

Supporting Information

A modular vector toolkit with a tailored set of
thermosensors to regulate gene expression in
Thermus thermophilus.

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Table S1. Strain list

| Strain | Genotype | Phenotype/use |
|---|---|---------------------------|
| <i>E. coli</i> DH5 α | F ⁻ <i>endA1 hsdR17 supE44 thi-1 recA1 gyrA96 relA1</i> Δ (<i>argF-lacZYA</i>)U169 Φ 80d <i>lac</i> Δ M15 | Cloning Promoter assay |
| <i>Thermus thermophilus</i> HB27 | ATCC BAA-163/DSM7039 | Wild type |
| <i>Thermus thermophilus</i> HB27 Δ Ago | Deletion of <i>argonaute</i> ¹ | Enhanced competence |

Table S2. Plasmid list

| Plasmid | Description | Use | Reference |
|---------------|--|---|--------------|
| pEM2S (pMK18) | plasmid with the Tth minimal replication origin from pMY1 | Source of replication origin | ² |
| pMKnqosIFP | pMK derivative with IFP expression driven by Pnqo | IFP expression test | ³ |
| pTT8 | Endogenous plasmid of Tth HB8. | Source of the pTT8 replication origin | ⁴ |
| pSEVA1311 | pSEVA plasmid with the broad host range pBBR1 replication origin | Source of the pBBR1 E.c. replication origin | ⁵ |
| pMoTK110 | pMY Tth origin, Kanamycin, pUC <i>E.coli</i> origin, Pnqo driving IFP gene (NQO) | Basis for pMoT series | This work |
| pMoTK100 | pMoTK110 with linker A cloned between NheI and HindIII | pMoTK cloning | This work |

| | | | |
|------------|---|--------------------------------------|-----------|
| pMoTK111 | pMoTK110 with linker 1 cloned between NdeI and HindIII | pMoTK Pnqo library cloning | This work |
| pMoTK112 | pMoTK110 with linker 2 cloned between NdeI and HindIII | pMoTK Pnqo library cloning | This work |
| pMoTK113 | pMoTK110 with linker 3 cloned between NdeI and HindIII | pMoTK Pnqo library cloning | This work |
| pMoTK115 | pMoTK110, with lac promoter driving IFP (LAC) | Plac test | This work |
| pMoTK116 | pMoTK110 cut with NheI and HindIII and filled-in and religated to eliminate Pnqo-IFP sequence (pMoTK) | ☐ Pnqo-IFP control | This work |
| pMoTK117 | pMoTK110 cut with NheI and EcoRI filled-in and religated to delete Pnqo | ☐ Pnqo control | This work |
| pMoTK118 | pMoTK110, with 16S rRNA promoter driving IFP (16S) | P16S test | This work |
| pMoTK119 | pMoTK110, with <i>Haloferax volcanii</i> promoter driving IFP (3K) | 3K test | This work |
| pMoTK1103A | pMoTK110, with 3A variant of Pnqo driving IFP (K3A) | Pnqo 3A test | This work |
| pMoTB1103A | pMoTK1103A, with bleomycin resistance cassette cloned between PacI and AscI | pMoT with bleomycin expression test | This work |
| pMoTH1103A | pMoTK1103A, with hygromycin resistance cassette cloned between PacI and AscI | pMoT with hygromycin expression test | This work |
| pMoTK2103A | pMoTK1103A, with pTT8 replication origin cloned between PstI and AatII (Ori2-3A) | Origin pTT8 expression test | This work |
| pMoTK120 | pMoTK110 with nqo91 variant of Pnqo | Pnqo91 assay | This work |
| pMoTK121 | pMoTK110 with nqo72 variant of Pnqo | Pnqo72 assay | This work |
| pMoTK122 | pMoTK110 with nqo57 variant of Pnqo | Pnqo57 assay | This work |

| | | | |
|----------|--|---------------------------|-----------|
| pMoTK123 | pMoTK110 with nqo33 variant of Pnqo | Pnqo33 assay | This work |
| pMoTK124 | pMoTK110 with nqo91-35 variant of Pnqo | nqo91-35 assay | This work |
| pMoTK125 | pMoTK110 with nqo91 E-10 variant of Pnqo | nqo91 E-10 assay | This work |
| pMoTK126 | pMoTK110 with nqo91-10 variant of Pnqo | nqo91-10 assay | This work |
| pMoTK127 | pMoTK110 with nqo91-7 variant of Pnqo | nqo91-7 assay | This work |
| pMoTK128 | pMoTK110 with nqo72 E-10 variant of Pnqo | nqo72 E-10 assay | This work |
| pMoTK129 | pMoTK110 with nqo72 -10 variant of Pnqo | nqo72 -10 assay | This work |
| pMoTK130 | pMoTK110 with nqo72-7 variant of Pnqo | nqo72-7 assay | This work |
| pMoTK140 | pMoTK110 with 3Knqo72 variant of Pnqo | 3Knqo72 assay | This work |
| pMoTH100 | pMoTH1103A with linker A cloned between NheI and HindIII | pMoTH cloning | This work |
| pMoTH150 | pMoTH with IFP expressed cotranscriptionally with the hygromycin resistance gene. DF, direct fusion, control construct without thermosensor. See Sequence File | DF construct IFP assay | This work |
| pMoTH151 | pMoTH150 with polyA stretch cloned between XbaI and NdeI | polyA construct IFP assay | This work |
| pMoTH152 | pMoTH150 with thermosensor TS0 cloned between XbaI and NdeI | thermosensor TS0 assay | This work |
| pMoTH153 | pMoTH150 with thermosensor TS10 cloned between XbaI and NdeI | thermosensor TS10 assay | This work |
| pMoTH154 | pMoTH150 with thermosensor TS8 cloned between XbaI and NdeI | thermosensor TS8 assay | This work |

| | | | |
|----------|---|------------------------|-----------|
| pMoTH155 | pMoTH150 with thermosensor TSP cloned between XbaI and NdeI | thermosensor TSP assay | This work |
| pMoTH156 | pMoTH150 with thermosensor TS1 cloned between XbaI and NdeI | thermosensor TS1 assay | This work |
| pMoTH157 | pMoTH150 with thermosensor TS2 cloned between XbaI and NdeI | thermosensor TS2 assay | This work |
| pMoTH158 | pMoTH150 with thermosensor TS4 cloned between XbaI and NdeI | thermosensor TS4 assay | This work |
| pMoTH159 | pMoTH150 with thermosensor TS6 cloned between XbaI and NdeI | thermosensor TS6 assay | This work |

Table S3. Oligonucleotides list

| Name of oligonucleotide | Sequence | Utilization |
|-------------------------|--|--|
| EM2SRepAat | TGTTGGCGACGTCCTCAGTTGACC | Primer Fw to amplify TthRep1 from pEM2S with AatII site |
| EM2SRepPstRev | AAACTGCAGGTTATACTCCCCCGGG | Primer Fw to amplify TthRep1 from pEM2S with PstI site |
| PslAPacPst | AAACTGCAGTTAATTAAGAAACCTTAAGGCCGACCG | Primer Fw to amplify PslpA-kanamycin resistance cassette from pMK184 with PstI and PacI sites |
| KatAscNheRv | GTCGTGCTAGCTACAGGCGGCCACGTCGTGACTGGGAAAAC | Primer Rev to amplify PslpA-kanamycin resistance cassette from pMK184 with AscI and NheI sites |
| ColENheHind | CACAGCTAGCTACGAAGCTTAGCCGGAAGCATAAAG | Primer Fw to amplify pUC Ec replication origin with NheI and HindIII sites |
| ColE1AatNot | CTGACGTCAAGCGGCCGAGTTTACTCATATATACTTTAGA | Primer Rev to amplify pUC Ec replication origin with AatII and Not sites |
| TermSLNheFw | [Phos]CTAGTAAAGTTCTAACCCTGGCGGTCCCGCCCCGCCCTTTGGGGGGGGGGGGTTTTTGTCTCCCG | Primer Fw to generate the SlpA terminator (to NheI site) |
| TermSLNheRv | [Phos]CTAGCGGGGAGAAACAAAACCCCCGCCGCCCAAGGGGGCGGGGGCGGGACCGCCAGGGGTTAGAACTTTA | Primer Rv to generate the SlpA terminator (to NheI site) |
| PnqoNhe | AAAAGCTAGCTCCAGGGGCCTTCTTTCC | Primer Fw to amplify promoter nqo with NheI site |
| PnqoEcoRv | AAACATATGCCCTCCTTTCGTGCACGAAAGAATTCTTCTCACGAAAC | Primer Rv to amplify promoter nqo with sites EcoRI and NdeI |
| sIFPFw | GGGCATATGAGCAAAGGAGAAGAA | Primer Fw to amplify sIFP gene with site NdeI |
| sIFPRv | CCGCAAGCTTTTATTATTGTAGAGCTC | Primer Rv to amplify sIFP gene with site HindIII |

| | | |
|---------------|---|---|
| NdeltoVspIpsI | CATTATTAATCCTCACACCTCCTTAAGGGT | Primer Rv to amplify PlspA with site VspI |
| NdeHph17 | GAGGTGTGAGGCATATGAAAAAGCC | Primer Fw to amplify Hph17 gene with site NdeI |
| EndHph17 | CATAACCTGAAGGAGGCGCCATCTCTATTCTT | Primer Rv to amplify Hph17 gene with site Ascl |
| NdeBleo | GGTGTGAGGCATATGGCCAAGT | Primer Fw to amplify Bleomycin resistance gene with site NdeI |
| EndBleo | CCCGGGGATGGCGCCCTAGATTAGTC | Primer Rv to amplify Bleomycin resistance gene with site Ascl |
| pTT8Fw | CGCTCACCTGAGCG | Primer Fw to amplify pTT8 replication origin |
| pTT8PstRv | AAAAACTGCAGGCTATGGAGGGTTCTCCCTG | Primer Rv to amplify pTT8 replication origin with site PstI |
| OripBBRFw | ATAGGTGAAGCTTAGGCCACCCG | Primer Fw to amplify pBBR replication origin from pSEVA with HindIII site |
| OripBBRNot | AAAGCGGCCGCCCTACGGGCTT | Primer Rv to amplify pBBR replication origin from pSEVA with Not site |
| MKsIFPFW | AGAAGAATTCTTTCGTGCACGAAAGGAGGGGC | Primer Fw to amplify the sIFP CDS from pMKnqosIFP with EcoRI site |
| MKsIFPRv | TGCGGCCGCAAGCTTTTATTATTGTAG | Primer Rv to amplify the sIFP CDS from pMKnqosIFP with HindIII site |
| PlacNheFw | GCAGCTAGCACGACAGGTTTCC | Primer Rv to amplify lac promoter from pUC18 with NheI site |
| PlacEcoRv | TTGTTATGAATTCTACAATCCACAC | Primer Rv to amplify lac promoter from pUC18 with EcoRI site |
| P16SFw | AAAGCTAGCTTACC GCGAGTTTGACTCGTGAGCGCTTCATTGAGGAGATC CGTGCCCTCGCAA | Primer Fw to generate 16S promoter with NheI site |
| P16SRv | TTTGAATTCAGAAAAGCCATGCTATCAATCCCCCTCTTTTGTCAAGGCTTG CGAGGGCAGCGGATCT | Primer Rv to generate 16S promoter with EcoRI site |
| MoTLinkerFw | [Phos]CTAGCAGATATCATCTAGAAAACATATGAGGTACCAGAATCACTAG TGGATCCA | Oligonucleotide Fw to generate polylinker (to sites NheI and HindIII) |
| MoTLinkerRv | [Phos]AGCTTGGATCCACTAGTGAATTCTGGTACCTCATATGTTTTCTAGATG ATATCTGCTAG | Oligonucleotide Rv to generate polylinker (to sites NheI and HindIII) |
| Linker1Fw | [Phos]TATGGTCGACAAGATATCAGGATCCATCTAGAA | Oligonucleotide Fw to generate Linker1 (to sites NdeI and HindIII) |
| Linker1Rv | [Phos]AGCTTTCTAGATGGATCCTGATATCTTGTGACCA | Oligonucleotide Rv to generate Linker1 (to sites NdeI and HindIII) |
| Linker2Fw | [Phos]TATGGGTCGACAAGATATCAGGATCCATCTAGAA | Oligonucleotide Fw to generate Linker2 (to sites NdeI and HindIII) |
| Linker2Rv | [Phos]AGCTTTCTAGATGGATCCTGATATCTTGTGACCCA | Oligonucleotide Rv to generate Linker2 (to sites NdeI and HindIII) |
| Linker3Fw | [Phos]TATGGGGTCGACAAGATATCAGGATCCATCTAGAA | Oligonucleotide Fw to generate Linker3 (to sites NdeI and HindIII) |
| Linker3Rv | [Phos]AGCTTTCTAGATGGATCCTGATATCTTGTGACCCCA | Oligonucleotide Rv to generate Linker3 (to sites NdeI and HindIII) |
| nqo91 | GTAGGCTAGCCCCCTTGC GCC | Primer Fw to generate promoter nqo91 |
| nqo72 | TTGCGCGCTAGCCCGGGTGAT | Primer Fw to generate promoter nqo72 |
| nqo57 | GGTGCTAGCATGGGCAGGAAATGA | Primer Fw to generate promoter nqo57 |

| | | |
|---------------|---|---|
| nqo33 | AGGCCGGCTAGCTTTGGGGCTTTG | Primer Fw to generate promoter nqo33 |
| 3Knqo | CCAACGCAGCACTAGTGACGACGCTGGGT | Primer Rv to fuse promoter 3K to nqo 72 with site SpeI |
| sIFP_Rv | AAAGAATTCTTATTGTAGAGCTCATCCATGC | Primer Rv to amplify sIFP gene from pMoT with site EcoRI |
| TS_Parental | AAATCTAGACCTCCTTCAAAAAAGAAGGAGGATATACATATGAGCAAAGGA GAAGAACTTTTCA | Primer Fw to amplify sIFP gene adding thermosensor Parental (sites XbaI, EcoRI) |
| TS_Var_0 | AAATCTAGAACTCCTTCAAAAAAGAAGGAGGATATACATATGAGCAAAGGA GAAGAACTTTTCA | Primer Fw to amplify sIFP gene adding thermosensor ts0 |
| TS_Var_1 | AAATCTAGACCTCCTTAAACAAAAAGTTAAGGAGGATATACATATGAGCAA GGAGAACTTTTCA | Primer Fw to amplify sIFP gene adding thermosensor ts1 |
| TS_Var_2 | AAATCTAGACCTCCTTAGCAAAAAAGCTAAGGAGGATATACATATGAGCAA AGGAGAAGAACTTTTCA | Primer Fw to amplify sIFP gene adding thermosensor ts2 |
| TS_Var_4 | AAATCTAGACCTCCTTACGCAAAAAAGCGTAAGGAGGATATACATATGAGC AAAGGAGAAGAACTTTTCA | Primer Fw to amplify sIFP gene adding thermosensor ts4 |
| TS_Var_6 | AAATCTAGACCTCCTTACGCGCAAAAAAGCGCTAAGGAGGATATACATAT GAGCAAAGGAGAAGAACTTTTCA | Primer Fw to amplify sIFP gene adding thermosensor ts6 |
| TS_Var_8 | AAATCTAGACCTCCTTAAAGCAAAAAAGCTAAGGAGGATATACATATGAGC AAAGGAGAAGAACTTTTCA | Primer Fw to amplify sIFP gene adding thermosensor ts8 |
| TS_Var_10 | AAATCTAGACCTCCTTCAAAAAAAAAAAAAAAAAAGAAGGAGGATATACATA TGAGCAAAGGAGAAGAACTTT | Primer Fw to amplify sIFP gene adding thermosensor ts10 |
| TS_Var_PolyA | AAATCTAGAAAAAAACAAAAAGAAGGAGGATATACATATGAGCAAAGG AGAAGAACTTTTCA | Primer Fw to amplify sIFP gene adding thermosensor PA |
| TS_Var_ContC | AAATCTAGAAAGGAGGATATACATATGAGCAAAGGAGAAGAACTTTTCA | Primer Fw to amplify sIFP gene adding thermosensor ContC |
| Bgal_TS8_Fw | AAATCTAGACCTCCTTAAAGCAAAAAAGCTAAGGAGGATATACATATGTCGA CCGTTTGTACTACC | Primer Fw to amplify beta-galactosidase gene adding thermosensor ts8 |
| Bgal_EcoRI_Rv | TTTTGAATTCTCATGTCTCCTCCACACGGC | Primer Rv to amplify beta-galactosidase gene with site EcoRI |

Table S4. List of component sequences

Relevant restriction sites are in italics and underlined. Relevant ATGs and thermosensors in bold.

> Tth origin of replication derived from pMY1

*GCGGCCGCTTGACGTCAGCGCAGAAGTGGTCAGCTTGCATGCCTGTTGGCATGCCTCAGTTGA
CCCCATTGACCCTTCTCTCTGGAGGTGGTAGCTAGAGGGGAATGGTCCCACCCTGGAGCCCCA
CCCCTTGCCAAACCCTGGGCCCGGTATAATGCCGGGCAATGGGAAGACCGGGCCCCCTGGGGG
CCCAAGGCGGCAGGGCCGCGTGAGGACCAAAAAGAGAGGCTCCCGCTGGGAGGGAACCCAA
ATTCCAGCAACCTTTTTCTTCGCCTAAGGTTATCCCCTCCCAGCGGGAAAGGCAATAGGCGCCA
AGGCGGCGCGGGGGAGTGCCGTCTCCGACGCGGAAACGGGAGGGGTTTATGCCCAAG
AAGGAGTTGAAGGGAAGAGCTATAAAGAACAGCCGGAAGGACTACCGAGCGGGGCCGGG
CCTTCCTCGAGGCCCTGGCCCGGAGGAAGCGGGCGAGGGGCGAGGAGCTTCCGCCCTCTAC
CTGAAGCTCCTGGAGGGGGCCGCGCCCCCGGAGAAGGGGGTCCATGAGCCCCCGCCGAGG
AGGCGCCTCCCCGGAGGACCTCCGCCGGAAGCTCCGGGAGGCGCCGCTTCCCCGGCCCTC
CCCAACCGACAGGAGCTCTCCGCCTCCCCGCCCCCGCCGAGATGAGGCGGGACGCCTGGAG
CCTCGCCGACCGCCTCCTGGAGGAGGGGGAGCGGCGGGGCACCCTGCCAGGGCTTCCGAG
CGGGAGCGGAGGGTGTACCGGACCCTCCTCGCCCTGGGCCTCGAGGTCCTGGCCCGGAGGCT
GGGCCCGGGGAGGCCCTGCCAGGAACCTGTCCAGGTCTCCTTCTCGCCGTGAACGACG*

CCCTGGCCGTGGCCCTGGGAATCCCCCGGCCTCCCTCTACCGGGTCTGGCCTCCCTGGAGG
CCAAGGGGCTCATCCGCCGGAGGGCTGGCGCACCCCGGCCACCCTCAAGGGCCGGACGGG
GGTCTACGCCGGCGGGACCCTCTACGCCGTGCGCCTGCCCCACCGGGAGGCCCGCCCCGCC
TGGACCCGGAGGACTTCCGCCACCCCTGGCGGGACCTGGAGGGGGACGCCCGTCAGGGGCG
CACCGCCTGGAGCTTGAGAGAGTCATATACAAGTCCTCCTAAGGAGGACTCCGGGGTCTCCA
GCTCCTCCTTCGGTTTTCGTTATCCCCTGGCGAAGCCGAAACTCCGTTAGCTTTAGACTCTCTCA
CCGCCCTCTCCGGGCCCGCCCCGCCGAGCGCCGGGCCCTGGTGGAGGCCCTCGCCCTCTCCC
TGGCCCCGGGAGTTCCGGGATCGCGGGAGCGTGCCTTCTACGCCTGGGTCTCTGGAACGCC
CTCCGGGCCGAGCTCTACGGCCTGATGGAGGGGGCCCTCGAGGCCGTGCGCTGGGCGGTCC
GGCGGGCGCGGGAGGCCGCGGCCAAGTCTCTGGAGCCCGAGGGGCGAGGGGGTCCGGC
GCCCGGGGCCCTCCTCGCCACCTCCTCCGGGAGCGGGCCCTCCTGGAGCTTCCGCCAGG
CCCCTCAGTGGCGGGTGGCGTAGGGCTCCCGCCGGGCTAAGCTGGAGGTAGCCCATCCAGG
GCCTCTGGGGGGTGGCTTGTGTATCAAGACTGGCTGTTCAAGGACTGCCAGGGGGCTGGCG
CCTCCGCTACCGCCGAACGGGGCGGCTACGAGGCCTCCCGGGACGGTGGAGAGGTGGGAG
AAGTCCCCTCGTACGGAGGTTACGGGGCGCCTGAACAAGAACCTTCAGGACTGGGCCGT
GCGCCAGGTGGTGGAGTACCTCCAGGGGGAGCTGGTCCCCCGGAGATCCCCGGGGGAGTA
TAACCTGCTGCAGTTAATTAA

> Tth origin of replication derived from pTT8

GCGGCCGCTTGACGCTACCGCTAGGGACCCCGGGGGGTGAGCGCCGCTCAGCGCCCGGG
GTGAGCGGTGAGCGCTCACCTGAGCGCCTGGGTAGACTGGGGTCTGGAGCACCCGGAAG
GGGAAGACCCGACCCTCCTGCCCCGGCCTTGGCCGCCCGGGCCCTGGGGGTCTCCCCGCC
ACCCTCCGGCGCTACGCCGCCCTGTGGGAGCGGTTGGTAGGTCCCCTGCCCGGGATCCCCG
GGGGGGACGGCTCTGGCCCAAGGAGGCCCTGGCCCGCCTCCGGGCGGCCCGGGAGGCCAC
CTGCGGGAAGGCCTCCCCCTGGAGGAGGCCCTGGCCCGGTCCAGGGAGACTTCCCGGCCCT
CGCCCTCCCCTCCGAGGGGGAGGCCCTGGCCCTCCTCCGGGCCCTGGCCGAGCGGCTGGAGC
GGGTGGAGGGGGAGCTAAGGGCCCTGCGGGAGGAGAACGCCCGCCTCCGGGAGGCCCTGA
AGGCCCTGGAGCCCCCGGGAGGCGGCCCTGGTGGCGGTTCTGGGGGCATTGACCTGGGGA
CTACCGCTTGGTCCCGGGGCTTGTGCTAGAAATGGGGGGCATGAATGAAACAGACCTCGGCC
GGGTTAACCGCCGAGGCCTCTCCTCCCAGGAGGCCTCCATCCTAACCCGGCCGGGGGGAAG
GAGGCAAGAGATGGAGCACCAACCCGGCCGGAAGCGCAAATCCCGGACACCCTCGCTAAA
ATCGCAGGACTCTTCCAGATAAACCCCGACCTGGGGGAGGTGGTTCTTCGCGCCTACGCCGCC
CTGCGCGCCTCTCCCCGGAGGCCCTCCGCGCCACCTCCTGGCCCTCCCCTCCGCCCGGAG
CGGGCCCGGGAGGCCTTCCAGCGGCCCTACCTCGCCACTTCGCCCAGACTCTCCCCGCTAC
CCCTACGCCACGGACGACCCCAAGGAGGGGGTGCATCTACAAGCGGGAGAACGCCCTGA
AGCGGGTCCACGTCCAGGTGGGCCACTACCCACGCCGTCTTGGCGGTGGTGGTGGACGTG
GACCTCCCCTGGCCCCAGGTGGAGGAGCGGATCCACGCCCTCCCCCTCCCTGGTCTGGTC
AACCCGAGATCGGGCCACTTCCACGCCTGGTACGAGCTGGACCCCATCCCCCTCACGCCCCG
CCCGGGCGGGAGGGGAGCCTGAAGGGGGCCCTGGCCCTTCTCGCGGAGGTGGAGGCCCTGC
TGGAGGCCTACTACGGGGCGGACCCGGGCTACAACGGTCTCCTCTCCCGAAACCCCTTCTCC
ACCCCGGAGTGGACCTGGGGCGGGGGGAAGCGGTGGAGCCTGCGGGACCTCCACCGGG
AGCTCCGGGGGCTCCTTCCCTCCGGGACCCGGAGGCGGGTGGACCCGGGCCTGGCCTCTAC
GGGCGGAACAACGCCCTGTTTGACCGCCTGCGGGCGGAGGCCTACGCCACGTGGCCCTCT
CCGGGGCGTCCCCGGGGGGGAGGAGGCCTTCCGGGCCTGGGTGGAGCAGAGGGCCACGC
CCTGAACCAGTCCCTTCCGGGACCCCAAGGGGCCCTTGACCCCGGGAGGTCCACCA
CACGGCGAAGAGCGTGGCCAAGTGGACCTACCGGAACTACCGGGGGGGCGAGGGTCTACCCG
GTCTCCTCACGGGGAGGCCGGACCGGAGCCGCCTCTCCCCAGGCCCGGGCCCTGATCCC

GCCCCTCCAGGGCCAGGAGCTCCAGGAGGCGGTGCGGGAGGGCGGAAGGCGGCGGGATC
CCGGCGCAGGCAGGAGGCGGAGGAGAAAGCTCACGGAGGCCCTGAAGCGCCTCCAGGCCCGG
GGGGAGCGGGTCACGGCCAGGGCCCTGGCCCGGAGGCGGGGGTCAAGCCCCATACCGCCT
CCAAGTGGTTGAAGAGGATGCGGGAGTAGCGTCCAGGACGGCACAACCCCAAAAATGTGC
CCCAAGCTGGTGGCTATCAGGGTATACAGGCGGGGGTGCAGGGGGTGAACCCCGCCAGCC
CCGAAGGGGCTGTATTGGAGAACCTAGAGAGTCTCCTTAGCGTAGTTGACGGCTAAAGTCCC
CCTTCTTTTTCTGGAAGCCGCAAAAATTTTGTTCCTTGGGGTAGACACCTTGAGATCCGAG
AGAAAGTGAGATAACTCACATATTTACCCCGCCCGCACGCAATTTTCATCGTCCAGGACGGCA
AAAACACTGATTTTCCCCCTCGAGGACCGCCCCTAAAACGGCTTTTAGGCGGGGGGGTGGGCG
GCGGGACCAATCCACCCTCCCCCGCCCCCGCCTTCTAGGGGCCTTAGGACGGGTGAGA
GGGTCTTCTGGGGCAGGGTCCGGAGCCCGCTCTGGCCCGGGGGCTGGACCGGCCCTCG
GCGCTCAAGCGTAGACAGGGAGACCCCTCCATAGCCTGCAGTTAATTAA

> Ec replication origin derived from pUC

AAGCTTAGCCGGAAGCATAAAGTGTAAGCCTGGGGTGCCTAATGAGTGAGCTAACTCACAT
TAATTGCGTTGCGCTACTGCCCGCTTCCAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATG
AATCGGCAACGCGCGGGGAGAGGCGGTTTGCATTTGGGCGCTCTCCGCTTCCCTCGCTCAC
TGACTCGCTGCGCTCGGTGTTCCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAA
TACGGTTATCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAGGCCAGCA
AAAGGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCTCCGCCCCCTGA
CGAGCATCACAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGACTATAAAGAT
ACCAGGCGTTTTCCCCCTGGAAGCTCCCTCGTGCCTCTCCTGTTCCGACCCTGCCGTTACCGG
ATACCTGTCCGCTTTCTCCCTTCGGGAAGCGTGGCGCTTTCTCAAAGCTCACGCTGTAGGTAT
CTCAGTTCGGTGTAGGTGCTTCGCTCCAAGCTGGGCTGTGTGCACGAACCCCGTTCAGCCC
GACCGCTGCGCCTTATCCGTAACACTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCG
CCACTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAG
AGTTCTGAAGTGGTGGCCTAACTACGGCTACACTAGAAGAACAGTATTTGGTATCTGCGCTC
TGCTGAAGCCAGTTACCTTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCACCG
CTGGTAGCGGTGGTTTTTTTTGTTTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAG
AAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGAAAACACTCACGTTAAGGGA
TTTTGGTCATGAGATTACAAAAGGATCTTACCTAGATCCTTTTAAATTAATAAAGTTT
TAAATCAATCTAAAGTATATATGAGTAAACTGCGGCCGCTTGACGTC

> E.c. replication origin derived from pBBR (pSEVA1311)

AAGCTTAGGCCACCCGCGAGCGGGTGTTCCTTCTCACTGTCCCTTATTCGCACCTGGCGGTG
CTCAACGGGAATCCTGCTCTGCGAGGCTGGCCGTAGGCCGGCCCTACCGGCGCGGCAGCGTT
ACCCGTGTCGGCGGCTCCAACGGCTCGCCATCGTCCAGAAAACACGGCTCATCGGGCATCGG
CAGGCGCTGCTGCCCGCGCCGTTCCATTCTCCGTTTCGGTCAAGGCTGGCAGGTCTGGTTC
CATGCCCGAATGCCGGGCTGGCTGGGCGGCTCCTCGCCGGGGCCGGTCCGGTAGTTGCTGCT
CGCCCGGATACAGGGTCCGGATGCGGCGCAGGTGCCATGCCCAACAGCGATTCTGCTCTGG
TCGTGCTGATCAACCACCGGCGGCACTGAACACCGACAGGCGCAACTGGTCCGCGGGGCTG
GCCCCACGCCACGCGGTCAATTGACCACGTAGGCCGACACGGTGCCGGGGCCGTTGAGCTTCA
CGACGGAGATCCAGCGCTCGGCCACCAAGTCCTTACTGCGTATTGGACCGTCCGCAAAGAA
CGTCCGATGAGCTTGAAAGTGTCTTCTGGCTGACCACCGGCGTTCTGGTGGCCCATCTGC
GCCACGAGGTGATGCAGCAGCATTGCCGCGTGGGTTTCTCGCAATAAGCCCGGCCACGC
CTCATGCGCTTTGCGTTCCGTTTGCACCCAGTGACCGGGCTTGTTCCTGGCTTGAATGCCGATT
TCTCTGGACTGCGTGGCCATGCTTATCTCCATGCGGTAGGGGTGCCGCACGGTTGCGGCACCA

TGCGCAATCAGCTGCAACTTTTCGGCAGCGGACAACAATTATGCGTTGCGTAAAAGTGGCAG
TCAATTACAGATTTTTCTTTAACCTACGCAATGAGCTATTGCGGGGGGTGCCGCAATGAGCTGT
TGCGTACCCCCCTTTTTAAGTTGTTGATTTTTAAGTCTTTCGCATTTCCGCCTATATCTAGTTCT
TTGGTGCCCAAAGAAGGGCACCCCTGCGGGGTTCCCCACGCCTTCGGCGCGGCTCCCCCTCC
GGCAAAAAGTGGCCCCTCCGGGGCTTGTGATCGACTGCGCGGCCTTCGGCCTTGCCCAAGG
TGCGCTGCCCCCTTGAACCCCGCACTCGCCGCGTGAGGCTCGGGGGGCAGGCGGGCG
GGCTTCGCCCTTCGACTGCCCCACTCGCATAGGCTTGGGTCGTTCCAGGCGCGTCAAGGCCA
AGCCGCTGCGCGGTGCTGCGCGAGCCTTGACCCGCTTCCACTTGGTGTCCAACCGGCAAGC
GAAGCGCGCAGGCCGAGGCCGAGGCTTTTTCCCCAGAGAAAATTAATAAAAATTGATGGGG
CAAGGCCGAGGCCGCGCAGTTGGAGCCGGTGGGTATGTGGTGAAGGCTGGGTAGCCGGT
GGGCAATCCCTGTGGTCAAGCTCGTGGGCAGGCGCAGCCTGTCCATCAGCTTGTCCAGCAGG
GTTGTCCACGGGCCGAGCGAAGCGAGCCAGCCGGTGGCCGCTGCGGCCATCGTCCACATAT
CCACGGGCTGGCAAGGGAGCGCAGCGACCGCGCAGGGCGAAGCCGGAGAGCAAGCCCGT
AGGGGGGGCGGCCGCTTGACGTC

> SlpA promoter - Kanamycin resistance gene - SlpA terminator

CTGCAGTTAATTAAGAAACCTTAAGGCCGACCGTTTGACAAGGGCGCGTGAGGTTTTTACG
ATAGCGCCGGATGCGGGGAAAAGGGCTCCTTTTGGGGGGTTTTCCCCGCACCGGGCGGACC
TGGGCGGAGAGGAAACGCGGCAACTCGCCGTCTCGGGTCCC GCCACGACCCTTAAGGAG
GTGTGAGGATT**ATG**AATGGACCAATAATAATGACTAGAGAAGAAAGAATGAAGATTGTTCA
GAAATTAAGGAACGAATATTGGATAAATATGGGGATGATGTTAAGGCTATTGGTGTATGG
CTCTCTTGGTGCAGACTGATGGGCCCTATTCGGATATTGAGATGATGTGTGCATGTCAAC
AGAGGAAGCAGAGTTCAGCCATGAATGGACAACCGGTGAGTGGAAGGTGGAAGTGAATTT
TATAGCGAAGAGATTCTACTAGATTATGCATCTCAGGTGGAATCAGATTGGCCGCTTACACAT
GGTCAATTTTTCTCTATTTTGCCGATTTATGATTCAGGTGGATACTTAGAGAAAGTGTATCAA
CTGCTAAATCGGTAGAAAGCCAAAAGTTCCACGATGCGATTTGTGCCCTTATCGTAGAAGAGC
TGTTTGAATATGCAGGCAAATGGCGTAATATTCGTGTGCAAGGACCGACAACATTTCTACCAT
CCTTGACTGTACAGGTAGCAATGGCAGGTGCCATGTTGATTGGTCTGCATCATCGCATCTGTT
ATACGACGAGCGCTTCGGTCTTAAGCAAGCAGTTAAGCAATCAGATCTTCCCTCAGGTTATG
ACCATCTGTGCCAGTTCGTAATGTCTGGTCAACTTCCGACTCTGAGAACTTCTGGAATCGCT
AGAGAAATTTCTGGAATGGGATTGAGGAGTGACAGAACGACACGGATATATAGTGGATGTGT
CAAACGCATACCATTTTGAACGGAATTTATGCGGTGTGAAATACCGCACAGATGCGTAAGG
AGAAAATACCGCATCAGGCGCCATTGCCATTGAGGCTGCGCAACTGTTGGGAAGGGCGATC
GGTGCGGCCCTTTCGCTATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAA
GTTGGGTAACGCCAGGGTTTTCCAGTCACGACGTGGCGCGCTGTAGCTAGTAAAGTTCTAA
CCCCTGGCGGTCCC GCCCCCGCCCCCTTTGGGGGGGCGGGGGGTTTTTGTCTCCGCTAGC

> SlpA promoter - Hygromycin resistance gene - SlpA terminator

CTGCAGTTAATTAAGAAACCTTAAGGCCGACCGTTTGACAAGGGCGCGTGAGGTTTTTACG
ATAGCGCCGGATGCGGGGAAAAGGGCTCCTTTTGGGGGGTTTTCCCCGCACCGGGCGGACC
TGGGCGGAGAGGAAACGCGGCAACTCGCCGTCTCGGGTCCC GCCACGACCCTTAAGGAG
GTGTGAGGATT**ATG**AAAAAGCCTGAACTACCGCGACGTCTGTTGAGAAGTTTCTGATCGAA
AAGTTCGGCAGCGTCTCCGACCTGATGCAGCTCTCGGAGGGCGAAGAATCTCGTGCTTTCAGC
TTCGATGTAGGAGGGCGTGGATATGTCTGCGGGTAAATAGCTGCGCCGATGGTTTCTACAA
AGATCATTATGTTTATCGGCACTTTGCATCGGCCGCGCTCCCGATTCCGGAAGTGCTTGACATT
GGGGAATTTAGCGAGGGCCTGACCTATTGCATCTCCCGCGTGCACCGGGTGTACGTTGCAA
GACCTGCCTGAAACCGAACTGCCCGCTGTTCTGCAACCGGTGCGGAGGTTATGGATGCGATC

GCTGCGCCGATCTTAGCCAGACGAGCGGGTTCGGCCCATTCGGACCGCAAGGAATCGGTCA
ATACACTACATGGCGTGATTTTCATCTGCGCGATTGCTGATCCCCATGTGTATCACTGGCAA
GTGATGGACGACACCGTCAGTGCCTCGTCGCGCAGGCTCTCGATGAGCTGATGCTTTGGG
CGAGGACTGCCCCGAAGTCCGGCACCTCGTGCACGCGGATTCGGCTCCAACAATGTCCTGAC
GGACAATGGCCGCATAACAGCGGTCAATTGACTGGAGCGAGGCGATGTTTCGGGGATCCCCTAT
ACGAGGTCGCCAACATCTTCTTCTGGAGGCCGTGGTTGGCTTGTATGGAGCAGCAGGCGCGC
TACTTCGAGCGGAGGCATCCGGAGCTTGCAGGATCGCCGCGGCTCCGGGCGTATATGCTCCG
CATTGGTCTTGACCAACTCTATCAGAGCTTGGTTGACGGCAATTCGATGATGCAGCATGGGC
GCAGGGTTCGATGCGACGCAATCGTCCGATCCGGAGCCGGGACTGTCGGGCGTACACAAATCG
CCCGCAGAAGCGCGGCCGTCTGGACCGATGGCTGTGAAGAAGTACTCGCCGATAGTGAAAC
CGACGCCCCAGCACTCGTCCGAGGGCAAAGGAATAGAGATGGCGCGCTGTAGCTAGTAAAG
TTCTAACCCCTGGCGGTCCCGCCCCCGCCCCCTTGGGGGGGCGGGGGGTTTTTGTCTCCCG
CTAGCC

> SlpA promoter - Bleomycin resistance gene - SlpA terminator

CTGCAGTTAATTAAAGAAACCTTAAGCCCCGACCGTTTGACAAGGGCGCGTGAGTTTTTACG
ATAGCGCCGGATGCGGGGAAAAGGGCTCCTTTTGGGGGGTTTTCCCGCACCGGGCGGACC
TGGGCGGAGAGGAAACGCGGCAACTCGCCGTCTCGGGTCCCGCCCACGACCCTTAAGGAG
GTGTGAGGATT**ATG**GCCAAGTTGACCAGTGCCGTTCCGGTGCTCACCGCGCGCAGCTCGCC
GGAGCGGTGAGTTCTGGACCGACCGGCTCGGGTCTCCCGGGACTTCGTGGAGGACGACTT
CGCCGGTGTGGTCCGGGACGACGTGACCCTGTTTCATCAGCGCGGTCCAGGACCAGGTGGTGC
CGGACAACACCCAGGCCTGGGTGTGGGTGCGCGGCCTGGACGAGCTGTACGCCGAGTGGTC
GGAGGTCGTGTCCACGAACCTCCGGGACGCCTCCGGGCCGGCCATGACCGAGATCGTCGAGC
AGCCGTGGGGGCGGGAGTTCCGCCCTGCGCGACCCGGCCGGCAACTGCGTGCCTTCGTGGCC
GAGGAGCAGGACTAATCTAGGCGCGCTGTAGCTAGTAAAGTTCTAACCCCTGGCGGTCCCG
CCCCGCCCCCTTGGGGGGGCGGGGGGTTTTTGTCTCCCG**GCTAGCT**

> SlpA promoter - RBS - ATG

CTGCAGTTAATTAAAGAAACCTTAAGCCCCGACCGTTTGACAAGGGCGCGTGAGTTTTTACG
ATAGCGCCGGATGCGGGGAAAAGGGCTCCTTTTGGGGGGTTTTCCCGCACCGGGCGGACC
TGGGCGGAGAGGAAACGCGGCAACTCGCCGTCTCGGGTCCCGCCCACGACCCTTAAGGAG
GTGTGAGGATT**ATG**

> SlpA terminator

GGCGCGCTGTAGCTAGTAAAGTTCTAACCCCTGGCGGTCCCGCCCCCGCCCCCTTGGGGGG
GCGGGGGGTTTTTGTCTCCCG**GCTAGC**

> Pnqo

GCTAGCTCCAGGGCCTTCTTCCGCCCCCAAGGTGTGGAGCAGCCTGGTGCGCCTCACCC
CCACGGGCGCCCCGGACGACCCCGCCTCTCCGCCTCGTTGAGGCCGCTTCGGAAAGCGG
CGGAAGACCCTCTTAAACGCCCTGGCCGCGGCCGGCTACCCCAAGGCGCGGGTGGAGGAGG
CCCTGAGGGCCTTGGGCCTTCCCCTAGGGTGCGGGCCGAGGAGCTGGACCTCGAGGCCTTC
CGCCGGCTGAGGGAGGGCCTCGAGGGGGCGGTGTAGGCCCGCCTCCCTTCCCTACACCGTTC
CTTCTCCTTCTGTGTAGCCACCCCTTGCGCCACCCCGGGGTGATAAGATGGGCAGGAA
ATGAGGCCGGCCCCCTTGGGGCTTGGGGACCGGTTTCGTGAAGAAGAATTCTTCGTGCACG
AAAGGAGGGG**CATATG**

> Pnqo3A, deleted region indicated

GCTAGCCTCCAGGGCCTTCTTTCCGCCCCCAAGGTGTGGAGCAGCCTGGTGCGCCTCACCC
CCACGGGCGCCCCGGACGACCCCGCCTCTCCGCCTCGTTGAGGCCGCCTTCGAAAGCGG
CGGAAGACCCTCTTAAACGCCCTGGCCGCGGCCGGCTACCCCAAGGGCGGGGTGGAGGAGG
CCCTGAGGGCCTTGGGCCTTCCCCCTAGGGTGGGGCCGAGGAGCTGGACCTCGAGGCCTTC
CGCCGGCTGAGGGAGGGCCTCGAGGGGGCGGTGTAGGCCCGCCTCCCTTCCCTACACCGTTC
CTTCTCCTTCTGTGTAGCCCACCCCTTGCGCCACCCGGGGTGATAAGATGGGCAGGAA
ATGAGGCCGGCCCTTTGGGGCTTTGGGGACCGGTTTCGTGAAGAAGAATTCTTTCGTGCACG
AAAGGAGGGGCATATG

> IFP gene

CATATGAGCAAAGGAGAAGAAGCTTTTCACTGGAGTTGTCCCAATTCTTGTTGAATTAGATGGT
GATGTTAATGGGCACAAATTTCTGTCCGTGGAGAGGGTGAAGGTGATGCTACAAACGGAAA
ACTCACCTTAAATTTATTTGCACTACTGAAAACACTGTTCCGTGGCCAACACTTGTCACTA
CTCTGACCTATGGTCTTATGTGCTTTTCCGTTATCCGGATCACATGAAACGGCATGACTTTTTC
AAGAGTGCCATGCCCGAAGGTTATGTACAGGAACGCACTATATCTTTCAAAGATGACGGGAC
CTACAAGACGCGTGCTGAAGTCAAGTTTGAAGGTGATACCCTTGTTAATCGTATCGAGTTAAA
GGGTATTGATTTAAAGAAGATGGAAACATTCTTGACACAAACTCGAGTACAACCTTTAACTC
ACACAATGTATACATCACGGCAGACAAACAAAAGAATGGAATCAAAGCTAACTCAAATTCG
CCACAACGTTGAAGATGGTTCCGTTCACTAGCAGACCATTATCAACAAAATACTCCAATTGG
CGATGGCCCTGTCTTTTACCAGACAACCATTACCTGTCGACACAATCTGTCCTTTCGAAAGAT
CCCAACGAAAAGCGTGACCACATGGTCTTCTTGTAGTTTGTAACTGCTGCTGGGATTACACAT
GGCATGGATGAGCTCTACAATAATAAAAGCTT

> Plac

GCTAGCACGACAGGTTTCCCGACTGGAAAGCGGGCAGTGAGCGCAACGCAATTAATGTGAGT
TAGCTCACTCATTAGGCACCCCAGGCTTTACACTTTATGCTTCCGGCTCGTATGTTGTGTGGAA
TTGTGAGAATTCTTTCGTGCACGAAAGGAGGGGCATATG

> P3K

GCTAGCGTTTAAACTCTTACAGCTTGACGGCGTTTTTGTCTTGAGCGATTCAGGGACGTTTCGC
CCAGTAGGTGGTTTTTCGAGCCGACTGCGTGGTCAACCTGCACCTCGTACCAGCGCC
GAGCTTGAGCTGACGCCCGCGGTACATGATGACCTTGATGCGGTCGGtCTGCTGGACCA
GCACGTTGTCGTTGTTCCCTTGATCGAGGTGGTATTGAACTCCTCACCCAGCGTCGTCTTGTC
AGCTGCGTTGGCGTGCGTTAGGGTGGTGGCTTTGTCCGGGCCGATCCACTGGATCGTGGATT
TGCTGAGGTTGATGTTGTCGGCTCCGGCGGCTGGCGCACGGTGAGGTTACGTAAGTCGACT
CTAGAGAATTCTTTCGTGCACGAAAGGAGGGGCATATG

> P16S

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GCCTTGACAAAAGGAGGGGGATTGATAGCATGGCTTTCTGAATTCTTTCGTGCACGAAAG
GAGGGGCATATG

> Pnar

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AGGGCGAACCAGGGCCTTTCCAGGAAGGCCTCCCGGTGGGGGCCGTCGTAAGCGGTGAGGA

AGAGGTGCCGTCCTCAGGGTCGGCGAGGAAGAGCTCCGCCGCCTCCATGCATGGCGAGGG
CCTCCCGGAGGGCCCGGAGGAAGTCTGGACTCCTTAGGCGAAGGCCTCCGGGTGCCGGAG
CAGGTGCTGGGTGAGGTAGGAGGCTTCCCGCAGGAGGTCGGGCCCTTGGTCCGCTCGGGG
TAGAGCTCCACCAAGAACCCTTCCCCCTCCCGGTAGGCCTGGCACCGGAGCCTGCGCCCGTTC
ACCACGAGGGGCGTGCTGGCGCGGTAGGCGCCCCGCTTCAGCTCCCGGAGCGCCGGGCACC
GGGCGCAAAGCGGCCGCGCAAGGGTCTTTCCCTGGACCAGGTGGGCGCAGGGCACGCC
CGCGGCGCGAAACCCAGGGAGGCTTCGGCCTCTACCTCCCGCACGAGGCCTTGACCGTCAA
GGCGGATCCGCGCCAGGGCTTCCATCCGCCCTCAGCCTAGGGGGGTTCCGGTAGGACAGATG
TCCCGGGGCTTGCCAGTGGGCGCGAGGAGGCCCGGGCACACTGGGGCCGGAGGTGACCATG
GGCAGCAGCCATCATCATCATCACAGCAGCGGCCTGGTGCCGCGCGGCAGCCATATG

> LinkerA

GCTAGCGAGATATCATCTAGAAAACATATGAGGTACCAGAATTCCTAGTGGATCCAAGCTT

> Linker1

CATATGGTTCGACAAGATATCAGGATCCATCTAGAAAGCTT

> Linker2

CATATGGTTCGACAAGATATCAGGATCCATCTAGAAAGCTT

> Linker3

CATATGGGTCGACAAGATATCAGGATCCATCTAGAAAGCTT

Pnqo variants

> Pnqo209

GCTAGGGTGCGGGCCGAGGAGCTGGACCTCGAGGCCTTCCGCCGGCTGAGGGAGGGCCTCG
AGGGGGCGGTGTAGGCCCGCCTCCCTTCCCTACACCGTTCTTCTCCTTCTGTGTAGCCCACCC
CCCTTGCGCCCCACCCGGGGTGATAAGATGGGCAGGAAATGAGGCCGGCCCTTTGGGGCT
TTGGGGACCGTTTTCGTGAAGAAGAATTCTTTCGTGCACGAAAGGAGGGGCATATG

> Pnqo91

GCTAGCCCCCTTGCGCCCCACCCGGGGTGATAAGATGGGCAGGAAATGAGGCCGGCCCTT
TGGGGCTTTGGGGACCGTTTTCGTGAAGAAGAATTCTTTCGTGCACGAAAGGAGGGGCATAT
G

> Pnqo72

GCTAGCACCCCGGGTGATAAGATGGGCAGGAAATGAGGCCGGCCCTTTGGGGCTTTGGG
GACCGTTTTCGTGAAGAAGAATTCTTTCGTGCACGAAAGGAGGGGCATATG

> Pnqo57

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GAATTCTTTCGTGCACGAAAGGAGGGGCATATG

> Pnqo33

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GGGCATATG

>3Knqo72

GCTAGCGTTTAAACTCTTACAGCTTGACGGCGTTTTTGTCTTGAGCGATTCAGGGACGTTTCGC
CCAGTAGGTGGTTTTTCGAGCCGTAAGTGCCTGACCTCGTCACCAGCGCC
GAGCTTGGAGCTGACGCCGCCGGCGTACATGATGACCTTGATGCGGTCTGCTGGACCA
GCACGTTGTCGTTGTTCCCTTGATCGAGGTGGTATTGAACTCCTCACCAGCGTCGTactagC
CCGGGGTGATAAGATGGGCAGGAAATGAGGCCGCCCTTTGGGGCTTTGGGGACCGTTTT
CGTGAAGAAGAATTCTTTTCGTGCACGAAAGGAGGGGCATATG

> Casette Hygromycin resistance gene - Thermosensor PA-sIFP- terminator

CTGCAGTTAATTAAGAAACCTTAAGGCCGACCGTTTTGACAAGGGCGCGTGAGGTTTTTACG
ATAGCGCCGGATGCGGGGAAAAGGGCTCCTTTTGGGGGTTTTCCCGCACCGGGCGGACC
TGGGCGGAGAGGAAACGCGGCAACTCGCCGTCTCGGGTCCCGCCCACGACCCTTAAGGAG
GTGTGAGGATTATGAAAAAGCCTGAACTCACCGCGACGTCTGTTGAGAAGTTTCTGATCGAA
AAGTTCGGCAGCGTCTCCGACCTGATGCAGCTCTCGGAGGGCGAAGAATCTCGTGCTTTCAGC
TTCGATGTAGGAGGGCGTGGATATGTCCTGCGGGTAAATAGCTGCGCCGATGGTTTTCTACAA
AGATCATTATGTTTATCGGCACTTTCATCGGCCGCGCTCCCGATTCCGGAAGTGCTTGACATT
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GACCTGCCTGAAACCGAACTGCCCGCTGTTCTGCAACCGGTCGCGGAGGTTATGGATCGCTGC
GGCCGATCTTAGCCAGACGAGCGGGTTCGGCCATTTCGGACCGCAAGGAATCGGTCAATACA
CTACATGGCGTGATTTTCATCTGCGCGATTGCTGATCCCCATGTGTATCACTGGCAAACGTGAT
GGACGACACCGTCAAGTGCCTCGCGCAGGCTCTCGATGAGCTGATGCTTTGGGTGAGG
ACTGCCCCGAAGTCCGGCACCTCGTGCACGCGGATTTCCGGTCCAACAATGTCCTGACGGACA
ATGGCCGCATAACAGCGGTCATTGACTGGAGCGAGGCGATGTTCCGGGACCCCTATACGAG
GTCGCCAACATCTTCTTGGAGGCCGTGGTTGGCTTGATGGAGCAGCAGGCGCGCTACTTC
GAGCGGAGGCATCCGGAGCTTGCAGGATCGCCGCGGCTCCGGGCGTATATGCTCCGCATTGG
TCTTGACCAACTCTATCAGAGCTTGGTTGACGGCAATTTTCGATGATGCAGCATGGGCGCAGGG
TCGATGCGACGCAATCGTCCGATCCGGAGCCGGGACTGTCGGGCGTACACAAATCGCCGCA
GAAGCGCGGCCGTGGACCGATGGCTGTGAAGAAGTACTCGCCGATAGTGGAACCGACG
CCCCAGCACTCGTCCGAGGGCAAAGGAATAGAGATCAGCTTGCATGCCTGCAGGTGCAGCTCT
AGAAAAAAAAACAAGGAAGGAGGATATACATATGAGCAAAGGAGAAGAACTTTTCACT
GGAGTTGTCCCAATTCTTGTGAATTAGATGGTGATGTTAATGGGCACAAATTTTCTGTCCGTG
GAGAGGGTGAAGGTGATGCTACAAACGGAAAACCTCACCTTAAATTTATTTGCACTACTGGAA
AACTACCTGTTCCGTGGCCAACACTTGTCACTACTCTGACCTATGGTCTTATGTGCTTTTCCCGT
TATCCGGATCACATGAAACGGCATGACTTTTTCAAGAGTGCCATGCCGAAGGTTATGTACAG
GAACGCACTATATCTTTCAAAGATGACGGGACCTACAAGACGCGTGCTGAAGTCAAGTTTGAA
GGTGATAACCCTGTTAATCGTATCGAGTTAAAGGGTATTGATTTTAAAGAAGATGGAAACATT
CTTGGACACAACTCGAGTACAACCTTAACTCACACAATGTATACATCACGGCAGACAAACAA
AGAATGGAATCAAAGCTAACTTCAAATTCGCCACAACGTTGAAGATGGTTCCGTTCAACTA
GCAGACCATTATCAACAAAATACTCCAATTGGCGATGGCCCTGTCCTTTTACCAGACAACCATT
ACCTGTGCACACAATCTGTCTTTGAAAGATCCCAACGAAAAGCGTGACCACATGGTCCTTCT
TGAGTTTGTAAGTCTGCTGGGATTACACATGGCATGGATGAGCTCTACAAATAAGAAATTCAG
GGTTTTCCAGTCACGACGTGGCGCGCCTGTAGCTAGTAAAGTTCTAACCCCTGGCGGTCCCG
CCCCGCCCTTTGGGGGGGGCGGGGGTTTTTGTCTCCCGCTAGC

Thermosensors

> DF

TCTAGACCTCCTTCAAAAAAGAAGGAGGATATACATATG

> PA

TCTAGAAAAAAACAAAAAGAAGGAGGATATACATATG

> TS0

TCTAGAACTCCTTCAAAAAAGAAGGAGGATATACATATG

> TS1

TCTAGACCTCCTTAACAAAAAGTTAAGGAGGATATACATATG

> TS2

TCTAGACCTCCTTAGCAAAAAAGCTAAGGAGGATATACATATG

> TS4

TCTAGACCTCCTTACGCAAAAAAGCGTAAGGAGGATATACATATG

> TS6

TCTAGACCTCCTTACGCGCAAAAAAGCGGTAAGGAGGATATACATATG

> TS8

TCTAGACCTCCTTAAAGCAAAAAAGCTAAGGAGGATATACATATG

> TS10

TCTAGACCTCCTTCAAAAAAAAAAAAAAAAAAGAAGGAGGATATACATATG

> TSP

TCTAGACCTCCTTCAAAAAAGAAGGAGGTACCAGATG

> beta-galactosidase gene

CATATGTCGACCGTTTGTACTACCCGGAGCATTGGCCTGAAGAACGTTGGGAAGAGGACTTT
AAGGCCATGCGGGCTCTGGGGCTCCGCTATGTGCGCCTGGGGGAGTTTGCCTGGAGCGCCCT
CGAGCCCACCCAGGTGCTCTCCGCTGGGGTTGGCTGGATCGGGTTCTGGATCTGGCGCAGA
AGGAGGGCCTAGCCGTAGTCCTCGGTACCCCCACCGCTACCCCCCAAGTGGTTGGTGGACC
GGTACCCGGAGATCCTCCCTGTGGACCGGGAGGGGCGGAGGCGGAACCTTTGGTGGGCGGCG
GCACTACTGCTTCTCCAGCCCCGCTACCGGGAAGAGACTGCCCGCATCGTGGCCCTTCTAGC
GGAGCGTTACGGCCGCCACCCGCGGTGGTAGGGTTCCAGGTGGACAACGAGTTTGGCTGTC
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TCTGGACTACTACGTTTTCGCATCAGACCAGGTGCGGGCCTACAATCGTTTCCAAGTGGACCT
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GACCCCTTTGCCTTGGCCGAGGACCTGGATTTT GCCGCTGGGACAGCTACCCCTTGGGCTTC
ACCGACCTGATGCCCTTCCCCAGGAGGAGAAGGTTTCAGTGGGCCCGCACGGGCCACCCCGA
TGTGGCCGCTTCCACCACGACCTTTACCGGGGGTAGGGCGGGGGCGTTTTTGGGTCATGG
AGCAGCAGCCGGGTCCCGTGAAC TGGCCCCCACAAATCCAAGCCCGGCGCCCGGGATGGTA
CGCCTTTGGACTTGGGAGGCCATAGCCCATGGGGCAGAGGTGGTTTCTACTTCCGCTGGCG
CCAAGCGCCCTTTGCCAAGAGCAGATGCAAGCGGGTTCAACCGTCCAGATTTCCAGCCGG
AAGTGGCCTTTTTTGAAGTGCAGCGGTAGCAGAGGAGCTTCCGCCCTCCCCCTTCTCCCG
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CCAAGGGGCCGAGTGGAAATACCTGACCCTGGTGTTCCTTTTATAGCGTTTTCCGGCGCCT
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GTAATTGTCGGACCCCGTTCGGGGAGCAAACGGAGAAATTCCAGATCCCCCGAAATCCCT
CCGGGCGCGCTCCAAGCCCTCCTTCCCCTTAAGGTAGTGCGGGTGGAGAGCCTGCCCCGGG
GCTTTTGGAGGAGCCGAAGGACCCTGGGGCCGCTTTCCTTCGGGGTTTGGCGGGAGTGG
GTGGAGACCGATCTTCCCCCTTGCTCCGTTTTACGGATGGTGGGGGAATCCTTTCCGCAGG
GGTCGCTACCTGTACCTGGCGGCTTGCCAGCCAGAACTTCTCTTTGCCCTTTGCCAGTCT
TGGCCGAGGAGGCGGGCTTGCATCCCCGCTTCTCCCTGAGGGCCTGCGTTTACGGAGACGG
GGCCCGTTGGTGTTCCTTTAACTATGGCCCCGAGGTGGTGGAGGCACCTGCCCTCCAGGG
GTGCGGTTTCTCTTGGGGGATAGGCGTATCCCCCATGACCTTGCCGTGTGGGAGGAGAC
ATGAGGCTTGTACTGGGCGGCTTGGAGGTGCCCTGAAGGCCAAGGGGTGGAGGTCTTGG
AAGACGGGGCCCTCCTATGGGGCTCAGAGGTACGGGTCCACGCTCCTTTTCGGGCAGAGGGA
CCTGCAGCCAAGCTT

> CrtB, phytoene synthase

CATATGAAAATGCCCGCAAGTATGGAGCCCGACTGGAAAGCCCTCCTCCGCGTCTCCGCGCC
CACTCCGCCACCTTCTACCTGGGAAGCCTCCTTCCCCAAGGAGGCCCGCAAGGGGGCCTGG
GCGGTCTACGCCGCTGCCGCTGGGGGACGAGGCGGTGGACGGGGAGGGCGGGGGCCCG
GAGGCCCTCGAGGCCTGGTGGGCGGGGGTGGAGCGGGCCTACCGGGGAAGGCCCTCGCC
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CTCCTCCGGTACTGCTACCAGGTGGCGGGCACCGTGGGGCGGATGATGGCCCCATCGCCGG
GGGCGGCAAGGAGGCGGAAGCCCGGGCGGTGAAGCTGGGGCAGGCCATGCAGCTACCAA
CATCCTCCGGGACGTGGGGGAGGACCTGGAGCGGGACCGGGTCTACCTGCCCTTGGACCTCC
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CTAGGCCACCTCAAGGTGGGCCGGGCGGCCATCGCCCTTGCCGCTTGCAGTACCGGGGAT
CCTGGACAAGCTCAGGCTTTCAGGCTACGACAACCTGGGAAGGCGGGCCACCTCAAGGCCT
GGGAACGGGCCCTCCTCCTCCCCAAGGCCTTCTCGCCGCCGCTTCCCCCAAGGCCGGAGG
GAAGCCCCTGAtgggcccccttctcgtctggcaccggggcgacctccgctccacgaccaccggccctctggaggcc
ctcggccggggccagtggtgggctcgtggtcctggacccaacaacctgaagaccaccccgaggcgggcctggttc
ct**AAGCTT**

> Alkaline phosphatase

CATATGAAGCGAAGGGACATCCTGAAAGGTGGCCTGGCTGCGGGGGCCCTGGCCCTCCTGCC
CCGGGGCCATACCCAGGGGGCTCTGCAGAACCAGCCTTCTTGGGAAGGCGGTACCGCAACC
TCATCGTCTTCGTCTACGACGGGTTTTCTGGGAGGACTACGCCATCGCCAGGCCTACGCC
GGAGGCGGCAGGGCCGGGTCTGGCCCTGGAGCGCCTCCTCGCCCGCTACCCCAACGGGCTC
ATCAACACCTACAGCCTCACCAGCTACGTACCGAGTCCAGCGCCGCGGGGAACGCCTTCTCC
TGCGGGGTGAAGACGGTGAACGGGGGGCTCGCCATCCACGCCGACGGGACCCCCCTCAAGC
CCTTCTTCGCCGCGGCCAAGGAGGCGGGGAAGGCCGTGGGGCTCGTGACCACCACCACCGTC
ACCCACGCCACCCAGCGAGCTTCGTGGTGTCCAATCCCGACCGGAACGCCGAGGAGAGGAT
CGCCGAGCAGTACCTGGAGTTCGGGGCCGAGGTGTACCTTGGGGGCGGGGACCGCTTTTTCA
ACCCCGCCAGGCGCAAGGACGGGAAGGACCTTACGCCGCTTCGCCGCAAGGGGTACGG
GGTGGTGCGCACCTCGAGGAGCTCGCCGTTCCAACGCCACCCGGCTCCTGGGCGTCTTCGC
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AGGAAATGGTCCAGGCCGCTTTGCCCGGCTTGGCGCCACCGCGGGGGCTTCGTCTTTCAG
GTGGAAGCGGGGCGGATTGACCACGCCAACCATTTGAACGACGCCGGGGCCACCCCTTGGGA
CGTGCTGGCGGCGGACGAGGTCTGGAGCTCCTACCGCCTTCGTGGACCGGAACCCGGACA

CCCTCCTCATCGTGGTCTCGGACCACGCCACCGGGGTAGGGGGGCTTTACGGGGCCGGGCGG
 AGCTACCTGGAGAGCTCCCCGAGGGGTGGACCTCCTGGAGCCGACGCGGGCGAGCTTTGAGC
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 ATGAAGGGGGTGGACCTCGAGGACGCCGAGGCGGAAAGGGTGGTGCGGGCCATCCGGGAG
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 CTCGTGGACAACACCCACGTCTCCGCCTCATGGGGGAGGCCCTTGGCCTCCGCTACCAGAAC
 CCGGTGATGAGCGAGGAGGAGGCCCTGGAGATCCTCAAGGCCAGGCCCCAGGGGATGCGCC
 ACCCCGAGGACGTCTGGGCCTAAgtcgacgcgtAAGCTT

> *Pseudomonas fluorescens* esterase I variant 34

CATATGAGCACATTTGTTGCAAAGACGGTACCCTGATCTATTTCAAGGACTGGGGCAGCGGT
 AAACCGGTGTTGTTCAGCCACGGTTGGCTACTGGATGCCGACATGTGGGAATACCAGATGGA
 GTACCTCAGCAGCCGCGGCTATCGCACCATCGCCTTTGACCGCCGCGGCTTTGGCCGCTCGGA
 CCAACCCTGGACCGGCAACGACTACGACACCTTCGCCGACGACATCGCCAGTTGATCGAACA
 CCTGGACCTCAAGGAGGTGACCCTGGTGGGCTTCTCCATGGGCGGCGGCATGTGGCCCGCT
 ACATCGCCCGCCACGGCAGCGCACGGGTGGCCGGCCTGGTGCTGCTGGGCGCCGTCACCCCG
 CTGTTCGGCCAGAAGCCCGACTATCCGCAGGGTGTCCCGCTCGATGTGTTGCAAGGTTCAAG
 ACTGAGCTGCTGAAGGATCGCGCGCAGTTTCATCAGCGATTTCAACGCACCGTTCTATGGC
 ATCAAGGGCCAGGTGCTCTCCAAGGCGTGCAGACCCAGACCCTGCAAATCGCCCTGCTGGC
 CTCGCTCAAGTCCACGGTGGATTGCGTCACCGCGTTCGCCGAAACCGACTTCCGCCCGGATAT
 GGCCAAGATCGACGTACCCACCTGGTATCCATGGCGATGGCGACCAGATCGTGCCGTTG
 AGACCACCGGCAAAGTGGCGGCGGAGTTGATCAAGGGCGCCGAACTGAAGGTGTACAAGGA
 CGCGCCCCACGGTTTCGCGGTGACCCACGCCAGCAGTTGAACGAAGACCTGTTGGCGTTCTT
 GAAACGCGGATCCCATCATCATCATCATTGACTGCAGCCAAGCTT

Table S5. References for component parts

pEM2S (pMY1)², pTT8⁴, kanamycin resistance gene⁶, pUC18, pSEVA131⁵,
 hygromycin resistance gene⁷(and Bosch, Berenguer and Hidalgo, submitted),
 bleomycin B resistance gene⁸, Pnqo⁹, PslpA¹⁰, Pnar¹¹, GFP from pMKnqosGFP³, IFP
 (Berenguer and Hidalgo unpublished), beta-galactosidase¹², CrtB (phytoene synthase)
¹³, alkaline phosphatase¹⁴, *Pseudomonas fluorescens* esterase I variety 34 (Mate,
 Berenguer, Hidalgo, submitted). The sIFP gene combines the mutations of the
 superfolder GFP (sGFP)¹⁵ with those of the Citrine version of GFP¹⁶.

Supporting Methods

Bacterial strains and growth conditions

Growth of *T. thermophilus* strains was carried out at the indicated temperatures (55 to 65°C) with rotational shaking (150 rpm) in TB media. *T. thermophilus* colonies were grown aerobically on TB agar (1.5 % w/v agar) and the plates were supplemented with the relevant antibiotics. For liquid or solid selection, kanamycin (final concentration, 30 mg L⁻¹) hygromycin B (100 mg L⁻¹) and/or bleomycin (15 mg L⁻¹) were included as indicated.

Escherichia coli strain DH5α was the host for plasmid construction and promoter testing. *E. coli* was grown at 37 °C in liquid or solid LB media, with kanamycin (30 mg L⁻¹), ampicillin (100 mg L⁻¹), hygromycin B (100 mg L⁻¹) or bleomycin (3 mg L⁻¹) added when required.

E. coli competence was induced following Inoue's method ¹⁷. Transformation of *T. thermophilus* was achieved by natural competence as described ¹⁸.

Plasmid Construction

The DNA sequence of the Promoter 3K was synthesized by Genscript. The nqo promoter mutants were constructed using the oligonucleotides designed in Oligonucleotides List as Fw primers and the sIFPRv as Rv primer and cloned between sites NheI and HindIII in pMoTK110. The linkers were constructed by hybridization of the corresponding pair of oligonucleotides and cloning between sites NheI and HindIII of pMoTK110 for linker A and between sites NdeI and HindIII for linkers 1, 2, and 3, and the 16S promoter was also generated by hybridization and cloning of the corresponding phosphorylated oligonucleotides between sites NheI and EcoRI of pMoTK110 plasmid. All the thermosensor bearing genes were cloned into vector

pMoTH150 (see sequence and map below, Scheme S1) as XbaI-EcoRI fragments, that include the IFP gene, obtained by PCR performed with primers indicated in

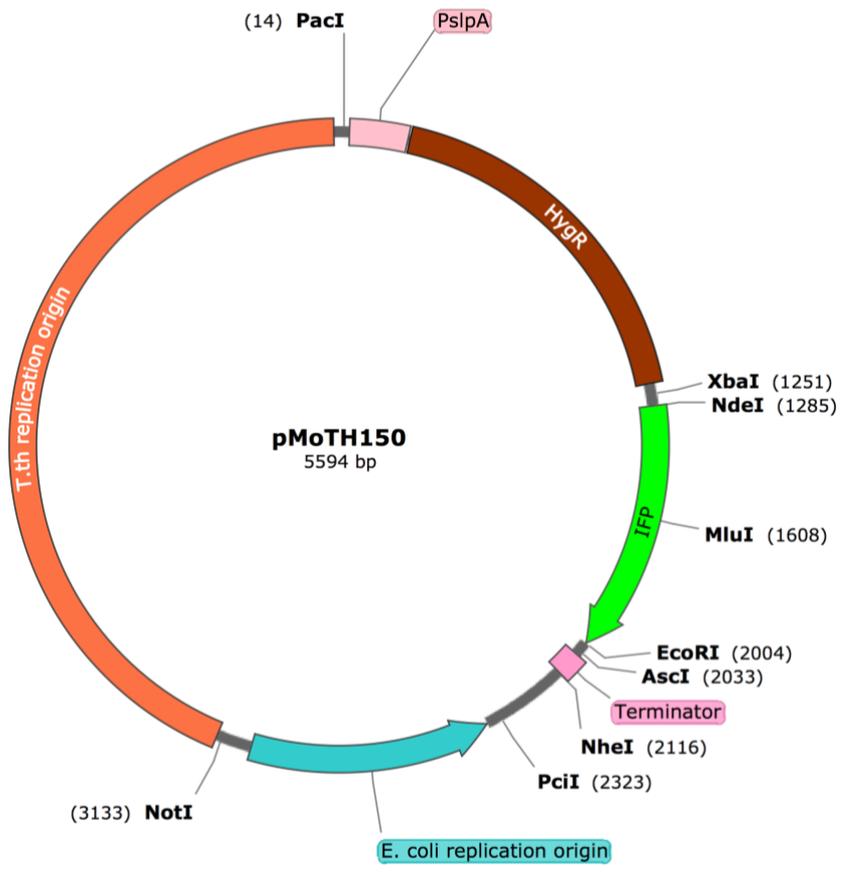
Oligonucleotides List.

> pMoTH150

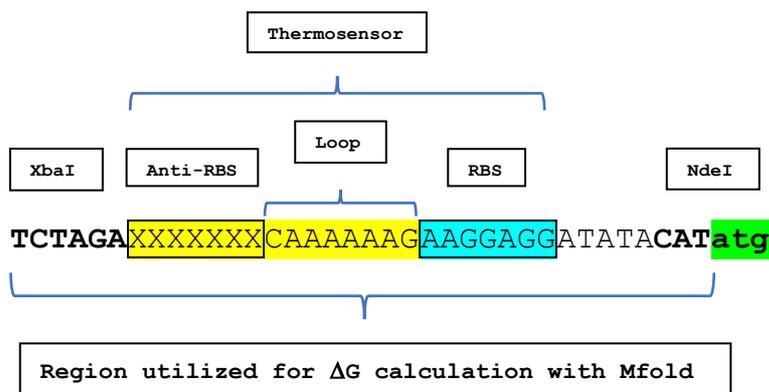
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Scheme S1. Map of plasmid pMoTH150



TS0 ($\Delta G = -11.10$ kcal/mol)

TCTAGAACTCCTTCAAAAAAGAAGGAGGATATACATatg

TS10 ($\Delta G = -12.50$ kcal/mol)

TCTAGACCTCCTTCAAAAAAAAAAAAAAAGAAGGAGGATATACATatg

TS8 ($\Delta G = -13.50$ kcal/mol)

TCTAGACCTCCTTAAAGCAAAAAAGCTAAGGAGGATATACATatg

TSP ($\Delta G = -14.20$ kcal/mol)

TCTAGACCTCCTTCAAAAAAGAAGGAGGATATACATatg

TS1 ($\Delta G = -16.20$ kcal/mol)

TCTAGACCTCCTTAACAAAAAGTTAAGGAGGATATACATatg

TS2 ($\Delta G = -18.60$ kcal/mol)

TCTAGACCTCCTTAGCAAAAAAGCTAAGGAGGATATACATatg

TS4 ($\Delta G = -21.10$ kcal/mol)

TCTAGACCTCCTTACGCAAAAAAGCGTAAGGAGGATATACATatg

TS6 ($\Delta G = -26.90$ kcal/mol)

TCTAGACCTCCTTACGCGCAAAAAAGCGCTAAGGAGGATATACATatg

PA

TCTAGAAAAAAACAAAAAGAAGGAGGATATACATatg

DF

TCTAGAAAGGAGGATATACATatg

Figure S1. Scheme of thermosensors. Different regions of the thermosensors are color-coded. ΔG values and RNA folding calculations were performed with the Mfold software¹⁹.

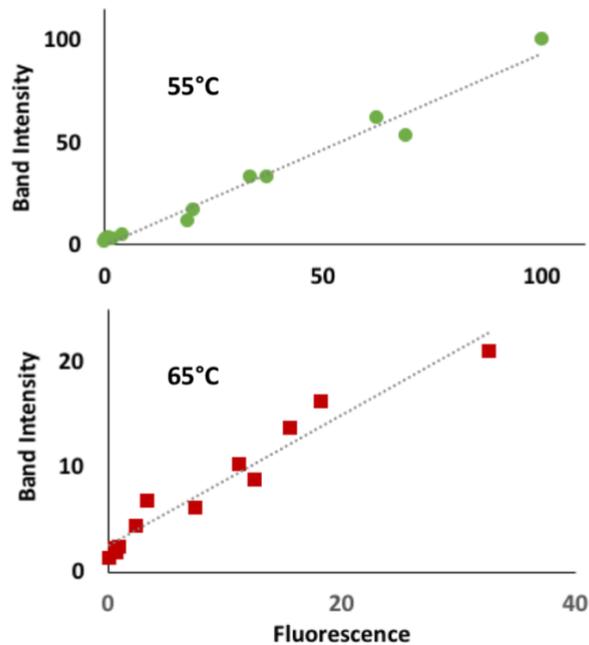


Figure S2. Correlation between sIFP fluorescence measurements and Western detection of the band corresponding to sIFP. Experiments performed at 55°C (green circles) and at 65°C (red squares) have been analyzed. The values of fluorescence intensity are those presented in Fig. 4A, quantification of sIFP band intensities as detected by Western blotting are from Fig. 4D. Dotted line represents a linear regression data fit.

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