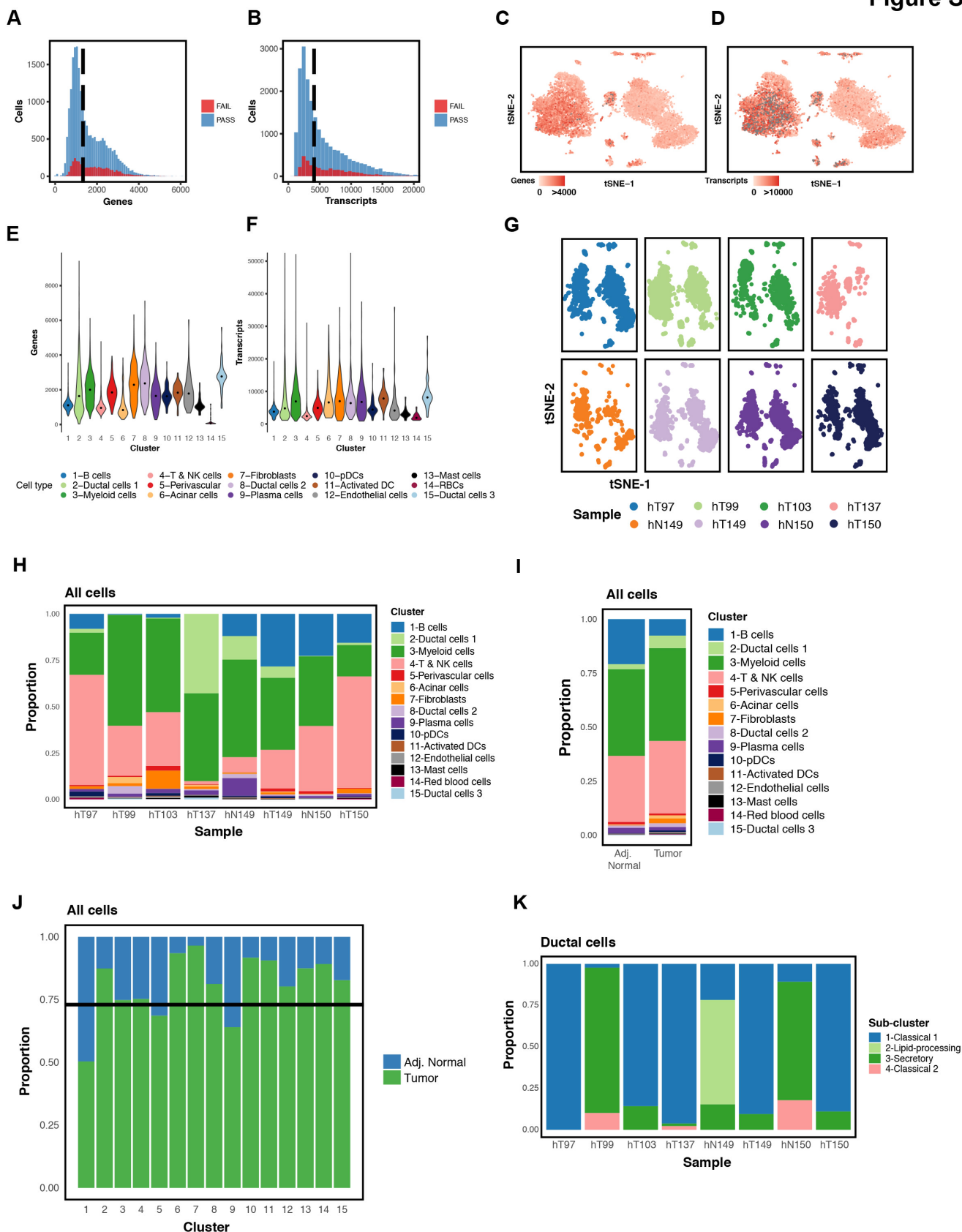
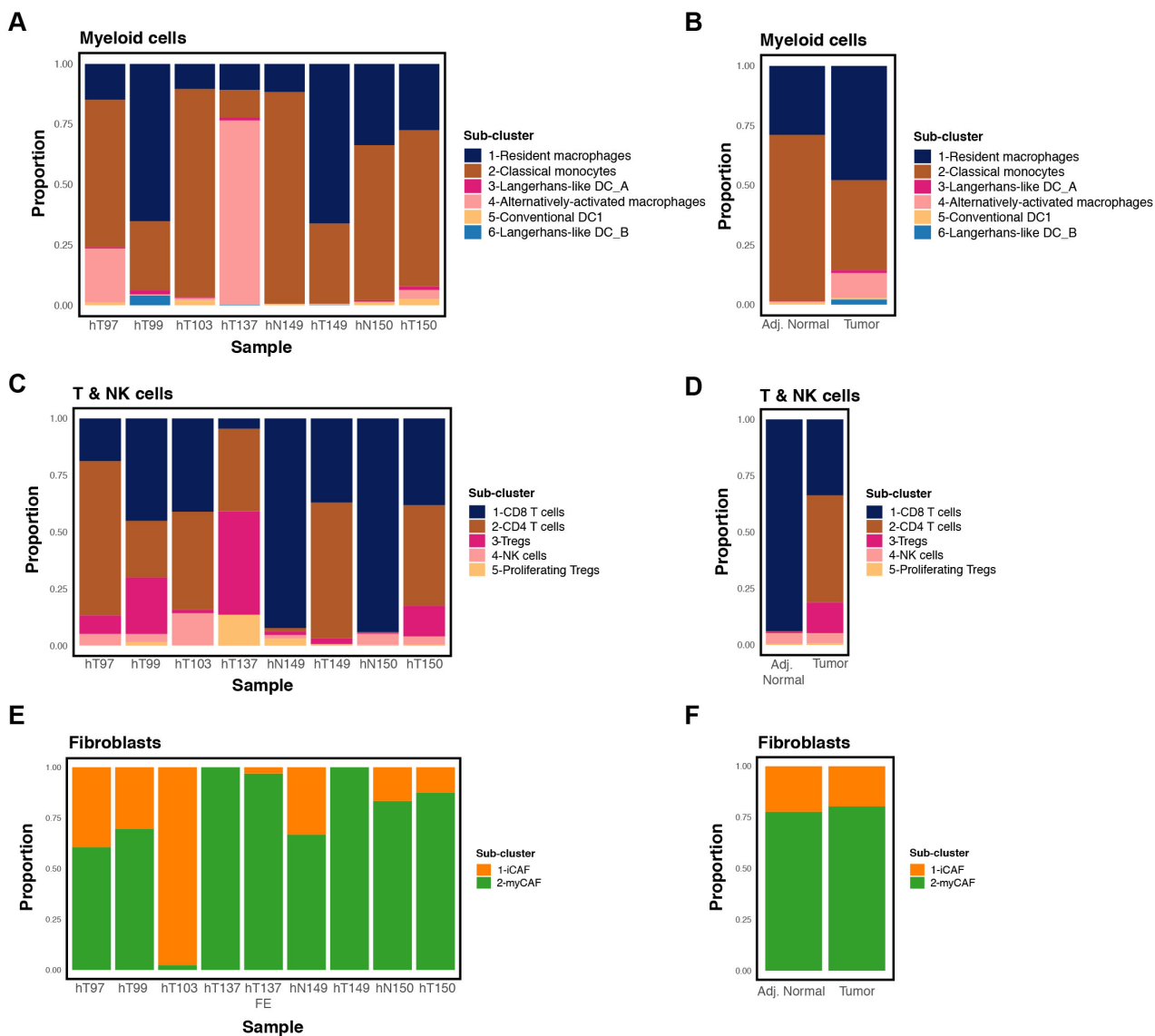


Figure S1



Supplementary Figure S1: Data quality and clustering metrics of the human PDAC single cell transcriptomes. A-B. Histograms showing the number of genes (**A**) and transcripts (**B**) that were detected per cell in the human PDAC dataset. Blue histogram shows cells that passed quality control (QC) and red histogram shows cells that failed QC. The dotted line represents the median value, which is 1,279 genes/cell and 3,938 transcripts/cell. **C-D.** Number of genes (**C**) and transcripts (**D**) detected per cell, represented as a t-SNE plot. Legend shows a color gradient corresponding to genes/transcripts number per cell. Gray color represents cells with greater than 4,000 genes or 10,000 transcripts, respectively. **E-F.** Distribution of genes (**E**) and transcripts (**F**) detected in each cluster in the human PDAC dataset. **G-H.** Patient contribution to each cluster, represented as separate t-SNE plots (**G**) and as bar plot (**H**) showing proportions of the different clusters in each sample. **I-J.** Bar plots comparing proportions of the different clusters between adjacent-normal and tumor samples, shown as clusters in each sample type (**I**) or sample types in each cluster (**J**). The horizontal black line in **J** represents the input contribution of adjacent-normal or tumor tissues into the dataset. **K.** Bar plot showing proportions of the different ductal cell sub-clusters in each human sample.

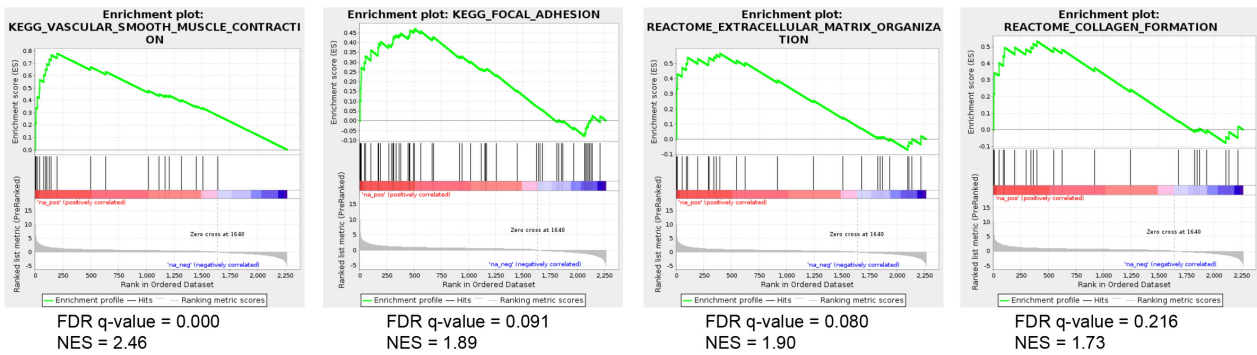
Figure S2



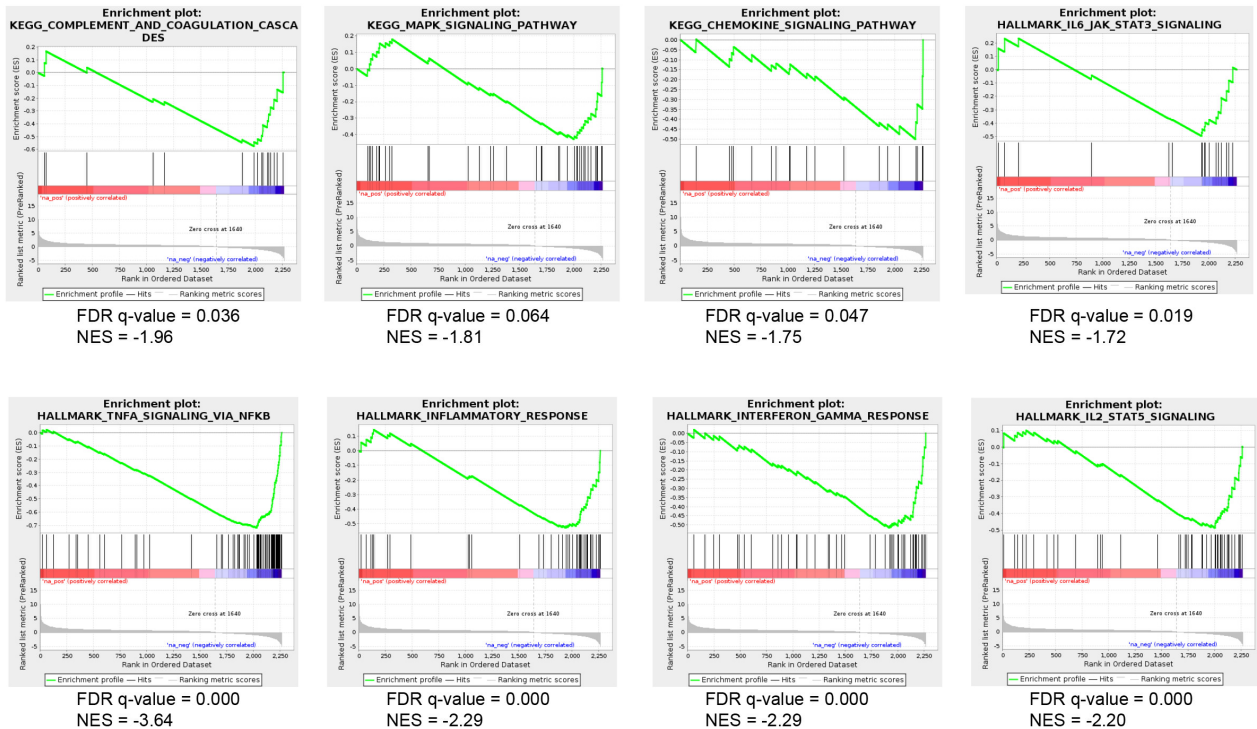
Supplementary Figure S2: Distribution of sub-clusters of cells throughout the human dataset. **A.** Bar plot showing proportions of the different myeloid cell sub-clusters in each human sample. **B.** Bar plot comparing proportions of the different myeloid cell sub-clusters between adjacent-normal and tumor samples. **C.** Bar plot showing proportions of the different T & NK cell sub-clusters in each human sample. **D.** Bar plot comparing proportions of the different T & NK cell sub-clusters between adjacent-normal and tumor samples. **E.** Bar plot showing proportions of the different fibroblast sub-clusters in each human sample. FE, fibroblast-enriched population. **F.** Bar plot comparing proportions of the different fibroblast sub-clusters between adjacent-normal and tumor samples.

Figure S3

A Pathways upregulated in human myCAFs vs. iCAFs

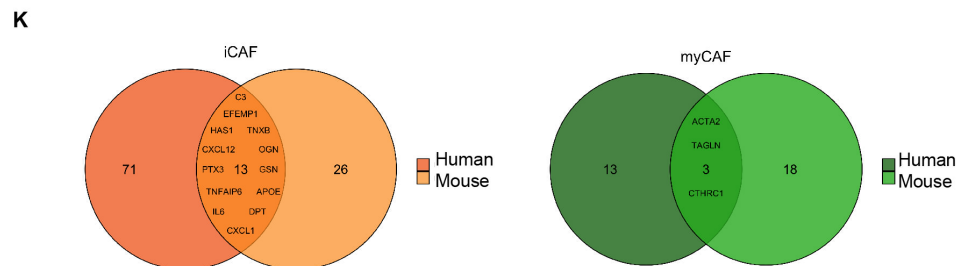
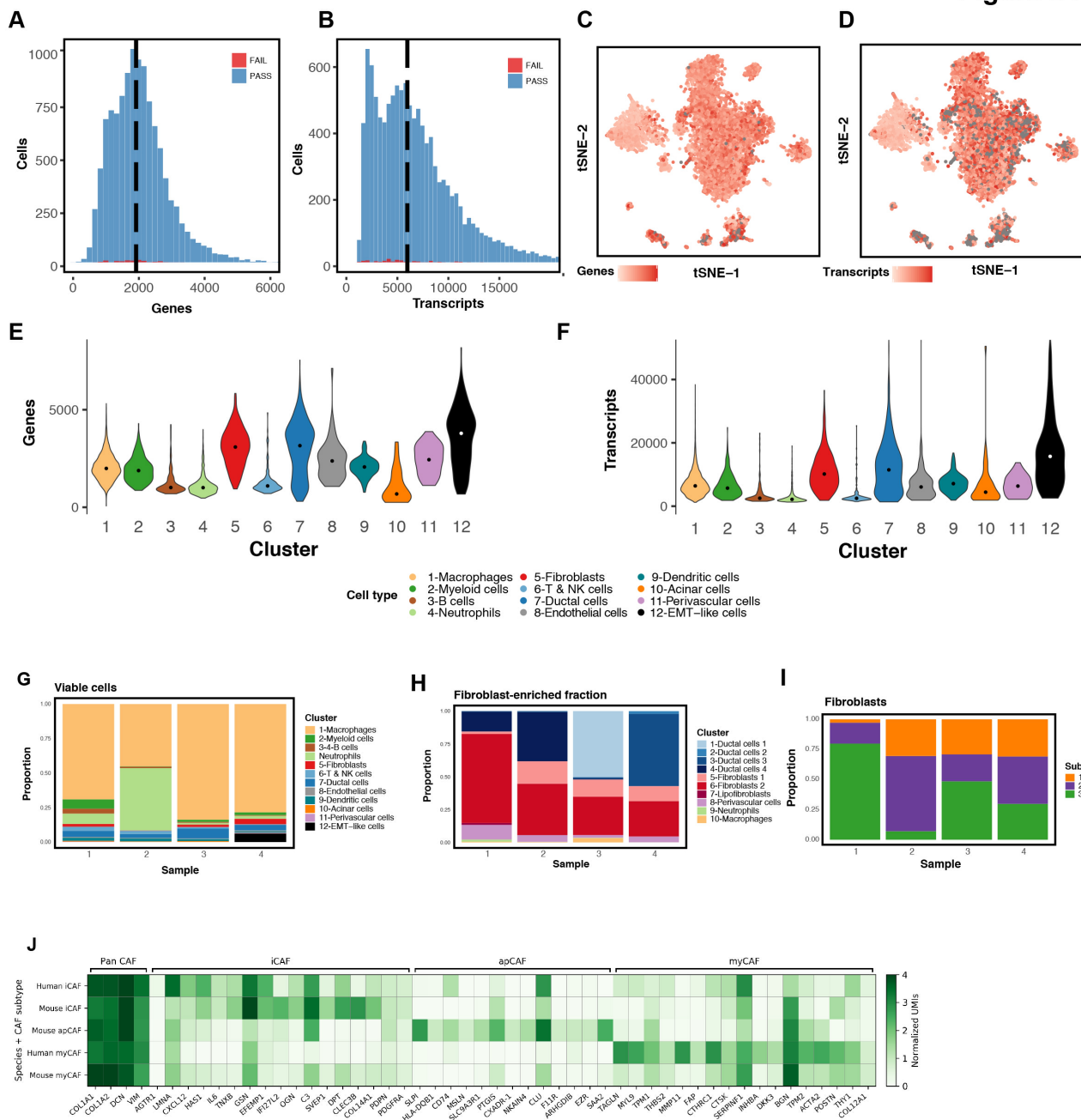


B Pathways downregulated in human myCAFs vs. iCAFs



Supplementary Figure S3: Pathway analysis in human iCAFs and myCAFs. Gene set enrichment analysis (GSEA) of human myCAFs vs. human iCAFs. Shown are pathways significantly upregulated in myCAFs (A) or significantly downregulated in myCAFs (B), compared to iCAFs.

Figure S4

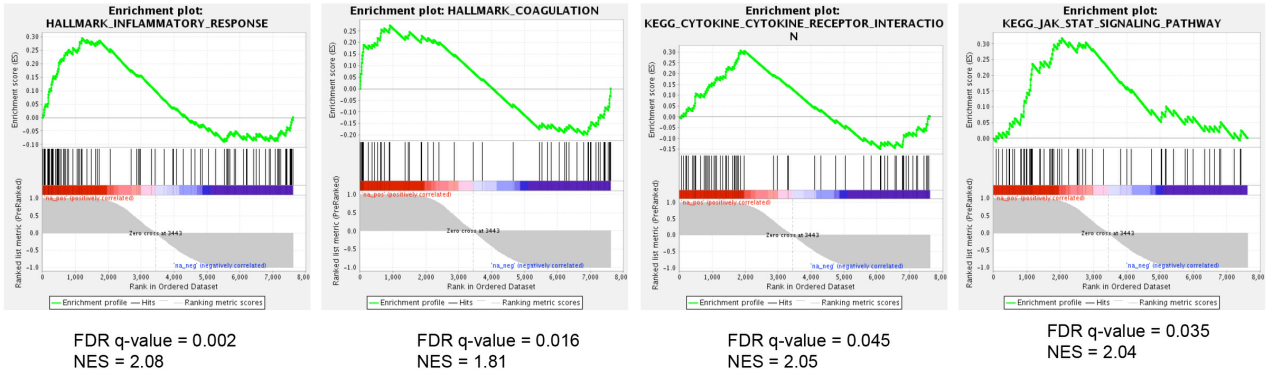


Supplementary Figure S4: Data quality and clustering metrics of the KPC mouse PDAC single cell transcriptomes. A-B. Histograms showing the number of genes (**A**) and transcripts (**B**) that were detected per cell in the mouse PDAC dataset. Blue histogram shows cells that passed QC and red histogram shows cells that failed QC. The dotted line represents the median value, which is 1,916 genes/cell and 5,989 transcripts/cell. **C-D.** Number of genes (**C**) and transcripts (**D**) detected per cell, represented as a t-SNE plot. Legend shows a color gradient corresponding to gene/transcript number per cell. Gray color represents cells with greater than 4,000 genes or 10,000 transcripts, respectively. **E-F.** Distribution of genes (**E**) and transcripts (**F**) detected in each cluster in the mouse PDAC dataset. **G.** Bar plot showing proportions of the different clusters in each KPC mouse sample in the 'viable cell fraction' analysis. **H.** Bar plot showing proportions of the different clusters in each KPC mouse sample in the 'fibroblast-enriched fraction' analysis. **I.** Bar plot showing proportions of the different fibroblast sub-clusters in each KPC mouse sample. **J.** Heatmap showing orthologous genes expressed in the CAF subtypes across different species. Raw counts matrices from human and mouse CAFs were concatenated along a set of common orthologues (provided by Ensembl, described here: <http://www.ensembl.info/2009/01/21/how-to-get-all-the-orthologous-genes-between-two-species/>) and were normalized and log transformed jointly. **K.** Venn diagram of the common markers of iCAFs and myCAFs between human and mouse datasets, informed by marker genes for each species subtypes (from Supplementary Tables S13 and S22).

Figure S5

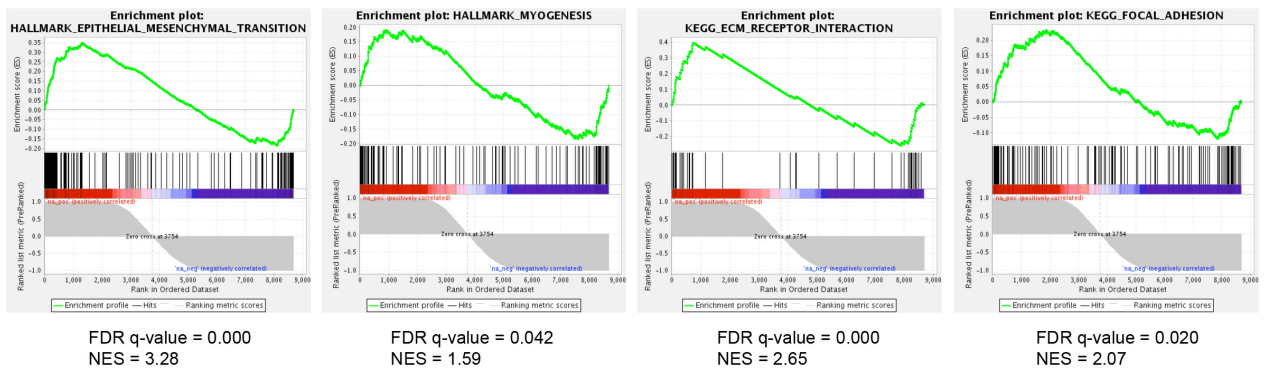
A

Pathways upregulated in mouse iCAFs vs. myCAFs & apCAFs



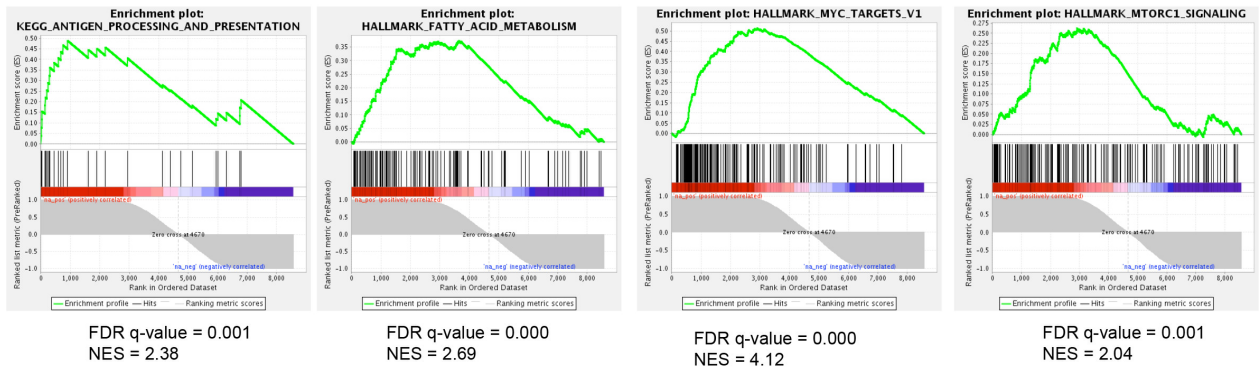
B

Pathways upregulated in mouse myCAFs vs. iCAFs & apCAFs



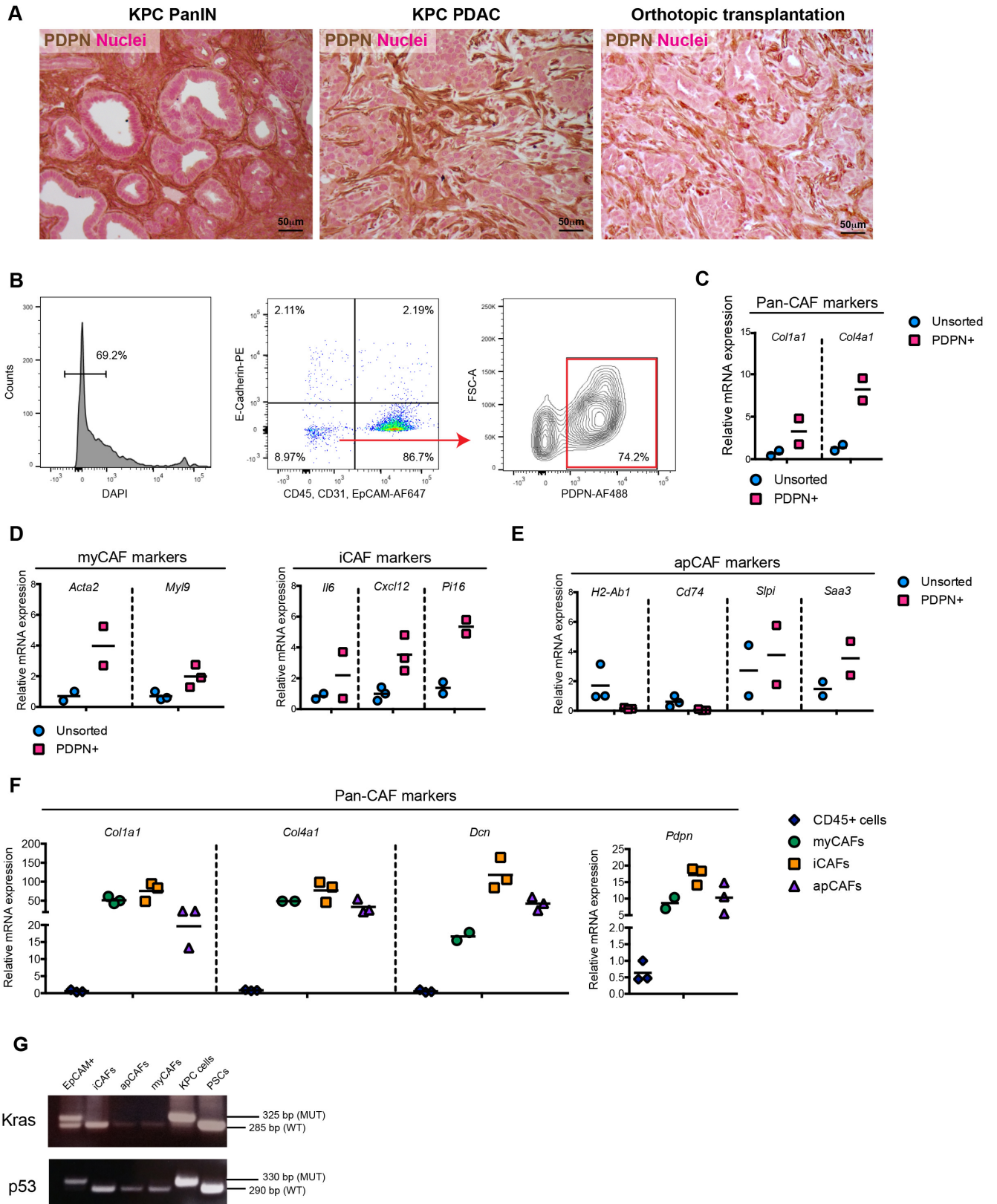
C

Pathways upregulated in mouse apCAFs vs. iCAFs & myCAFs



Supplementary Figure S5: Pathway analysis in different KPC CAFs. Gene set enrichment analysis (GSEA) of KPC mouse CAFs, showing upregulated pathways in iCAFs (**A**), myCAFs (**B**) and apCAFs (**C**). Each CAF subtype was compared to each of the other two subtypes, separately, and significant pathways that came up in both comparisons were selected to be shown. Histogram and significance of one of the comparisons is shown for convenience.

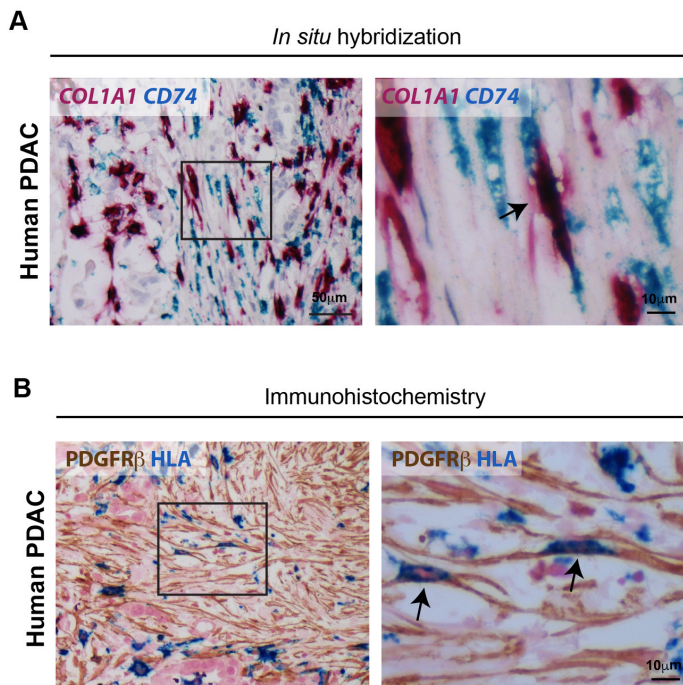
Figure S6



Supplementary Figure S6: Podoplanin is a pan-fibroblast marker in murine PDAC.

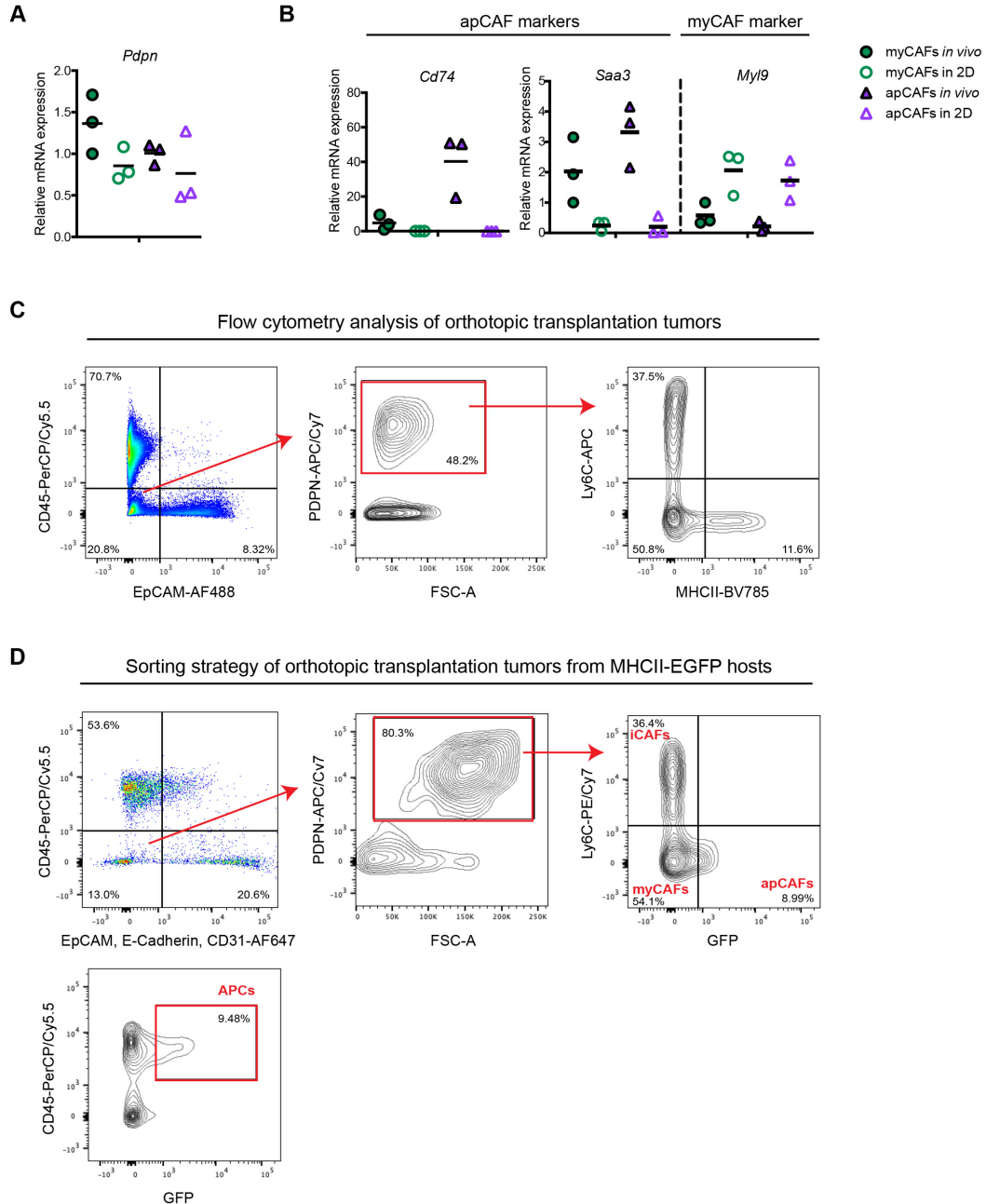
A. IHC of podoplanin (PDPN) in a preneoplastic and neoplastic mouse tissue sections: KPC pancreatic intraepithelial neoplasia (PanIN, left panel), KPC PDAC (middle panel) and an orthotopic transplant model of PDAC (right panel). Nuclear Fast Red is used as counterstain. **B.** A representative FACS sorting strategy of PDPN-positive fibroblasts from a KPC tumor. Cells that were viable, negative for CD45, CD31, EpCAM and E-Cadherin, and positive for PDPN were sorted. **C-E.** qPCR analysis of pan-CAF markers (**C**) and specific CAF subtype markers (**D, E**) in PDPN⁺ sorted cells compared to unsorted population of a KPC tumor (n≥2 biological replicates). Black horizontal line represents mean value of data points. All transcripts were normalized to *Hprt*. **F.** qPCR analysis of pan-CAF markers in the three CAF subtypes sorted from KPC tumors, compared to CD45⁺ cells (n≥2 biological replicates). Black horizontal line represents mean value of data points. All transcripts were normalized to *Hprt*. **G.** PCR analysis of *Kras* and *p53* genetic status in the different CAF subtypes, compared to EpCAM⁺ population sorted from KPC tumors. Shown is a representative example of 3 biological replicates. Bands corresponding to wild-type and recombined alleles are detected according to their different sizes. KPC cells and pancreatic stellate cells (PSCs) serve as positive controls for recombined alleles and wild-type alleles, respectively.

Figure S7



Supplementary Figure S7: Validation of apCAFs in human PDAC. **A.** Duplex *in situ* hybridization of *COL1A1* and *CD74* in human PDAC sections. The black square on the left panel is magnified in the right panel. Arrow indicates an apCAF. **B.** IHC of PDGFR β and HLA-DR/DP/DQ in human PDAC sections. The black square on the left panel is magnified in the right panel. Arrows indicate apCAFs.

Figure S8



Supplementary Figure S8: Characterization of apCAFs in KPC and orthotopic transplantation models. A-B. qPCR analysis of *Pdpn* (A) and CAF marker genes (B) in apCAFs sorted from KPC tumors (apCAFs *in vivo*) compared to the same population following culture in two-dimensional monolayer (apCAFs in 2D). myCAFs sorted from the same KPC tumors (myCAFs *in vivo*) and myCAFs grown in 2D (myCAFs in 2D) were used as positive controls for myCAF genes (n=3 biological replicates). Black horizontal line represents mean value of data points. All transcripts were normalized to *Hprt*. **C.** A representative flow cytometry analysis of cell suspension from an orthotopic transplantation tumor. Forward- and side-scatter were used to eliminate debris, and DAPI staining was used to eliminate dead cells. After gating for CD45- and EpCAM-negative cells, PDPN-positive cells were stained for Ly6C and MHCII, and showed distribution into three CAF subpopulations, similar to the KPC tumors. **D.** A representative sorting strategy of orthotopic transplantation tumors from MHCII-EGFP host mice. Forward- and side-scatter were used to eliminate debris, and DAPI staining was used to eliminate dead cells. After gating for CD45-, EpCAM-, E-Cadherin- and CD31-negative cells, PDPN-positive cells were plotted for Ly6C and GFP. GFP-positive CAFs were sorted as apCAFs. In parallel, CD45-positive GFP-positive cells were sorted as professional APCs.