

Table S1. sGC β 1 mutants, expression vectors, and DNA primers used in this study.

	Vector Mutant	pCMV5, pET20b (rat sGC, rat TC-sGC)	pMAL (bovine sGC)
PAS domain mutants	Δ 284-301	CTTTGTA CTGAGAAGCAAGGAAGGGAGCTGC CTCCGTCTCAAAGGCCAAA	GAGAAGCAAGGAAGGGAGCTGCTTACGGCTCA
	Δ 265-271	CTCTCTGGTCCGCCCTCATATTGACATCAGTAT CAATACCGTCTTTGTA CTGAGAAGCAAGG	CGTCCTCATATTGACATCAGTATCAATACAGTCTTC GTGTTG
	R335S/R336S/D342S	CTTGATGACCTAACAAGCAGCGGCCTGTACC TGAGTAGCATCCCTCTCCATGATG	CCTGGACGACCTGACCAGCAGCGGGCTGTACCTGA GTAGCATCCCTCTGCACGATG
	L340D/I343D/L345D	CCTAACAAGAAGAGGGCCTGTACGATAGTGACG ACCCTGACCATGATGCTACACGAGACCTG	ACCCGGCGAGGGCTGTACGATAGTGACGACCCTG ATCACGATGCCACTCGGGAC
	H266A/H271A/I272D	CCTCATATTGACATCAGTTTCGCCGGGATTCTT TCAGCCGACAATACCGTCTTTGTA CTGAGAAG	CCTCATATTGACATCAGTTTCGCTGGAATCCTCTCA GCCGACAATACAGTCTTCGTGTTGAGAAG
	L269D/I272S/V275D	TCAGTTTCCACGGGATTGATTCACACAGCAATA CCGACTTTGTA CTGAGAAGCA	TCAGTTTCCATGGAATCGACTCACACAGCAATACA GACTTCGTGTTGAGAAGCA
	E291A/E295A/L296D	CTGGATGTTGAGAACTTGCATGTGAGGATGC AGATACTGGGGCAGAGATTAGCTG	GGATGTAGAGAAATCAGCGTGTGAGGATGCGGAT ACGGGCACTGAAATCAGCTGC
	L333D/L338D/L340D	AGTGTGATGAACTTGGATGACGATACAAGAA GAGGCGATTACGATAGTGACATCCCTCTCCAT GATGCTAC	ATGAACCTGGACGACGATACCCGGCGAGGGGATT ACGATAGTGACATCCCTCTGCACG
Heme Binding mutants	H105F	CCTCGACGCCCTGTTCGACCACCTCGCC	
	Y135A R139A	GGGCTCATTCTGCACGCCTACTCGGAAGCAGA GGGGCTTCAGGA	

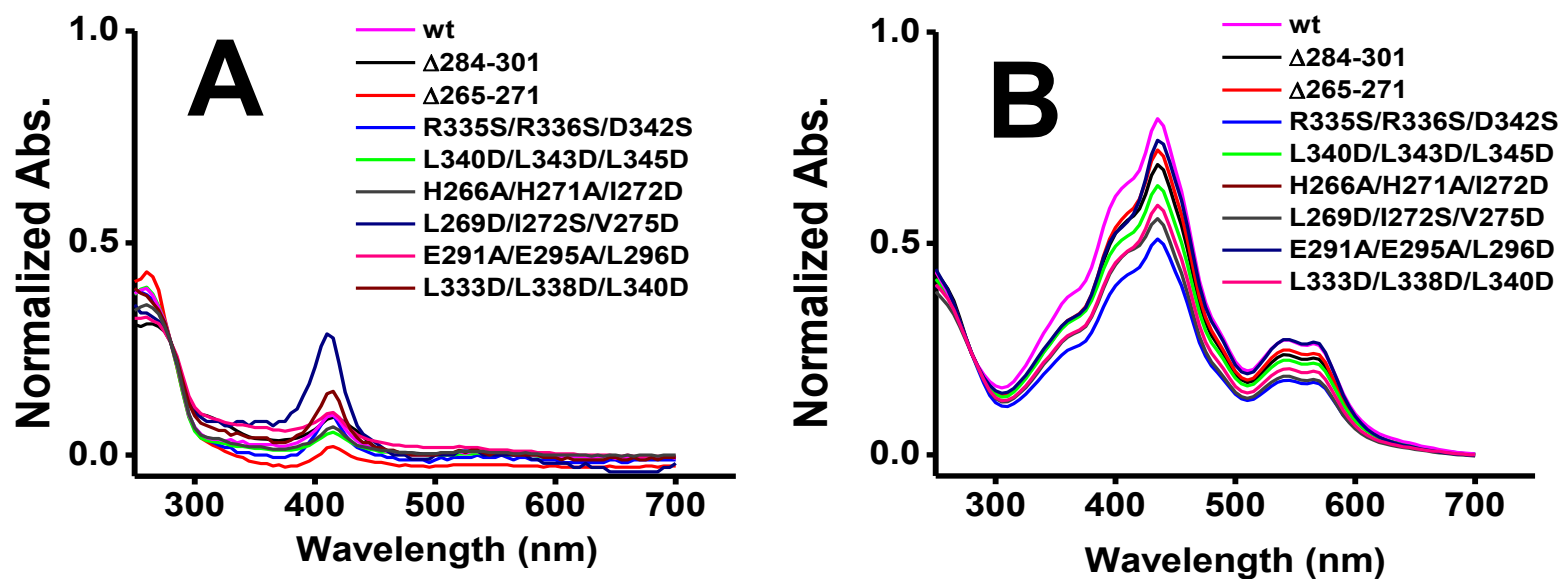


Fig. S1. *UV-Vis spectrum of purified recombinant sGC β (1-358) proteins.* Spectra are normalized to the absorption values at 280 nm. *Panel A*, spectra of proteins after purification. *Panel B*, spectrum of proteins after undergoing heme reconstitution.

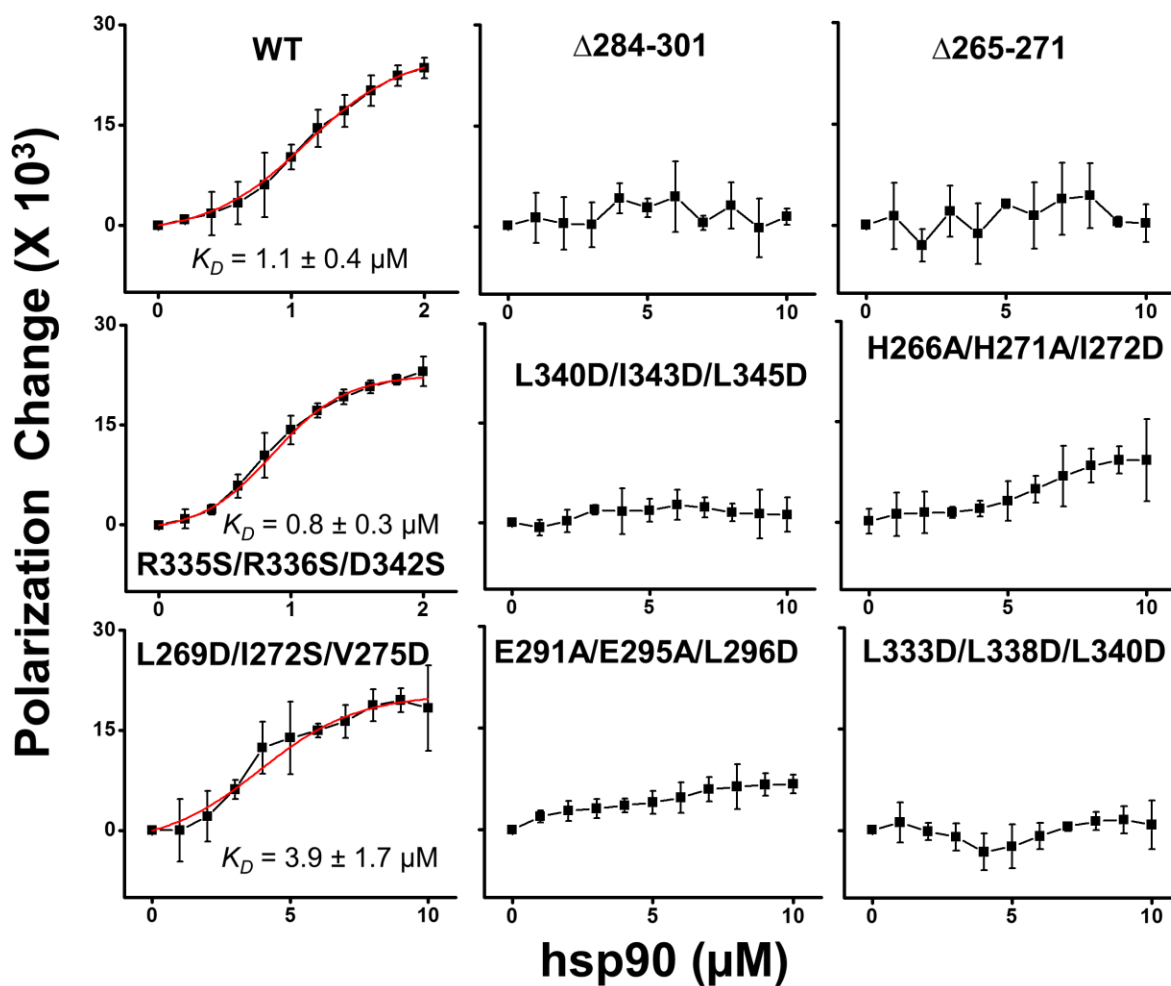


Fig. S2. Binding interactions of purified apo-sGC β 1(1-358) mutant proteins with FITC labeled hsp90. Each indicated FITC-labeled hsp90 (0.5 μM) was titrated with increasing concentrations of sGC β proteins and the residual polarization change was recorded and plotted. The points depict mean \pm S.D. for three samples, and are representative of two independent experiments. Red lines are computer fits derived using a single site binding equation, and gave the indicated estimates for k_d .

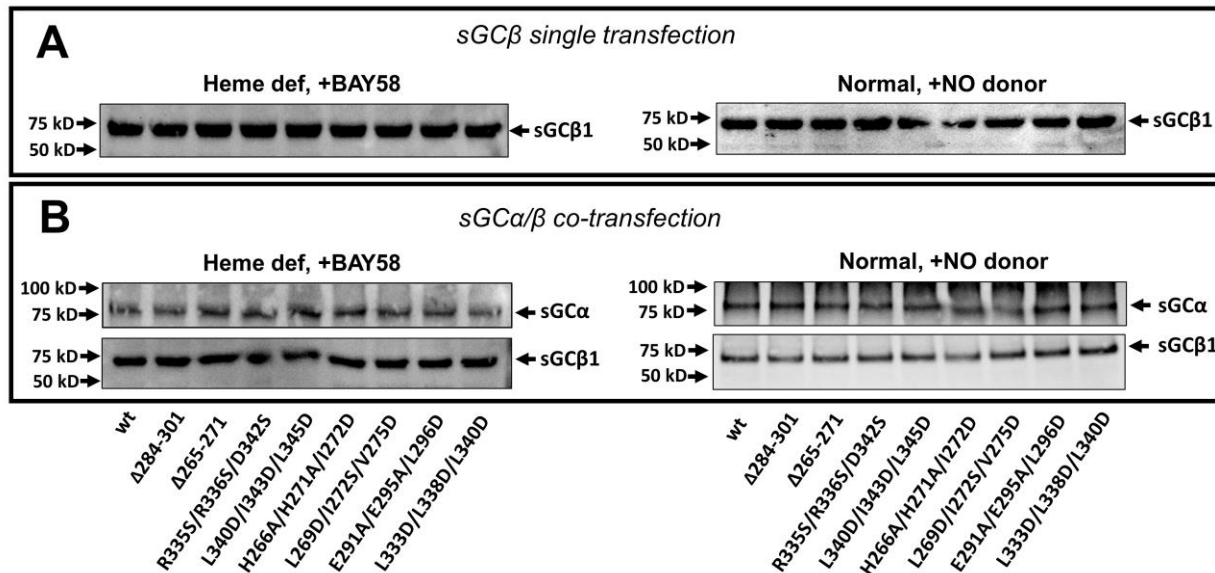


Fig. S3. Expression levels of *sGCβ*1(1–619) proteins in cell supernatants used for guanylyl cyclase activity quantification. Panel A shows a representative Western blot indicating expression levels of the *sGCβ* proteins in COS-7 cell that were only transfected with *sGCβ* constructs. Panel B shows a representative Western blot indicating expression levels of *sGCβ* proteins and *sGCα* in COS-7 cell that transfected with both *sGCα* and *sGCβ* constructs. The analysis used the same supernatants that were used in Fig. 5 A and B.

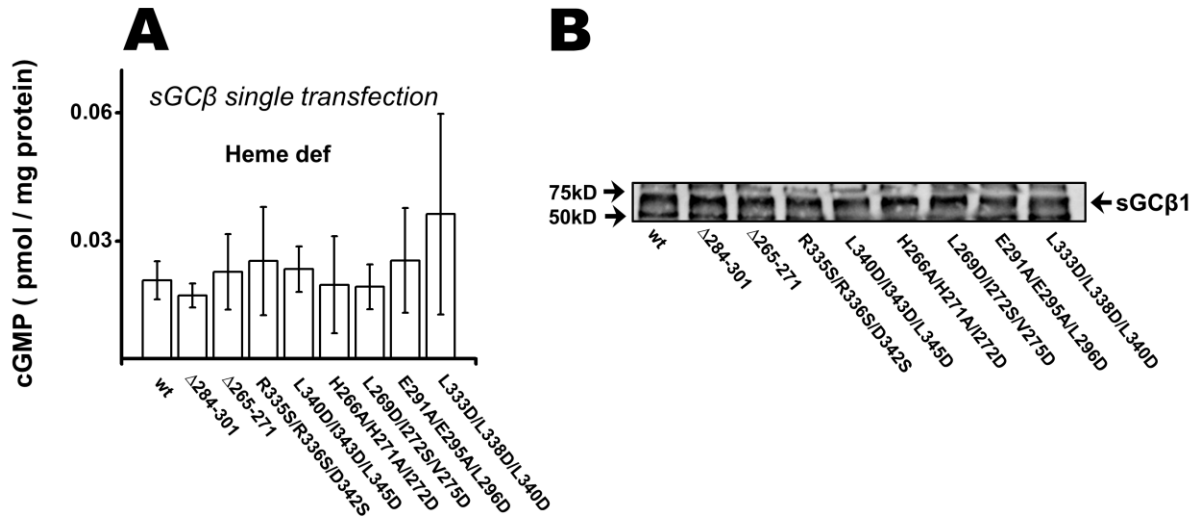


Fig. S4. NO-dependent guanylyl cyclase activity of the *sGCβ1*(1-619)-containing supernatants from heme-depleted cells. COS-7 cells were cultured in heme-depleted conditions (heme def) and transfected to express each *sGCβ1*(1-619) protein. Their supernatants were collected and used for GTP cyclase activity measurements in response to the NO donor NOC18. *Panel A* compares the activities. Values are the mean \pm SD of three measurements, and representative of two experiments each. *Panel B* shows a representative Western blot comparing the expression levels of the *sGCβ* proteins in the supernatants (equal protein loaded).

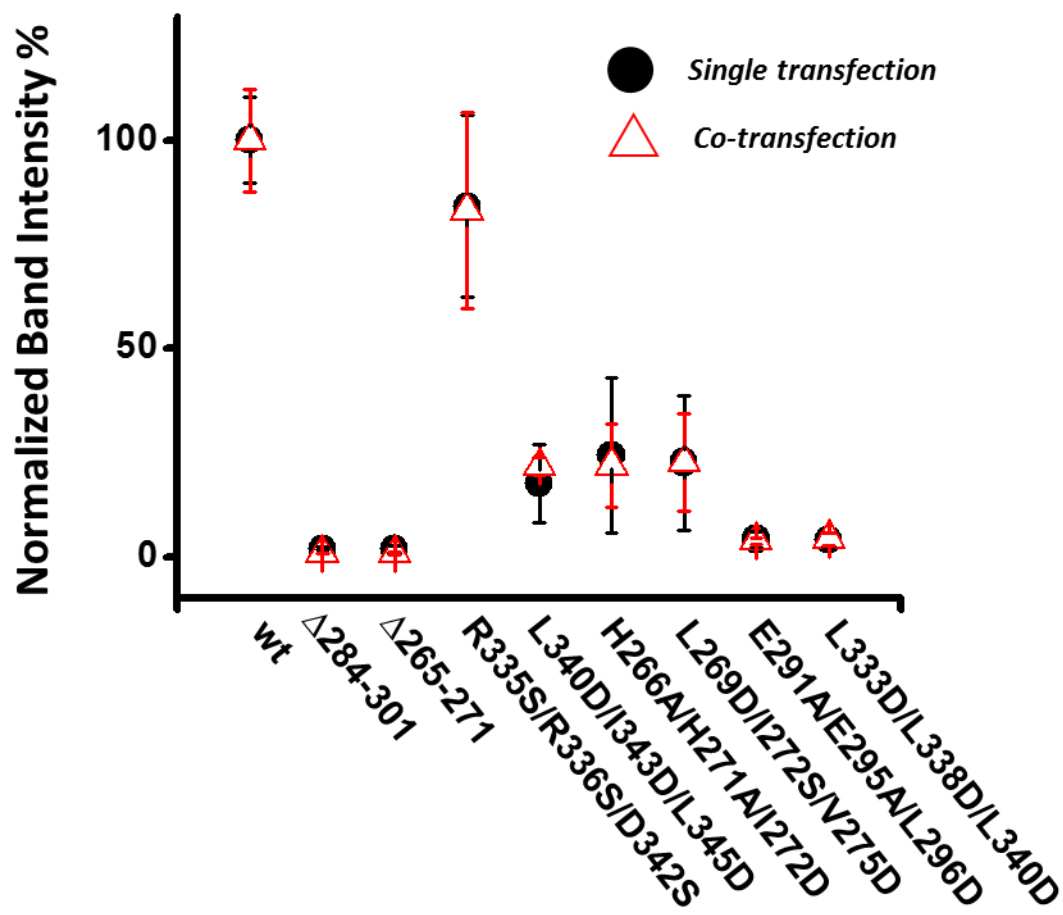


Fig. S5. Effect of sGC α co-expression on the hsp90 association levels of the sGC β 1(1-619) proteins expressed in heme-depleted cells. The figure plots the normalized hsp90 band intensity levels that are reported in Figs. 3 and 6. Band intensities were normalized relative to the hsp90 band intensity for wild type sGC β 1(1-619) under the two different culture conditions.