

Supplemental Materials

Molecular Biology of the Cell

Mukherjee et al.

SUPPLEMENTAL INFORMATION

Rtn4a promotes exocytosis in mammalian cells

Richik Nilay Mukherjee, Daniel L. Levy

SUPPLEMENTAL FIGURE LEGENDS

Figure S1: Overexpression of Rtn4a, Rtn4b, and REEP5 changes ER morphology and decreases ER sheet volume.

HeLa cells were transiently transfected with plasmids expressing GFP-NLS as a control, Rtn4a-GFP, GFP-Rtn4b, or mCherry-REEP5. **(A)** Cells were immunostained for Rtn4a. **(B)** Rtn4a immunofluorescence intensity was quantified for 47-51 cells per condition. **(C)** Whole cell lysates were immunoblotted for Rtn4a. It is typical for Rtn4a to run as a collection of bands ranging from 130 kDa to over 260 kDa (Li *et al.*, 2004; Osborne *et al.*, 2004). **(D)** Rtn4a band intensities were normalized to β -actin loading controls. Averages from three independent experiments are shown. **(E)** Whole cell lysates were immunoblotted for Rtn4. **(F)** Rtn4b band intensities were normalized to β -actin loading controls. Averages from three independent experiments are shown. **(G)** Cells were immunostained for REEP5. **(H)** REEP5 immunofluorescence intensity was quantified for 20-27 cells per condition. **(I)** Cells were immunostained for the total ER marker KDEL. **(J)** To quantify the distribution of KDEL staining, intensity values were measured along 25 μ m long straight lines drawn from the nuclear envelope to the cell periphery. Pixel intensities along KDEL line scans were averaged for 10-12 cells per condition and 3 lines per cell. **(K)** Cells were immunostained for the ER sheet marker CLIMP63. **(L)** Quantitation of mean ER sheet volume based on CLIMP63 immunofluorescence from 3D reconstructed confocal z-stacks. 54-57 cells were quantified per condition.

Scale bars are 10 μ m. Representative images are maximum intensity projections of confocal z-stacks. Error bars represent SD. **** $p \leq 0.0001$; *** $p \leq 0.001$.

Figure S2: Validation of Integrin β 1 and HLA-A surface staining (A-C).

Overexpression of Rtn4a, Rtn4b, and REEP5 does not alter the total levels of

Integrin β 1 and HLA-A (D-I). Rtn4 RNAi decreases Rtn4a and Rtn4b levels without affecting ER morphology (J-N). (A-C) HeLa cells were transiently transfected with

plasmids expressing Rtn4a-GFP, GFP-Rtn4b, or mCherry-REEP5. Non-permeabilized cells were stained for surface-localized Integrin β 1 or HLA-A. Single confocal z-planes of transfected cells are shown from the same maximum intensity projected images in Fig. 1A-B. (D-E) HeLa cells were transiently transfected with plasmids expressing GFP-NLS

as a control, Rtn4a-GFP, GFP-Rtn4b, or mCherry-REEP5. Whole cell lysates were immunoblotted for Integrin β 1 and HLA-A. Integrin β 1 and HLA-A band intensities were normalized to β -actin loading controls. Averages from three independent experiments are shown. (F-I) HeLa cells were transiently transfected with plasmids expressing GFP-NLS

as a control or Rtn4a-GFP. Permeabilized cells were stained for total Integrin β 1 or HLA-A. Total Integrin β 1 fluorescence staining intensity was quantified for 39-49 cells per condition. Total HLA-A fluorescence staining intensity was quantified for 48-52 cells per condition. (J-L) HeLa cells were transiently co-transfected with siRNA against Rtn4

and Block-iT fluorescent control or with Block-iT alone. Whole cell lysates were immunoblotted for Rtn4. It is typical for Rtn4a to run as a collection of bands ranging from 130 kDa to over 260 kDa (Li *et al.*, 2004; Osborne *et al.*, 2004). Rtn4a and Rtn4b band intensities were normalized to β -actin loading controls. Averages from three independent experiments are shown. (M-N) HeLa cells were transiently co-transfected

with siRNA against Rtn4 and Block-iT fluorescent control or with Block-iT alone. Cells were immunostained for the ER sheet marker CLIMP63. Mean ER sheet volume based on CLIMP63 immunofluorescence was quantified from 3D reconstructed confocal z-stacks. 35-57 cells were quantified per condition.

Scale bars are 10 μ m. Unless otherwise noted, representative images are maximum intensity projections of confocal z-stacks. Error bars represent SD. ** $p \leq 0.01$; NS not significant.

Figure S3: Rtn4a overexpression increases cell surface localization, but not total levels, of Integrin β 1 and HLA-A in MRC-5 cells (A-H). Overexpression of Rtn4a

increases the secretion of endogenous FBLN5 and TSP1 (I-N), without causing ER stress (O-R). (A-H) MRC-5 cells were transiently transfected with plasmids expressing GFP-NLS as a control or Rtn4a-GFP. (A-B) Non-permeabilized cells were stained for surface-localized Integrin β 1 or HLA-A. (C) Integrin β 1 surface fluorescence staining intensity was quantified for 25-31 cells per condition. (D) HLA-A surface fluorescence staining intensity was quantified for 20-25 cells per condition. (E-F) Permeabilized cells were stained for total Integrin β 1 or HLA-A. (G) Total Integrin β 1 fluorescence staining intensity was quantified for 20-29 cells per condition. (H) Total HLA-A fluorescence staining intensity was quantified for 27-33 cells per condition. (I-N) HeLa cells were transiently nucleofected with plasmids expressing GFP-NLS as a control or Rtn4a-GFP. Media was collected and whole cell lysates were prepared 12 hours after transfection. Samples were subjected to sandwich ELISA. Data are presented from three independent experiments. (I,L) Amounts of secreted FBLN5 and TSP1 present in the media were quantified by ELISA and normalized to the number of live cells. (J,M) Amounts of intracellular FBLN5 and TSP1 present in whole cell lysates were quantified by ELISA and normalized to the number of live cells. (K,N) Total amounts of FBLN5 and TSP1 were calculated by summing the secreted and intracellular amounts of each protein per cell. (O-R) HeLa cells were transiently transfected with plasmids expressing GFP-NLS as a control or Rtn4a-GFP. Whole cell lysates were immunoblotted for ER stress-related chaperones calnexin, ERp72, and GRP78. Band intensities were normalized to β -actin loading controls, and data from three independent experiments are shown. Scale bars are 10 μ m. Representative images are maximum intensity projections of confocal z-stacks. Error bars represent SD. *** $p \leq 0.001$; ** $p \leq 0.01$; * $p \leq 0.05$; NS not significant.

Figure S4: Rtn4a overexpression and knockdown in Neuro-2a cells. (A-D)

Undifferentiated Neuro-2a cells were transiently co-transfected with plasmids expressing Rtn4a-myc and mCherry or with mCherry only as a control. (A) Cells were immunostained for the myc epitope and Rtn4a. (B) Rtn4a immunofluorescence intensity was quantified for 36-39 transfected and non-transfected cells per condition. (C) Whole

cell lysates were immunoblotted for Rtn4a. **(D)** Rtn4a band intensities were normalized to β -actin loading controls. Averages from three independent experiments are shown. **(E-H)** Undifferentiated Neuro-2a cells were transiently co-transfected with siRNA against Rtn4a and Block-iT fluorescent control or with Block-iT alone. **(E)** Whole cell lysates were immunoblotted for Rtn4a. **(F)** Rtn4a band intensities were normalized to β -actin loading controls. Averages from three independent experiments are shown. **(G)** Cells were immunostained for Rtn4a. **(H)** Rtn4a immunofluorescence intensity was quantified for 29-36 transfected and non-transfected cells per condition. Scale bars are 10 μ m. Representative images are maximum intensity projections of confocal z-stacks. Error bars represent SD. **** $p \leq 0.0001$; * $p \leq 0.05$; NS not significant.

SUPPLEMENTAL REFERENCES

- Li, W., Walus, L., Rabacchi, S.A., Jirik, A., Chang, E., Schauer, J., Zheng, B.H., Benedetti, N.J., Liu, B.P., Choi, E., Worley, D., Silvian, L., Mo, W., Mullen, C., Yang, W., Strittmatter, S.M., Sah, D.W., Pepinsky, B., and Lee, D.H. (2004). A neutralizing anti-Nogo66 receptor monoclonal antibody reverses inhibition of neurite outgrowth by central nervous system myelin. *The Journal of biological chemistry* 279, 43780-43788.
- Osborne, S.L., Corcoran, S.L., Prinjha, R.K., and Moore, S.E. (2004). Nogo A expression in the adult enteric nervous system. *Neurogastroenterol Motil* 16, 465-474.

Figure S1

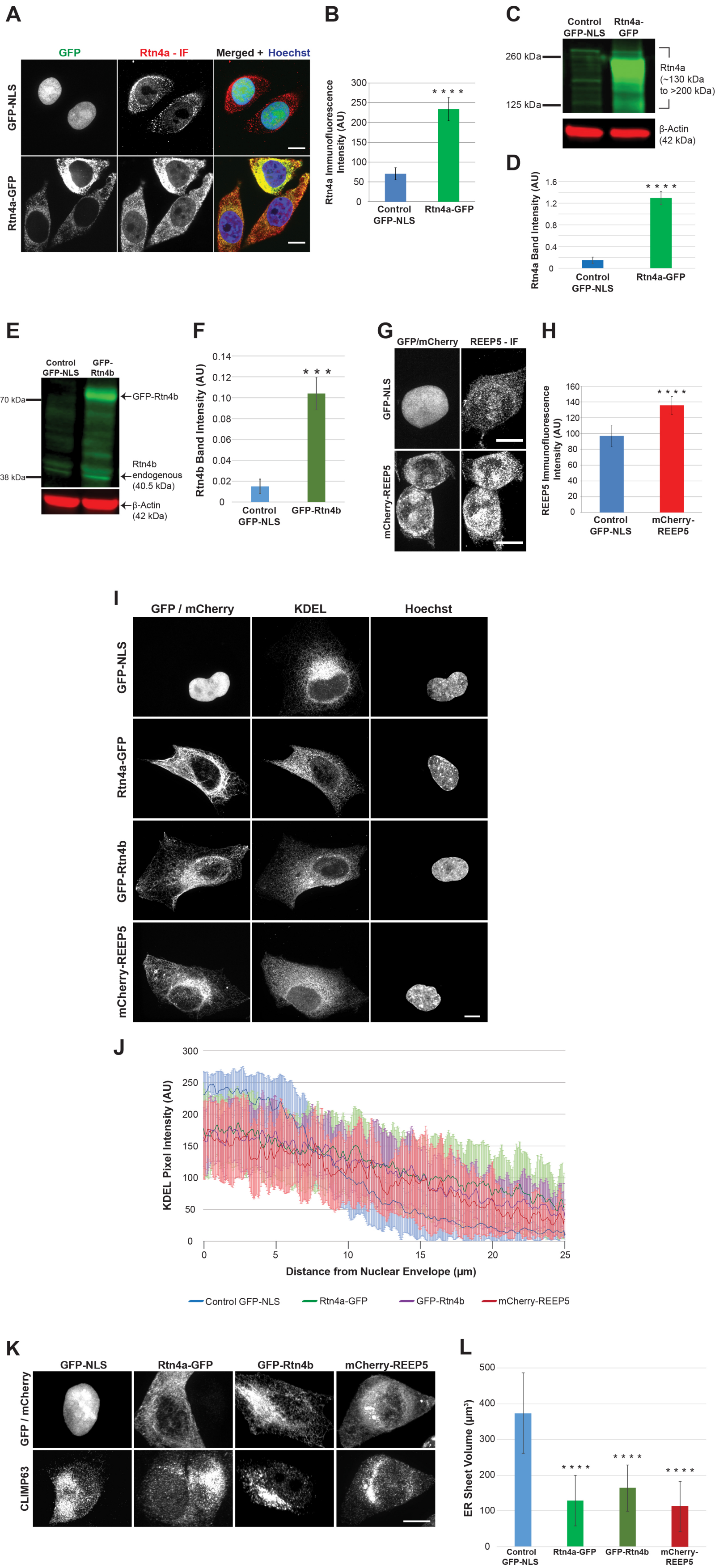


Figure S2

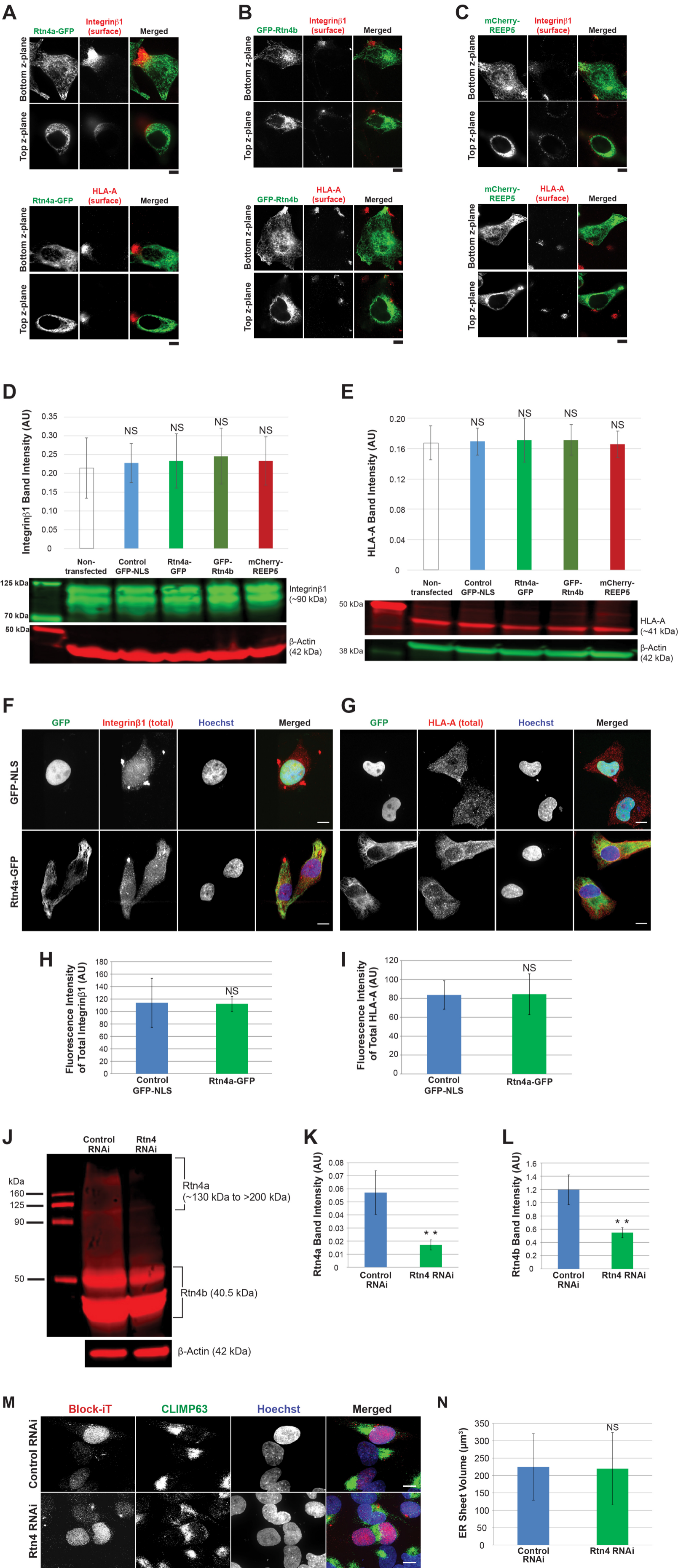


Figure S3

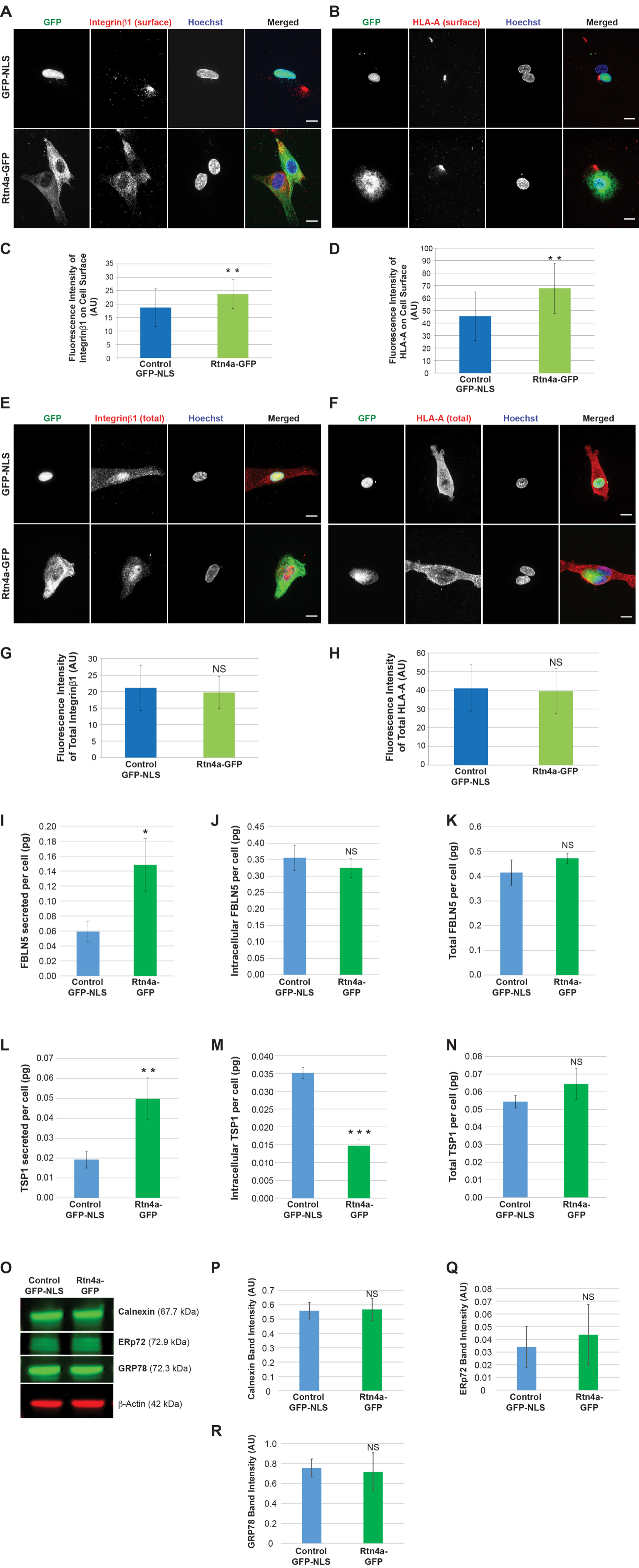


Figure S4

