С Sh-Con Α В Sh-pHIV7-GFP Sh-PHF2-I H3Kme2 levels(A.U.) 3 1.5 Sh-PHF2-II PHF2/Tubulin 1.0 2 0.5 1 H3KAmel 0.0 ShCon HF21 ShPHF21 0 H3K9mel H3K27mel D n.s. n.s. n.s. Sh-Con Sh-PHF2 250 Distance moved(m) 800 20 Number of entries Time centers(s) 200 15 0 600 150 10 400 00 100 5 200 a 50 Sh-Con Sh-PHF2 0 0 Sh-PHF2 sh-Cor Sh-Con PHF2



Figure EV1.

Expanded View Figures

Figure EV1. Mice lacking PHF2 show no difference in basal behavior tests.

- A Stereotaxic injection of GFP-expressing lentiviral particles into bilateral hippocampus CA1 in WT mice. Scale bar, 200 μm.
- B Quantification of the immunoblot analysis. PHF2 protein levels of hippocampal tissues in sh-PHF2 (I, II) mice were normalized against tubulin and quantified as fold change relative to that seen in sh-Con mice.
- C The H3Kme2 protein levels in sh-PHF2 (I, II) mice were normalized against H3 and quantified as fold change relative to that seen in sh-Con mice.
- D Open field test shows that basal anxiety and locomotor activity in sh-PHF2 mice were not altered in comparison with sh-Con mice. Path traces of single-trial open field tests were recorded for all sh-Con and sh-PHF2 mice (left). Distance moved, time in center, and number of entries were scored (right).
- E $\;$ Rotarod test showed no differences in motor activity for sh-Con and sh-PHF2 mice.

Data information: In (B, C), data are presented as the mean \pm SD (n = 6). *P < 0.05 (unpaired, two-sided Student's t-test). In (D, E), data are presented as the mean values \pm SEM (n = 8). Data were analyzed using unpaired, two-sided Student's t-test.

Figure EV2. PHF2 t/g mice show no difference in basal behavior tests.

- A Quantification of the immunoblot analysis. PHF2 protein levels of PHF2 t/g mice were normalized against tubulin and quantified as fold change relative to that seen in WT littermate mice.
- B Immunofluorescence of hippocampal coronal sections of WT littermate and Flag-PHF2 t/g mice stained with Flag (green), MAP2 (red), and DAPI (blue). Ectopic expression of Flag-PHF2 in the CA1 region of the hippocampus of PHF2 t/g compared to WT littermate mice. Scale bar, 200 μm.
- C H3Kme2 protein levels in PHF2 t/g mice were normalized against H3 and quantified as fold change relative to levels in WT mice.
- D Path traces of single-trial open field tests for WT littermate and PHF2 t/g mice (left). No differences in distance moved, time spent in the center, or number of entries to the center area were observed between the two groups (right).
- E No difference in rotarod motor activity was observed between WT littermate and PHF2 t/g mice.
- F No difference in swimming ability was observed between WT littermate and PHF2 t/g mice.

Data information: In (A, C), data are presented as the mean \pm SD (n = 6). * P < 0.05 (unpaired, two-sided Student's t-test). In (D–F), data are presented as the mean values \pm SEM (n = 9). Data were analyzed using unpaired, two-sided Student's t-test.



Figure EV2.

Figure EV3. BDNF-HFS mimics the protein synthesis-dependent enhancement of LTP observed in PHF2 t/g mice.

- A Input–output of the SC-CA1 synapse of sh-Con or sh-PHF2 expressing mice is represented by fiber volley and fEPSP slope, respectively. Each point represents a mean for a narrow range of fiber volley amplitudes. Representative raw traces are shown for each fiber volley amplitude. Scale bar, 1 mV/5 ms.
- B Relationship of PPR of the SC-CA1 synapses in the hippocampus of sh-Con (white) and sh-PHF2 (blue) mice. Example raw traces for PPR are shown for 60-ms intervals. Scale bar: 1 mV/20 ms.
- C SC-CA1 synapses of WT and PHF2 t/g mice show no differences in input—output relationship. Representative raw traces are shown for each fiber volley amplitude. Scale bar, 1 mV/5 ms.
- D PPR is shown for PHF2 t/g mice in comparison with WT mice. Example raw traces for PPR are shown for 60-ms intervals. Scale bar: 1 mV/20 ms.
- E WT hippocampal slices were pre-incubated with BDNF (20 ng/ml) for > 1 h before HFS induction. BDNF-HFS-induced LTP enhancement was blocked by simultaneous application of CHX (60 μM). Example traces of baseline (1: black) and 40 min after LTP induction (2: red) are shown for BDNF-treated and BDNF + CHX-treated WT hippocampal slices (Middle). Scale bar, 1 mV/5 ms. Comparison of LTP at 40 min post-HFS induction (right).

Data information: In (A–E), data are presented as the mean values \pm SEM (sh-Con, n = 8; sh-PHF2, n = 6; WT, n = 11; PHF2 t/g, n = 12, BDNF on WT slices, n = 8; BDNF+CHX on WT slices, n = 6). **P < 0.01 (unpaired, two-sided Student's t-test).



Figure EV3.



Figure EV4. Transient ectopic expression of PHF2 facilitates long-term potentiation (LTP) in the hippocampal Schaffer collateral-CA1 synapses.

- A Field excitatory postsynaptic potentials (fEPSPs) induced by a single 1-s training at 100 Hz HFS. LTP was significantly greater in slices derived from transiently Flag-PHF2 expressing mice compared to those from control mice (P < 0.05). Measurements correspond to the time points indicated on the time course graph in this figure. Error bars show the standard error of the mean (**P = 0.0055, 2-way ANOVA). Scale bar, 0.25 mV/5 ms.
- B LTP of control mice was 111 \pm 4.11% of baseline during the 10 min before and after time point 2 (n = 5 slices from 3 mice). In Flag-PHF2 overexpressing mice, fEPSPs were 120.5 \pm 2.75% of baseline during the 10 min before and after time point 2 (n = 9 slices from 4 mice). Error bars show the standard error of the mean (*P = 0.0465, 2-way ANOVA).

Figure EV5. Basal synaptic transmission of NMDA- and AMPA-mediated EPSCs do not differ by PHF2 expression levels.

A, B Recording of NMDAR- (A) and AMPAR-mediated (B) PPR at 50, 100, 150, and 200 ms intervals. Insets indicate raw trace of PPR (Top). Scale bars, 100 pA/150 ms. C, D Recording of NMDAR- (C) and AMPAR-mediated (D) PPR at 50, 100, 150, and 200 ms intervals. Insets indicate raw trace of PPR (Top). Scale bars, 100 pA/150 ms. Data information: In (A-D), data are presented as mean values \pm SEM (sh-Con, n = 14; sh-PHF2, n = 13; wild type, n = 16; PHF2 t/g, n = 26).



Figure EV5.