# Supplementary protocol:

# Live imaging of mRNA using RNA-stabilized fluorogenic proteins

# Procedure Cell culture

- Culture U2OS cells in a T75 cell culture flask with culture media (DMEM supplemented with 10% fetal bovine serum, 100 U ml<sup>-1</sup> penicillin and 100 µg ml<sup>-1</sup> of streptomycin) at 37°C with 5% CO<sub>2</sub>.
- Prewarm culture media, 1xPBS, and TrypLE Express in a 37°C water bath.
- Aspirate culture media, gently rinse cells with 1xPBS, aspirate 1xPBS, then add 2 ml TrypLE Express to the culture flask, and incubate under 37°C for 5 to 10 minutes to detach cells from the flask.
- Add 8 ml of culture media to the culture flask, and gently pipette up and down to make sure all cells are fully detached from the flask.
- Transfer resuspended cells from the culture flask to a 15 ml tube, then centrifuge resuspended cells in the 15 ml tube at 300 g for 3 minutes.
- Aspirate supernatant, and resuspended cell pellet with fresh culture media, pipette up and down gently, to make sure cells are fully resuspended.
- Count the number of cells using a hemocytometer.
- To each 35 mm imaging dish (poly-D-lysine-coated), seed 2 x  $10^5$  cells in 2 ml of culture media. Gently shake the imaging dish to make sure cells are evenly spread. Keep the imaging dish at  $37^{\circ}$ C with 5% CO<sub>2</sub> overnight.

#### **Transfection**

- On the next day, transfect U2OS cells. We suggest to determine the optimal amount of plasmid encoding (mNeonGreen)<sub>4</sub>-tDeg in the transfection. An optimal amount of (mNeonGreen)<sub>4</sub>-tDeg plasmid will give highest mRNA fluorescent puncta signal-to-noise ratio possible in fluorescence imaging. In our setup, we use 1.4 µg of miniCMV-(mNeonGreen)<sub>4</sub>-tDeg or 0.28 µg of UbC-(mNeonGreen)<sub>4</sub>-tDeg with 1.12 µg of pUC19 (as a diluent DNA) as the fluorogenic protein in the transfection. The amount of plasmid encoding (mNeonGreen)<sub>4</sub>-tDeg should be titrated and optimized case by case.
- For transfection, prepare a 1.7 ml tube for each imaging dish. To this tube, add 125 μl of Opti-MEM, 1.4 μg of mRNA reporter plasmids, an optimal amount of plasmid encoding (mNeonGreen)<sub>4</sub>-tDeg, and 9.6 μl of FuGENE® HD reagent. Mix thoroughly by pipetting up and down, then incubate at room temperature for 10 to 15 minutes.
- After incubation, add this mixture drop by drop to the imaging dish with cells. Keep the imaging dish under 37°C with 5% CO<sub>2</sub> overnight.
- On the next day, prewarm 1xPBS and culture media.
- Aspirate culture media from the imaging dish, gently rinse cells with 1xPBS, aspirate 1xPBS, add 2 ml fresh culture media to cells. Culture cells under 37°C with 5% CO<sub>2</sub> overnight.

## **Imaging**

- Prior to live-cell imaging, prewarm 1xPBS, and imaging media (phenol red-free DMEM supplemented with 10% fetal bovine serum, 100 U ml<sup>-1</sup> penicillin and 100 µg ml<sup>-1</sup> of streptomycin, 1x GlutaMAX<sup>™</sup>, and 1 mM sodium pyruvate).
- Aspirate culture media from the imaging dish, gently rinse cells with 1xPBS, aspirate 1xPBS, add 2 ml of imaging media to the imaging dish.
- Move the imaging dish to the microscope in the prewarmed environmental chamber.

- Add sufficient immersion oil (N = 1.520) on a  $100 \times /1.4$ -NA oil objective, and use this objective under phase illumination to focus on adhered cells.
- Switch from phase to fluorescence acquisition with FITC filter sets (with excitation filter 475  $\pm$  14 nm, dichroic mirror with a reflection band of 481-502 nm, and a transmission band of 506-543 nm, and emission filter 525  $\pm$  25 nm).
- Determine the proper exposure time by acquiring images with an exposure time between 50 ms to 200 ms. A proper exposure time will allow for highest possible signal without any pixel saturation.
- Set up software to acquire fluorescence images.

# **Materials**

Reagents

| DMEM                                       | Thermo Fisher Scientific 11995-065 |
|--|------------------------------------|
| Fetal bovine serum                         | Corning 35-010-CV                  |
| Penicillin-Streptomycin                    | Thermo Fisher Scientific 15140122  |
| 1xPBS                                      | Thermo Fisher Scientific 10010023  |
| TrypLE™ Express Enzyme (1x), no phenol red | Thermo Fisher Scientific 12604013  |
| Opti-MEM                                   | Thermo Fisher Scientific 31985070  |
| FuGENE HD                                  | Promega 2311                       |
| Phenol red-free DMEM                       | Thermo Fisher Scientific 31053-028 |
| GlutaMAX™                                  | Thermo Fisher Scientific 35050-061 |
| Sodium Pyruvate (100 mM)                   | Thermo Fisher Scientific 11360-070 |
| Immersion oil (N = 1.520)                  | Applied Precision                  |

**Equipment** 

| <u> </u>                                     |                                  |
|--|----------------------------------|
| T75 cell culture flask                       | Corning 430641U                  |
| Precision™ shaking water bath                | Thermo Fisher Scientific         |
| 15 ml tube                                   | VWR 82050-276                    |
| 35 mm imaging dishes                         | Mattek Corporation P35GC-1.5-14C |
| 1.7 ml tube                                  | Thomas Scientific C2170          |
| Epifluorescence inverted microscope          | Olympus IX-70                    |
| Evolve® 512 EMCCD OEM camera                 | Photometrics                     |
| Insight SSI 7 color solid state illumination | Applied Precision                |
| system                                       |                                  |