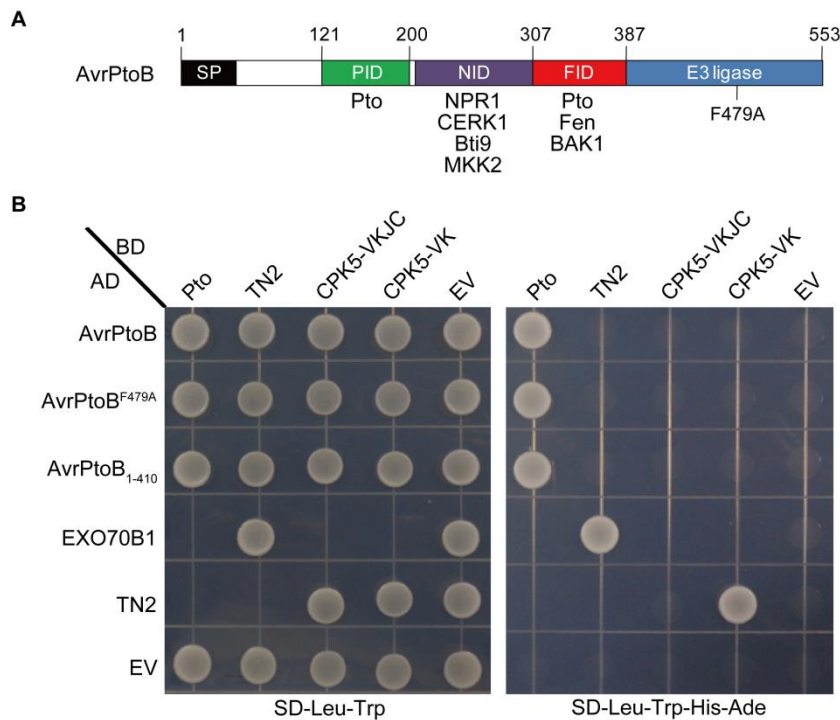
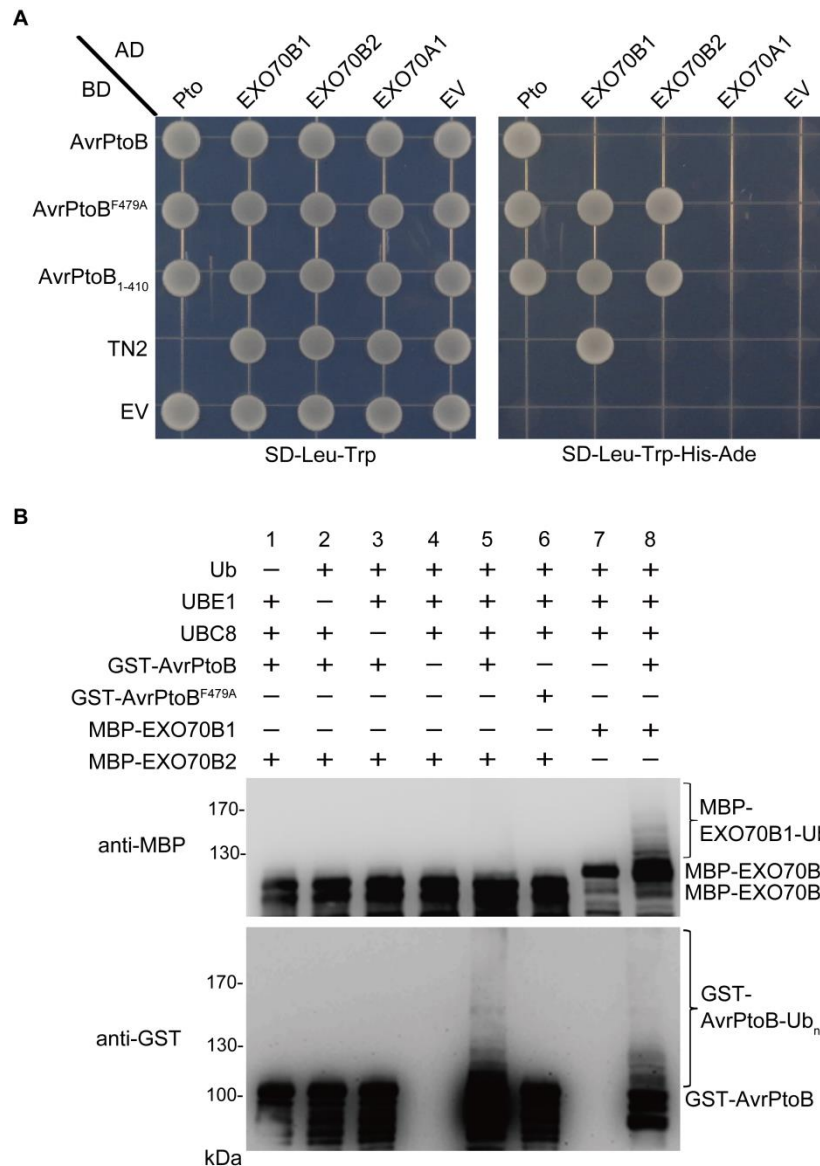


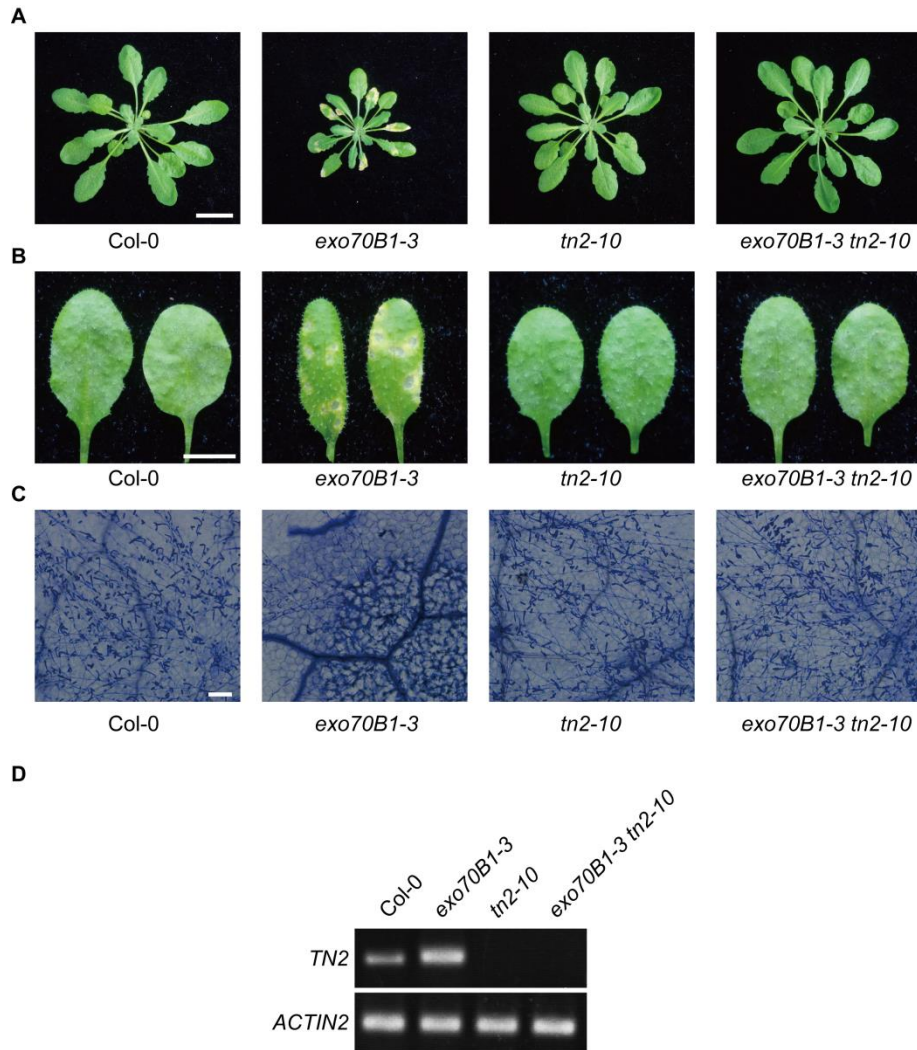
## Supplementary Figures



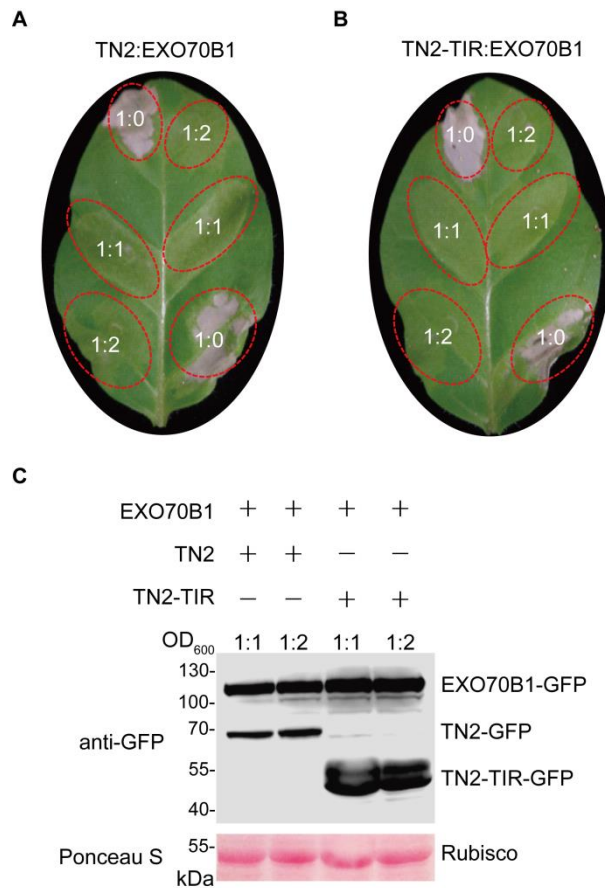
**Figure S1. No interaction was observed between AvrPtoB and TN2 or CPK5 in Y2H assays.** (A) A schematic diagram of the AvrPtoB. Each box indicates the specific domain of AvrPtoB, including the predicted signal peptide (SP), Pto-interacting domain (PID), NPR1-interacting domain (NID), Fen-interacting domain (FID) and E3 ubiquitin ligase region. F479A indicated the point mutation of AvrPtoB in E3 ligase domain. (B) AvrPtoB, AvrPtoB<sup>F479A</sup>, and AvrPtoB<sub>1-410</sub> were fused to the AD, and TN2, CPK5-VKJC and CPK5-VK were fused to the BD. Different pairs of constructs were cotransformed into AH109. Yeast cells containing the indicated plasmids were spotted onto SD-Leu-Trp or SD-Ade-His-Leu-Trp media. Photographs were taken 5 days after incubation. No yeast cells were dropped on the blank areas of SD-Leu-Trp media and these same areas on SD-Ade-His-Leu-Trp media. This experiment was repeated three times with similar results.



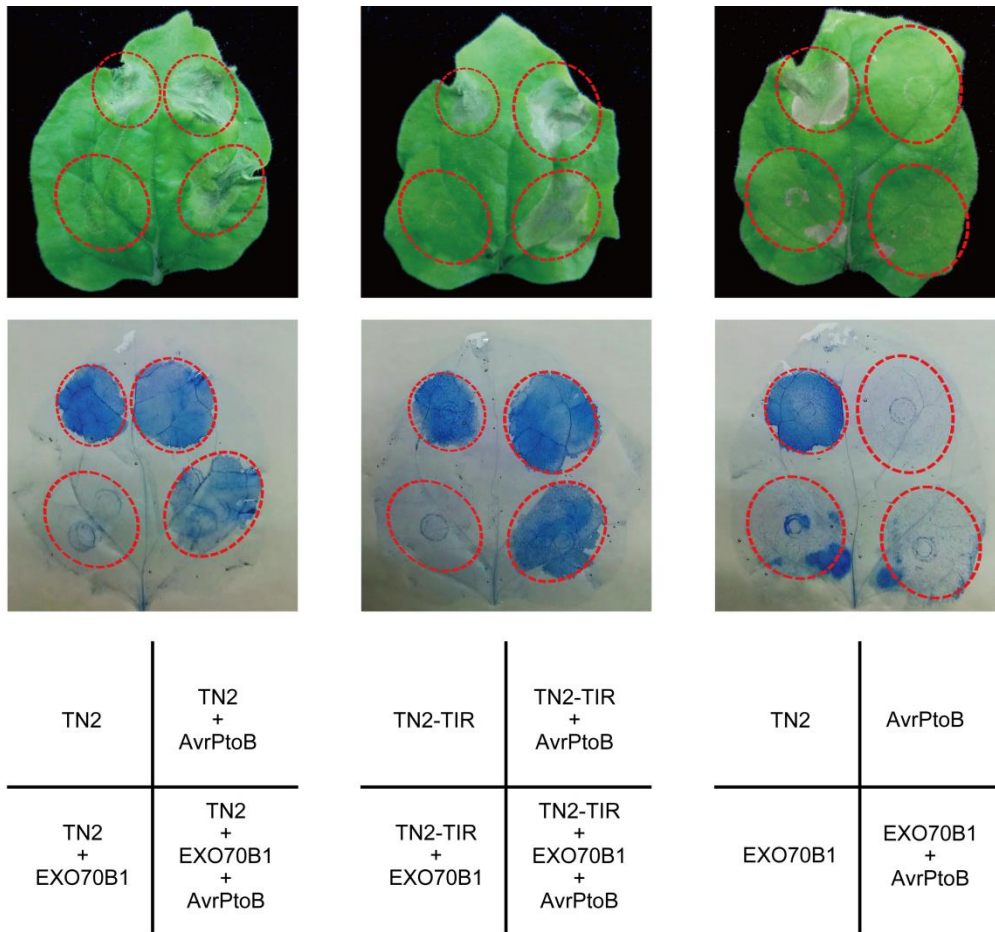
**Figure S2. EXO70B2 is not a substrate of AvrPtoB.** (A) EXO70B1, EXO70B2 and EXO70A1 were fused to the AD, and AvrPtoB, AvrPtoB<sup>F479A</sup>, and AvrPtoB<sub>1-410</sub> were fused to the BD. Different pairs of constructs were cotransformed into AH109. Yeast cells containing the indicated plasmids were spotted onto SD-Leu-Trp or SD-Ade-His-Leu-Trp media. Photographs were taken 5 days after incubation. No yeast cells were dropped on the blank areas of SD-Leu-Trp media and these same areas on SD-Ade-His-Leu-Trp media. (B) *In vitro* ubiquitination assay. Recombinant MBP-EXO70B2 incubated in ubiquitination buffer in the presence or absence of ubiquitin, E1 (UBE1), E2 (UBC8) or E3 (AvrPtoB or AvrPtoB<sup>F479A</sup>). MBP-EXO70B1 was used for a positive control. The reaction mixtures were subjected to immunoblot analysis with anti-MBP and anti-GST antibodies. Ubiquitinated MBP-EXO70B1 was detected by anti-MBP antibody. These experiments were repeated three times with similar results.



**Figure S3. The *tn2-10* mutation suppresses *exo70B1-3*-mediated cell death and resistance to *G. cichoracearum*.** (A) Five-week-old plants were photographed under short-day conditions. The *exo70B1-3* mutants displayed hypersensitive responses, but no cell death was observed in Col-0, *tn2-10* and *exo70B1-3 tn2-10* mutants. Bar = 1.0 cm. (B) Four-week-old plants were infected with *G. cichoracearum*. Representative leaves were detached and photographed at 8 days post infection. The *exo70B1-3* mutant was more resistant to *G. cichoracearum*, while the *tn2-10* and *exo70B1-3 tn2-10* mutants showed wild-type like responses to *G. cichoracearum*. Bar = 0.5 cm. (C) Leaves at 8 days post infection were stained with trypan blue to observe fungal structures and dead cells. Few spores were produced in the *exo70B1-3* mutant, whereas many fungal spores were produced in Col-0, *tn2-10*, and *exo70B1-3 tn2-10* plants. Bar = 100 μm. (D) The transcript level of *TN2* was examined by RT-qPCR. Total RNA was isolated from the indicated five-week-old plants shown in (A). The transcripts of *TN2* were up-regulated in *exo70B1-3* mutants and were not detected in *tn2-10* and *exo70B1-3 tn2-10* plants. These experiments were repeated three times with similar results.



**Figure S4. EXO70B1 suppresses TN2- or TN2-TIR-triggered cell death in *N. tabacum*.** (A) and (B) TN2- or TN2-TIR-triggered cell death was suppressed by EXO70B1 in *N. tabacum*. *Agrobacterium* GV3101 cells carrying different constructs were mixed prior to adding infiltration buffer. The concentration of agrobacteria expressing *TN2* or *TN2-TIR* was brought to  $OD_{600} = 0.5$  and the concentration of agrobacteria expressing *EXO70B1* was brought to  $OD_{600} = 0, 0.5,$  or  $1.0$ . Five-week-old *N. tabacum* leaves were used for transient expression. Cell death was observed 36 to 48 h later. Pictures were taken 4 days post infiltration. Dashed lines mark the infiltrated areas. (C) Accumulation of the indicated recombinant proteins from the experiments in (A), which failed to trigger a HR. Proteins were detected by anti-GFP antibody and equal loading is shown by Rubisco. These experiments were repeated three times with similar results.



**Figure S5. AvrPtoB rescues TN2- or TN2-TIR-triggered cell death, which is suppressed by EXO70B1, in *N. benthamiana*.** *Agrobacterium* GV3101 cells carrying different constructs were infiltrated into five-week-old *N. benthamiana* leaves. Pictures were taken 4 days post infiltration (upper panel). Leaves at 4 days post infiltration were stained with trypan blue to visualize cell death (lower panel). Dashed lines mark the infiltrated areas. These experiments were repeated three times with similar results.

**Supplementary Table 1. Primers used in this study.**

<b>Primer name</b>	<b>Purpose</b>	<b>Sequence (5'-3')</b>
AvrPto PF	Yeast two-hybrid	GGAATTCCATATGATGGGAAATATA TGTGTCGG
AvrPto PR	Yeast two-hybrid	CGGAATTCTCATTGCCAGTTACGGT ACGG
AvrPtoB PF	Yeast two-hybrid	GGAATTCCATATGATGGCGGGTATC AATAGAGC
AvrPtoB PR	Yeast two-hybrid	CGGAATTCTCAGGGGACTATTCTAA AAGCATAC
AvrPtoB 1-410 PR	Yeast two-hybrid	CGGAATTCTCACCCCGGGTTCAGGT TAAG
AvrPtoB 1-205 PR	Yeast two-hybrid	CGGGATCCTCATGACGCCGCCTGTT GGT
AvrPtoB 1-307 PR	Yeast two-hybrid	CGGGATCCTCATAACATGTCTTTCAA GGGCC
AvrPtoB 1-387 PR	Yeast two-hybrid	CGGGATCCTCACACCCGCAATCGTG TTG
AvrPtoB 308-533 PF	Yeast two-hybrid	GGAATTCCATATGATGCAGCGCCTC CCTATC
AvrPtoB 308-533 PR	Yeast two-hybrid	CGGGATCCTCAGGGGACTATTCTAA AAGCATAC
Pto PF	Yeast two-hybrid	GGAATTCCATATGATGGGAAGCAA GTATTCTAAGG
Pto PR	Yeast two-hybrid	CGGAATTCTTAAATAACAGACTCTT GGAGACGAA
EXO70A1 PF	Yeast two-hybrid	CGGAATTCATGGCTGTTGATAGCAG AATG
EXO70A1 PR	Yeast two-hybrid	AACTGCAGTTACCGGCGTGGTTCAT T
EXO70B2 PF	Yeast two-hybrid	CGGAATTCATGGCTGAAGCCGGTGA C
EXO70B2 PR	Yeast two-hybrid	AACTGCAGTCAACTTGAGCTTTCCT TGA
avrPtoB M PF	Site-directed mutagenesis	TGACGCACTGTCTTGCTGGCGGAGA ATTGT
avrPtoB M PR	Site-directed mutagenesis	ACAATTCTCCGCCAGCAAGACAGTG CGTCA
AvrPtoB PF	To clone into pGEX-4T-1	CGGAATTCATGGCGGGTATCAATAG AGC

AvrPtoB PR	To clone into pGEX-4T-1	TTGCGGCCGCGGGGACTATTCTAAA AGCATAAC
AvrPtoB 1-410 PR	To clone into pGEX-4T-1	TTGCGGCCGCTCACCCCGGGTTCAG GTAAAG
EXO70B1 PF	To clone into PUC19	CGAGCTCATGGCGGAGAATGGTGA A
EXO70B1 PR	To clone into PUC19	GCGTCGACTTTTCTTCCCCTGGTAG
AvrPtoB PF	To clone into pSuper1300-MYC	GCGTCGACATGGCGGGTATCAATAG AGC
AvrPtoB PR	To clone into pSuper1300-MYC	GGGGTACCGGGGACTATTCTAAAA GCATAAC
AvrPtoB 1-410	To clone into pSuper1300-MYC	GGGGTACCCCCCGGGTTCAGGTAA G
EXO70B1 PF	To clone into pSuper1300-GFP	GCGTCGACATGGCGGAGAATGGTG AA
EXO70B1 PR	To clone into pSuper1300-GFP	GGGGTACCTTTTCTTCCCCTGGTAG
LBb1.3	Genotyping	ATTTTGCCGATTTTCGGAAC
TN2 Salk PF	Genotyping	TTCCAACAAAATCACCAGCTC
TN2 Salk PR	Genotyping	AAATCCCATCTGGGATTTGTC
TN2 PF	Real-time PCR	GGCTCATGAGTCAGAAAG
TN2 PR	Real-time PCR	GAAGATTCAGTCCCGGAT
PR1 PF	Real-time PCR	TTCACAACCAGGCACGAGGAG
PR1 PR	Real-time PCR	CTAACCCACATGTTACGGCG
PR2 PF	Real-time PCR	GAATCAAGGAGCTTAGCCTCACC
PR2 PR	Real-time PCR	GTAGAGCCGCATTCGCTGGAT
PAD4 PF	Real-time PCR	CTTTCTTCAGTTAAAGATCAAGGAA GG
PAD4 PR	Real-time PCR	GGCAGAAGTTGTGTGCTAAACG
NPR1 PF	Real-time PCR	TAGAGTTGCACTTGCTCAACGTC
NPR1 PR	Real-time PCR	GTTTCCCGAGTTCCACGGT
EDS5 PF	Real-time PCR	GAACTCGCTGCTCTTGACC
EDS5 PR	Real-time PCR	CAGCCCAAGGACCGAATAATC
ACTIN2 PF	Real-time PCR	TCTCCCGCTATGTATGTCGCC
ACTIN2 PR	Real-time PCR	GTCACGTCCAGCAAGGTCAAGA
EXO70B1 PF	Real-time PCR	GAAACCTATCGGCATCTGTC
EXO70B1 PR	Real-time PCR	CCGAAGGGAGAGCATCAAT