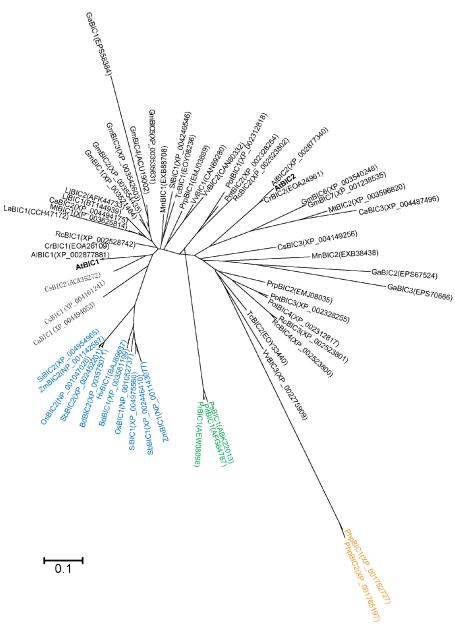
Supporting Information - Figures

A CRY-BIC negative feedback circuitry regulating blue light sensitivity of Arabidopsis

Xu Wang^{1,7,8}, Qin Wang^{1,7,8}, Yun-Jeong Han^{2,8}, Qing Liu¹, Liangfeng Gu¹, Zhaohe Yang¹, Jun Su¹, Bobin Liu¹, Zecheng Zuo¹, Wenjin He^{3,7}, Jian Wang⁴, Bin Liu⁵, Minami Matsui⁶, Jeong-II Kim^{2*}, Yoshito Oka^{1*}, and Chentao Lin^{7*}





Moss Php: *Physcomitrella patens*

Gymnosperm Pr: *Pinus radiata* Ps: *Picea sitchensis* Pit: *Pinus taeda*

Angiosperm(Dicot) Al: Arabidopsis lyrata At: Arabidopsis thailana Ca: Cicer arietinum Cr: Capsella rubella Cs: Cucumis sativus Ga: Genlisea aurea Gm: Glyaine max La: Lupinus angustfolius Lj: Lotus japonicus Mn: Morus notabilis Mt: Medicago truncatula Prp: Prunus persica Pot: Populus trichocarpa Rc: Ricinus communis SI: Solanum lycopersicum Tc: Theobroma cacao Vv: Vtit si vinifera

Angiosperm(Monocot) Bd: Brachypodium distachyon Hv: Hordeum vulgare Os: Oryza sativa Sb: Sorghum bicolor Si: Setaria italica Zm: Zea mays

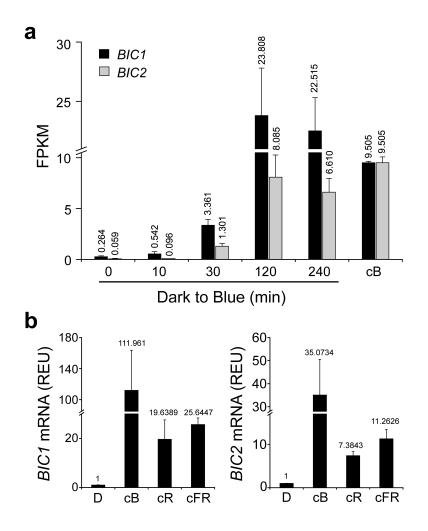
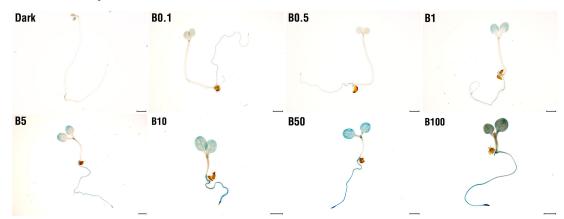


Figure S3



a 489BIC1pro::GUS

b 2100BIC2pro::GUS

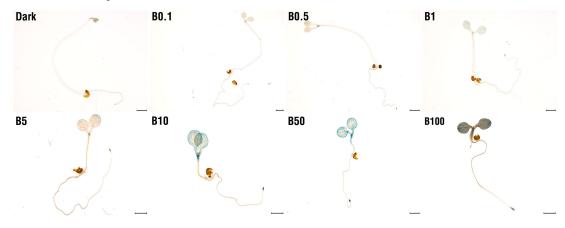


Figure S4

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Chr 3

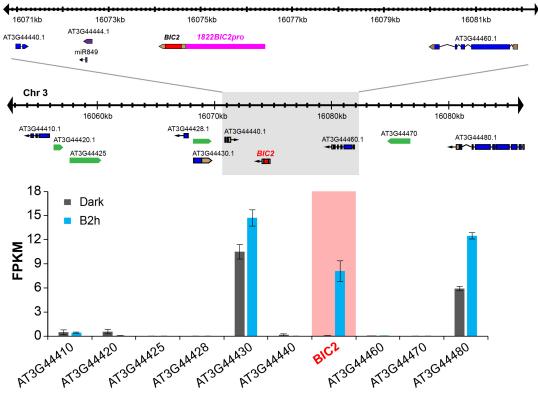
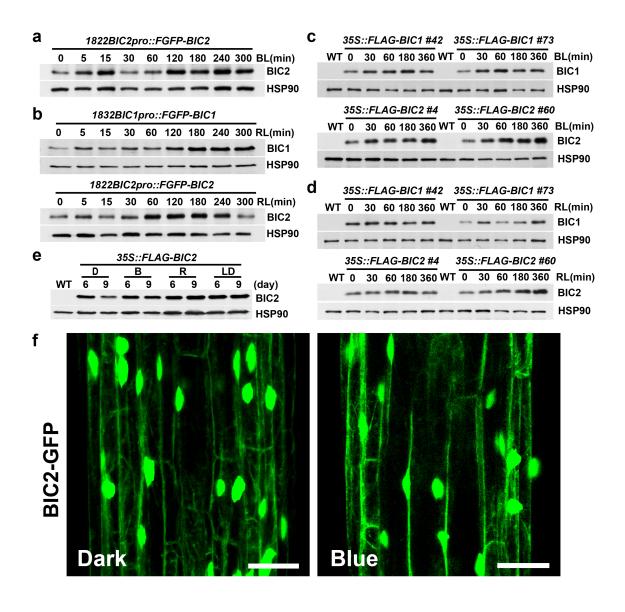
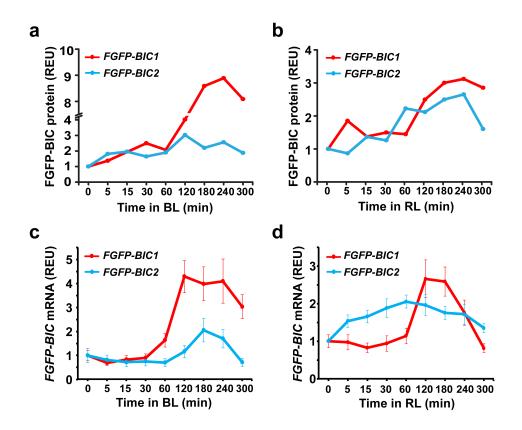


Figure S5





Supporting Information - Table

A CRY-BIC negative feedback circuitry regulating blue light sensitivity of Arabidopsis

Xu Wang^{1,7,8}, Qin Wang^{1,7,8}, Yun-Jeong Han^{2,8}, Qing Liu¹, Liangfeng Gu¹, Zhaohe Yang¹, Jun Su¹, Bobin Liu¹, Zecheng Zuo¹, Wenjin He^{3,7}, Jian Wang⁴, Bin Liu⁵, Minami Matsui⁶, Jeong-II Kim^{2*}, Yoshito Oka^{1*}, and Chentao Lin^{7*}

| Time (min) | Fluence rate (µmol·m ⁻² ·s ⁻¹) | BIC1 REU | S.E. | BIC2 REU | S.E. |
|---------------|---|------------|-----------|-----------|---------|
| 0 | 0.1 | 1 | 0.23526 | 1 | 0.18915 |
| 20 | 0.1 | 4.65893 | 0.97559 | 2.38392 | 0.47806 |
| 40 | 0.1 | 42.84001 | 5.93908 | 9.06307 | 1.82179 |
| 60 | 0.1 | 47.61464 | 4.27414 | 18.85227 | 3.39669 |
| 120 | 0.1 | 273.7408 | 60.58904 | 26.35491 | 4.91537 |
| 0 | 0.5 | 1 | 0.23526 | 1 | 0.18915 |
| 20 | 0.5 | 4.78991 | 1.04816 | 2.89454 | 0.48194 |
| 40 | 0.5 | 40.91272 | 7.71456 | 11.3924 | 3.65014 |
| 60 | 0.5 | 61.67726 | 5.44981 | 22.05958 | 3.59029 |
| 120 | 0.5 | 507.28991 | 95.7389 | 55.71524 | 7.94898 |
| 0 | 1 | 1 | 0.23526 | 1 | 0.18915 |
| 20 | 1 | 5.61778 | 1.0917 | 4 | 0.67555 |
| 40 | 1 | 39.48775 | 7.05874 | 11.2876 | 2.08133 |
| 60 | 1 | 51.50593 | 4.59981 | 23.53387 | 3.61337 |
| 120 | 1 | 514.37142 | 94.28539 | 116.16245 | 16.9215 |
| 0 | 5 | 1 | 0.23526 | 1 | 0.18915 |
| 20 | 5 | 7.04533 | 1.54067 | 9.38268 | 1.59526 |
| 40 | 5 | 50.06651 | 9.33013 | 15.49073 | 2.31373 |
| 60 | 5 | 50.0975 | 4.66786 | 27.66519 | 5.24691 |
| 120 | 5 | 588.13356 | 100.023 | 133.74428 | 21.5093 |
| 0 | 10 | 1 | 0.23526 | 1 | 0.18915 |
| 20 | 10 | 16.37398 | 3.22283 | 18.63574 | 2.93425 |
| 40 | 10 | 43.41134 | 7.91556 | 18.9834 | 3.80151 |
| 60 | 10 | 78.9755 | 7.49194 | 36.9286 | 6.23327 |
| 120 | 10 | 636.19858 | 109.4778 | 137.187 | 28.6800 |
| 0 | 50 | 1 | 0.23526 | 1 | 0.18915 |
| 20 | 50 | 15.67072 | 3.16481 | 16.91229 | 3.29628 |
| 40 | 50 | 52.34573 | 10.79796 | 32.37182 | 6.24525 |
| 60 | 50 | 154.3434 | 12.18343 | 56.62367 | 10.9685 |
| 120 | 50 | 805.27254 | 135.5645 | 244.41618 | 24.8442 |
| 0 | 100 | 1 | 0.23526 | 1 | 0.18915 |
| 20 | 100 | 12.52437 | 2.53298 | 16.99062 | 2.94425 |
| 40 | 100 | 124.21251 | 25.29764 | 32.50475 | 6.84529 |
| 60 | 100 | 217.26822 | 36.6526 | 67.80564 | 9.69295 |
| 120 | 100 | 1392.37619 | 250.31219 | 271.22255 | 43.7270 |

Table S1. The relative expression levels of *BIC1* and *BIC2* in response to bluelight shown in Figure 2.

Supporting Information Legends

Figure S1. Phylogenetic analysis of BIC proteins.

The unrooted phylogenetic tree was generated by the neighbor-joining method. Genus/species abbreviations are shown in the figure. The scale bar indicates substitution per site.

Figure S2. BIC mRNA expression in response to lights.

a, RNA-seq results showing the absolute expression levels of *BIC* genes in response to blue light. 5-day-old wild-type seedlings grown under continuous blue light (cB; 20 μ mol·m⁻²·s⁻¹) and 5-day-old wild type etiolated seedlings exposed to blue light (20 μ mol·m⁻²·s⁻¹) for indicated time (min) were used for RNA-seq. The mean FPKM (Fragments Per Kilobase of transcript per Million mapped reads) values of three repeats for each sample are indicated on the top of each column. Standard deviations (n=3 biological replicates) are shown.

b, qPCR showing the levels of mRNA expression of *BIC1* and *BIC2* under different light conditions. RNA was isolated from the wild-type seedlings grown in the dark (D), blue light (cB; 8 μ mol·m⁻²·s⁻¹), red light (cR; 10 μ mol·m⁻²·s⁻¹) and far-red light (cFR; 6 μ mol·m⁻²·s⁻¹) for 5 days. The relative expression unit (REU) was calculated by re-normalization of the normalized qPCR signal in plants exposed to light against the normalized qPCR signal in plants kept in the dark. *PP2A* (At1g69960) was used as the reference gene for qPCR normalization. The mean REU values of three repeats for each sample are revealed on the top of each column. Standard deviations (n=3) are shown.

Figure S3. Blue light-dependent activity of the *BIC1* and *BIC2* promoters. Histochemical images of GUS stain of transgenic seedlings expressing the *489BIC1pro::GUS* or *2100BIC2pro::GUS* reporters. *Arabidopsis* seedlings expressing the *489BIC1pro::GUS* (**a**) or *2100BIC2pro::GUS* (**b**) were grown in the dark or in blue light (0.1, 0.5, 1, 5, 10, 50 or 100 μ mol·m⁻²·s⁻¹) for 5 days and stained with X-gluc. Scale bar=1 mm.

Figure S4. The chromatin segments where *BIC2* gene resides in is not transcription-repressed region in the dark.

Genome browser view of the BIC2 locus on chromosome 3 in A. thaliana accession Col. The 10kb region (top) enlarged from the ~40kb (middle) areas around the BIC2 gene and all genes within this ~40kb region are shown. Blue and golden boxes represent exons and untranslated regions are shown. BIC2 has no intron and it is highlighted by red color. Green boxes represent non-protein-coding RNAs genes. Purple boxes represent microRNA genes. Arrows indicate the direction of transcription. The promoter sequence used for making transgenic lines in Figure. 3f and Figure S5a-b (1822BIC2pro::FGFP-BIC2) are represented by a pink box. The relative levels of mRNA expression derived from a RNA-seq experiment for all the genes in the ~40kb region are shown (bottom). In this RNA-seq experiment, etiolated wild-type seedlings were kept in the dark (Dark) or exposed to blue light (20 µmol·m⁻²·s⁻¹) for 2 hr (B2h) were analyzed. FPKM stands for Fragments Per Kilobase of transcript per Million mapped reads. Three biological repeats were performed for RNA-seq analysis. The error bar indicates the standard deviation of FPKM values of each gene for three repeats. Levels of BIC1 mRNA expression is highlighted, the absolute values of BIC1 mRNA expression is included in Figure S2a.

Figure S5. Light slightly enhances BICs protein stabilities without affecting subcellular localization of BIC2.

a and **b**, Immunoblots showing that levels of FGFP-BIC1 and FGFP-BIC2 recombinant proteins in transgenic plants expressing the *1832BIC1pro::FGFP-BIC1* or *1822BIC2pro::FGFP-BIC2* transgene driven by the *BIC* promoters. 7-day-old etiolated seedlings kept in the dark or exposed to blue (a; BL; 30 μmol·m⁻²·s⁻¹) and red light (b; RL; 30 μmol·m⁻²·s⁻¹) for indicated time (min) before sample collection. The recombinant FGFP-BIC1 (Flag-GFP-BIC1) and BIC2 (Flag-GFP-BIC2) protein were analyzed by

immunoblot probed with the anti-Flag antibody. HSP90 was used as the loading control. The upstream regions of *BIC1* (1832 bp including 5'-UTR) and *BIC2* (1822 bp including 5'-UTR) genes were used as the *BIC* promoters.

c and **d**, Expression of the constitutively transcribed recombinant BICs protein in response to blue (c) and red light (d). 7-day-old etiolated seedlings of wild type (WT), *35S::Flag-BIC1* (#42 and #73) and *35S::Flag-BIC2* (#4 and #60) were exposed to blue (c; BL; 30 μ mol·m⁻²·s⁻¹) and red light (d; RL; 30 μ mol·m⁻²·s⁻¹) for the indicated time (min). The recombinant BIC1 and BIC2 proteins were detected by immunoblot using anti-Flag antibody. HSP90 was used as the loading control.

e, Expression of the constitutively transcribed recombinant BIC2 protein in 6-day-old or 9-day-old seedlings. Seedlings of *35S::Flag-BIC2* were grown in the dark, continuous blue light (30 μmol·m⁻²·s⁻¹), continuous red light (30 μmol·m⁻²·s⁻¹), or long day conditions for 6 or 9 days. The recombinant Flag-BIC2 protein were detected by immunoblot using anti-Flag antibodies. HSP90 was used as the loading control.

f, Lack of light effect on BIC2 subcellular localization. 3-day-old etiolated seedlings of 35S::BIC2-GFP were kept either in the dark (Dark) or blue light (10 µmol·m⁻²·s⁻¹) for 3 hr (Blue). BIC2-GFP subcellular distributions in hypocotyl were observed under a confocal microscope. Scale bar= 50 µm.

Figure S6. Protein and mRNA expression of the *1832BIC1pro::FGFP-BIC1* and *1822BIC2pro::FGFP-BIC1* transgenes in response to light.

a and **b**, Densitometry analysis of the immunoblots shown in Figure. 3c (for the *1832BIC1pro::FGFP-BIC1* transgene), and Figure S5 a-b (for the *1822BIC2pro::FGFP-BIC2* transgene). The band intensities of FGFP-BIC1 and FGFP-BIC2 were digitized, normalized against that of HSP90, then re-normalized to the value of time 0 to obtain the Relative expression units (REU). Two independent protein extracts were used for the immunoblot analysis. The mean value of the REU from two repeats were shown in the Figure S6 a and b.

c and **d**, qPCR showing the mRNA levels of *FGFP-BIC1* and *FGFP-BIC2* transgenes in response to blue (**c**) or red light (**d**). The RNAs isolated from the same tissue samples used for the immunoblot analyses shown in Figure 3c (for the *1832BIC1pro::FGFP*-

BIC1 transgene) or Figure S5a-b (for the *1822BIC2pro::FGFP-BIC1* transgene) were used in this experiment. The relative expression unit (REU) was calculated by renormalization of the normalized qPCR signal of all the samples to the normalized qPCR signals of the sample kept in the dark. *PP2A* (At1g69960) was used as the reference gene for qPCR normalization. Three biological repeats were used for each experiment and the error bars indicate the standard error of three repeats.

Table S1. The relative expression levels of *BIC1* and *BIC2* in response to blue lightshown in Figure 2.