Developmental Cell, Volume 50

Supplemental Information

Selective Autophagy of Mitochondria

on a Ubiquitin-Endoplasmic-Reticulum Platform

Maria Zachari, Sigurdur R. Gudmundsson, Ziyue Li, Maria Manifava, Ronak Shah, Matthew Smith, James Stronge, Eleftherios Karanasios, Caterina Piunti, Chieko Kishi-Itakura, Helena Vihinen, Eija Jokitalo, Jun-Lin Guan, Folma Buss, Andrew M. Smith, Simon A. Walker, Eeva-Liisa Eskelinen, and Nicholas T. Ktistakis



Figure S1 related to Figure 1

IVM induces mitophagy. (A) Structure of IVM. (B-D)) HEKcells were treated with 15 μ M IVM for 90 min, and examined for LC3 response by iffl and blots. (E)) HEKcells treated with PP242 for 1 hr or with IVM for 1 and 2 hr were lysed and immunoblotted for phospho-S6K, phopsho-ULK1 or phospho-4EBP1. (F) HEKcells stably expressing GFP-DFCP1 (top panels) or parental HEKcells (bottom panels) were treated with 15 μ M IVM +/- 100 nM wortmannin and examined for DFCP1 or LC3. (G) HEKcells stably expressing GFP-DFCP1 were treated with 20 μ M IVM for 90 min and immunolabelled for ATG13. (H) HEKcells treated with 15 μ M IVM were fixed for EM examination. Arrow marks phagophore engulfing a single mitochondrion. (I) HEKcells were treated with the indicated amounts of IVM for 8 hr or 16 hr and then immunoblotted for TOM20 and FIP200 . (J-K)HEKcells were transfected with siRNAs against PINK1 or with non-targeting control (NT) and treated with IVM, CCCP or antimycin A before immunoblotting for MITOFUSIN 2 (J) or PINK1 (K). Asterisks indicate higher mobility forms of MITOFUSIN 2. (L) HEKcells were transiently transfected with mCherry-Parkin and treated for 90 min with CCCP or with IVM as indicated beforeTOM20 immunolabellimg (M) HeLa cells were treated with IVM for 90 min before immunolabelling with WIPI2 and TOM20.





control

IVM

control

IVM



Figure S2 related to Figure 1.

IVM does not induce mitochondrial permeabilization; IVM—induced mitophagy depends on ubiquitination. (A) HEK-293 cells were treated with 15 IM IVM for the times indicated and immunolabelled for TOM20 and cytochrome C as indicated. (B) HEK-293 cells were incubated with 50 ng/ml of Mitotracker Red FM or Mitotracker Red CMXRos as indicated for 10 min followed by addition of IVM for an additional 60 min. (C) Cells treated as in A were immunolabelled with antibodies to ubiquitin. (D) HEK-293 cells were treated with IVM in the presence or absence of PYR-41 and lysates were immunoblotted for LC3. (E) Triplicate experiment as shown in D is quantitated.

merged

V. Contraction of the second s

90

42

Ž

GPDH

Figure S3 related to Figures 2-3

Requirement for ATG9, Optineurin and Tax1bp in IVM-induced mitophagy. (A) Wild type MEF cells or ATG9 KOs were treated for 90 min with 15 mM IVM or with 1 mM PP242 and immunolabelled for ubiquitin, phospho-TBK1, Optineurin and WIPI2. Number of puncta for each condition are shown in the graphs. (B) MEF cells deleted for FIP200 were treated for 90 min with 15 mM IVM or with 1 mM PP242 and immunolabelled for ubiquitin and ATG9. (C) Wild type MEF cells or optineurin KOs were treated for 90 min with 15 μ M IVM or with 1 μ M PP242 and immunolabelled for ubiquitin and ATG9. (C) Wild type MEF cells or optineurin KOs were treated for 90 min with 15 μ M IVM or with 1 μ M PP242 and immunolabelled for ubiquitin, phospho-TBK1, Optineurin and WIPI2. Number of puncta for each condition is shown in the graphs. (D) Wild type and Optineurin KO MEFs as in A were transfected with siRNAs against Tax1bp or with a non-targeting oligonucleotide for 72 hrs. The cells were then treated with 15 μ M IVM and were immunolabelled with ubiquitin, phospho-TBK1, optineurin and WIPI2. The number of puncta for each condition is shown in the graph or Optineurin KO MEFs transfected with siRNAs to Tax1bp or with a non-targeting control (NT) and immunoblotted for the indicated proteins.

A 1613	0	32"	1'04''	1'36"	2'08''	2'40''	3'12''	3'44''	4'16"	4'48''	5'20''	5'52"	6'24''	6'56''	7'28''
PTN A					7									10	-
nerged N				5	2						1				•
ATG13 T	8′	8'32''	9'04''	9'36''	10'08''	10'40''	11'12''	11'44''	12'16''	12'48''	13'20''	13'52''	14'24''	14'56''	15'28''
OPTN	•		U.	. • .			•			- 0-	9		ø		N
MITO															
merged	ø,	•	0	•		2	9,	3			•	0			

Dynamics of optineurin and ubiquitin during mitophagy. (A) Wide field live-imaging of HEK-293 cells stably expressing GFP-ATG13 and transiently transfected with mCherry-optineurin and CFP-MITO (A) or transiently transfected with GFP-optineurin, CFP-LC3 and mCherry-MITO (B) orstably expressing GFP-ATG13 and transiently transfected with mCherry-ubiquitin and CFP-MITO (C). IVM was added for 10 min and then cells were imaged for an additional 60 min. The bar represents 1 μ m.

Figure S5 related to Figure 5.

(A) An expanded set of the images shown in Figure 8 D. Four-color SIM images of mitochondrial fragments (blue) during IVMinduced mitophagy and associated with ATG13 (green), optineurin (purple), WIPI2 (purple), phospho-TBK1 (purple) Tax1BP (purple), NDP-52 (purple) and ER (red) as indicated. Shown are the four individual channels, the four-color overlay and twocolor overlays of the various components as indicated. (B) Four-color SIM images of mitochondrial fragments (blue) during IVM-induced mitophagy in HEK-293 cells lacking ATG13. Staining for ER (red) and other components is as indicated. (C) Dynamics of mitophagy as revealed in this work. Several components are involved in the engulfment of the mitochondrial target (top diagram): some cover it smoothly (ubiquitin, ER, omegasomes, LC3, continuous lines) whereas others appear to oscillate on and off and translocate to multiple sites on the target (optineurin and ATG13 shown as an example, swirling lines). We imagine that these components co-ordinate the translocation of the rest of the machinery, including the lipidation complex and LC3. One way to explain these dynamics is that the ubiquitination step leads to some stable interaction of the mitochondrial target with an ER cradle, which provides the spatial clues for organizing the translocation of the autophagy/mitophagy machinery (bottom diagram).

Figure S6 related to Figures 4-5.

Dynamics of mitophagy induced by Antimycin A/Oligomycin (A/O) treatment. (A) HEK-293 cells expressing GFP-ATG13 and mCherry-MITO were treated with A/O for 8 h and then imaged every 12 sec.. (B-C) HEK-293 cells expressing GFP-ATG13, mCherry-MITO and CFP-ER were treated with A/O for 8 h (C) or 18 (D) h and imaged every 12 sec as shown in the selected panels. Arrows indicate the position of ATG13 and ER structures during the mitochondrial engulfment process. The bar in all panels represents 1 μ m.

La specific par

Figure S7 related to Figure 6.

Correlative light-electron microscopy images of IVM-induced mitophagy. (A) A cell with 3 simultaneous mitophagosomes (i, ii, iii). Events i and ii show a forming phagophore tightly next to the mitochondrion while event iii shows an ER-like structure next to the mitochondrion, although an ATG13 signal is evident in live imaging in all three events. The absence of a phagophore when ATG13 is seen to translocate to the mitochondrion is reminiscent of the event shown in Fig 9 M-P and probably represents the earliest intermediates in mitophagy pathway. (B) A mitophagy event showing how ER (yellow arrows) and phagophore (green arrows) wrap around the targeted mitochondrial fragment.