

Supplemental Information

Biophysical Characterization of Human Lipoproteins, their Subclasses and their Lipids by ^1H NMR Spectroscopy

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Tables

Supplemental Table S1. ^1H -NMR resonances of lipids isolated in $\text{CDCl}_3/\text{methanol-d}_4$.^a

Chemical shift ppm]	Compound	Chemical group	Position
0.6949	C	CH_3	C-18
0.6974	CE	CH_3	C-18
0.8635	C / CE	CH_3	C-26, C-27
0.8681	C / CE	CH_3	C-26, C-27
0.8747	C / CE / FA	CH_3	C-26, C-27 / FA- CH_3
0.8790	C / CE / FA	CH_3	C-26, C-27 / FA- CH_3
0.8871	FA	CH_3	FA- CH_3
0.8996	FA	CH_3	FA- CH_3
0.9110	FA	CH_3	FA- CH_3
0.9229	C / CE	CH_3	C-21
0.9337	C / CE	CH_3	C-21
0.9438	C / CE	CH , CH_2	C-9, C-14, C-22
0.9547	C / CE	CH , CH_2	C-9, C-14, C-22
0.9667	C / CE	CH , CH_2	C-9, C-14, C-22
0.9733	C / CE	CH , CH_2	C-9, C-14, C-22
0.9813	C / CE	CH , CH_2	C-9, C-14, C-22
0.9920	C / CE	CH , CH_2	C-9, C-14, C-22
1.0013	C / CE	CH , CH_2	C-9, C-14, C-22
1.0090	C / CE	CH , CH_2	C-9, C-14, C-22
1.0125	C / CE	CH , CH_2	C-9, C-14, C-22
1.0177	C	CH_3	C-19
1.0236	C / CE	CH , CH_2	C-9, C-14, C-22
1.0295	C / CE	CH , CH_2	C-9, C-14, C-22
1.0406	CE	CH_3	C-19
1.0523	C / CE	CH , CH_2	C-1, C-12, C-15, C-17, C-23, C-24
1.0616	C / CE	CH , CH_2	C-1, C-12, C-15, C-17, C-23, C-24
1.0711	C / CE	CH , CH_2	C-1, C-12, C-15, C-17, C-23, C-24
1.0816	C / CE	CH , CH_2	C-1, C-12, C-15, C-17, C-23, C-24
1.0920	C / CE	CH , CH_2	C-1, C-12, C-15, C-17, C-23, C-24
1.1021	C / CE	CH , CH_2	C-1, C-12, C-15, C-17, C-23, C-24
1.1104	C / CE	CH , CH_2	C-1, C-12, C-15, C-17, C-23, C-24
1.1199	C / CE	CH , CH_2	C-1, C-12, C-15, C-17, C-23, C-24

1.1286	C / CE	CH, CH ₂	C-1, C-12, C-15, C-17, C-23, C-24
1.1349	C / CE	CH, CH ₂	C-1, C-12, C-15, C-17, C-23, C-24
1.1452	C / CE	CH, CH ₂	C-1, C-12, C-15, C-17, C-23, C-24
1.1535	C / CE	CH, CH ₂	C-1, C-12, C-15, C-17, C-23, C-24
1.1677	C / CE	CH, CH ₂	C-1, C-12, C-15, C-17, C-23, C-24
1.1770	C / CE	CH, CH ₂	C-1, C-12, C-15, C-17, C-23, C-24
1.1873	C / CE	CH, CH ₂	C-1, C-12, C-15, C-17, C-23, C-24
1.1996	C / CE	CH, CH ₂	C-1, C-12, C-15, C-17, C-23, C-24
1.2090	C / CE	CH, CH ₂	C-1, C-12, C-15, C-17, C-23, C-24
1.2674	FA / C / CE	CH, CH ₂	FA-CH ₂ / C-16, C-20, C-22, C-23
1.2808	FA / C / CE	CH, CH ₂	FA-CH ₂ / C-16, C-20, C-22, C-23
1.2929	FA / C / CE	CH, CH ₂	FA-CH ₂ / C-16, C-20, C-22, C-23
1.3070	FA / C / CE	CH, CH ₂	FA-CH ₂ / C-16, C-20, C-22, C-23
1.3202	FA / C / CE	CH, CH ₂	FA-CH ₂ / C-16, C-20, C-22, C-23
1.3427	C / CE / FA	CH, CH ₂	C-16, C-20, C-22, C-23 / FA-CH ₂
1.3528	C / CE / FA	CH, CH ₂	C-16, C-20, C-22, C-23 / FA-CH ₂
1.3654	C / CE / FA	CH, CH ₂	C-16, C-20, C-22, C-23 / FA-CH ₂
1.3777	C / CE / FA	CH, CH ₂	C-16, C-20, C-22, C-23 / FA-CH ₂
1.4052	C / CE / FA	CH, CH ₂	C-16, C-20, C-22, C-23 / FA-CH ₂
1.4471	C / CE	CH, CH ₂	C-7, C-8, C-11, C-15, C-25
1.4516	C / CE	CH, CH ₂	C-7, C-8, C-11, C-15, C-25
1.4638	C / CE	CH, CH ₂	C-7, C-8, C-11, C-15, C-25
1.4818	C / CE	CH, CH ₂	C-7, C-8, C-11, C-15, C-25
1.4910	C / CE	CH, CH ₂	C-7, C-8, C-11, C-15, C-25
1.5148	C / CE	CH, CH ₂	C-7, C-8, C-11, C-15, C-25
1.5262	C / CE	CH, CH ₂	C-7, C-8, C-11, C-15, C-25
1.5368	C / CE	CH, CH ₂	C-7, C-8, C-11, C-15, C-25
1.5478	C / CE	CH, CH ₂	C-7, C-8, C-11, C-15, C-25
1.5599	C / CE	CH, CH ₂	C-7, C-8, C-11, C-15, C-25
1.5797	C / CE / FA	CH ₂ / CH ₂ -CH ₂ -COO	C-7, C-8, C-11, C-15, C-25 / FA-CH ₂
1.5849	C / CE / FA	CH ₂ / CH ₂ -CH ₂ -COO	C-7, C-8, C-11, C-15, C-25 / FA-CH ₂
1.6049	C / CE / FA	CH ₂ / CH ₂ -CH ₂ -COO	C-7, C-8, C-11, C-15, C-25 / FA-CH ₂
1.6233	FA	CH ₂ -CH ₂ -COO	FA-CH ₂
1.6352	FA	CH ₂ -CH ₂ -COO	FA-CH ₂
1.67-1.78	FA	CH=CH-CH ₂ -CH ₂ -CH ₂ -COO	FA-CH ₂
1.8171	C / CE	CH ₂	C-1, C-2, C-16
1.8278	C / CE	CH ₂	C-1, C-2, C-16
1.8337	C / CE	CH ₂	C-1, C-2, C-16
1.8444	C / CE	CH ₂	C-1, C-2, C-16
1.8547	C / CE	CH ₂	C-1, C-2, C-16
1.8672	C / CE	CH ₂	C-1, C-2, C-16
1.8750	C / CE	CH ₂	C-1, C-2, C-16
1.8804	CE	CH ₂	C-2
1.8967	CE	CH ₂	C-2
1.9028	CE	CH ₂	C-2
1.9152	CE	CH ₂	C-2
1.9644	C / CE	CH ₂	C-7, C-12
1.9734	C / CE	CH ₂	C-7, C-12

1.9807	C / CE	CH_2	C-7, C-12
2.0034	C / CE / FA	$\text{CH}_2 / \text{CH}_2\text{-CH=CH}$	C-7, C-12 / FA- CH_2
2.0192	C / CE / FA	$\text{CH}_2 / \text{CH}_2\text{-CH=CH}$	C-7, C-12 / FA- CH_2
2.0262	C / CE / FA	$\text{CH}_2 / \text{CH}_2\text{-CH=CH}$	C-7, C-12 / FA- CH_2
2.0407	C / CE / FA	$\text{CH}_2 / \text{CH}_2\text{-CH=CH}$	C-7, C-12 / FA- CH_2
2.0673	FA	$\text{CH=CH-CH}_2\text{-CH}_2\text{-CH}_2\text{-COO}$	FA- CH_2
2.0770	FA	$\text{CH=CH-CH}_2\text{-CH}_2\text{-CH}_2\text{-COO}$	FA- CH_2
2.0993	FA	$\text{CH=CH-CH}_2\text{-CH}_2\text{-CH}_2\text{-COO}$	FA- CH_2
2.1266	FA	$\text{CH=CH-CH}_2\text{-CH}_2\text{-CH}_2\text{-COO}$	FA- CH_2
2.1383	FA	$\text{CH=CH-CH}_2\text{-CH}_2\text{-CH}_2\text{-COO}$	FA- CH_2
2.1662	FA	$\text{CH=CH-CH}_2\text{-CH}_2\text{-CH}_2\text{-COO}$	FA- CH_2
2.1798	FA	$\text{CH=CH-CH}_2\text{-CH}_2\text{-CH}_2\text{-COO}$	FA- CH_2
2.1923	FA	$\text{CH=CH-CH}_2\text{-CH}_2\text{-CH}_2\text{-COO}$	FA- CH_2
2.2028	C	CH_2	C-4
2.2262	C	CH_2	C-4
2.2388	C	CH_2	C-4
2.2431	C	CH_2	C-4
2.2471	C	CH_2	C-4
2.2505	C	CH_2	C-4
2.2542	C	CH_2	C-4
2.2602	C / CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.2620	C / CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.2716	C / CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.2776	C / CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.2826	C / CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.2950	CE / FA	$\text{CH}_2\text{-COO}$	FA- CH_2
2.3072	CE / FA	$\text{CH}_2\text{-COO}$	FA- CH_2
2.3188	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3275	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3295	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3323	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3386	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3417	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3450	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3496	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3518	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3574	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3586	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.3620	CE / FA	$\text{CH}_2 / \text{CH}_2\text{-COO}$	C-4, FA- CH_2
2.4125	FA	$\text{CH=CH-CH}_2\text{-CH}_2\text{-COO}$	FA- CH_2
2.7688	FA	$\text{CH=CH-CH}_2\text{-CH=CH}$	FA- CH_2
2.7800	FA	$\text{CH=CH-CH}_2\text{-CH=CH}$	FA- CH_2
2.7916	FA	$\text{CH=CH-CH}_2\text{-CH=CH}$	FA- CH_2
2.8147	FA	$\text{CH=CH-(CH}_2\text{-CH=CH-)}_x$	FA- CH_2
2.8231	FA	$\text{CH=CH-(CH}_2\text{-CH=CH-)}_x$	FA- CH_2
2.8341	FA	$\text{CH=CH-(CH}_2\text{-CH=CH-)}_x$	FA- CH_2
2.8411	FA	$\text{CH=CH-(CH}_2\text{-CH=CH-)}_x$	FA- CH_2
2.8514	FA	$\text{CH=CH-(CH}_2\text{-CH=CH-)}_x$	FA- CH_2

2.8607	FA	$\text{CH}=\text{CH}-(\text{CH}_2-\text{CH}=\text{CH}-)_x$	FA- CH_2
2.8682	FA	$\text{CH}=\text{CH}-(\text{CH}_2-\text{CH}=\text{CH}-)_x$	FA- CH_2
3.0919	PE	CH_2	PE-2'
3.1026	PE	CH_2	PE-2'
3.1099	PE	CH_2	PE-2'
3.2140	SM	$\text{N}^+(\text{CH}_3)_3$	SM- CH_3
3.2246	PC	$\text{N}^+(\text{CH}_3)_3$	PC- CH_3
3.4402	C / MeOH	CHOH	C-3
3.4485	C / MeOH	CHOH	C-3
3.4590	C / MeOH	CHOH	C-3
3.4669	C / MeOH	CHOH	C-3
3.4748	C / MeOH	CHOH	C-3
3.4853	C / MeOH	CHOH	C-3
3.4936	C / MeOH	CHOH	C-3
3.5974	Choline	CH_2	PC-2'
3.6038	Choline	CH_2	PC-2'
3.6125	Choline	CH_2	PC-2'
3.7616	PI	CH	PI-6'
3.7772	PI	CH	PI-6'
3.7938	PI	CH	PI-6'
3.8684	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.8727	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.8791	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.8847	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.8907	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.8963	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.9015	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.9118	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.9174	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.9251	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.9316	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.9389	HG	CH, CH_2	PLA-1 ^d ,1 ^u , CAR-1',2',3', SPH-2,3 ^u , PI,1'
3.9932	HG	CH_2	PLA-3, PC-3, PE-3
4.0038	HG	CH_2	PLA-3, PC-3, PE-3
4.0139	HG	CH_2	PLA-3, PC-3, PE-3
4.1467	HG	CH_2	PI-1 ^u , PC-1 ^u , PE-1 ^u , PI-1', TG-1 ^u , DG-1 ^u , SPH-3 ^d
4.1579	HG	CH_2	PI-1 ^u , PC-1 ^u , PE-1 ^u , PI-1', TG-1 ^u , DG-1 ^u , SPH-3 ^d
4.1672	HG	CH_2	PI-1 ^u , PC-1 ^u , PE-1 ^u , PI-1', TG-1 ^u , DG-1 ^u , SPH-3 ^d
4.1778	HG	CH_2	PI-1 ^u , PC-1 ^u , PE-1 ^u , PI-1', TG-1 ^u , DG-1 ^u , SPH-3 ^d
4.1871	HG	CH_2	PI-1 ^u , PC-1 ^u , PE-1 ^u , PI-1', TG-1 ^u , DG-1 ^u , SPH-3 ^d
4.2511	Choline	CH_2	PC-1'
4.3243	TG	CH_2	TG-1 ^d
4.3310	TG	CH_2	TG-1 ^d
4.3439	TG / DG	CH_2	TG-1 ^d , DG-1 ^d
4.3508	TG / DG	CH_2	TG-1 ^d , DG-1 ^d
4.3615	PLA	$-\text{CH}_2-\text{CH}=\text{CH}-\text{O}-$	PLA-b
4.3713	PLA	$-\text{CH}_2-\text{CH}=\text{CH}-\text{O}-$	PLA-b
4.4156	HG	CH_2	PC-1 ^d , PE-1 ^d , PI-1 ^d

4.4206	HG	CH₂	PC-1 ^d , PE-1 ^d , PI-1 ^d
4.4354	HG	CH₂	PC-1 ^d , PE-1 ^d , PI-1 ^d
4.4407	HG	CH₂	PC-1 ^d , PE-1 ^d , PI-1 ^d
4.5716	CE / MeOH	CHOR	C-3
4.5788	CE / MeOH	CHOR	C-3
4.5898	CE / MeOH	CHOR	C-3
4.5993	CE / MeOH	CHOR	C-3
4.6078	CE / MeOH	CHOR	C-3
4.6169	CE / MeOH	CHOR	C-3
4.6260	CE / MeOH	CHOR	C-3
5.2225	HG	CH	PC-2, PE-2, PI-2
5.2279	HG	CH	PC-2, PE-2, PI-2
5.2324	HG	CH	PC-2, PE-2, PI-2
5.2374	HG	CH	PC-2, PE-2, PI-2
5.2431	HG	CH	PC-2, PE-2, PI-2
5.2485	HG	CH	PC-2, PE-2, PI-2
5.2521	HG	CH	PC-2, PE-2, PI-2
5.2577	HG	CH	PC-2, PE-2, PI-2
5.2764	TG	CH	TG-2
5.2845	TG	CH	TG-2
5.2925	TG	CH	TG-2
5.3086	FA	CH=CH	FA-CH
5.3205	FA	CH=CH	FA-CH
5.3268	FA	CH=CH	FA-CH
5.3389	C / FA	CH, CH=CH	C-6, FA-CH
5.3448	C / FA	CH, CH=CH	C-6, FA-CH
5.3494	C / FA	CH, CH=CH	C-6, FA-CH
5.3543	C / FA	CH, CH=CH	C-6, FA-CH
5.3625	FA	CH=CH	FA-CH
5.3680	FA	CH=CH	FA-CH
5.3780	CE / FA	CH, CH=CH	C-6, FA-CH
5.3848	CE / FA	CH, CH=CH	C-6, FA-CH
5.3923	CE / FA	CH, CH=CH	C-6, FA-CH
5.4049	FA	CH=CH	FA-CH
5.4423	SM	-CH₂-CH=CH-CHOH-	SPH-a
5.4550	SM	-CH₂-CH=CH-CHOH-	SPH-a
5.4683	SM	-CH₂-CH=CH-CHOH-	SPH-a
5.6887	SM	-CH₂-CH=CH-CHOH-	SPH-b
5.7011	SM	-CH₂-CH=CH-CHOH-	SPH-b
5.7129	SM	-CH₂-CH=CH-CHOH-	SPH-b
5.7259	SM	-CH₂-CH=CH-CHOH-	SPH-b
5.7383	SM	-CH₂-CH=CH-CHOH-	SPH-b
5.9181	PLA	-CH₂-CH=CH-O-	PLA-a
5.9279	PLA	-CH₂-CH=CH-O-	PLA-a

^aSamples were contained in a mixture of CDCl₃/methanol-d₄ (2:1, by volume). In addition, 0.03 vol% tetramethylsilane (TMS) and 14.11 mM pyrazine was added as reference. The spectra were recorded at 293 K at 600.12 MHz and referenced to internal TMS. For obtaining approximate chemical shifts relative to DSS in D₂O a value of 0.074 ppm has to be added to the given values (see Materials

and Methods). However, the size of the particles also determine the chemical shifts. In serum at high DSS concentrations (32 mM) the mean position of the cholesterol C18 signal is 0.722 ppm. With the above correction the mean size dependent shift would be -0.05 ppm. C, cholesterol; CAR, cardiolipin; CE, cholesterol ester; DG; diacylglycerol; FA, fatty acid; HG, head group; MeOH, methanol; PC, phosphatidylcholine; PE, phosphatidylethanolamine; PI, phosphatidylinositol; PLA, plasmalogen; SM, sphingomyelin; SPH, sphingosine; TG, triacylglycerol. The CH protons of the sphingosine and plasmalogene double bonds are designated as a (next to head group) and b (next to alkyl chain). The diastereotopic CH₂ protons with two separated signals are designated as u (upfield) and d (downfield). Note, that the signal positions of the resolved multiplet components are given.

Supplemental Table S2: T₁ relaxations times of internal standards and selected functional groups of lipids.^a

	Pyrazine	TMS	DSS	Choline	Fatty acid	Fatty acid	Fatty acid	Cholesterol
	-(CH) ₄	-(CH₃) ₄	-(CH₃) ₃	-N(CH₃) ₃	-(CH=CH) -CH₂- (CH=CH)-	acid	acid	-C¹⁸H₃
						-CH₂-	-CH₃	
Lipid extracts								
δ [ppm]	8.636	0.00	-	3.202	2.779	1.260	0.883	0.688
T ₁ [s]	6.85	5.65	-	0.82	1.54	1.26	3.16	0.85
Serum								
δ [ppm]	8.642	-	0.00 ^b	3.247	2.759	1.306	0.893	0.67
T ₁ [s]	9.66	-	3.95	0.39	0.47	0.43	0.55	0.48
HDL								
δ [ppm]	-	-	-	3.209	2.697	1.220	0.829	0.644
T ₁ [s]	-	-	-	0.53	0.59	0.56	0.76	0.64

^aThe protons observed are given in bold letters. 600 MHz spectra of lipid extracts were measured at 293 K, of serum and HDL at 310 K. The serum and the HDL sample were from different patients.

^bNote that the DSS signal shifts with its concentration, at 0.1 mM DSS it is located at -0.029 ppm.

Supplemental Table S3. T₁ relaxations times of internal standards at different temperatures in human blood serum.^a

T [K]	Pyrazine -(CH) ₄	Dioxane -(CH) ₄	DSS -(CH) ₃
	T ₁ [s]		
283	5.44	3.30	2.23
293	6.92	4.11	278
303	8.55	4.97	3.54
310	9.66	5.63	3.95
313	10.20	5.91	3.98
323	12.36	7.02	4.81

^a T₁ relaxation times were measured at 600 MHz. The protons observed are given in bold letters.

Supplemental Table S4. NMR signals usable for a quantitative analysis of lipids in NMR spectra of extracts^a

Mean position [ppm]	Integration range [ppm]	Group	Application
0.641	0.550 - 0.732	C, CE (C-18)	Total cholesterol (C _M)
0.841	0.732 - 0.949	C, CE (C-21, C-26, C-27), FA (- CH ₃)	CH ₃ -groups of cholesterol, fatty acids of CE, TG, PL, SL, PLA, and of side chains of SL and PLA.
1.011	0.996 - 1.026	C (C-19)	Cholesterol (C _M)
1.042	1.026 - 1.057	CE (C-19)	Cholesterol esters (C _M)
1.320	1.216 - 1.424	C, CE (C-16, C-20, C-22, C-23), FA (- CH ₂ -)	Estimation of mean fatty acid chain length
1.663	1.552 - 1.672	C, CE (C-7, C-8, C-11, C-15, C-25), FA (- CH ₂ -CH ₂ -COO-)	Total fatty acid residues (C _M)
2.092	1.949 - 2.235	C (C-4), FA (- CH ₂ -CH=CH-)	Double bounds of fatty acid residues (control)
2.339	2.236 - 2.389	C, CE (C-4) / FA (- CH ₂ -COO-)	Total fatty acid residues (control)
2.761	2.720 - 2.801	FA (-CH=CH- CH ₂ -CH=CH-)	Estimation of polyunsaturated fatty acid ratio (diunsaturated:polyunsaturated)
2.823	2.721 - 2.926	FA (-CH=CH-(CH ₂ -CH=CH-) _x), FA (-CH=CH- CH ₂ -CH=CH-)	Estimation of polyunsaturated fatty acid ratio (diunsaturated:polyunsaturated)

2.864	2.801 - 2.927	FA (-CH=CH-(<u>CH</u> ₂ -CH=CH-) _x)	Estimation of polyunsaturated fatty acid ratio (diunsaturated:polyunsaturated)
3.102	3.070 - 3.133	PE (PE-2')	Ethanolamine head group, concentration of PE (C _M)
3.224	3.194 - 3.255	SM (-Choline), PC (-Choline), MeOH- ¹³ C-Satellite	Choline head group of mainly PC and SM (C _M)
3.468	3.422 - 3.513	MeOH- ¹³ C-Satellite, C (C-3)	Used for quantification and correction of MeOH- ¹³ C-Satellite
3.598	3.567 - 3.629	PC (PC-2'), PI (PI-4')	PC and PI (control)
3.778	3.737 - 3.819	PI (PI-6')	Inositol head group of PI (C _M)
4.171	4.141 - 4.200	PI (PI-1 ^u , PI-1'), PC (PC-1 ^u), PE (PE-1 ^u), TG (TG-1 ^u), SPH (SPH-3d)	PI, PC, PE, TG and SPH (control)
4.256	4.217 - 4.295	PC (PC-1')	Choline head group (PC/SM) (control)
4.341	4.302 - 4.380	TG (TG-1 ^d)	Triacylglyceride (TG)
5.233	5.200 - 5.266	PC (PC-2), PE (PE-2), PI (PI-2)	PC, PE and PI (control)
5.283	5.266 - 5.300	TG (TG-2)	TG (control)
5.296	5.042 - 5.551	C, CE (C-1), FA (-CH=CH-), PC (PC-2), PE (PE-2), PI (PI-2)	Estimation of fatty acid saturation degree
5.466	5.436 - 5.496	SL (SPH-a)	Sphingosine (control)
5.720	5.682 - 5.758	SL (SPH-b)	Sphingosine (C _M)
5.921	5.887 - 5.955	PLA (PLA-a)	Plasmalogene (C _M)

^aThe diastereotopic CH₂ protons with two separated signals are designated as u (upfield) and d (downfield). The CH protons of the shingosine and plasmalogene double bonds are designated as a (next to head group) and b (next to alkyl chain). (C_M) useful for calculation of the molar concentration. C = Total Cholesterol, CE = Cholesterol ester, FA = Fatty acid, TG = Triacylglyceride, PC = Phosphatidylcholine, PE = Phosphatidylethanolamine, PI = Phosphatidylinositol, PLA = Plasmalogene, SL = Sphingolipids, SM = Sphingomyelin, SPH = Sphingosine.

Supplemental Table S5. Distribution of fatty acid species in different lipoprotein samples^a

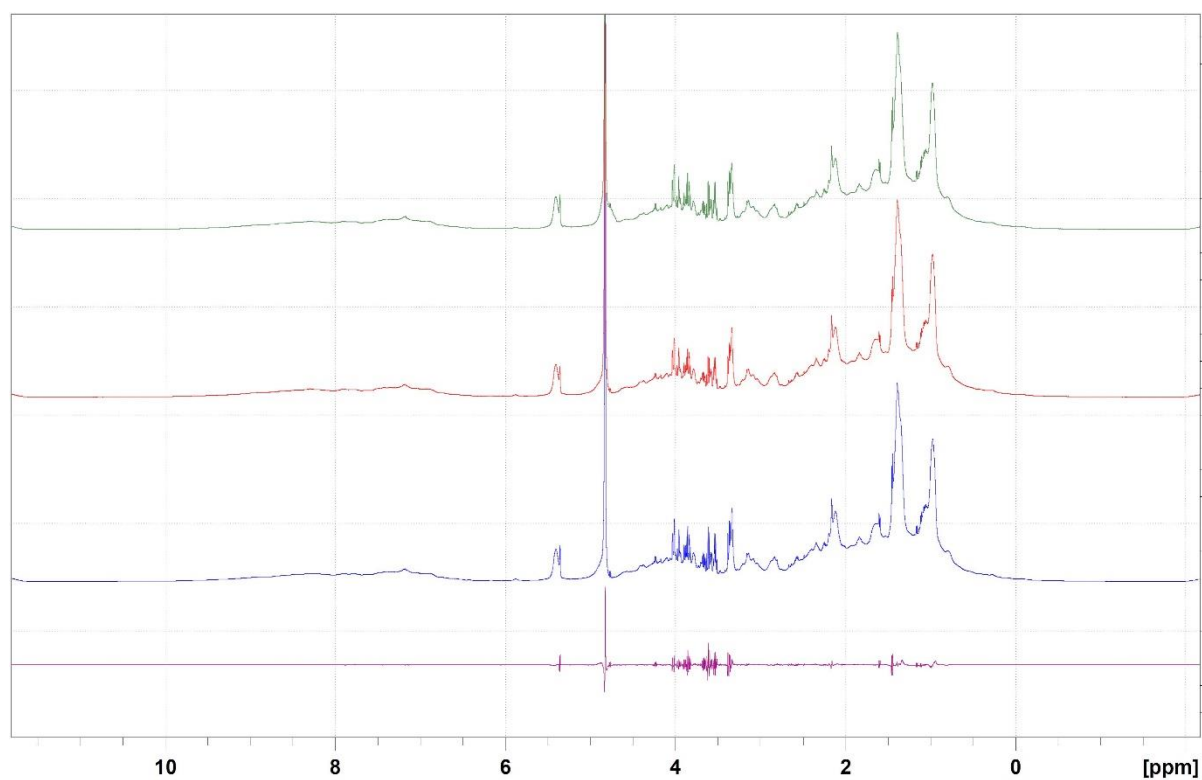
Fatty acid	Serum	VLDL	IDL	LDL	HDL2	HDL3
Relative molar concentration [%]						
16:0	24.0	33.1	29.6	21.9	27.3	33.3

18:0	7.7	7.2	11.5	8.8	9.1	11.6
20:0	0.0	0.0	2.7	1.8	2.1	0.7
22:0	0.0	0.0	1.6	2.9	3.0	1.6
24:0	0.0	0.0	0.0	3.9	4.0	2.1
18:1	16.1	27.1	11.4	6.8	4.8	1.9
24:1	8.0	1.4	2.3	5.6	2.6	0.2
18:2	31.9	22.8	30.3	37.0	34.4	33.6
20:3	3.0	2.6	2.8	2.0	0.2	0.4
20:4	5.1	2.5	2.8	2.7	1.0	2.5
20:5	2.8	1.9	2.2	2.6	4.7	6.4
22:5	0.6	0.9	2.9	4.1	7.2	5.7
22:6	0.8	0.5	0.0	0.0	0.1	0.0

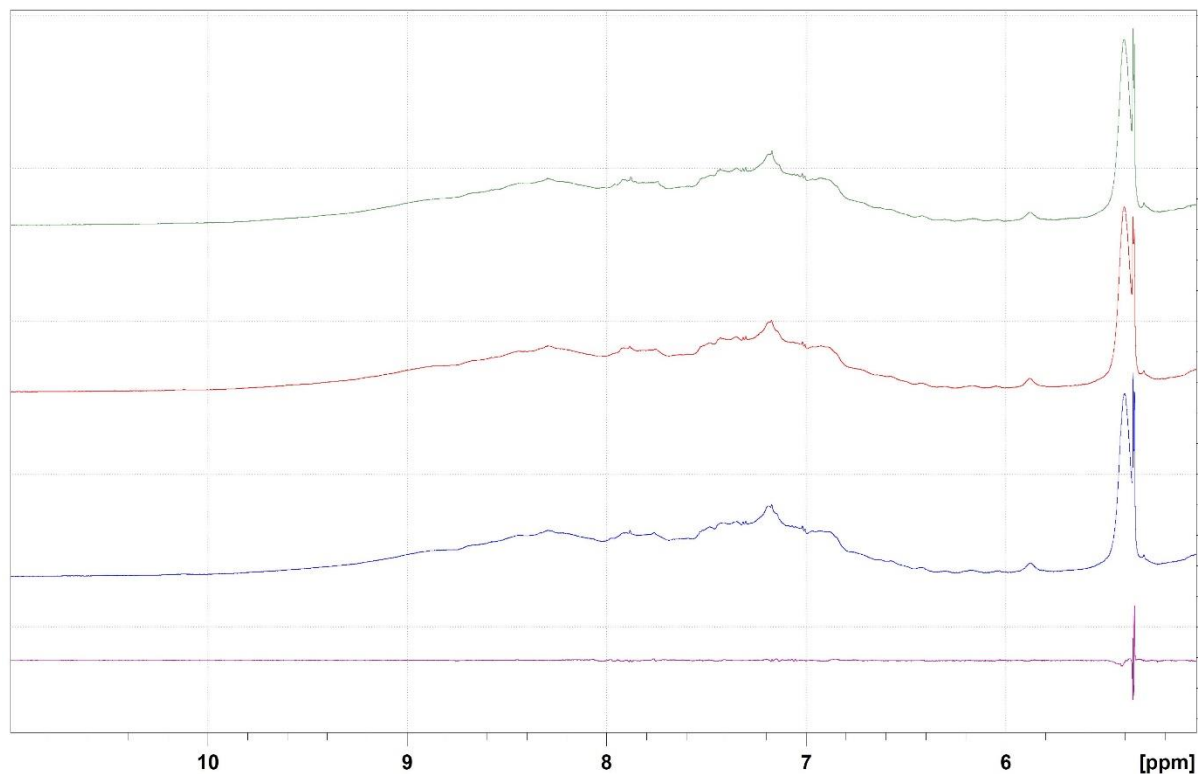
^aThe distribution was obtained by a multidimensional of the NMR spectra of lipoprotein extracts assuming a continuous length distribution.

Figures

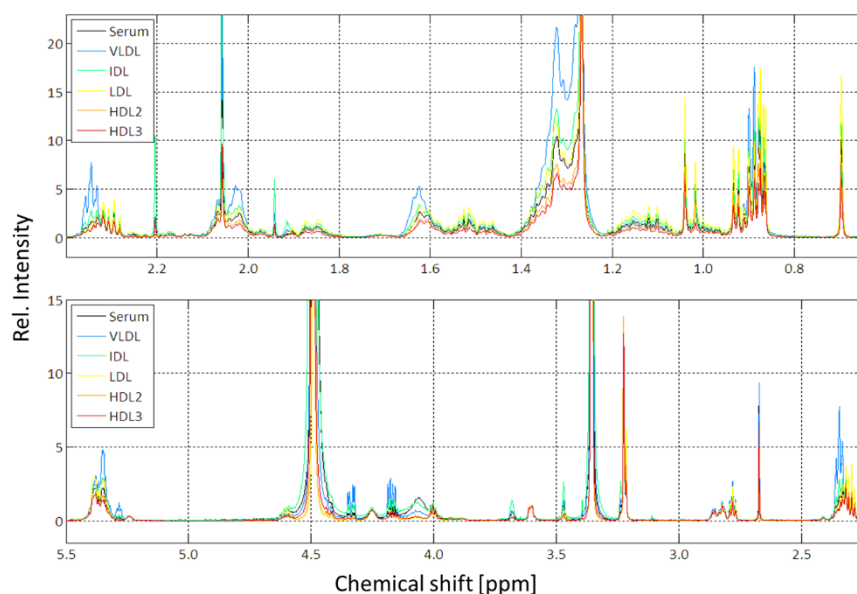
Supplemental Figure S1. Effect of freezing and thawing on ¹H NMR spectra of human blood serum. 600 MHz 1D-NOESY spectra of human blood serum. (from top to bottom) fresh sample, sample after freezing, storing at 253 K for 5 h and thawing, sample after four freezing-thawing cycles, difference between first and last spectrum.



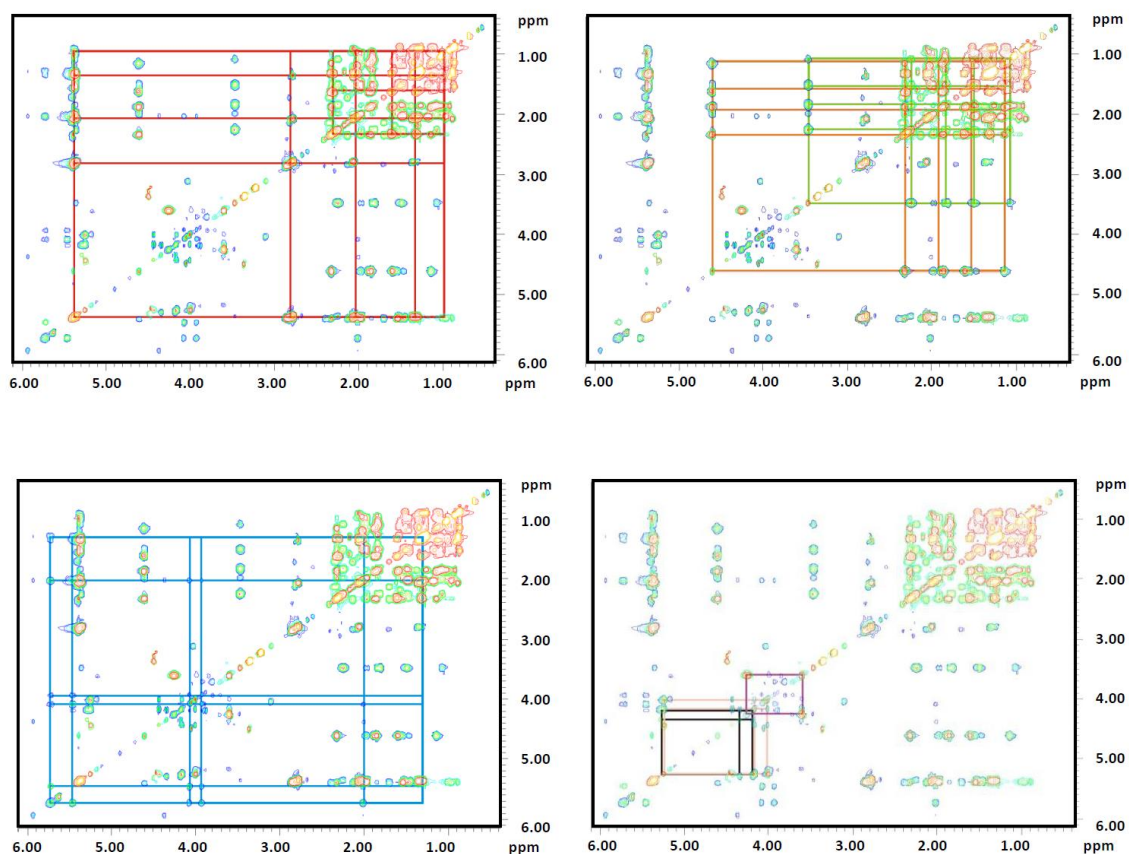
Supplemental Figure S2. Effect of freezing and thawing on ^1H NMR spectra of the proteins of human blood serum. 600 MHz 1D-NOESY spectra of human blood serum, downfield part of the spectra shown in Fig. S1 scaled up 6-times. (from top to bottom) fresh sample, sample after freezing, storing at 253 K for 5 h and thawing, sample after four freezing-thawing cycles, difference between first and last spectrum.



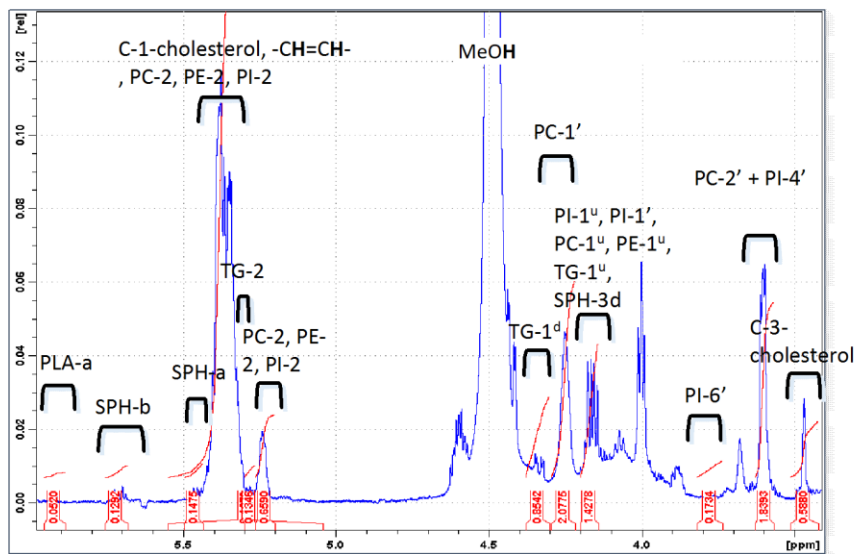
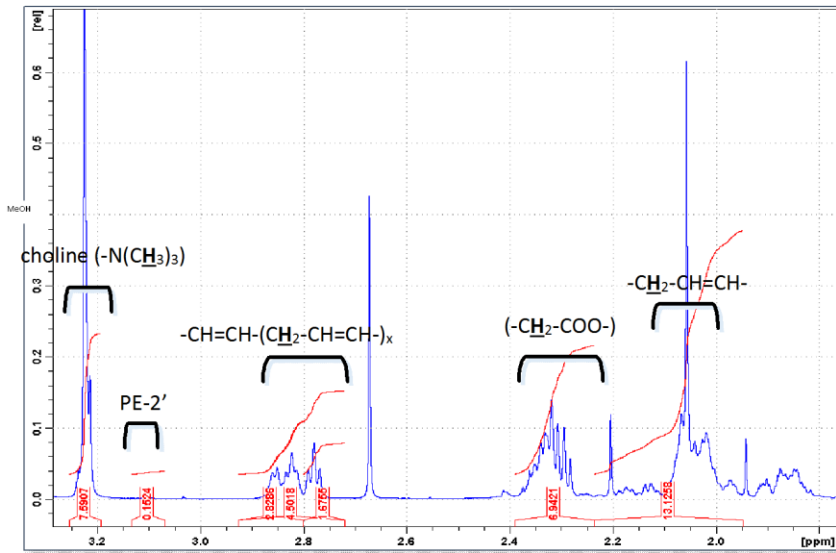
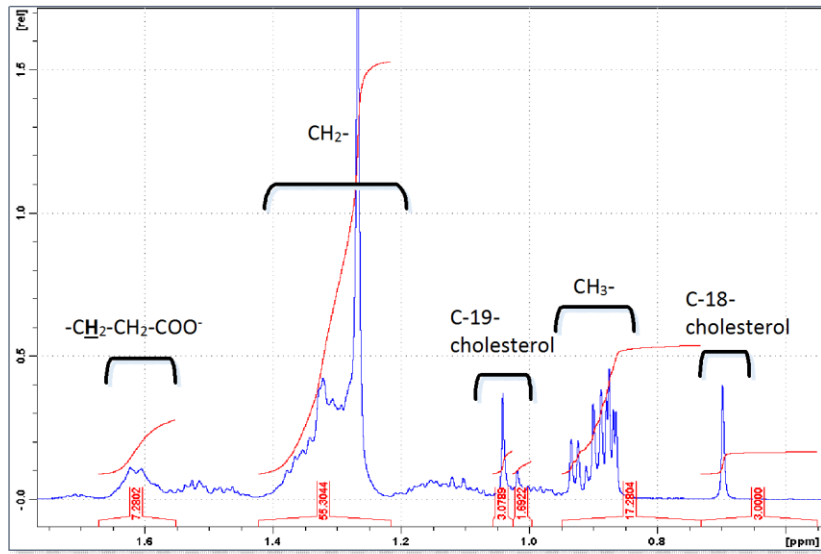
Supplemental Figure S3. Overlay of the 600 MHz ^1H NMR spectra of chloroform-methanol extracts of different lipoprotein fractions. The different lipoprotein fractions obtained by analytical ultracentrifugation and shown in Fig. 2 were extracted with perdeutero chloroform and perdeutero methanol (2:1) as described in Materials and Methods. 600 MHz ^1H NMR spectra, 293 K, reference TMS.



Supplemental Figure S4: Identification of lipid components of LDL extracts. 600 MHz ^1H NMR spectra of LDL extracted as described in Materials and Methods. Temperature 293 K, mixing time 60 ms. The signals of deuterio methanol and chloroform were reduced by suitable processing. (Top, left) signals of fatty acids. (Top, right) signals of cholesterol and cholesterol esters. (Bottom, left) signals of sphingosine. (Bottom, right) signals of choline, glycerol in phosphoglycerides and triacylglycerols. For assignments see Supplemental Table S1.



Supplemental Figure S5. ^1H NMR spectra of a chloroform-methanol extract of HDL3. HDL3 was obtained by analytical ultracentrifugation and extracted with perdeutero chloroform and perdeutero methanol (2:1) as described in Materials and Methods. 600 MHz ^1H NMR spectra, 293 K, reference TMS. The integration ranges of Table S4 are indicated.



Supplemental Figure S6: CH₃ group and cholesterol signal quantification. (A) ¹H-NMR spectra of neat cholesterol in CDCl₃/MeOD with line broadenings LB adapted to the line width of the C-18H₃ resonance of cholesterol of native LDL at 323 K and 293 K. (B) Spectra obtained after subtracting the artificial cholesterol spectra (A) from the CH₂/CH₃ region of LDL.

