

## Supplementary Material

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**‘Germinality does not necessarily define mAb expression and thermal stability’**

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# Amino acid alignments of V<sub>H</sub> and V<sub>L</sub> of mature antibodies

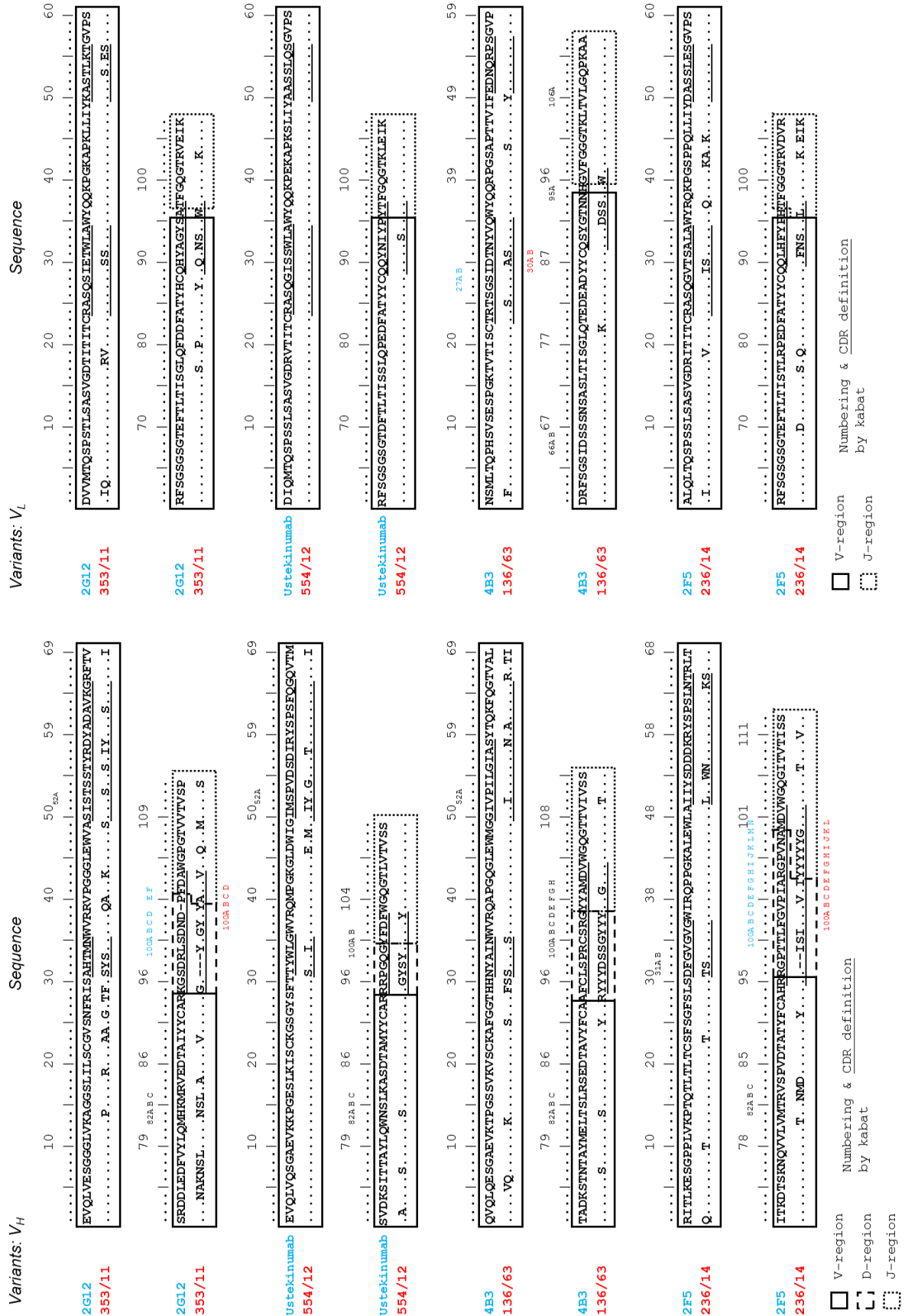


Figure S1: Amino acid alignments of  $V_H$  and  $V_L$  of mature antibodies (2G12, Ustekinumab, 4B3 and 2F5) to their nearest related germline variants (353/11, 554/12, 136/63, 236/14). Numbering and CDRs were defined by Kabat using the abYsis tool (Kabat *et al.*, 1991; Swindells *et al.*, 2017). V(D)J regions were determined by IMGT/DomainGapAlign (Lefranc, 2014) and alignments of D-regions were performed by EMBOSS (Rice *et al.*, 2000).

## Establishment of the host cell line CHO RMCE I3

Created with SnapGene®

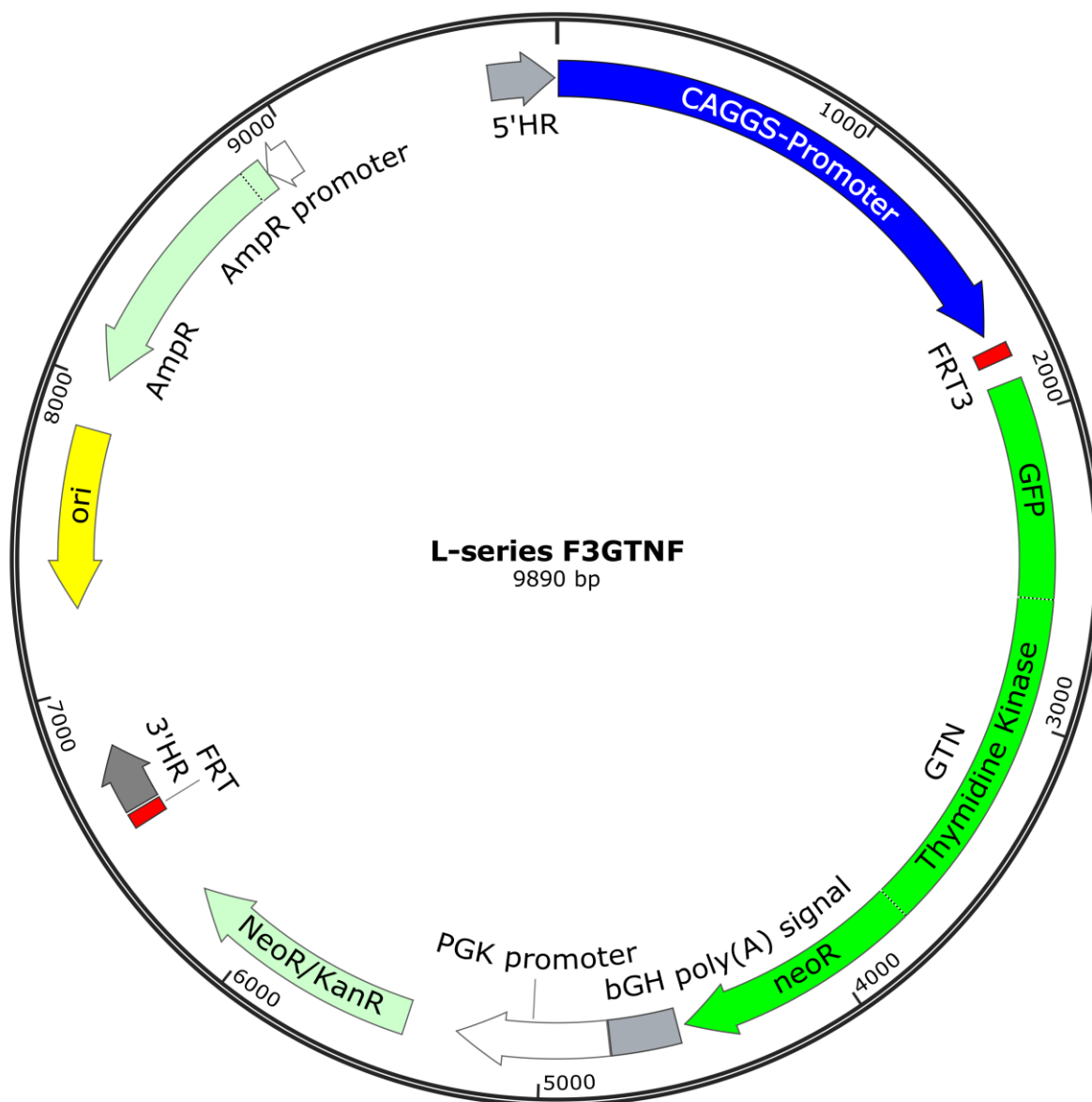


Figure S2: Plasmid map for host cell line development of CHO RMCE I3. To establish a recombinase-mediated cassette exchange (RMCE) system in CHO K1 cells (ATCC CCL-61) a CAGGS promoter was placed 5' upstream of the flippase recognition target (FRT) site to form a promoter trap in the parental RMCE cassette. The fusion protein GTN (gfp/thymidine-kinase/neomycin-phosphotransferase) serves as positive and negative selection marker and

includes the reporter GFP. GTN is located between the two heterospecific FRT sites to be exchanged during the integration of the target gene.

## Cultivation of isogenic IgG-producing cell lines

Established CHO cell lines were routinely cultivated in CD CHO supplemented with 4 mM glutamine, 15 mg/mL phenol red and 2  $\mu$ M GCV in 125 mL Erlenmeyer shake flasks (Corning) at a working volume of 24 mL. Cultivation was done in an ISF1-X incubator (Kuhner) operated at 37°C, 140 rpm, 7% CO<sub>2</sub> and 90% relative humidity. They were seeded at a cell concentration of  $2 \times 10^5$  c/mL and split every three to four days.

## Flow cytometric analysis of intracellular HC and LC

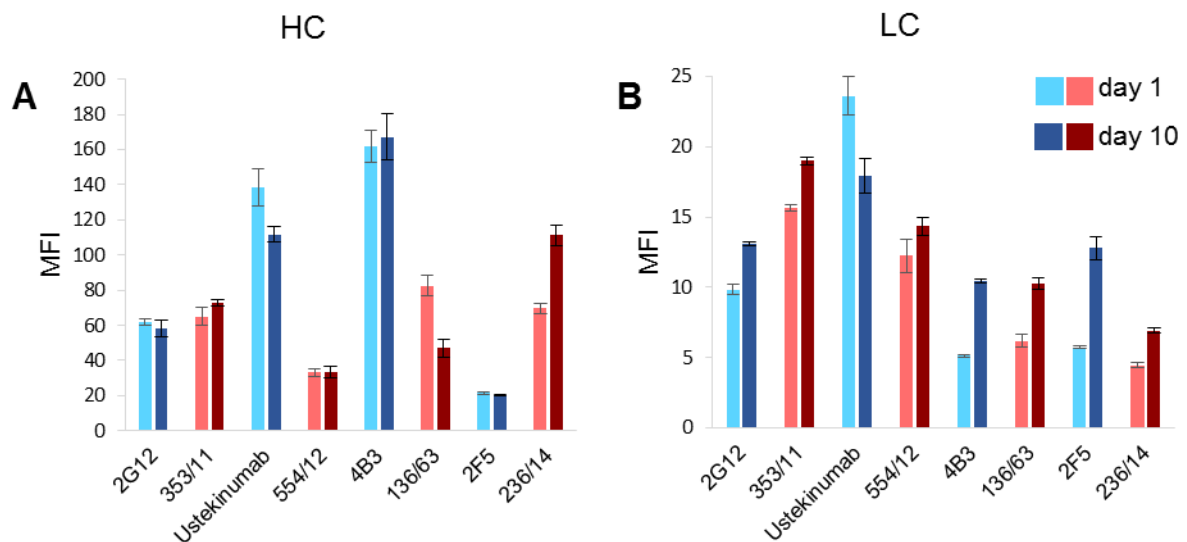


Figure S3: Flow cytometric analysis of intracellular heavy (A) and light chain (B) of eight RMCE cell lines in the semi-continuous perfusion experiment. Samples were taken on day 1 and day 10. CHO RMCE I3 host cell line was used as negative control. Mean values of

intracellular heavy or light chain of IgG producing clones are shown by median fluorescence intensities (MFI).

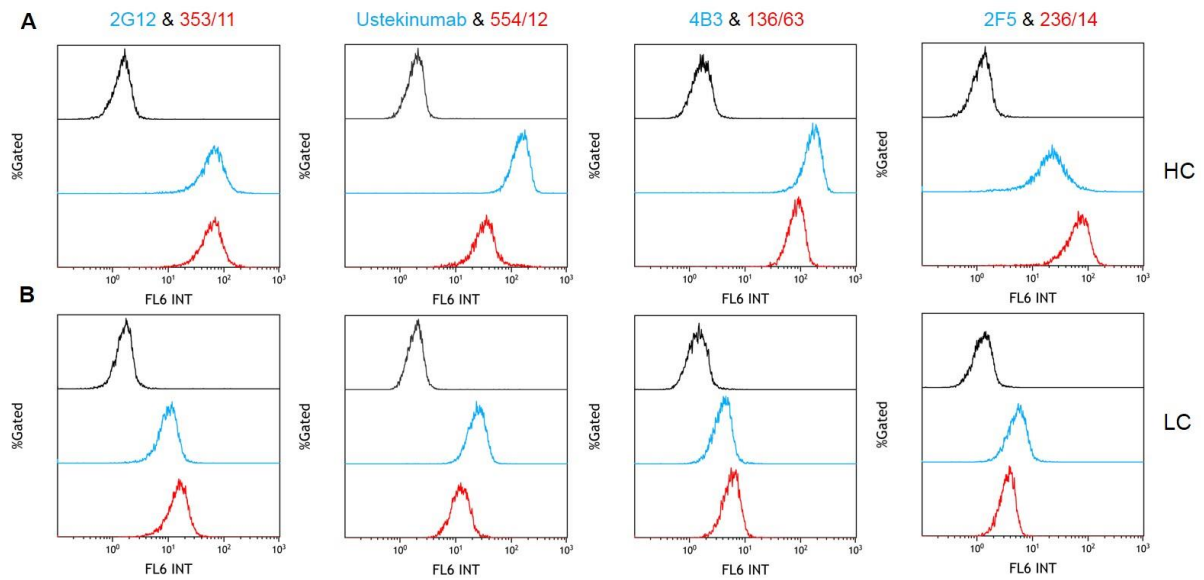


Figure S4: Histograms of flow cytometric analysis of intracellular (A) heavy chain and (B) light chain of eight RMCE cell lines in semi-continuous perfusion. Samples of mature (blue) and germline (red) antibodies.

## Hydropathic index of $V_H$ and $V_L$

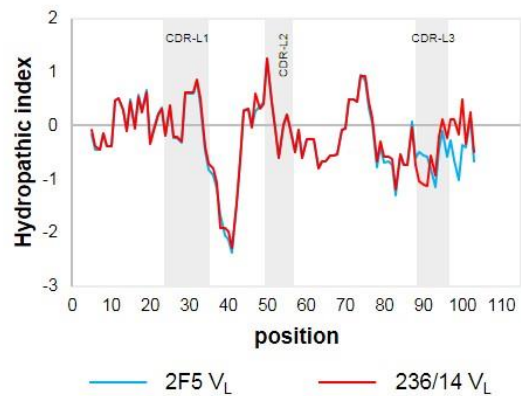
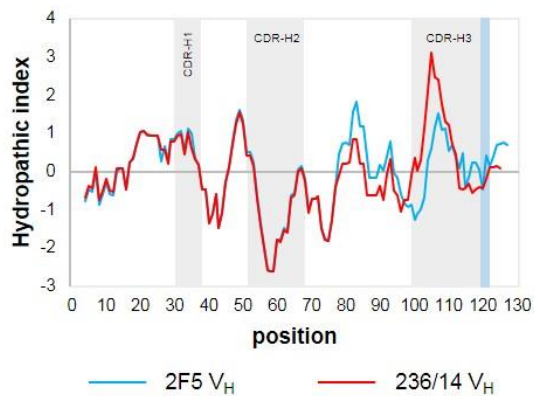
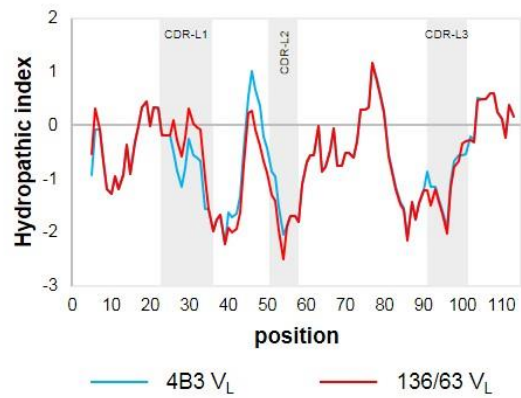
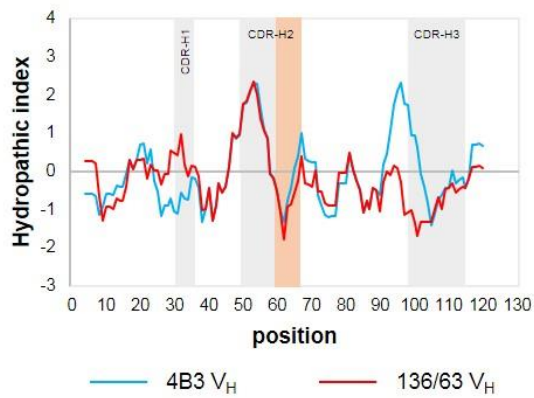
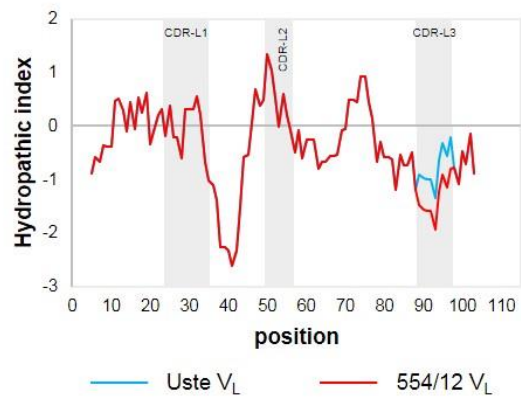
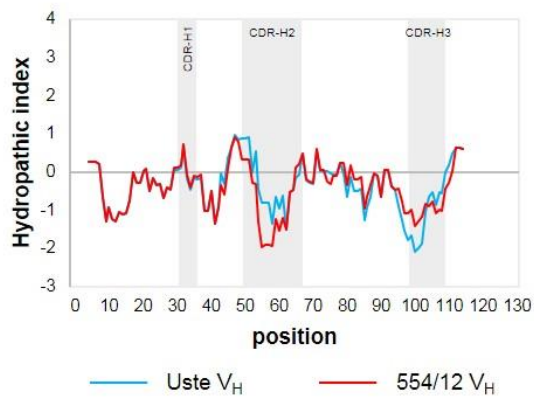
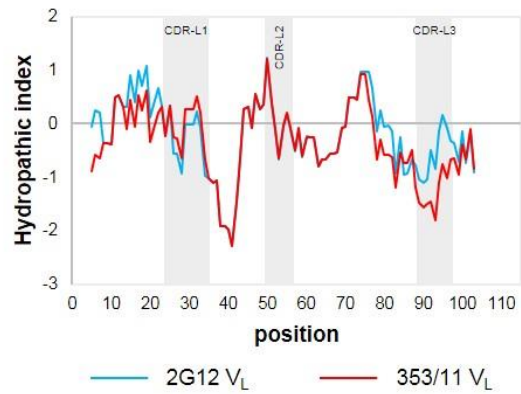
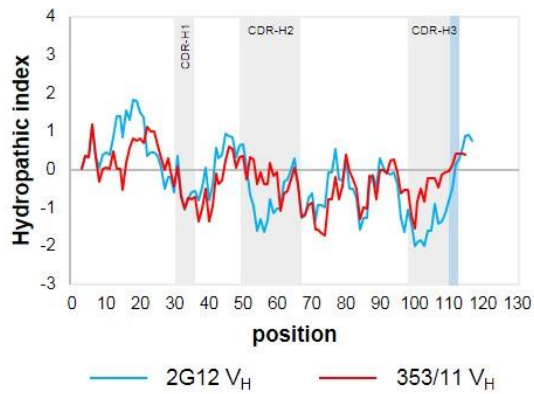


Figure S5: Kyte-Doolittle hydropathy plot of  $V_H$  and  $V_L$  (Kyte and Doolittle, 1982). Kyte-Doolittle analysis using ProtScale (<http://web.expasy.org/protscale/>) (Gasteiger *et al.*, 2005). Parameters were set to: window size: 9, relative weight: 100%, linear model, no normalization. The CDRs are marked in grey for the mature and the germline variant. Residues within the CDRs of only the mature variant are shown in light blue and of the germline variant in light red.

## Comparison of IgG and scFv-Fc antibodies in stable CHO cell lines and transient transfections

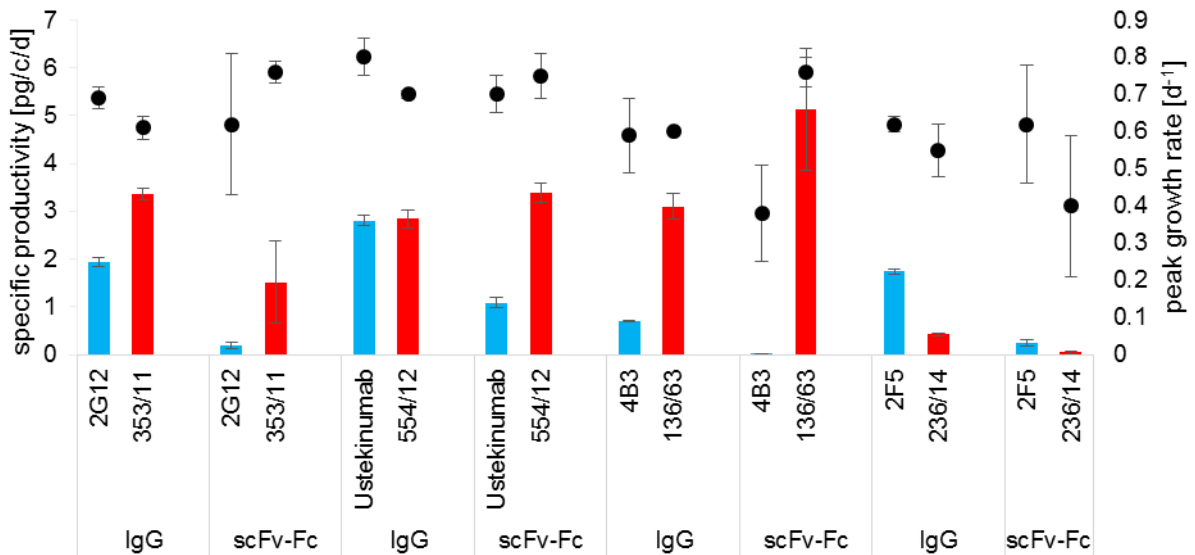


Figure S6: Peak growth rate and peak specific productivity of IgG and scFv-Fc antibodies. IgG's were produced in semi-continuous perfusion experiment with stable CHO cell lines. Same mAb variants were expressed in the scFv-Fc format in HEK293 transient transfection cell pools. Values represent the mean of at least three replicates. Black dots indicate peak growth rate.



## Specificity mature and germline antibodies

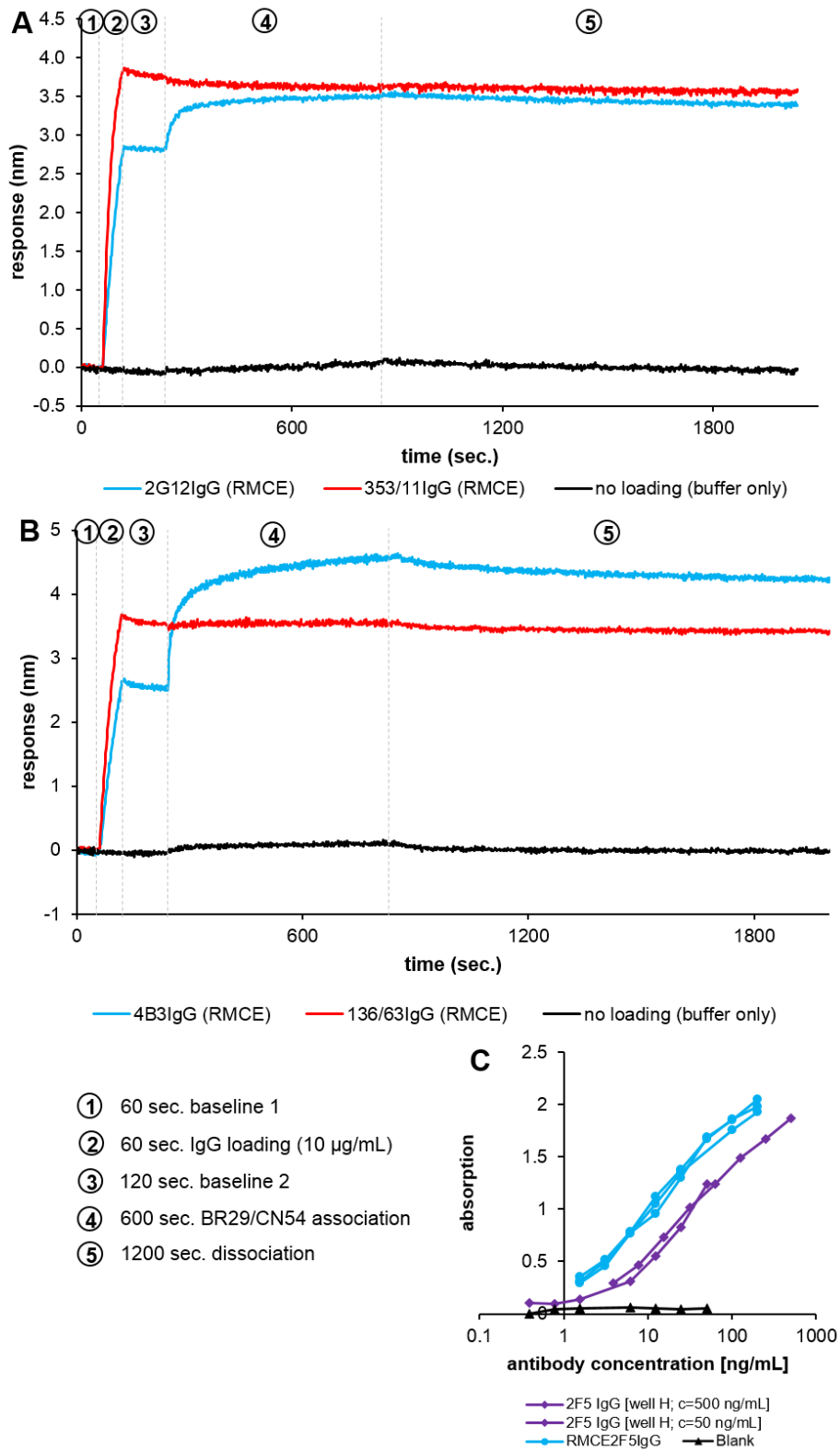


Figure S7: Evaluation of specificity of (A) 2G12 & 353/11 to BR29 (188  $\mu$ g/mL), (B) 4B3 & 136/63 to CN54 (368  $\mu$ g/mL) and (C) 2F5 IgG to UG37.

Table S1: Number of selected amino acids in variable region of mature and germline antibody variants. Histidine, proline and phenylalanine is increased in mature variants and tyrosine, serine and tryptophan is increased in germline variants as described in Clark *et al.*, 2006.

<b>V<sub>H</sub> &amp; V<sub>L</sub></b>	increased in mature variant			increased in germline variant		
	<b>H</b>	<b>P</b>	<b>F</b>	<b>Y</b>	<b>S</b>	<b>W</b>
2G12	4	8	9	10	26	5
353/11	0	7	8	16	40	6
Ustekinumab	0	11	8	14	33	7
554/12	0	10	7	18	37	7
4B3	4	11	7	11	30	4
136/63	2	10	5	19	39	5
2F5	3	17	10	8	28	4
236/14	1	12	8	14	30	5

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