Supplemental Material

Mitigation of T-cell dependent immunogenicity by reengineering Factor VIIa analogue

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Supplemental Methods

Supplemental methods to Figure S1

Creation of a single score for each positional change:

The scoring function used to determine the viability of all amino acid substitutions at each position incorporated three factors: (i) A population-based score judging the in-silico probability of an immune response. (ii) A score judging the evolutionary pressure for conservation of an amino acid at each position in the sequence. (iii) A score assessing functional consequence of each amino acid substitution based on a third-party algorithm, EVmutation 1 .

Immunogenicity Score:

The immunogenicity score is equal to for any amino acid change defined as:

$$
\sum_{p \in P} \sum_{a \in A} f_a \cdot T_t
$$

Where:

P is the set of all possible peptides containing the positional change

A is the set of all HLA-DRB1 alleles considered

is the frequency of allele a in the population of interest

is a binary operator testing whether the predicted binding affinity of peptide p and allele a is less than threshold t

The Immunogenicity score is then compared to the Immunogenicity score for the original protein over the same positions but with no amino acid substitutions. If the amino acid substitution has a lower score than the original protein, it is considered a candidate for substitution.

Conservation score:

For each column in the sequence of interest, a score was calculated by examining the divergence between a baseline probability distribution and the probability distribution of amino acids in that column. The baseline amino acid distribution was derived from the Uniprot-Knowledge-base- SwissProt database. The distribution is the frequency of amino acids within the database (553,474 proteins). This background distribution represents an expected distribution of amino acids with no evolutionary pressure.

The set of sequences examined included a wide distribution of species. The sequences for FVII in these species were aligned using MUSCLE and the 31 amino acid sequences of interest were extracted from these sequences. Jensen Shannon Divergence:

$$
\lambda = \frac{1}{2}
$$

\n
$$
RE_{p_c,q} = \sum_{\alpha \in A.A.} p_c(\alpha) \log \frac{p_c(\alpha)}{q(\alpha)}
$$

\n
$$
r = \lambda p_c + (1 - \lambda)q
$$

\n
$$
D_c^{JS} = \lambda RE_{p_c,r} + (1 - \lambda)RE_{(q,r)}
$$

Where:

 $p_c(\alpha) =$ probability distribution of amino acids of the samples $q(\alpha)$ = probability distribution of amino acids UniProtKB – Swiss – Prot c is the column being observed

 α is one particular amino acid being observed

The divergence between the background distribution and the observed distribution was evaluated using the Jensen-Shannon-Divergence score. This score has similar properties to the Kullback-Leibler divergence that is often used to evaluate the divergence of probability distance²; however, it is symmetric and bounded by 0 and 1. It was found to be a favorable tool for identifying functionally important residues in proteins.

A position in the peptide is considered to be not conserved if the amino acids at that position could be seen as more likely to occur as random chance rather than as a result of evolutionary pressure. A position with a random assortment of amino acids would have a lower Jensen-Shannon-Divergence score than a position that was more homogeneous as it is closer to the baseline random distribution of peptides in the proteome. These low scoring positions can be considered to have less evolutionary pressure for amino acid conservation.

A Conservation score was calculated for each position on the peptide. During analysis, only a subset of positions in the region of interest was considered. Positions which are greater than one mean absolute deviation below the median are considered to be the less conserved positions within the region of interest and are retained for further consideration.

EVmutation Score

The EVmutation score was obtained by using the precomputed single mutation scores available at the EVmutation website [https://marks.hms.harvard.edu/EVmutation/index.html]. Scores of roughly 0 or higher are considered stable mutations. For our analysis, we kept scores that were greater than -0.2 in order to have a small amount of leeway around 0.

Overall Consideration:

Using this method, the number of possible changes to be made was reduced from 285 to 123 after the immunogenicity step, 42 after the conservation step and 5 after the EVmutation step. The python and R code used for the selection of the optimal positions is available upon request

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Supplemental Figures

Figure S1. The Deimmunization algorithm (*RID*). *RID* is a set of steps intended to filter the possible search space for amino acid changes. There are three filters: 1. The use of the promiscuity scores to determine whether an amino acid change will result in reduced engagement with HLA variants at the population level (depicted in green on the workflow). 2. The use of a conservation score applied to amino acid substitutions that result in lower promiscuity scores, to identify primary sequence locations that should not be altered as they are likely to be functionally important (depicted in orange on the workflow). 3. The determination of an EVmutation score²⁵ for amino acid substitutions that result in lower promiscuity scores and are *not* conserved to identify amino acid changes that could be functionally deleterious (depicted in blue on the workflow).

Figure S2. Sequence alignment. Amino acid sequence alignment of wild-type FVIIa, vatreptacog alfa (VA), and the 6 variants analyzed. Substitutions inherent to VA are shown in red. Additional substitutions are shown in blue. Larger fonts are used to highlight the substitution in VA at position 158 as well as the substitutions made in the immunogenic region of interest surrounding positions 296 and 298.

Figure S3. Frequencies of HLA-DRB1 variants used in this study. The relative frequency of individual HLA-DRB1 variants used in predicting immunogenicity using RID (left) and the relative frequencies of HLA-DRB1 variants identified in the 50 subjects used in each of the two assays (center and right). The total coverage of alleles in the world population is also indicated.

Figure S4. T-cell Responses. Cells from 50 subjects were subjected to a 3H-thymidine incorporation T-cell proliferation assay and IL-2-ELISpot assay. A stimulation index > 1.9 is considered a positive responder for either assay. Positive donors are shown in red for the T-cell Proliferation assay and in blue for the ELISpot assay.

Figure S5. Quantification of FVIIa-like activity by clotting assay. (A) FVIIa samples were serially diluted in Tris-BSA buffer (pH 7.4) and added to a commercially available plasma from congenital hemophilia A patients pre-mixed with 4 µM of phospholipid (PS:PC) vesicles. Clot formation was monitored through optical density (clot turbidity) at 412nm. (B) Clot time values were calculated for each dilution of the FVIIa samples from the kinetic data as the point when optical density reached half of its maximal value. (C) Calibration curve used to calculate relative activities of FVIIa variants by comparing the clot times for several vatreptacog alfa (VA) serial dilutions.

Figure S6. Validation of the selected screening candidates. (A) Selected mutations N301D and N301E (blue squares), predicted to retain function comparable to VA and the mutation L300D (orange square) predicted to be nonfunctional. (B) Activities of wild-type FVIIa, the vatreptacog alfa (VA) discontinues variant and the mutants L300E, N301D and N301E determined using the Thrombin Generation assay in the presence of phospholipid vesicles (4 µM).

Supplemental Tables

Table S1. Percentage of each HLA-DRB1 allele in different human populations. These frequency distributions were used in silico calculations of weighted promiscuity scores.

Table S2. Percentage of each HLA-DRB1 allele in each of the two T-cell proliferation experiments as well as the respective frequency of that allele in the World population.

Assay 1

VA/WT

Assay 2

Table S3. Percentage of each HLA-DRB1 allele in different human populations for each of the two T-cell proliferation experiments.

References

- 1. Hopf TA, Ingraham JB, Poelwijk FJ, et al. Mutation effects predicted from sequence co-variation. *Nat Biotechnol.* 2017;35(2):128-135.
- 2. Capra JA, Singh M. Predicting functionally important residues from sequence conservation. *Bioinformatics.* 2007;23(15):1875-1882.