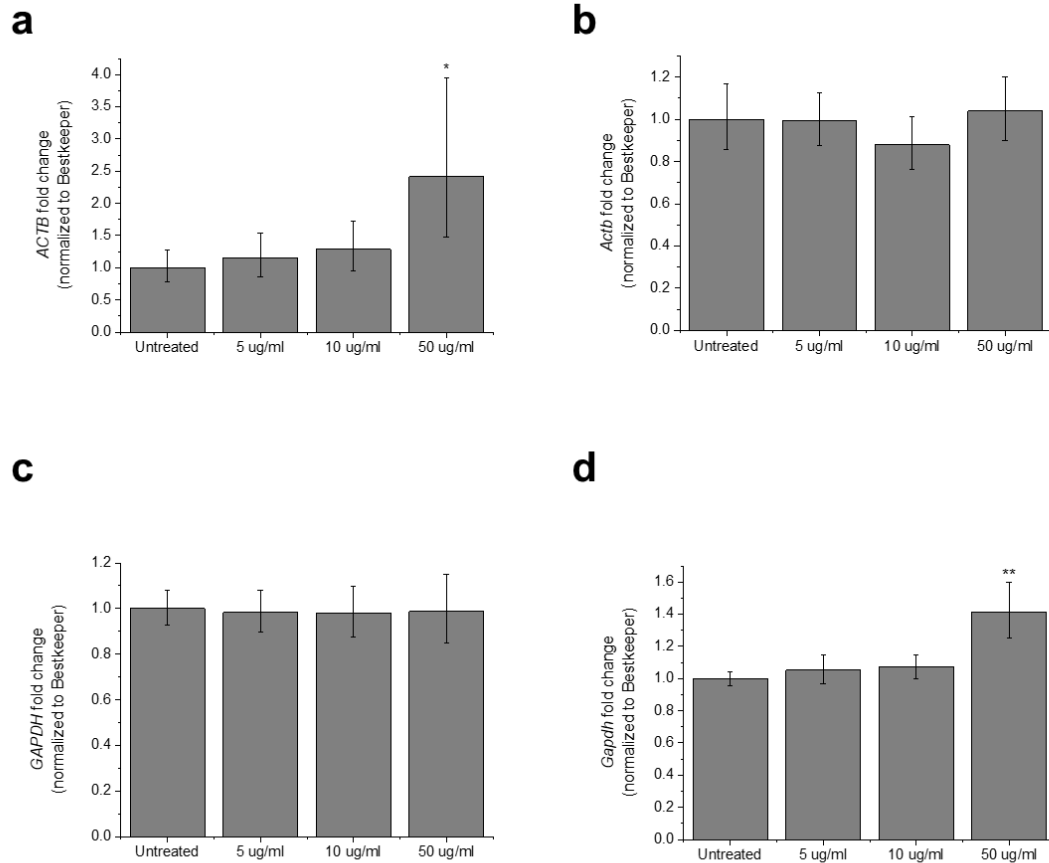
**Figure S1. Analysis of RNA integrity on the 2200 TapeStation system.** Gel electrophoresis shows the separation profile of individual (a) MCF7 and (b) MEF samples, including 28S, 18S, small rRNAs and lower marker. RIN numbers and 28S/18S ratios are provided in **Tables S3 and S4**. When gels have been cropped to organize the samples in consecutive order, cropping sites are delineated by grey dashed lines.

## Supporting Figure 2



**Figure S2. *RPL27* mRNA levels normalized to Bestkeeper index. (a) *RPL27* expression in MCF7 cells exposed to GO, normalized to Bestkeeper index. (b) *Rpl27* expression in MEFs exposed to GO, normalized to Bestkeeper index. Bars represent fold change; error bars represent propagation of standard error (SE). \*p<0.05, \*\*p<0.01 and \*\*\*\*p<0.0001, assessed by one-way ANOVA and Tukey's test, n=3.**

## Supporting Figure 3



**Figure S3. *ACTB* and *Gapdh* mRNA levels normalized to Bestkeeper index. (a) *ACTB* and (b) *GAPDH* expression in MCF7 cells exposed to GO, normalized to Bestkeeper index. (c) *Actb* and (d) *Gapdh* expression in MEFs exposed to GO, normalized to Bestkeeper index. Bars represent fold change; error bars represent propagation of standard error (SE). \* $p < 0.05$  and \*\* $p < 0.01$ , assessed by one-way ANOVA and Tukey's test,  $n = 3$ .**